

**UNIVERSITÉ DU QUÉBEC À MONTRÉAL**

**RÉACTIONS DE *N*-, *O*-, *S*-ARYLATION CATALYSÉES AU CUIVRE  
IMPLIQUANT DES ORGANOBIISMUTHINES FONCTIONNALISÉS ET  
APPLICATION À L'ARYLATION D'ACIDES AMINÉS ET DE PEPTIDES**

**MÉMOIRE**

**COMME EXIGENCE PARTIELLE**

**DE LA MAÎTRISE EN CHIMIE**

**PRÉSENTÉ PAR**

**MARTIN JEAN-GUY HÉBERT**

**DÉCEMBRE 2016**

UNIVERSITÉ DU QUÉBEC À MONTRÉAL  
Service des bibliothèques

Avertissement

La diffusion de ce mémoire se fait dans le respect des droits de son auteur, qui a signé le formulaire *Autorisation de reproduire et de diffuser un travail de recherche de cycles supérieurs* (SDU-522 – Rév.07-2011). Cette autorisation stipule que «conformément à l'article 11 du Règlement no 8 des études de cycles supérieurs, [l'auteur] concède à l'Université du Québec à Montréal une licence non exclusive d'utilisation et de publication de la totalité ou d'une partie importante de [son] travail de recherche pour des fins pédagogiques et non commerciales. Plus précisément, [l'auteur] autorise l'Université du Québec à Montréal à reproduire, diffuser, prêter, distribuer ou vendre des copies de [son] travail de recherche à des fins non commerciales sur quelque support que ce soit, y compris l'Internet. Cette licence et cette autorisation n'entraînent pas une renonciation de [la] part [de l'auteur] à [ses] droits moraux ni à [ses] droits de propriété intellectuelle. Sauf entente contraire, [l'auteur] conserve la liberté de diffuser et de commercialiser ou non ce travail dont [il] possède un exemplaire.»



## REMERCIEMENTS

Je tiens à remercier avant tout, mon directeur de recherche, le Pr. Alexandre Gagnon, pour m'avoir accueilli dans son groupe de recherche dynamique et fascinant. Grâce à vous, j'ai réussi à approfondir mes connaissances et ma passion pour la chimie organique. J'apprécie énormément les techniques que vous m'avez apprises dans le laboratoire et les notions d'écriture d'articles scientifiques que vous m'avez enseignées hors du laboratoire. Votre encadrement sera pour moi un atout tout au long de ma carrière en tant que chimiste. J'apprécie également les opportunités que vous m'avez offertes pendant mes deux années de maîtrise qui m'ont permis de grandir et de m'épanouir. Je vous remercie encore une fois pour votre excellente formation qui m'a poussé à poursuivre mes études en chimie organique.

Également, je tiens à remercier ma famille et mes amis qui m'ont soutenu durant mes études de maîtrise. Je voudrais particulièrement remercier mes parents, Jean-Guy et Marie-Thérèse, ainsi que mon frère, Denis, pour m'avoir encouragé tout au long de mes études.

Par ailleurs, je tiens aussi à remercier tous mes collègues de laboratoire que j'ai eu le privilège de rencontrer et connaître pendant mes deux années de maîtrise à l'UQÀM. Maxime, Emeline, Julien et Adrien, vous êtes les meilleurs et vous serez à jamais mes frères et sœurs. En plus de ma passion pour la chimie, vous avez été pour moi une raison de plus à vouloir entrer au labo avec un gros sourire aux lèvres. Vous êtes tous de bonnes personnes et je vous remercie infiniment pour avoir été là pour moi à chaque jour pendant les bons moments et les moments plus difficiles.

En outre, je voudrais remercier tous les professeurs qui m'ont formé au cours de mes deux années de maîtrise à l'UQÀM. En particulier, j'aimerais remercier le Pr. Alexandre Gagnon, le Pr. Sylvain Canesi, le Pr. René Roy, ainsi que le Pr. Marc

Lussier. Je voudrais aussi remercier Sonia, Mathieu, Odette, Julien et Luc pour votre aide administrative.

Finalement, j'aimerais remercier les organismes qui m'ont soutenu du côté financier pendant ma maîtrise. Notamment, j'aimerais remercier le Conseil de Recherche en Sciences Naturelles et Génie (CRSNG) et les Fonds de Recherche du Québec – Nature et Technologies (FRQNT) de m'avoir accordé des bourses d'études. Je voudrais également remercier le Conseil en Chimie Verte et Catalyse (CCVC) de m'avoir octroyé une bourse de voyage qui m'a permis de donner une présentation orale au 25<sup>th</sup> ISHC Congress à Santa-Barbara en Californie du 23-28 août 2015.

## **DÉDICACE**

Je dédie mon mémoire à mes parents, Jean-Guy et Marie-Thérèse, qui m'ont toujours soutenu tout au long de mes études. Je vous remercie énormément pour m'avoir encouragé à travers la réalisation de ce travail.

## AVANT-PROPOS

La présente recherche a été effectuée sous la direction du professeur Alexandre Gagnon qui a débuté sa carrière à l'UQÀM en 2011. Ses intérêts de recherche comprennent la chimie médicinale, la chimie organique, ainsi que la chimie organométallique et plus particulièrement la chimie du bismuth. Depuis l'année 2007, durant ses diverses expériences, il a pu développer de nombreuses méthodes d'alkylation et d'arylation catalysées au cuivre ou au palladium à base de trialkylbismuthines et de triarylbiismuthines.

En outre, le groupe Gagnon a développé des méthodologies de dérivation de groupes fonctionnels directement sur les triarylbiismuthines menant ainsi à des fonctionnalités plus élaborées et utiles en synthèse organique et en chimie médicinale.

En ce qui concerne les méthodologies développées par catalyse au cuivre, le groupe Gagnon a développé une méthode pour effectuer la *N*-cyclopropylation directe d'amides et d'hétérocycles azotés (indoles, pyrroles, etc.) en utilisant le tricyclopropylbismuth. Par la suite, les triarylbiismuthines ont été employés pour aryle des phénols, des aminoalcools, ainsi que des hétérocycles azotés.

Concernant les réactions catalysées au palladium, le groupe Gagnon a développé des méthodes pour transférer des aryles hautement fonctionnalisés et des groupements alkyles sur des halogénures (ou pseudohalogénures tels que des triflates) d'aryles et d'hétéroaryles.

Compte tenu des méthodologies développées par le professeur Gagnon, soit l'arylation de substrats tels que des indoles (présent dans le tryptophane), des imidazoles (présent dans l'histidine), des phénols (présent dans la tyrosine) et des aminoalcools (présent dans la sérine et la thréonine), celles-ci ont été transposées à l'arylation des chaînes latérales d'acides aminés.

La contribution de la présente recherche servira à approfondir nos connaissances sur la modification des chaînes latérales d'acides aminés, en particulier la tyrosine, en employant des triarylbismuthines, ce qui n'a jusqu'à présent pas été rapporté dans la littérature. Ce travail a comme but de développer une méthode chimiosélective qui nous permettrait de marquer un résidu d'acide aminé spécifique pour être en mesure de suivre ces peptides à travers des processus biochimiques.



## TABLE DES MATIÈRES

<b>REMERCIEMENTS</b> .....	ii
<b>DÉDICACE</b> .....	iv
<b>AVANT-PROPOS</b> .....	v
<b>LISTE D'ÉQUATIONS</b> .....	x
<b>LISTE DE SCHÉMAS</b> .....	xiv
<b>LISTE D'ABRÉVIATIONS ET ACRONYMES</b> .....	xv
<b>LISTE DES SYMBOLES ET DES UNITÉS</b> .....	xxi
<b>RÉSUMÉ</b> .....	xxiii
<b>CHAPITRE I</b>	
<b>BISMUTH</b> .....	1
1.1. Historique du bismuth.....	1
1.2. Trialkylbismuthines .....	1
1.3. Triarylbiismuthines.....	2
1.4. Synthèse d'organobismuthines hautement fonctionnalisés et dérivatisation.....	5
1.5. Triarylbiismuthines pentavalents.....	9
<b>CHAPITRE II</b>	
<b>COUPLAGES-CROISÉS CATALYSÉS AU CUIVRE</b> .....	12
2.1. Couplages-croisés catalysés au Cu(0).....	12
2.2. Ligands employés lors des couplages-croisés catalysés au cuivre.....	13
2.3. Couplages-croisés catalysés au Cu(I).....	14
2.4. Couplages-croisés catalysés au Cu(II).....	15
2.4.1. Formation de liens C–O via des couplages-croisés catalysés au Cu(II) utilisant des organobismuthines .....	18
2.4.2. Formation de liens C–N via des couplages-croisés catalysés au Cu(II) utilisant des organobismuthines .....	20
<b>CHAPITRE III</b>	
<b>COUPLAGES-CROISÉS CATALYSÉS AU PALLADIUM</b> .....	23
3.1. Sources organométalliques.....	23
3.2. Formation de liens C–C via des couplages-croisés catalysés au palladium.....	23
<b>CHAPITRE IV</b>	
<b>MODIFICATION D'ACIDES AMINÉS, DE PEPTIDES ET DE PROTÉINES</b> .....	26
4.1. Modification d'acides aminés.....	26
4.1.1. Activation C–H.....	26
4.1.2. Arylation du N-terminal d'acides aminés.....	27
4.1.3. Arylation de chaînes latérales d'acides aminés par catalyse au cuivre.....	28
4.2. Arylation de la tyrosine.....	30
4.2.1. Substitution nucléophile aromatique (S <sub>N</sub> Ar).....	30
4.2.2. Arylation catalysée au Cu(I).....	31
4.2.3. Arylation catalysée au Cu(II).....	32
4.2.4. Arylation catalysée au Pd(0).....	33

4.2.5. Iode hypervalent.....	34
4.3. Marquage d'acides aminés sur des protéines.....	35
4.3.1. Modification de la lysine.....	36
4.3.2. Modification de la tyrosine.....	37
4.3.3. Modification du tryptophane.....	39
4.3.4. Modification de la cystéine.....	40
<b>CONCLUSION.....</b>	<b>42</b>
<b>ANNEXE A</b>	
<b>CONTRIBUTIONS SCIENTIFIQUES</b>	
<b>Article 1: Synthesis of Highly Functionnalized Triarybismuthines by Functional Group Manipulation and Use in Palladium- and Copper-Catalyzed Arylation Reactions.....</b>	<b>44</b>
Mise en Contexte.....	44
Contributions personnelles.....	45
Contributions des coauteurs .....	45
<b>Manuscript 2: Chemoselective Copper-Catalyzed O-Arylation of Small Tyrosine-Containing Peptides using Functionnalized Organobismuth Reagents.....</b>	<b>47</b>
Mise en contexte.....	47
Contributions personnelles.....	48
Contributions des coauteurs .....	48
<b>ANNEXE B</b>	
<b>Article 1: Synthesis of Highly Functionnalized Triarybismuthines by Functional Group Manipulation and Use in Palladium- and Copper-Catalyzed Arylation Reactions.....</b>	<b>49</b>
<b>ANNEXE C</b>	
<b>Manuscript 2: Chemoselective Copper-Catalyzed O-Arylation of Small Tyrosine-Containing Peptides using Functionnalized Organobismuth Reagent.....</b>	<b>186</b>
<b>BIBLIOGRAPHIE.....</b>	<b>323</b>



## LISTE D'ÉQUATIONS

Éq 1 : Synthèse de trialkylbismuthines et emploi dans des réactions de couplages-croisés catalysés au Pd(0) ou au Cu(II).....	2
Éq 2 : Synthèse de triarylbiomuthines par ajout de réactifs organomagnésiens, organozinciques, organolithiens ou organosodiques sur un sel de biomuth.....	3
Éq 3 : Synthèse de triarylbiomuthines portant des groupes fonctionnels sensibles en employant les conditions de Knochel.....	3
Éq 4 : Hydrolyse de la fonction acétal du dérivé 2-diéthoxyméthyle en 2-formyle.....	7
Éq 5 : Hydrolyse de la fonction acétal du dérivé 3-diéthoxyméthyle en 3-formyle.....	7
Éq 6 : Dérivatisation de la fonction ester en alcool tertiaire.....	8
Éq 7 : Synthèse de triarylbiomuthines hautement fonctionnalisés via un échange halogène-métal catalysé au cobalt et au zinc.....	9
Éq 8 : Oxydation de triarylbiomuthines en ses homologues pentavalents par ajout de peroxyde, de peracide ou de perborate de sodium.....	9
Éq 9 : Oxydation de triarylbiomuthines en ses homologues pentavalents par ajout de XeF <sub>2</sub> , de Cl <sub>2</sub> ou de Br <sub>2</sub> .....	10
Éq 10 : Substitution d'ions de chlore ou de brome par d'autres anions.....	10
Éq 11 : Oxydation de triarylbiomuthines en ses homologues pentavalents par ajout d'iode hypervalent.....	11
Éq 12 : Synthèse de biaryles par couplage d'Ullmann catalysé au Cu(0).....	12

Éq 13 : Synthèse de biaryléthers et de biarylamine par couplage d'Ullmann catalysé au Cu(I).....	12
Éq 14 : Arylation de nucléophiles de type N-H, O-H ou S-H par couplage de Buchwald utilisant des halogénures d'aryles catalysés au Cu(I) avec l'ajout de ligands.....	14
Éq 15 : Arylation de nucléophiles de type N-H, O-H ou S-H par couplage de type Chan-Evans-Lam utilisant des acides boroniques, des trifluoroborates de potassium ou des triarylbismuthines (trivalents et pentavalents).....	16
Éq 16 : Arylation d'alcools aliphatiques par couplage-croisé catalysé au Cu(II) en utilisant des organobismuthines pentavalents.....	18
Éq 17 : Arylation d'alcools tertiaires par couplage-croisé catalysé au Cu(II) en utilisant des triarylbismuthines pentavalents.....	19
Éq 18 : Arylation d'alcools comportant des groupes électroattracteurs par couplage-croisé catalysé au Cu(II) en utilisant des triarylbismuthines trivalents et pentavalents.....	19
Éq 19 : Arylation de phénols par couplage-croisé catalysé au Cu(II) en utilisant des triarylbismuthines.....	20
Éq 20 : Arylation d'anilines par couplage-croisé catalysé au Cu(II) en utilisant des triarylbismuthines.....	20
Éq 21 : Arylation d'amines, d'hydrazines et d'hydrazones par couplage-croisé catalysée au Cu(II) en utilisant des triarylbismuthines.....	21
Éq 22 : Arylation d'amides par couplage-croisé catalysée au Cu(II) en utilisant des triarylbismuthines.....	21

<b>Éq 23 :</b> Arylation d'hétérocycles azotés par couplage-croisé catalysée au Cu(II) en utilisant des triarylbismuthines trivalents et pentavalents.....	22
<b>Éq 24 :</b> Activation C–H de la position C-2 du tryptophane avec des iodures d'aryles catalysé au Pd(OAc) <sub>2</sub> .....	27
<b>Éq 25 :</b> Activation C–H du CH <sub>3</sub> de l'alanine avec des iodure d'aryles catalysé au Pd(OAc) <sub>2</sub> .....	27
<b>Éq 26 :</b> Arylation du N-terminal d'acides aminés avec des organobismuthines pentavalents par catalyse au Cu(0) et au Cu(II).....	28
<b>Éq 27 :</b> Arylation du N-terminal d'acides aminés avec des iodures d'aryles catalysé au Cu(I).....	28
<b>Éq 28 :</b> Arylation d'histidine par couplage de type Chan-Evans-Lam.....	30
<b>Éq 29 :</b> Substitution nucléophile aromatique de F et de NO <sub>2</sub> par la fonction phénol de la tyrosine.....	30
<b>Éq 30 :</b> Arylation de la tyrosine par S <sub>N</sub> Ar via la formation d'un complexe métal-arène à base de Mn(I), Fe(II) ou Ru(II).....	31
<b>Éq 31 :</b> Arylation de la tyrosine via un couplage de type Buchwald catalysé au Cu(I).....	32
<b>Éq 32 :</b> Arylation de la tyrosine via un couplage de type Chan-Evans-Lam catalysé au cuivre.....	32
<b>Éq 33 :</b> Préparation de sels de diaryliodonium par emploi d'iodures d'aryles (ou d'I <sub>2</sub> ) et un aryle quelconque en présence d'un oxydant.....	35

Éq 34 : Préparation de sels de diaryliodonium par emploi d'iodures d'aryles, un oxydant et d'un acide boronique.....	35
Éq 35 : Arylation de phénols en utilisant des sels de diaryliodonium comme source d'aryle.....	35
Éq 36 : Amination réductrice de la lysine sur un lysozyme en utilisant du formiate de sodium par catalyse à l'Ir(I).....	37
Éq 37 : Alkylation de type Tsuji-Trost d'un résidu de tyrosine sur une protéine avec un acétate allylique comprenant une rhodamine fluorescente.....	38
Éq 38 : Marquage de résidus de tyrosine avec une rhodamine sur la Chymotrypsinogen A par réaction de Mannich.....	39
Éq 39 : Marquage de résidus de tryptophane sur la myoglobine par addition de carbénoïdes de rhodium.....	40
Éq 40 : Marquage de résidus cystéines sur la protéine MBP-ReacTR par addition d'un chloroacétamide comprenant une sonde fluorophore (TexasRed).....	41



## LISTE DE SCHÉMAS

<b>Schéma 1 :</b> Préparation d'organobismuthines fonctionnalisés.....	5
<b>Schéma 2 :</b> Dérivatisation de la fonction aldéhyde du dérivé 3-formyle.....	8
<b>Schéma 3 :</b> Exemples de ligands de type <i>N,N</i> -, <i>O,O</i> - et <i>N,O</i> - employés dans des couplages-croisés catalysés au cuivre.....	13
<b>Schéma 4 :</b> Cycle catalytique d'un couplage-croisé catalysé au Cu(I).....	15
<b>Schéma 5 :</b> Cycle catalytique d'un couplage-croisé catalysé au Cu(II) de type Chan-Evans-Lam.....	17
<b>Schéma 6 :</b> Couplages-croisés des halogénoaryles et d'halohétéroaryles avec des triarylbismuthines catalysés au palladium.....	24
<b>Schéma 7 :</b> Mécanisme de la réaction de couplage-croisé d'halogénoaryle avec un triarylbismuthine (et d'autres sources organométalliques) catalysé au palladium.....	25
<b>Schéma 8 :</b> Arylation de sérine, thréonine et tyrosine par couplage de type Chan-Evans-Lam.....	29
<b>Schéma 9 :</b> Formation de complexes métal-arène à base de Mn(I), Fe(II) ou Ru(II) et de chlorures d'aryle.....	31
<b>Schéma 10 :</b> Arylation chimiosélective de tryptophane, déhydroalanine et tyrosine via couplage-croisé au Pd(II), addition conjuguée catalysée au Rh(I) et couplage-croisé catalysé au Pd(0) respectivement.....	34

## LISTE D'ABRÉVIATIONS ET ACRONYMES

1° : Primaire

2° : Secondaire

3° : Tertiaire

(br) : Large (broad)

(l) : État liquide

[O] : Oxydant/Oxydation

Ac : Acétyl

acac : Acétylacétate

AcO/OAc : Acétate

AcOEt/EtOAc : Acétate d'éthyle (Ethyl acetate)

AcOH : Acide acétique

Ala : Alanine

All : Allyle

Ar : Aryle

Asn : Asparagine

Asp : Acide aspartique

BiAr<sub>3</sub> : Triarylbismuthine

BINAP : 2,2'Bis(diphénylphosphino)-1,1'-binaphtalène

BiR<sub>3</sub> : Trialkylbismuthine

Bn : Benzyle

Boc: *Tert*-butyloxycarbonyle

Bzl/Bz : Benzoyle

Bu : Butyle

CCM/TLC : Chromatographie sur couche mince (Thin layer chromatography)

Cbz : Carboxybenzyle

cPr : Cyclopropyle

Cy : Cyclohexyle

Cys : Cystéine

d : Doublet

dba : Dibenzylidèneacétone

DCM : Dichlorométhane

DCE : 1,2-Dichloroéthane

dd : Doublet de doublet

ddd : doublet de doublet de doublet

dhAla : Déhydroalanine



DiPEA : *N,N*-diisopropyléthylamine

DMA : *N,N*-Diméthylacétamide

DMAP : 4-(Diméthylamino)pyridine

DME : Diméthoxyéthane

DMF: *N,N*-Diméthylformamide

dppf : 1,1'-Bis(diphénylphosphino)ferrocène

dq : Doublet de quadruplet

dt : Doublet de triplet

Éq(s) : Équation(s)

éq/équiv/eq/equiv : Équivalent

Et : Éthyle

Et<sub>2</sub>O : Éther diéthylique

Et<sub>3</sub>N : Triéthylamine

Fmoc : Fluorénylméthoxycarbonyle

GF/FG: Groupe fonctionnel (Functional group)

Gln : Glutamine

Glu : Acide glutamique

HCTU : *O*-(6-Chlorobenzotriazol-1-yl)-*N,N,N',N'*-tétraméthyluronium  
hexafluorophosphate

Hex : Hexanes

His : Histidine

HMPA : Hexaméthylphosphoramide

*i*Pr : *Iso*-propyle

IR : Infrarouge

L : Ligand

Lys : Lysine

m : Multiplet

MALDI-MS : Spectrométrie de Masse – Désorption-Ionisation Laser Assistée par Matrice (Matrix Assisted Laser Desorption Ionisation – Mass Spectrometry)

mCPBA : Acide métachloroperbenzoïque (metachloroperbenzoic acid)

Me : Méthyle

MeCN : Acétonitrile

MeOH : Méthanol

MOM : Méthoxyméthyl acétal

mmol : Millimole

NMP : *N*-Méthyl-2-pyrrolidone

nuît/o.n. : Nuit (overnight)

pf/mp : Point de fusion (melting point)

Ph : Phényle

Piv : Pivaloyle

PivO/OPiv : Pivaloate

Pyr : Pyridine

q : Quadruplet

quint : Quintuplet

R : Substituant (alkyle, aryle, H, etc.)

*rac* : Racémique

R<sub>f</sub> : Rapport frontal

RMN/NMR : Résonance Magnétique Nucléaire (Nuclear Magnetic Resonance)

s : Singulet

Ser : Sérine

SMHR/HRMS : Spectrométrie de Masse à Haute Résolution (High Resolution Mass Spectrometry)

S<sub>N</sub>Ar : Substitution nucléophile aromatique

t : Triplet

TA/RT : Température ambiante (Room temperature)

*t*Bu : *Tert*-butyle

TCEP : *Tris*(2-carboxyéthyl)phosphine

td : Triplet de doublet

TFA : Acide trifluoroacétique ou trifluoroacétate

Tf : Triflyle

TfO/OTf : Triflate

TfOH : Acide trifluorométhanesulfonique

THF: Tétrahydrofurane

THP: Tétrahydropyrane

Thr : Thréonine

TM/MS : Tamis moléculaire (Molecular sieves)

TMG : *N,N,N',N'*-tétraméthylguanidine

Trp : Tryptophane

Trt : Trityle (Triphenylméthyle)

Ts : Tosyle

TsO/OTs : Tosylate

TsOH : Acide paratoluènesulfonique

Tyr : Tyrosine

Val : Valine

X : Hétéroatome (O, N, S, Cl, Br, I)

## LISTE DES SYMBOLES ET DES UNITÉS

° : Degré

°C : Degrés Celsius

$\alpha$  : Alpha

Å : Ångström ( $10^{-10}$  mètres)

$\beta$  : Béta

c : Centi ( $10^{-2}$ )

$\text{cm}^{-1}$  : Nombre d'onde (par centimètre)

$\delta$  : Delta (Déplacement chimique)

g : Gramme

h : Heure

Hz : Hertz

j/d : Jours (Days)

L : Litre

$\mu$  : Micro ( $10^{-6}$ )

M : Méga ( $10^6$ )

M : Mole par litre

m : Milli ( $10^{-3}$ )

mins : Minutes

Mm : Masse molaire (g/mol)

ppm : Parties par million

™ : Trademark



## RÉSUMÉ

Dans les dernières décennies, les triarylbismuthines sont des espèces organométalliques qui ont gagné beaucoup d'intérêt en synthèse organique. Ces réactifs sont utilisés dans des réactions de couplages-croisés catalysés au palladium, pour former des liens C-C à partir d'halogénoaryles, et au cuivre pour former des liens C-N, C-O et C-S à partir de nucléophiles de type N-H, O-H et S-H.

Dans la littérature, Barton et Finet ont été en mesure d'aryler toutes sortes de substrats en utilisant des triarylbismuthines par catalyse au cuivre. Cependant, les travaux effectués sur le développement de méthodes d'arylation des chaînes latérales des acides aminés demeurent limités.

Au cours des dernières années, notre groupe de recherche a été en mesure de synthétiser une librairie d'organobismuthines hautement fonctionnalisés par la manipulation de groupes fonctionnels. D'autre part, nous avons été capable d'aryler une grande variété de substrats de type N-H et O-H tels que des indoles, des imidazoles, des phénols, des aminoalcools, etc. Sachant que ces groupes fonctionnels sont présents dans la structure des acides aminés, nous avons transposé nos méthodes à l'arylation d'acides aminés utilisant des triarylbismuthines hautement fonctionnalisés.

La modification des chaînes latérales d'acides aminés peut s'avérer très utile en chimie médicinale ou en biochimie. En effet, plusieurs médicaments possèdent des motifs d'acides aminés dans leur structure. En biochimie, il peut être utile de marquer certains acides aminés dans des peptides ou des protéines pour être en mesure de les suivre avec des techniques analytiques à travers des processus biochimiques.

Nous rapportons dans cet ouvrage le développement d'une méthode d'arylation de chaînes latérales d'acides aminés en utilisant des triarylbismuthines par catalyse au cuivre. En ajout, il s'avère que notre méthode d'arylation démontre une sélectivité



envers l'arylation de la tyrosine, que ce soit dans un contexte d'un acide aminé simple ou d'un oligopeptide. Conséquemment, nous nous sommes concentrés sur l'arylation de ce résidu en transférant des aryles hautement fonctionnalisés et nous avons mené des études de chimiosélectivité en effectuant l'arylation de la tyrosine sur un peptide contenant un autre résidu possiblement réactif, notamment le tryptophane.

**Mots clés :**

- Triarylbismuthines
- Couplage-croisé
- Catalyse au cuivre
- Arylation d'acides aminés et de peptides

# CHAPITRE I

## BISMUTH

### 1.1. Historique du bismuth

Le bismuth est un métal lourd qui fait partie du 15<sup>e</sup> groupe du tableau périodique, soit le groupe de l'azote, autrement appelé les pnictogènes (N, P, As, Sb, Bi). Le bismuth métallique ( $\text{Bi}^0$ ) possède la configuration électronique  $[\text{Xe}] 4f^{14} 5d^{10} 6s^2 6p^3$  et il possède également deux formes d'oxydation stables, notamment le bismuth trivalent ( $\text{Bi}^{+3}$ ), où il y a la perte de ses 3 électrons dans les orbitales  $6p^3$  ( $[\text{Xe}] 4f^{14} 5d^{10} 6s^2 6p^0$ ), ainsi que le bismuth pentavalent ( $\text{Bi}^{+5}$ ), où il y a la perte subséquente de ses 2 électrons dans l'orbitale  $6s^2$  ( $[\text{Xe}] 4f^{14} 5d^{10} 6s^0 6p^0$ ).<sup>1</sup>

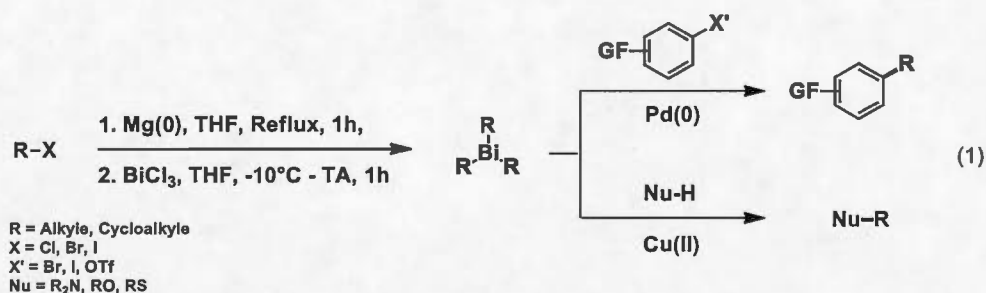
Ce métal a été découvert en 1753 par Claude Geoffroy le Jeune en le séparant du plomb.<sup>2</sup> La synthèse du triéthylbismuthine, soit un trialkylbismuthine, était le premier complexe organométallique de bismuth rapporté en 1850 par Löwig et Schweizer.<sup>3</sup> Par la suite, la première synthèse de triarylbiomuthine a été effectuée en 1887 par Michaëlis et Polis.<sup>4</sup> Finalement, la synthèse des premiers tétraarylbiomuthines et pentaarylbiomuthines a été publiée en 1952 par Wittig et Clauss.<sup>5</sup>

Comme il a été mentionné antérieurement, les organobismuthines se divisent principalement en trois catégories, notamment les trialkylbismuthines, les triarylbiomuthines et les triarylbiomuthines pentavalents.

### 1.2. Trialkylbismuthines

Malgré leur utilité en synthèse organique, les trialkylbismuthines comportent certains désavantages. Par exemple, ceux-ci sont très instables, car ils sont pyrophoriques, ce qui nécessite qu'ils soient manipulés sous atmosphère inerte.<sup>1</sup> Compte tenu de ceci, ces derniers doivent être préparés *in situ* en ajoutant un bromure d'alkylmagnésium sur du  $\text{BiCl}_3$ . Par la suite, le trialkylbismuthine peut être impliqué

dans une réaction de couplage-croisé catalysée au Pd(0), en rajoutant un halogénoaryle, ou au Cu(II), en rajoutant un nucléophile de type N-H, O-H ou S-H (voir Éq 1).<sup>6,7</sup>

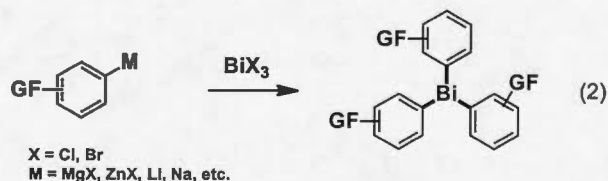


Éq 1 : Synthèse de trialkylbismuthines et emploi dans des réactions de couplages-croisés catalysés au Pd(0) ou au Cu(II)

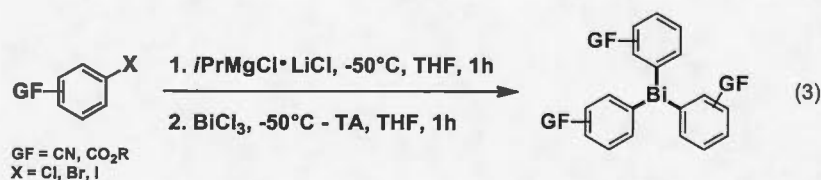
### 1.3. Triarylbismuthines

Les triarylbismuthines sont beaucoup plus stables que les trialkylbismuthines. En effet, ces derniers sont stables à l'eau et à l'air. De plus, ils peuvent être entreposés pendant plusieurs mois jusqu'à plusieurs années à température ambiante. En ajout, il est possible de les purifier par chromatographie sur colonne de silice ou par cristallisation. D'ailleurs, il a été rapporté que les sels de bismuth possèdent une faible toxicité en comparaison à d'autres métaux de transition utilisés pour effectuer des couplages-croisés tel que l'étain. En effet, on retrouve le bismuth dans certains médicaments qui sont vendus sur le marché, tels que le subsalicylate de bismuth (Peptobismol) ainsi que le subcitrate de bismuth (CBS; De-Nol).<sup>8</sup> En outre, il s'avère que les triarylbismuthines tolèrent une grande variété de groupes fonctionnels ce qui les rend très utiles en développement de méthodes.<sup>9</sup> En ajout, les réactifs d'organobismuth sont considérés comme étant très versatiles ce qui explique leur application en synthèse totale,<sup>10</sup> pour la préparation de complexes organométalliques comportant des métaux de transition,<sup>11</sup> en tant que catalyseurs pour des réactions de polymérisation,<sup>12</sup> ainsi qu'en chimie médicinale.<sup>13</sup>

En ce qui concerne la synthèse des triarylbismuthines,  $\text{BiAr}_3$ , ces derniers sont facilement accessibles en formant une espèce organométallique, tel qu'un réactif de Grignard, à partir d'un halogénoaryle et du magnésium métallique, et en l'ajoutant sur un sel de bismuth, souvent le  $\text{BiCl}_3$ .<sup>14</sup> Il est à noter que les triarylbismuthines peuvent également être synthétisés à partir d'autres sources organométalliques telles que des organozinciques,<sup>15</sup> des organolithiens,<sup>16</sup> (voir **Éq 2**) etc. Dans le cas où il y aurait des groupes fonctionnels (GF) sensibles au caractère nucléophile des réactifs de Grignard, tels qu'un ester ou un nitrile, il est préférable d'employer les conditions de Knochel afin de préparer ces organobismuthines respectifs (voir **Éq 3**).<sup>17</sup>



**Éq 2 :** Synthèse de triarylbismuthines par ajout de réactifs organomagnésiens, organozinciques, organolithiens ou organosodiques sur un sel de bismuth

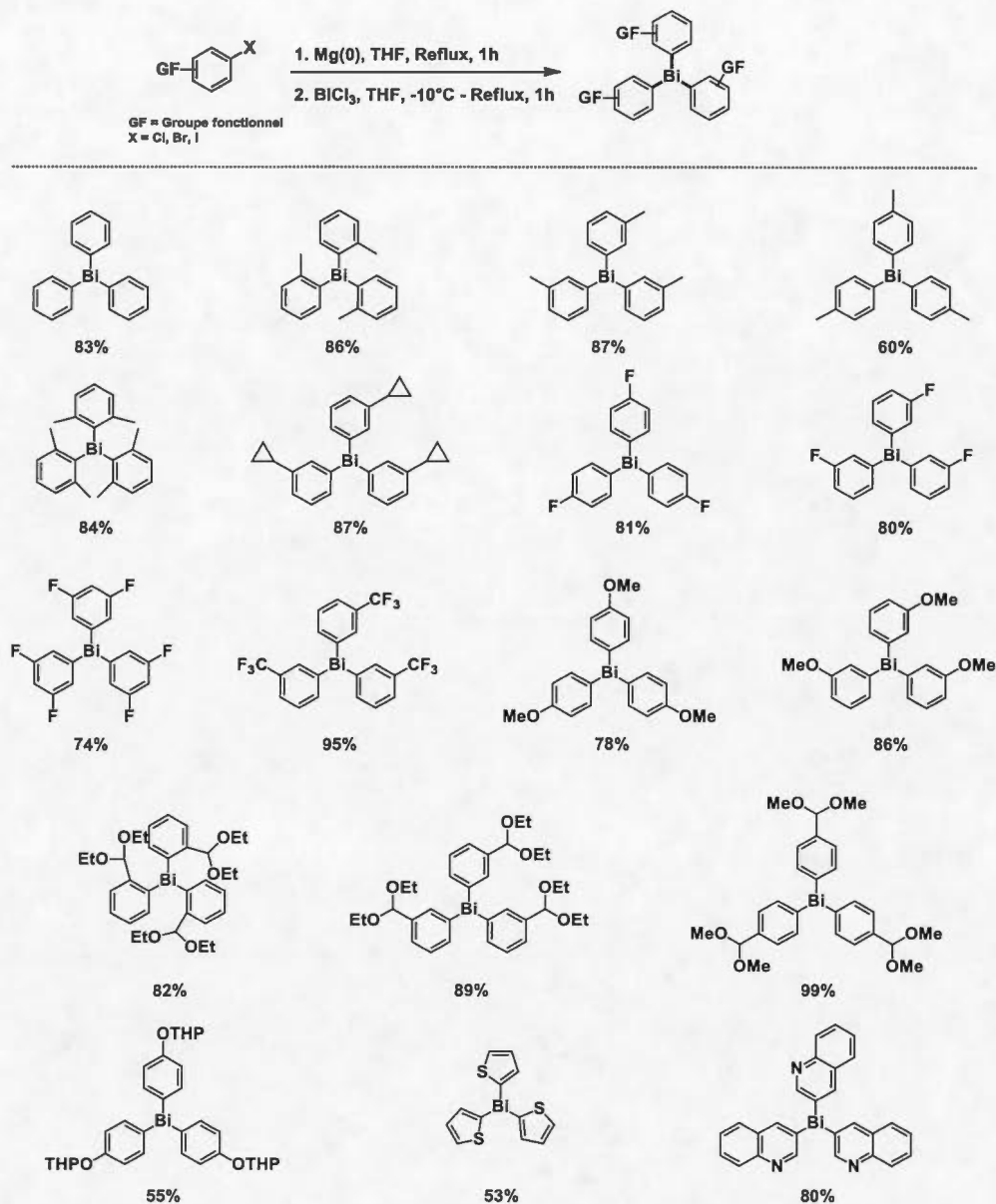


**Éq 3 :** Synthèse de triarylbismuthines portant des groupes fonctionnels sensibles en employant les conditions de Knochel

Dans notre laboratoire, nous avons été capables de préparer une variété d'organobismuthines en employant cette méthode (voir **Schéma 1**). En utilisant cette méthode de synthèse directe, nous avons accès à plusieurs groupes fonctionnels tels que des des alkyles, des cycloalkyles, des fluorures, des trifluorométhyles, des méthoxys, des acétals d'aldéhyde, des acétals de phénol, ainsi que des noyaux



hétéroaromatiques. D'ailleurs, il est possible d'obtenir des aryles substitués dans les positions *ortho*, *méta*, *para*, ainsi que des aryles polysubstitués. Il est à noter que les rendements des triarylbismuthines substitués en *ortho*, ainsi que les dérivés polysubstitués, sont plus faibles, tout probablement en raison de l'encombrement stérique présent sur ces molécules. D'autre part, lors de la purification par chromatographie des acétals d'aldéhydes, il est possible d'observer l'hydrolyse de la fonction acétal en aldéhyde en raison du caractère acide de la silice. Pour contourner ce problème, il peut être prudent de conditionner la colonne préalablement avec de la triéthylamine,  $\text{Et}_3\text{N}$ , dans l'éluant afin de réduire l'acidité de la silice. Ensuite, pour s'assurer que la colonne demeure relativement basique, il est mieux de continuer à ajouter un peu de triéthylamine dans l'éluant pendant la purification. En opposition, le phénol protégé par le groupement THP (tétrahydropyranne) est assez robuste pour résister à l'acidité de la silice, donc l'emploi de triéthylamine n'est pas nécessaire pour sa purification.



**Schéma 1 : Préparation d'organobismuthines fonctionnalisés**

#### 1.4. Synthèse d'organobismuthines hautement fonctionnalisés et dérivatisation

Comme il a été mentionné antérieurement, certains groupements fonctionnels contenant des entités électrophiles tels que des carbonyles (aldéhyde, cétone, ester) ou des nitriles ne peuvent pas être synthétisés directement en utilisant cette méthode en

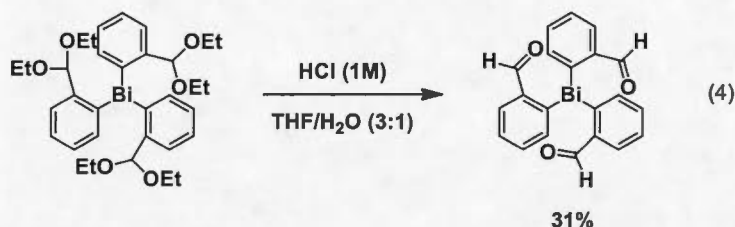
raison de leur sensibilité à la forte nucléophilie des réactifs de Grignard formés lors de cette transformation.

Or, il est possible de préparer des réactifs de Grignard contenant des groupes fonctionnels sensibles, tels que des esters et des nitriles, en employant les conditions de Knochel (voir **Éq 3**). D'abord, il est important de savoir que la réaction d'échange halogène-métal Br/Li est beaucoup plus rapide que l'échange Br/Mg.<sup>18</sup> De plus, l'échange Br/Li est effectué à basse température, alors que l'échange Br/Mg exige un chauffage, ce qui n'est pas compatible avec plusieurs groupes fonctionnels. Aussi, la réaction d'échange Br/Mg est particulièrement lente pour les halogénoaryles riches en électrons, ce qui fait en sorte que la réaction d'échange entre en compétition avec l'élimination de HBr du 2-bromopropane (formé lors de l'échange avec *i*PrMgCl). Pour contourner ces problèmes, on effectue la réaction d'échange à basse température (entre -50°C et -20°C) avec une solution contenant du *i*PrMgCl•LiCl, soit dans les conditions de Knochel. L'ajout du sel de lithium, soit le chlorure de lithium (LiCl), fait en sorte que la réaction d'échange halogène-métal est de type Br/Li et permet que celle-ci soit rapide. De plus, elle est effectuée à basse température ce qui permet que les fonctions sensibles (ester, nitrile) ne soient pas touchées. D'ajout, puisque la réaction d'échange Br/Li est beaucoup plus rapide que l'échange Br/Mg, on évite la compétition avec l'élimination de HBr du 2-bromopropane.<sup>17</sup> Également, des triarylbismuthines ayant des groupes fonctionnels comprenant de protons acides (OH, NH, SH) ne pourront pas être préparés par cette méthode dû au caractère basique des réactifs de Grignard formés pendant la réaction.

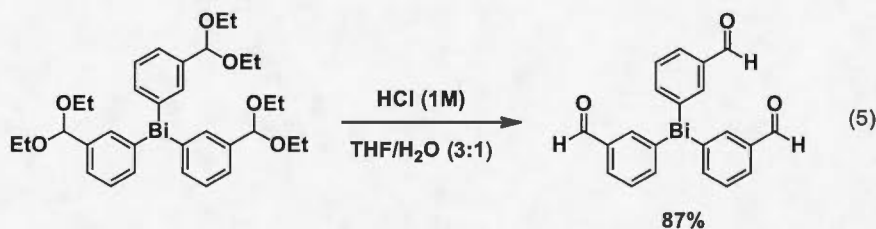
Comme il a été illustré au **Schéma 1**, la méthode de préparation directe d'organobismuthines donne accès à quelques fonctionnalités, mais celles-ci demeurent relativement simples et limitées. Cependant, il est possible d'effectuer une dérivatisation de groupe fonctionnel directement sur l'organobismuthine afin d'obtenir une fonction plus élaborée. Par exemple, en mettant les dérivés 2- ou 3-



diéthoxyméthyles en condition acide aqueuse, la fonction acétal s'hydrolyse cédant ainsi les dérivés 2- et 3-formyles (voir Éqs 4 et 5).

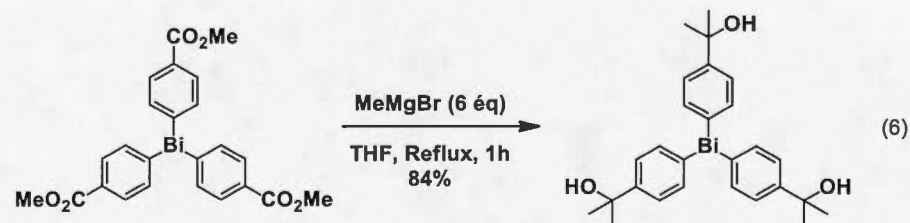
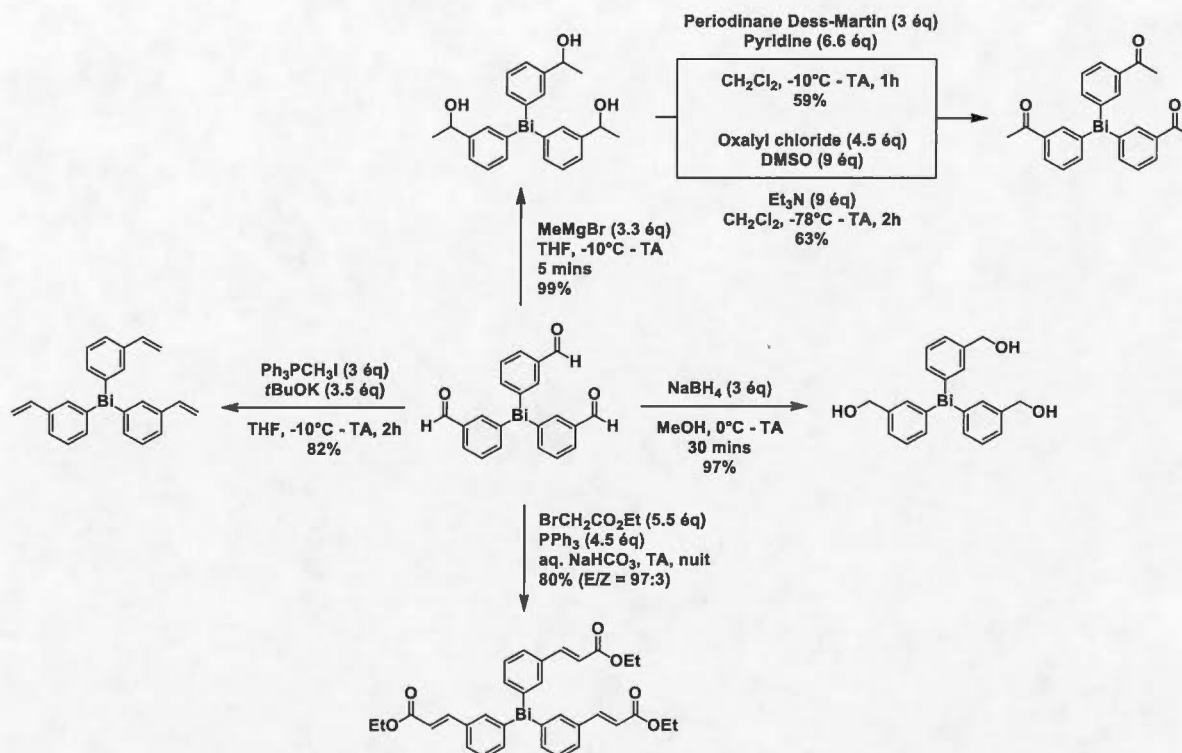


**Éq 4 :** Hydrolyse de la fonction acétal du dérivé 2-diéthoxyméthyle en 2-formyle



**Éq 5 :** Hydrolyse de la fonction acétal du dérivé 3-diéthoxyméthyle en 3-formyle

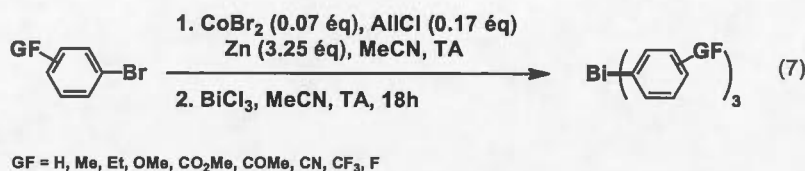
Avec les dérivés formyles en main, on peut prendre avantage de la versatilité de la fonction aldéhyde pour donner un accès à une variété de groupes fonctionnels intéressants. Premièrement, en ajoutant du bromure de méthylmagnésium sur le dérivé 3-formyle, le dérivé possédant un alcool secondaire est obtenu. Subséquemment, cet alcool peut être oxydé en cétone soit par le periodinane de Dess-Martin ou par oxydation de Swern. Deuxièmement, la réduction de l'aldéhyde en alcool primaire est effectuée grâce au borohydrure de sodium ( $\text{NaBH}_4$ ). Finalement, une réaction de Wittig peut être effectuée sur la fonction aldéhyde afin d'obtenir les dérivés ester  $\alpha,\beta$ -insaturé et vinyne (voir **Schéma 2**). D'autre part, l'addition de bromure de méthylmagnésium sur l'ester méthylique mène au dérivé comprenant un alcool tertiaire (voir **Éq 6**). En résumé, les organobismuthines sont des composés robustes qui sont capables de résister aux conditions acides, basiques, et ils peuvent tolérer la présence de nucléophiles, d'oxydants et de réducteurs (voir **Éqs 4-6** et **Schéma 2**).



Éq 6 : Dérivatisation de la fonction ester en alcool tertiaire

Condon a rapporté une méthode très puissante pour synthétiser des organobismuthines hautement fonctionnalisés en passant par un échange halogène-métal catalysé au cobalt et au zinc. Cette méthode permet de former des triarylbismuthines hautement fonctionnalisés directement à partir d'halogénures d'aryles fonctionnalisés (contenant des fonctions telles que des esters, des nitriles, des

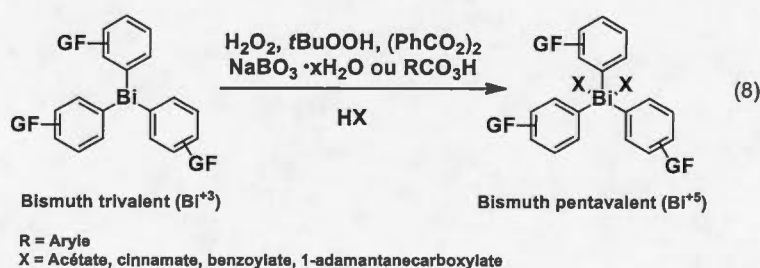
cétones, etc.), une quantité catalytique de bromure de cobalt(II), un excès de poudre de zinc et du chlorure d'allyle (voir Éq 7).<sup>15</sup>



**Éq 7 :** Synthèse de triarylbismuthines hautement fonctionnalisés via un échange halogène-métal catalysé au cobalt et au zinc

### 1.5. Triarylbismuthines pentavalents

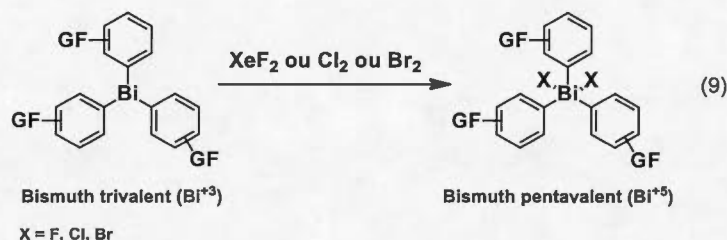
Les triarylbismuthines pentavalents peuvent être obtenus en effectuant une oxydation sur le triarylbismuthine correspondant. Pour ce faire, on doit ajouter un oxydant au triarylbismuthine, ainsi qu'un acide tel que l'AcOH. Dans la littérature, on retrouve souvent que cette réaction emploie des peroxydes, tels que le H<sub>2</sub>O<sub>2</sub>, le *t*BuOOH, le (PhCO<sub>2</sub>)<sub>2</sub> et le NaBO<sub>3</sub>·xH<sub>2</sub>O ou des peracides, RCO<sub>3</sub>H, en présence d'un acide HX (où R = Aryle, X = acétate, cinnamate, benzoates substitués et 1-adamantanecarboxylate) (voir Éq 8).<sup>19</sup>



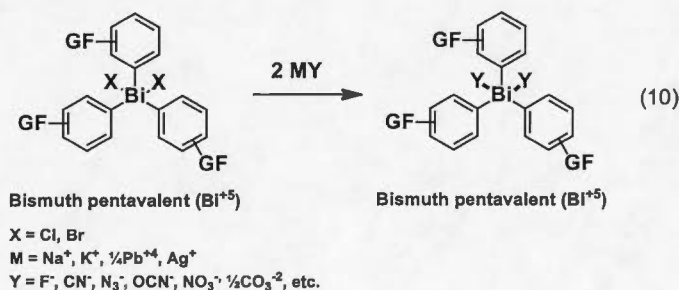
**Éq 8 :** Oxydation de triarylbismuthines en ses homologues pentavalents par ajout de peroxyde, de peracide ou de perborate de sodium

Par ailleurs, il est possible de synthétiser les difluoro-, dichloro- et dibromotriarylbismuthines en oxydant le triarylbismuthine correspondant avec soit du

$\text{XeF}_2$ , du  $\text{Cl}_2$  ou du  $\text{Br}_2$  (voir Éq 9).<sup>4,20</sup> Or, l'ajout d' $\text{I}_2$  sur un  $\text{BiAr}_3$  ne donnera pas son analogue diiodo- pentavalent, mais plutôt du  $\text{Ar}_2\text{BiI}$  et du  $\text{ArI}$ .<sup>20</sup> Une fois que les dérivés pentavalents dichloro- et dibromo- sont obtenus, il est possible de substituer le chlore ou le brome par d'autres anions tels que  $\text{F}^-$ ,  $\text{CN}^-$ ,  $\text{N}_3^-$ ,  $\text{OCN}^-$ ,  $\text{NO}_3^-$ ,  $\text{CO}_3^{2-}$ ,  $\text{AcO}^-$  et  $\text{TfO}^-$  en ajoutant des sels de sodium, potassium, plomb ou argent (voir Éq 10).<sup>21,22</sup> Les sels d'argent, tels que  $\text{AgOTf}$ , fonctionnent particulièrement bien car l'argent piège les halogènes en formant des précipités très peu solubles, soient  $\text{AgCl}$  et  $\text{AgBr}$ , ce qui permet la substitution du chlore et du brome par des anions moins nucléophiles tels que  $\text{TfO}^-$ .



Éq 9 : Oxydation de triarylbismuthines en ses homologues pentavalents par ajout de  $\text{XeF}_2$ , de  $\text{Cl}_2$  ou de  $\text{Br}_2$

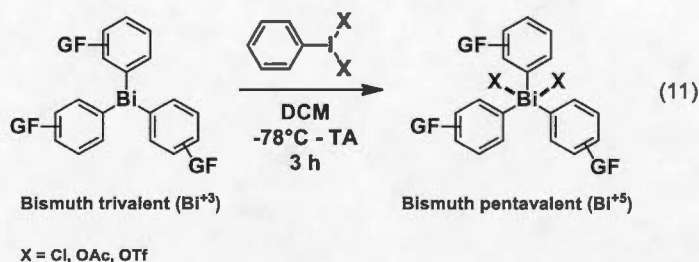


Éq 10 : Substitution d'ions de chlore ou de brome par d'autres anions

Par ailleurs, il est aussi possible de transformer le bismuth trivalent en bismuth pentavalent à l'aide d'iode hypervalent. Par exemple, on peut synthétiser du  $\text{BiAr}_3\text{X}_2$  en rajoutant du  $\text{PhIX}_2$ , où  $\text{X} = \text{Cl}, \text{OAc}$  ou  $\text{OTf}$ , au  $\text{BiAr}_3$  correspondant (voir Éq 11).



Cette réaction permet d'obtenir le produit désiré, ainsi que de l'iodobenzène qui peut être enlevé *in vacuo*.<sup>22</sup>



**Éq 11 :** Oxydation de triarylbismuthines en ses homologues pentavalents par ajout d'iode hypervalent

Les réactifs de bismuth pentavalent peuvent être difficiles, voire impossible à purifier sur silice, donc ils sont soit préparés *in situ* et ensuite utilisés pour effectuer des réactions de couplages-croisés, ou ils sont purifiés par recristallisation.

De plus, il est possible de réduire les organobismuthines pentavalents en leur analogues trivalents en utilisant des agents réducteurs tels que l'hydrazine hydrate ( $\text{NH}_2\text{NH}_2 \cdot x\text{H}_2\text{O}$ ), le dithionite de sodium ( $\text{Na}_2\text{S}_2\text{O}_4$ ), l'ammoniac liquide ( $\text{NH}_3(\text{l})$ ), l'hydrure d'aluminium et de lithium ( $\text{LiAlH}_4$ ), le borohydrure de sodium ( $\text{NaBH}_4$ ) et le sulfure de sodium ( $\text{Na}_2\text{S}$ ).<sup>20</sup>

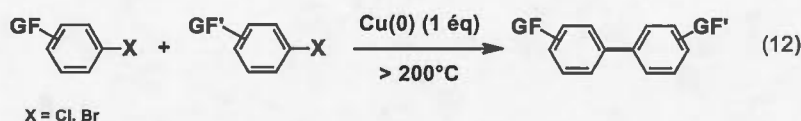


## CHAPITRE II

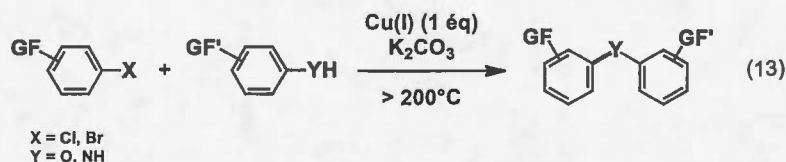
### COUPLAGES-CROISÉS CATALYSÉS AU CUIVRE

#### 2.1. Couplages-croisés catalysés au Cu(0)

C'est en 1901 que Fritz Ullmann a découvert que la synthèse de biaryles pouvait être effectuée en faisant réagir des halogénoaryles avec une quantité stœchiométrique de cuivre métallique en chauffant à plus de 200°C (voir Éq 12).<sup>23</sup> Par la suite, en 1903, Ullmann a rapporté les premiers exemples d'arylation de nucléophiles, soient des anilines et des phénols, encore une fois en employant une quantité stœchiométrique de Cu(I) en chauffant à plus de 200°C, mais cette fois-ci en ajoutant une base (K<sub>2</sub>CO<sub>3</sub>) (voir Éq 13).<sup>24</sup> En 1906, Irma Goldberg a démontré qu'il était possible d'effectuer cette dernière transformation en utilisant une quantité substœchiométrique de cuivre.<sup>25</sup>



Éq 12 : Synthèse de biaryles par couplage d'Ullmann catalysé au Cu(0)



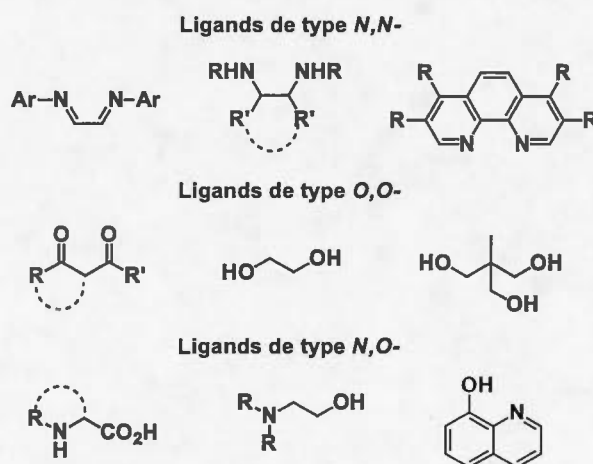
Éq 13 : Synthèse de biaryléthers et de biarylamines par couplage d'Ullmann catalysé au Cu(I)

La découverte du couplage d'Ullmann-Goldberg fût très importante pour la formation de liaisons C–N, C–O et C–S en chimie du cuivre. Toutefois, cette réaction nécessite des conditions relativement extrêmes, dont des températures de réaction au-dessus de 200°C. Conséquemment, plusieurs groupes de recherche se sont mis à travailler sur des moyens pour effectuer ces réactions de couplage en utilisant des conditions plus douces. À cet effet, la grande majorité des couplages de type N–H, O–

H et S–H catalysés au cuivre rapportés dans la littérature utilisent plutôt des catalyseurs de Cu(I) ou de Cu(II) avec ou sans l'ajout de ligands.

## 2.2. Ligands employés lors des couplages-croisés catalysés au cuivre

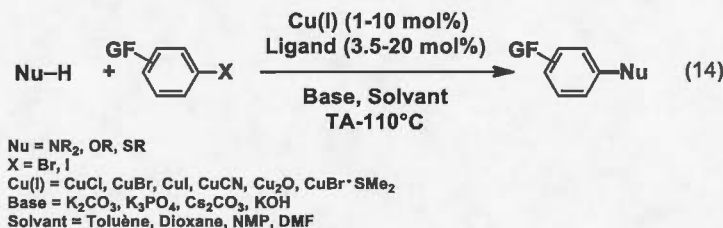
Le cuivre, se retrouvant à l'extrême droite de la première période des métaux de transition, est considéré comme un métal relativement dur, ce qui signifie que son rayon atomique est relativement petit et que sa charge est plus concentrée autour de son noyau (contrairement au palladium). Cela étant dit, il va préférer se lier à des ligands durs contenant des atomes d'oxygène et d'azote. Par exemples, des ligands de type *N,N*- (tels que des  $\alpha$ -diimines, des 1,2-diamines, des phénanthrolines, etc.), de type *O,O*- (tels que des 1,3-dicétones, des 1,2-diols, des 1,3-triols, etc.) et de type *N,O*- (tels que des dérivés d'acides aminés (glycine, proline), des 1,2-aminoalcools, des hydroxyquinolines etc.) sont capables d'activer les catalyseurs de cuivre en se liant à ce dernier (voir **Schéma 3**). La formation de ces complexes de cuivre *in situ* engendre souvent une augmentation du rendement de la réaction de couplage et cette variation peut aussi permettre d'effectuer la réaction dans des conditions plus douces.<sup>26</sup>



**Schéma 3** : Exemples de ligands de type *N,N*-, *O,O*- et *N,O*- employés dans des couplages-croisés catalysés au cuivre

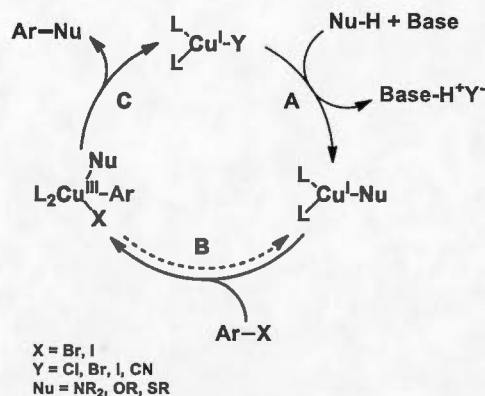
### 2.3. Couplages-croisés catalysés au Cu(I)

Typiquement, les couplages-croisés catalysés au cuivre utilisent soit une source de Cu(I) ou de Cu(II). En ce qui concerne la catalyse au Cu(I), les catalyseurs les plus fréquemment utilisés sont : CuCl, CuBr, CuI, CuCN, Cu<sub>2</sub>O et CuBr•SMe<sub>2</sub>. Les conditions typiques d'une réaction de couplage d'un nucléophile de type N-H, O-H ou S-H sont : une quantité catalytique de Cu(I) (1-10 mol%), un ligand (3.5-20 mol%), une base (K<sub>2</sub>CO<sub>3</sub>, K<sub>3</sub>PO<sub>4</sub>, Cs<sub>2</sub>CO<sub>3</sub>, KOH) et un halogénoaryle (Br, I). Ces réactions peuvent être effectuées à température ambiante, mais elles nécessitent souvent un chauffage (40-110°C) (voir Éq 14).



**Éq 14 :** Arylation de nucléophiles de type N-H, O-H ou S-H par couplage de Buchwald utilisant des halogénures d'aryles catalysés au Cu(I) avec l'ajout de ligands

Cette réaction est possible en utilisant une quantité catalytique de cuivre, car le Cu(I) est régénéré à la fin du couplage-croisé. D'abord, il y a substitution de l'halogène du cuivre, Y, par le nucléophile, Nu (voir étape A). Par la suite, il y a addition oxydante du Cu(I) dans la liaison Ar-X, formant ainsi un espèce de Cu(III) (voir étape B). Finalement, il y a élimination réductrice qui mène à la formation de la liaison entre le nucléophile et l'aryle, régénérant ainsi le Cu(I) (voir étape C). Il est à noter que l'étape d'addition oxydante est réversible, donc on peut aussi reformer l'halogénure d'aryle (Ar-X) comme réaction secondaire. Dans ce mécanisme, il est aussi possible de voir la formation d'un complexe entre les ligands bidendates (de type *O,O*-, *N,O*- et *N,N*-) mentionnés dans la section précédente et le cuivre (voir Schéma 4).<sup>27</sup>

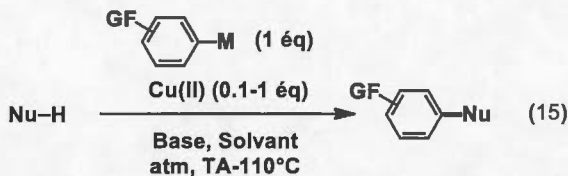


**Schéma 4 :** Cycle catalytique d'un couplage-croisé catalysé au Cu(I)

#### 2.4. Couplages-croisés catalysés au Cu(II)

Également, des nucléophiles de types N-H, O-H et S-H peuvent être arylés par une réaction de couplage-croisé catalysé au Cu(II) en utilisant des acides boroniques, des aryltrifluoroborates de potassium ou des organobismuthines (trivalents et pentavalents) comme source d'aryle, soit une réaction de type Chan-Evans-Lam. Le catalyseur le plus fréquemment utilisé pour effectuer cette transformation est le  $\text{Cu}(\text{OAc})_2$ , mais d'autres catalyseurs tels que  $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ ,  $\text{Cu}(\text{OTf})_2$ ,  $\text{Cu}(\text{OPiv})_2$ ,  $\text{Cu}(\text{TFA})_2$ ,  $\text{Cu}(\text{acac})_2$  et  $\text{Cu}(\text{NO}_3)_2$  peuvent être utilisés. Cette réaction est souvent effectuée dans le DCM ou le toluène dans des gammes de températures entre la température de la pièce jusqu'à 110°C. Cette réaction requiert aussi l'ajout d'une base, typiquement la pyridine ou la  $\text{Et}_3\text{N}$  afin de capturer le proton du nucléophile lors de la substitution d'un ligand sur le cuivre (voir Éq 15).





Nu = NR<sub>2</sub>, OR, SR  
 M = B(OH)<sub>2</sub>, BF<sub>3</sub>K, BiAr<sub>2</sub>, BiAr<sub>2</sub>(OAc)<sub>2</sub>  
 Cu(II) = Cu(OAc)<sub>2</sub>, Cu(OAc)<sub>2</sub> · H<sub>2</sub>O, Cu(OTf)<sub>2</sub>, Cu(OPiv)<sub>2</sub>, Cu(TFA)<sub>2</sub>, Cu(acac)<sub>2</sub>, Cu(NO<sub>3</sub>)<sub>2</sub>  
 Base = Pyridine, Et<sub>3</sub>N  
 Solvant = DCM, Toluène  
 atm = Air, O<sub>2</sub>

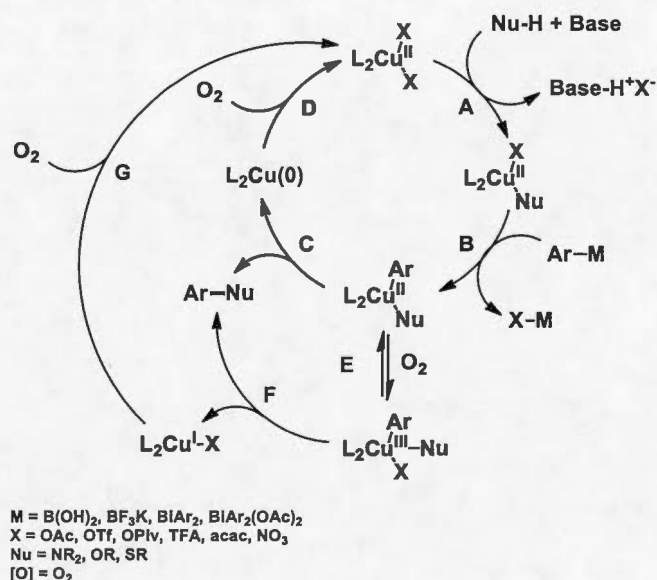
**Éq 15 :** Arylation de nucléophiles de type N–H, O–H ou S–H par couplage de type Chan-Evans-Lam utilisant des acides boroniques, des trifluoroborates de potassium ou des triarylbismuthines (trivalents et pentavalents)

Les réactions de couplage-croisé catalysées au Cu(II) passent par un mécanisme différent de celles catalysées au Cu(I). En effet, les réactions catalysées au Cu(I) utilisent des halogénoaryles comme source d'aryle et peuvent utiliser une quantité catalytique de cuivre, car il est régénéré à la fin de la réaction de couplage-croisé.

Or, dans le cas des réactions de couplage-croisé catalysées au Cu(II), celles-ci peuvent employer une quantité substœchiométrique de cuivre si elles sont effectuées sous air ou O<sub>2</sub>. Notamment, le mécanisme de réaction implique l'emploi d'une source de Cu(II), souvent Cu(OAc)<sub>2</sub>, où il y a substitution d'un acétate, X, par le nucléophile, Nu (voir étape A). Ensuite, il y a transmétallation d'un aryle sur le cuivre (voir étape B). Par la suite, l'espèce formée peut passer par deux différents parcours, soit le parcours C–D, soit le parcours E–G. D'une part, dans le parcours C–D, il y a élimination réductrice accompagnée de la formation du produit désiré, Ar–Nu, et du Cu(0) (voir étape C). Ensuite, le Cu(0) est oxydé en Cu(II), fermant ainsi le cycle catalytique (voir étape D). D'autre part, dans le parcours E–G, l'espèce de Cu(II) est oxydée en Cu(III) à l'aide d'O<sub>2</sub> (voir étape E). Après cela, le produit désiré est formé par élimination réductrice, livrant ainsi un espèce de Cu(I) (voir étape F) qui peut subséquemment se faire oxyder en Cu(II) en présence d'O<sub>2</sub> (voir étape G). Il est à noter qu'il n'y a toujours pas un accord définitif du mécanisme du couplage-croisé de type Chan-Evans-Lam



catalysé au Cu(II) parmi la communauté scientifique. Cependant, selon Lam,<sup>28</sup> l'étape d'élimination réductrice C serait très lente, alors que celle de l'étape F serait rapide, suggérant que le parcours E-G serait plus probable que le parcours C-D (voir Schéma 5).

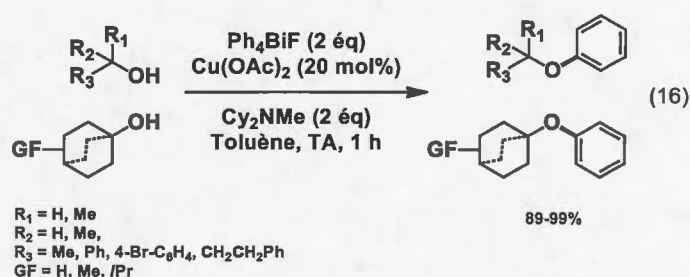


**Schéma 5 :** Cycle catalytique d'un couplage-croisé catalysé au Cu(II) de type Chan-Evans-Lam

Parallèlement, certains groupes de recherche ont rapporté l'arylation de nucléophiles catalysée au Cu(II) en utilisant des triarylbismuthines pentavalents préparés en avance ou *in situ*. En ce qui concerne la préparation *in situ* des triarylbismuthines pentavalents, on doit ajouter un oxydant, tel que de l'OXONE™ (2  $KHSO_5 \cdot KHSO_4 \cdot K_2SO_4$ ), un peracide ( $CH_3CO_3H$ ) ou de l'iode hypervalent ( $PhI(OAc)_2$ ), sur le triarylbismuthine trivalent correspondant pour oxyder le Bi(III) en Bi(V).<sup>29</sup> Cependant, cette approche n'est pas convenable pour des triarylbismuthines portant des fonctionnalités sensibles aux conditions oxydantes.

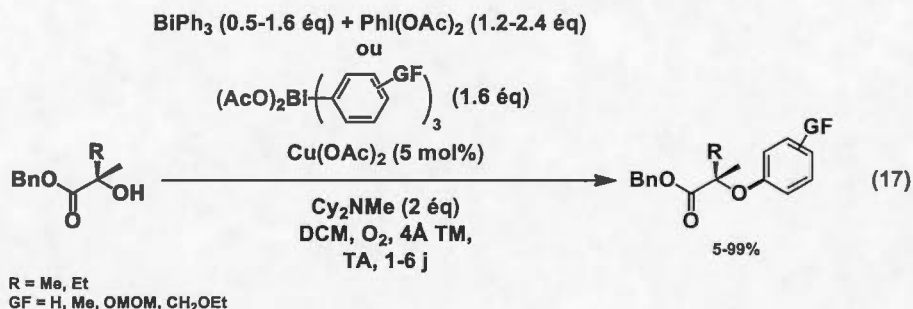
### 2.4.1. Formation de liens C–O via des couplages-croisés catalysés au Cu(II) utilisant des organobismuthines

Certains groupes de recherche ont développé des méthodes de *O*-arylation d'alcools aliphatiques en utilisant des réactifs d'organobismuth. Par exemple, le groupe de Mukaiyama<sup>30</sup> a été en mesure d'aryler des alcools 1°, 2° et 3° facilement en utilisant des tétraarylbismuthines par catalyse au Cu(II) (voir Éq 16).

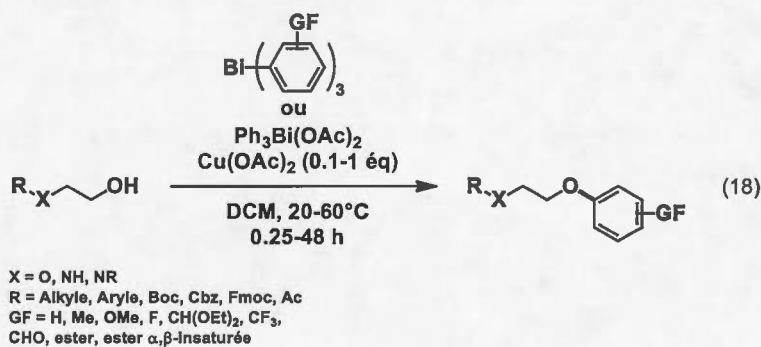


Éq 16 : Arylation d'alcools aliphatiques par couplage-croisé catalysé au Cu(II) en utilisant des organobismuthines pentavalents

De plus, d'autres groupes ont réussi à aryler des alcools ayant des groupes électroattracteurs à proximité de la fonction hydroxyle. Notamment, le groupe de Sato<sup>31</sup> a rapporté une méthode d'arylation d' $\alpha$ -hydroxy esters en utilisant des organobismuthines pentavalents préparés en avance, ou formés *in situ* en rajoutant du  $\text{PhI(OAc)}_2$  comme oxydant sur du  $\text{BiPh}_3$  (oxydation du Bi(III) en Bi(V)) et une quantité catalytique du catalyseur de Cu(II) (voir Éq 17). D'ailleurs, les groupes de Gagnon<sup>32a</sup> et Barton<sup>32b</sup> ont été en mesure d'aryler des 1,2-aminoalcools et des 1,2-alcoxyalcools en utilisant des triarylbismuthines (trivalents ou pentavalents) et une quantité substœchiométrique du catalyseur de Cu(II) (voir Éq 18). Selon les travaux du groupe de Gagnon, il semblerait y avoir un lien entre la tendance de l'alcool à se faire aryler et de la présence d'un groupe électroattracteur à proximité de ce dernier.<sup>32a</sup>

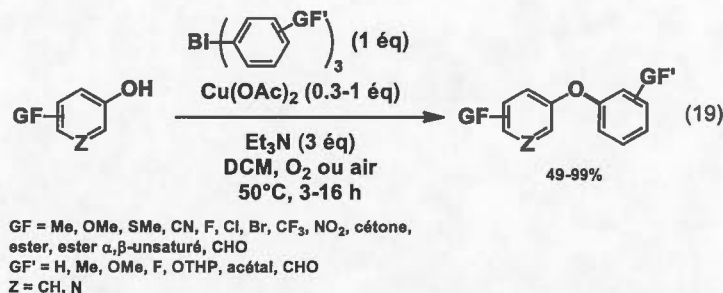


**Éq 17 :** Arylation d' $\alpha$ -hydroxy esters par couplage-croisé catalysé au Cu(II) en utilisant des triarylbismuthines pentavalents



**Éq 18 :** Arylation d'alcools comportant des groupes électroattracteurs par couplage-croisé catalysé au Cu(II) en utilisant des triarylbismuthines trivalents et pentavalents

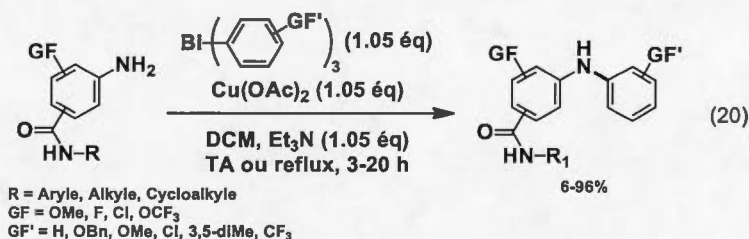
D'ajout, le groupe de Gagnon<sup>33</sup> a développé une méthode permettant la préparation de biaryléthers par arylation de phénols en utilisant des triarylbismuthines fonctionnalisés par catalyse au Cu(II) (voir **Éq 19**).



**Éq 19** : Arylation de phénols par couplage-croisé catalysé au Cu(II) en utilisant des triarylbismuthines

#### 2.4.2. Formation de liens C–N via des couplages-croisés catalysés au Cu(II) utilisant des organobismuthines

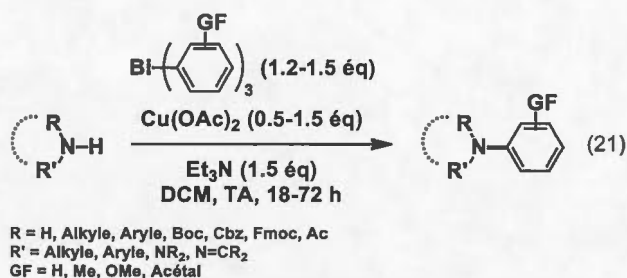
En ce qui concerne la *N*-arylation de substrats par catalyse au cuivre, Sorenson<sup>34</sup> a été en mesure d'aryler des anilines substituées avec des triarylbismuthines et une quantité supastœchiométrique d'acétate de Cu(II) (voir **Éq 20**).



**Éq 20** : Arylation d'anilines par couplage-croisé catalysé au Cu(II) en utilisant des triarylbismuthines

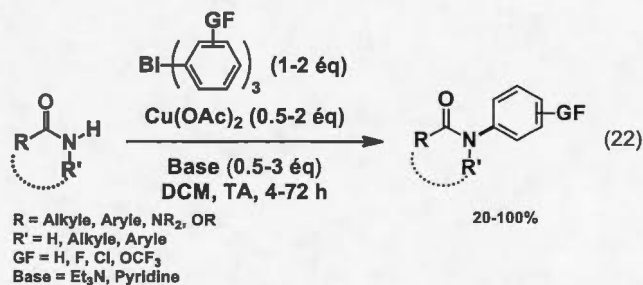
D'autres substrats tels que des amines<sup>35</sup>, des hydrazines<sup>36</sup> et des hydrazones<sup>37</sup> ont pu être *N*-arylés en employant des triarylbismuthines et de l'acétate cuprique (voir **Éq 21**).





**Éq 21 :** Arylation d'amines, d'hydrazines et d'hydrazones par couplage-croisé catalysée au Cu(II) en utilisant des triarylbismuthines

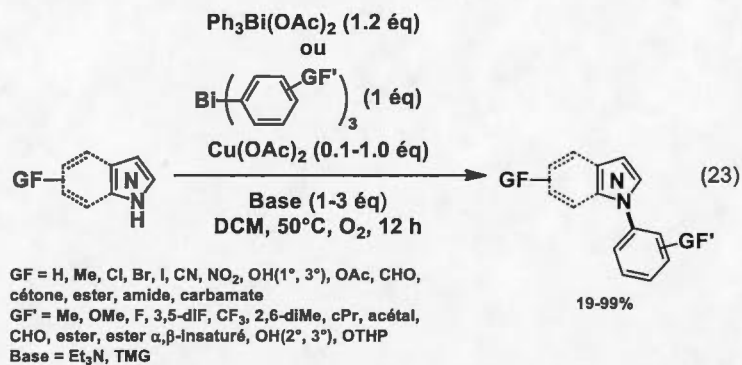
De même, il est possible d'effectuer l'arylation d'amides, de carbamates et d'urées en utilisant des triarylbismuthines par catalyse à l'acétate de cuivre (voir **Éq 22**).<sup>38</sup>



**Éq 22 :** Arylation d'amides par couplage-croisé catalysée au Cu(II) en utilisant des triarylbismuthines

Finalement, une variété d'hétérocycles azotés (incluant les indoles, les benzimidazoles, les indazoles, les pyrroles, les pyrazoles, les imidazoles, etc.) peuvent être *N*-arylés en employant des triarylbismuthines trivalents ou pentavalents par catalyse au Cu(II) (voir **Éq 23**).<sup>39</sup>





Éq 23 : Arylation d'hétérocycles azotés par couplage-croisé catalysée au Cu(II) en utilisant des triarylbismuthines trivalents et pentavalents

## **CHAPITRE III**

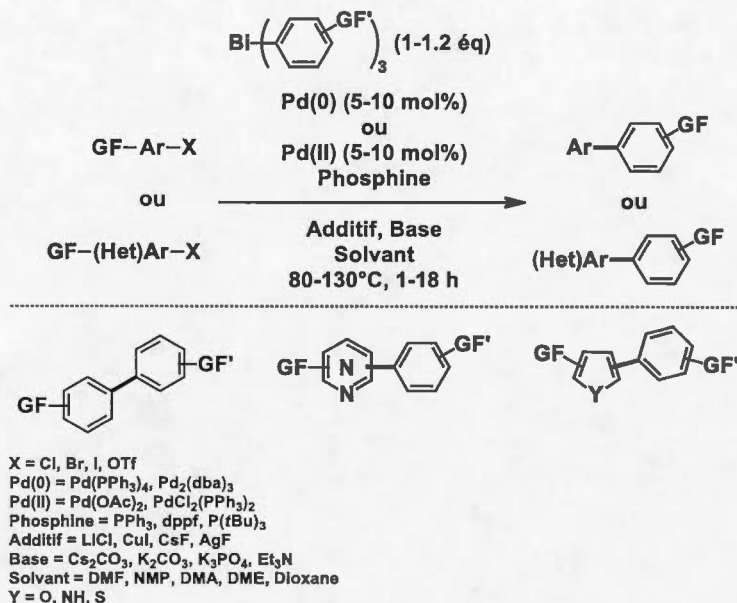
### **COUPLAGES-CROISÉS CATALYSÉS AU PALLADIUM**

#### **3.1. Sources organométalliques**

La catalyse au palladium a révolutionné la manière que l'on crée de nouveaux liens C-C. En effet, en combinant un halogénoaryle, un réactif organométallique (tel qu'un acide boronique, un organozincique, un organomagnésien, un organostannane ou un organobismuthine), ainsi qu'un catalyseur de palladium, il est possible d'effectuer la formation de liens C-C. L'emploi d'acides boroniques comme source organométallique est de loin celle la plus commune, soit un couplage de Suzuki-Miyaura.<sup>40</sup> L'utilisation d'espèces organométalliques telles que des organozinciques, soit un couplage de Negishi, exige des conditions inertes, car ces réactifs sont sensibles à l'air et à l'humidité.<sup>41</sup> Similairement au couplage de Negishi, le couplage de Kumada-Corriu exige aussi des conditions inertes, car cette méthode de couplage-croisé emploie des réactifs organomagnésiens qui sont également sensibles à l'humidité.<sup>42</sup> L'utilisation d'organostannanes comme source organométallique, soit un couplage de Stille, fonctionne relativement bien pour effectuer des réactions de couplage-croisé, mais ces réactifs possèdent le désavantage d'avoir une haute toxicité.<sup>43</sup> Finalement, les organobismuthines sont une excellente alternative comme source organométallique, car ceux-ci ne sont pas toxiques, ils ne sont pas sensibles à l'air et l'humidité et les organobismuthines peuvent transférer leurs trois groupements organiques.

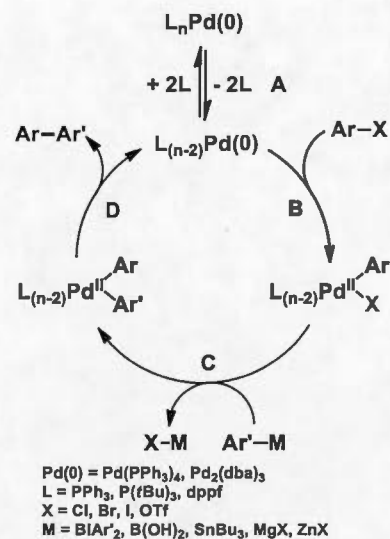
#### **3.2. Formation de liens C-C via des couplages-croisés catalysés au palladium**

Dans la littérature, il a été démontré que les organobismuthines sont capables d'effectuer des réactions de couplage-croisé sur toutes sortes de substrats tels que des chlorures d'acyle,<sup>44</sup> des halogénures allyliques et propargyliques,<sup>45</sup> des halogénures d'aryle (et d'hétéroaryle), (voir **Schéma 6**) etc.<sup>46</sup> En effet, Rao<sup>47</sup> a grandement contribué à l'approfondissement des connaissances des réactions de couplage-croisé catalysées au palladium impliquant des organobismuthines avec une grande variété de substrats.



**Schéma 6 :** Couplages-croisés des halogénoaryles et d'halohétéroaryles avec des triarylbismuthines catalysés au palladium

En ce qui a trait au mécanisme des couplages-croisés catalysés au palladium, le Pd(0) perd premièrement deux ligands, L (voir étape A), pour passer d'un complexe à 18 électron à un complexe à 14 électrons permettant ainsi l'addition oxydante du palladium dans le lien Ar-X, oxydant ainsi le Pd(0) en Pd(II) (voir étape B). Par la suite, on observe l'étape de la transméallation où il y a un transfert d'aryle provenant de l'espèce organométallique sur le Pd(II) (voir étape C). Finalement, il y a élimination réductrice qui permet la formation du produit désiré, régénérant ainsi la source de Pd(0), ce qui ferme le cycle catalytique (voir étape D) (voir **Schéma 7**).<sup>27</sup> Il est à noter que le Pd(0) peut être généré *in situ* en réduisant du Pd(II) (typiquement Pd(OAc)<sub>2</sub>) en Pd(0) par une phosphine (PPh<sub>3</sub> par exemple). En ajout, en catalyse au palladium, les trois aryales des triarylbismuthines peuvent être transférés lors d'une réaction de couplage-croisé, alors qu'en catalyse au cuivre, un seul aryle est transféré.



**Schéma 7 :** Mécanisme de la réaction de couplage-croisé d'halogénoaryle avec un triarylbismuthine (et d'autres sources organométalliques) catalysé au palladium

L'ajout de monoxyde de carbone dans le milieu réactionnel peut aussi mener à l'insertion de CO dans le lien  $Ar-Pd$  livrant ainsi des diarylcétones.<sup>48</sup>



## **CHAPITRE IV**

### **MODIFICATION D'ACIDES AMINÉS, DE PEPTIDES ET DE PROTÉINES**

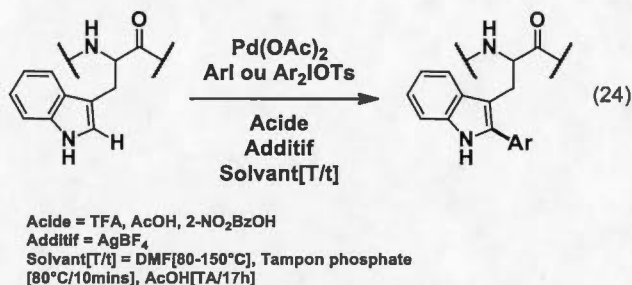
#### **4.1. Modification d'acides aminés**

##### **4.1.1. Activation C–H**

Dans la littérature, certains laboratoires ont développé des méthodologies permettant l'arylation de résidus d'acides aminés via l'activation d'un lien C–H, notamment sur le tryptophane<sup>49</sup> et l'alanine<sup>50</sup>.

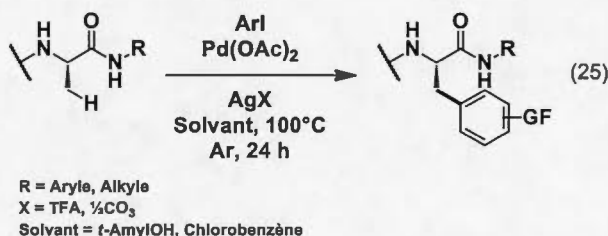
Dans le cas des activations C–H du tryptophane, ces réactions utilisent des iodures d'aryle ou des sels de diaryliodonium et elles sont catalysées au Pd(II), soit par le Pd(OAc)<sub>2</sub>. L'activation C–H en position C-2 peut être effectuée soit dans un solvant organique, soit en milieu aqueux (tampon phosphate salin). Également, l'activation C–H du tryptophane peut être effectuée à température ambiante, ainsi que par microonde. La réaction performée à température ambiante requiert un plus long temps de réaction, soit 17 h, alors qu'un chauffage entre 80-150°C par microonde accélère cette réaction de façon drastique et celle-ci est complète après 5-20 minutes (voir **Éq 24**). De plus, cette réaction est très chimiosélective en présence d'une variété d'acides aminés. Cependant, cette méthodologie ne fonctionne pas sur des peptides contenant des résidus d'acides aminés ayant un atome de soufre tel que la méthionine, notamment en raison de la forte affinité entre le palladium et le soufre (acide mou et base molle).





**Éq 24 :** Activation C–H de la position C-2 du tryptophane avec des iodures d'aryles catalysé au Pd(OAc)<sub>2</sub>

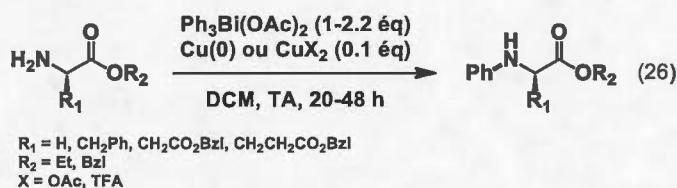
Dans le cas de l'activation C–H de l'alanine, encore une fois, celle-ci utilise toujours du Pd(OAc)<sub>2</sub>, des iodures d'aryle et un sel d'argent afin d'effectuer l'arylation du CH<sub>3</sub> de sa chaîne latérale (voir Éq 25).



**Éq 25 :** Activation C–H du CH<sub>3</sub> de l'alanine avec des iodure d'aryles catalysé au Pd(OAc)<sub>2</sub>

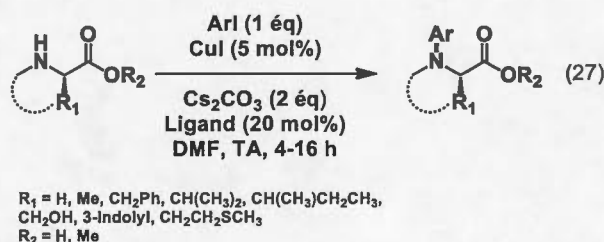
#### 4.1.2. Arylation du N-terminal d'acides aminés

Les groupes de Barton<sup>51</sup> et Jain<sup>52</sup> ont rapporté des méthodologies pour aryle le N-terminal de certains acides aminés par catalyse au cuivre. Le groupe de Barton a employé des organobismuthines pentavalents comme source d'aryle par catalyse au Cu(0) ou au Cu(II) (voir Éq 26).



**Éq 26 :** Arylation du N-terminal d'acides aminés avec des organobismuthines pentavalents par catalyse au Cu(0) et au Cu(II)

Le groupe de Jain a utilisé des conditions de Buchwald pour effectuer l'arylation du N-terminal des acides aminés. Ils utilisent des iodures d'aryle comme source d'aryle, un catalyseur de Cu(I), ainsi qu'un ligand de type 1,3-dicétone (voir Éq 27).



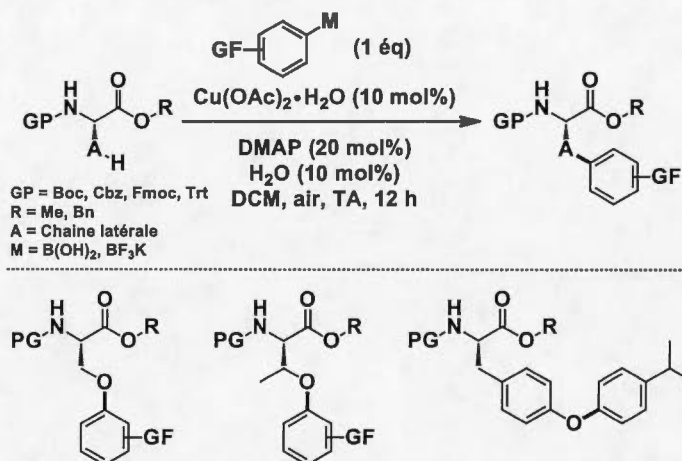
**Éq 27 :** Arylation du N-terminal d'acides aminés avec des iodures d'aryles catalysé au Cu(I)

#### 4.1.3. Arylation de chaînes latérales d'acides aminés par catalyse au cuivre

Certains groupes de recherche ont rapporté des méthodes d'arylation des chaînes latérales d'acides aminés avec l'emploi d'acides boroniques et de trifluoroborates de potassium comme source d'aryle par catalyse au Cu(II).

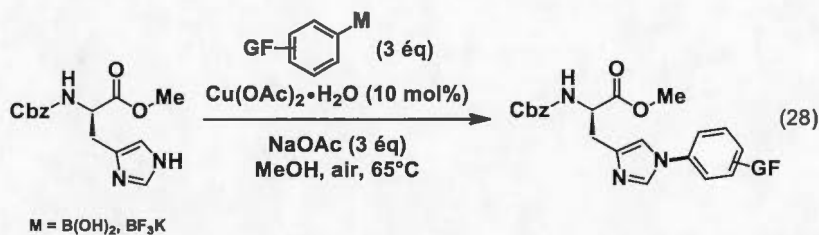
Notamment, le groupe de Molander<sup>53</sup> a effectué le transfert d'une variété d'aryles sur l'hydroxyle de la sérine et de la thréonine (N- et C-terminal protégés). De plus, il a réussi à aryler de façon chimiosélective l'alcool de la sérine du dipeptide Cbz-

Ser-Thr-OMe avec un rendement de 22%. En outre, il démontre aussi un exemple d'arylation de tyrosine avec un rendement moyen de 42% (voir **Schéma 8**).



**Schéma 8** : Arylation de sérine, thréonine et tyrosine par couplage de type Chan-Evans-Lam

De plus, le groupe de Campagne<sup>54</sup> a développé une méthode permettant l'arylation de la fonction imidazole de l'histidine en utilisant une variété d'acides boroniques et de trifluoroborates de potassium par catalyse au Cu(II). Or, ce groupe de recherche n'a arylé que des histidines N- et C-protégées contrairement à des substrats un peu plus complexes tels que des oligopeptides contenant plus d'un acide aminé (voir **Éq 28**). Le groupe de Hanzlik a rapporté une méthode d'arylation de l'histidine utilisant des iodures d'aryles par catalyse au Cu(II)<sup>55</sup> et plusieurs exemples d'arylation de l'histidine où ils utilisent des organoplombs (ArPb(OAc)<sub>3</sub>) comme source d'aryle par catalyse au Cu(II) sont aussi décrits.<sup>56</sup>

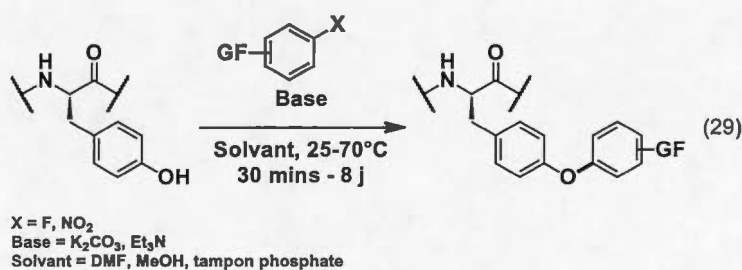


Éq 28 : Arylation d'histidine par couplage de type Chan-Evans-Lam

## 4.2. Arylation de la tyrosine

### 4.2.1. Substitution nucléophile aromatique ( $\text{S}_{\text{N}}\text{Ar}$ )

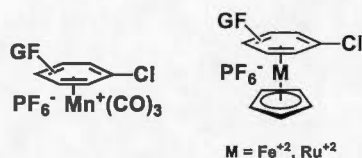
Une des méthodes d'arylation du phénol de la tyrosine couramment utilisée est par substitution nucléophile aromatique ( $\text{S}_{\text{N}}\text{Ar}$ ) de groupes tels que  $\text{F}^{57}$  et  $\text{NO}_2$ .<sup>58</sup> Il est important de savoir que les réactions de  $\text{S}_{\text{N}}\text{Ar}$  se déroulent plus facilement en appauvrissant le cycle aromatique de sa densité électronique. En absence de groupes électroattracteurs, notamment  $\text{NO}_2$ , cette réaction peut prendre plusieurs jours, mais en présence de plusieurs groupes électroattracteurs, la réaction peut être complète après seulement 30 minutes (voir Éq 29).



Éq 29 : Substitution nucléophile aromatique de F et de  $\text{NO}_2$  par la fonction phénol de la tyrosine

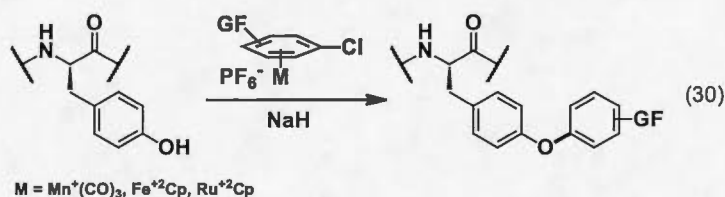
D'autre part, il est aussi possible d'effectuer une  $\text{S}_{\text{N}}\text{Ar}$  sur un chlorure d'aryle via la formation d'un complexe métal-arène avec des métaux tels que  $\text{Mn(I)}$ ,<sup>59</sup>  $\text{Fe(II)}$ <sup>60</sup> ou  $\text{Ru(II)}$  (voir Schéma 9).<sup>60b,61</sup>





**Schéma 9** : Formation de complexes métal-arène à base de Mn(I), Fe(II) ou Ru(II) et de chlorures d'aryle

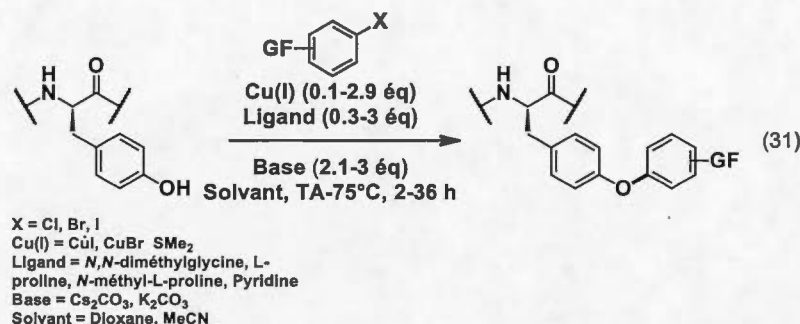
Parallèlement, le cycle aromatique est appauvri en électrons ce qui permet la substitution du chlorure par la fonction phénol de la tyrosine. Après la formation des complexes illustrés au **Schéma 9**, ceux-ci peuvent être employés pour aryle la tyrosine, souvent après la déprotonation de la fonction phénol (voir **Éq 30**).



**Éq 30** : Arylation de la tyrosine par  $S_NAr$  via la formation d'un complexe métal-arène à base de Mn(I), Fe(II) ou Ru(II)

#### 4.2.2. Arylation catalysée au Cu(I)

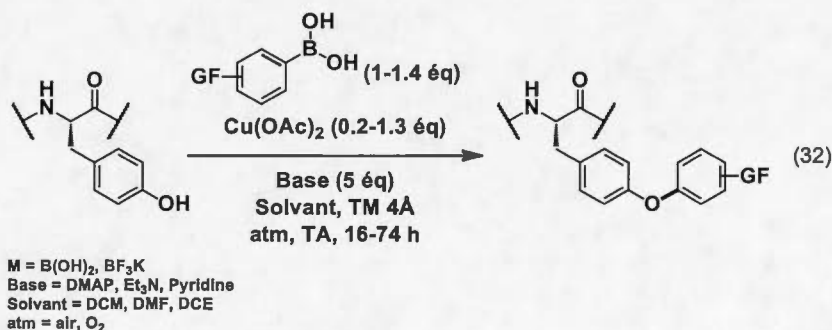
La méthodologie développée par Buchwald a été utilisée pour effectuer l'arylation de la fonction phénol de la tyrosine, soit en utilisant des halogénures d'aryles avec l'ajout d'une base, un ligand, ainsi qu'un catalyseur de Cu(I) (voir **Éq 31**). Ce couplage-croisé fonctionne mieux en utilisant des iodures d'aryles comme source d'aryle (versus ses homologues chlorés et bromés), car la réaction d'addition oxydante dans la liaison C-X se fait mieux avec les iodures. D'autre part, les ligands les plus employés dans ce type de réaction sont à base d'acides aminés, soient la *N,N*-diméthylglycine, la *L*-proline, ainsi que son dérivé protégé, la *N*-méthyl-*L*-proline.<sup>62</sup>



**Éq 31** : Arylation de la tyrosine via un couplage de type Buchwald catalysé au Cu(I)

#### 4.2.3. Arylation catalysée au Cu(II)

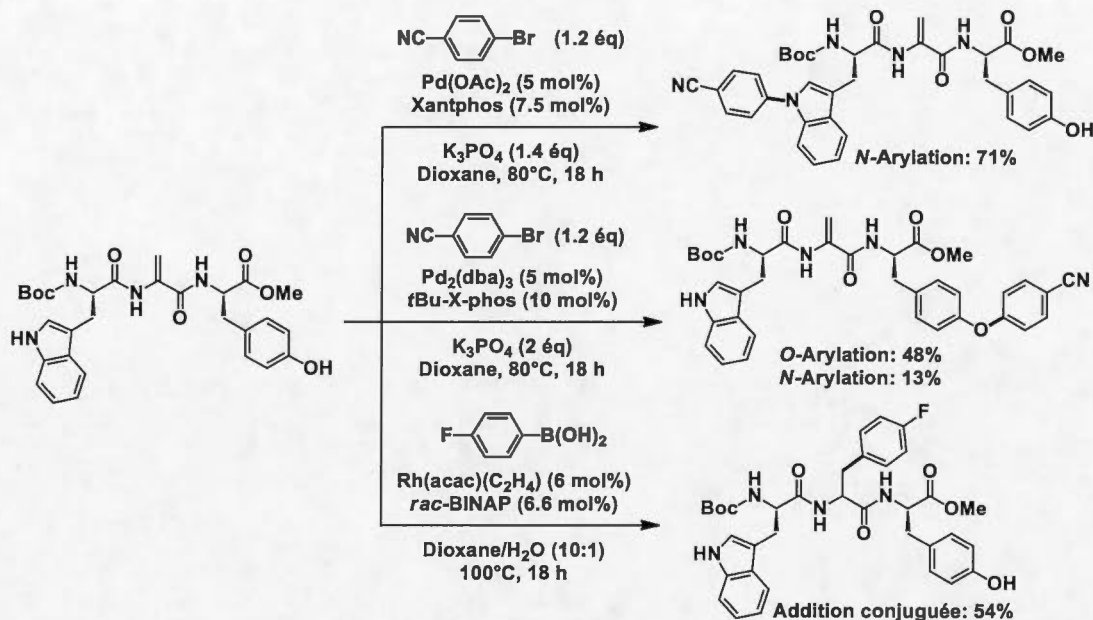
Certains laboratoires ont rapporté l'arylation de la tyrosine en effectuant un couplage-croisé de type Chan-Evans-Lam. Ces derniers ont effectué l'arylation par catalyse au Cu(II), soit le  $\text{Cu}(\text{OAc})_2$ , en utilisant des acides boroniques ( $\text{ArB}(\text{OH})_2$ ) comme source d'aryle et des bases azotées telles que la DMAP, la  $\text{Et}_3\text{N}$  et la pyridine (voir **Éq 32**).<sup>63</sup> Cette réaction est effectuée à température ambiante et pour être en mesure d'utiliser une quantité substœchiométrique de cuivre, une atmosphère d' $\text{O}_2$  peut être employée. Cependant, au meilleur de nos connaissances, aucun laboratoire n'a rapporté l'arylation de la tyrosine en employant des triarylbismuthines comme source d'aryle.



**Éq 32** : Arylation de la tyrosine via un couplage de type Chan-Evans-Lam catalysé au cuivre

#### 4.2.4. Arylation catalysée au Pd(0)

Willis et Frost<sup>64</sup> ont publié un article très intéressant où ils se concentrent sur l'arylation chimiosélective de trois acides aminés, soient la tyrosine, la déhydroalanine (dhAla) et le tryptophane. L'arylation de la tyrosine a été effectuée en utilisant une source de Pd(0), à savoir le Pd<sub>2</sub>(dba)<sub>3</sub>, une phosphine (*t*Bu-X-phos) au départ d'un bromure d'aryle comme source d'aryle. L'arylation de la déhydroalanine a été effectuée par une addition conjuguée catalysée au Rh(acac)(C<sub>2</sub>H<sub>4</sub>) avec du *rac*-BINAP et un acide boronique comme source d'aryle. L'arylation du tryptophane a été effectuée en utilisant du Pd(OAc)<sub>2</sub> comme catalyseur, la Xantphos comme ligand et un bromure d'aryle comme source d'aryle. L'arylation conjuguée sur la déhydroalanine, ainsi que l'arylation du tryptophane ont procédé de façon chimiosélective avec des rendements de 54% et 71% respectivement. Or, dans le cas de la tyrosine, la réaction a donné un rendement de 48% d'*O*-arylation et 13% de N-arylation. Cela étant dit, les méthodes d'arylation de tryptophane et d'addition conjuguée sur la déhydroalanine sont chimiospécifiques alors que l'arylation de la tyrosine est chimiosélective (voir **Schéma 10**).



**Schéma 10 :** Arylation chimiosélective de tryptophane, déhydroalanine et tyrosine via couplage-croisé au Pd(II), addition conjuguée catalysée au Rh(I) et couplage-croisé catalysé au Pd(0) respectivement

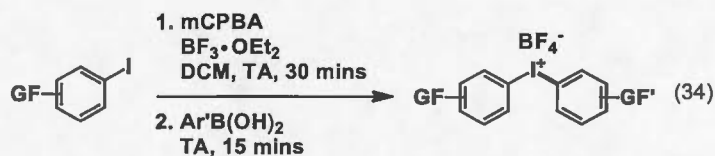
#### 4.2.5. Iode hypervalent

En outre, le groupe d'Olofsson<sup>65</sup> a rapporté une méthode d'arylation de la tyrosine via la formation de sels de diaryliodonium, soit une méthode qui ne requiert pas l'emploi de métaux. Le sel de diaryliodonium peut être formé en mélangeant un iodure d'aryle et un aryle quelconque (ou un aryle avec du I<sub>2</sub>) et en rajoutant un oxydant, soit mCPBA, en présence d'acide tel que TfOH, TsOH (voir Éq 33). Ce dernier peut aussi être synthétisé à partir d'un iodure d'aryle, un oxydant et un acide boronique (voir Éq 34).



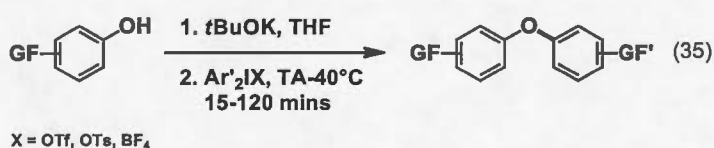


**Éq 33** : Préparation de sels de diaryliodonium par emploi d'iodures d'aryles (ou d'I<sub>2</sub>) et un aryle quelconque en présence d'un oxydant



**Éq 34** : Préparation de sels de diaryliodonium par emploi d'iodures d'aryles, un oxydant et d'un acide boronique

Une fois que les sels de diaryliodonium sont préparés, ils peuvent être utilisés dans une réaction d'arylation de phénols (incluant deux exemples d'arylation de la tyrosine) (voir **Éq 35**).



**Éq 35** : Arylation de phénols en utilisant des sels de diaryliodonium comme source d'aryle

#### 4.3. Marquage d'acides aminés sur des protéines

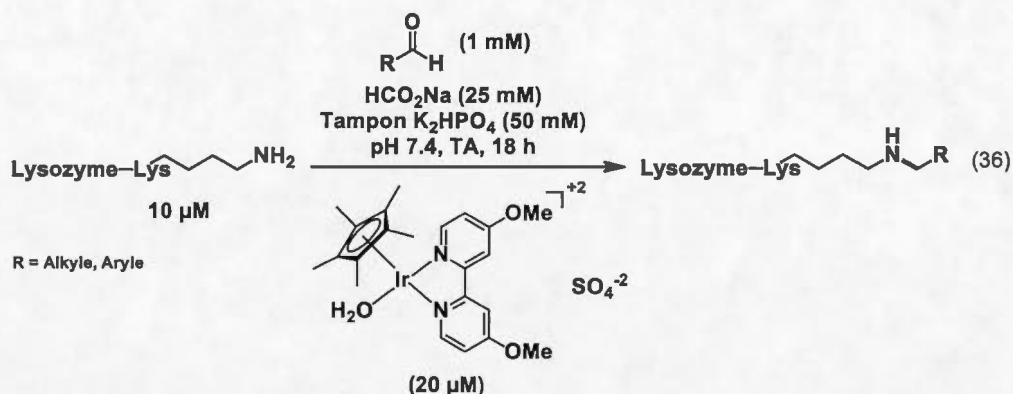
Certains groupes de recherche ont été en mesure de développer des méthodologies pour marquer des acides aminés spécifiques sur des protéines tels que la lysine, le tryptophane, la tyrosine et la cystéine. Par la suite, les modifications

effectuées sur les protéines peuvent être détectées par des techniques analytiques telles que la SMHR (par ajout d'un fragment sur la protéine ou par ajout d'atomes avec une abondance isotopique intéressante telle que le Cl et le Br), la RMN (par marquage isotopique au  $^2\text{D}$ ,  $^{13}\text{C}$  ou en ajoutant du fluor) ou encore la spectrométrie de fluorescence (en ajoutant des sondes fluorophores).

Dans la littérature, on retrouve plusieurs méthodes de modification de protéines.<sup>66</sup> Seulement quelques-unes entre elles seront présentées dans les sections qui suivent.

#### 4.3.1. Modification de la lysine

En ce qui concerne la modification de la lysine, une des stratégies employées est l'amination réductrice de sa fonction amine avec un aldéhyde ou une cétone. Il est possible d'effectuer cette réaction en utilisant le cyanoborohydrure de sodium ( $\text{NaBH}_3\text{CN}$ ) comme réducteur. Cependant, cette méthode nécessite l'utilisation d'un grand excès de réducteur ce qui entraîne la dénaturation de la protéine qui peut être toxique pour des cellules vivantes.<sup>67</sup> Pour contourner ce problème, le groupe de Francis<sup>68</sup> a développé une méthode d'amination réductrice la lysine du lysozyme catalysée à l'iridium en utilisant du formiate de sodium comme source d'hydrure (voir **Éq 36**). En effet, le formiate de sodium forme un hydrure (et un dégagement de  $\text{CO}_2$ ) qui vient substituer la molécule d'eau sur l'Ir(I) menant ainsi au complexe actif dans cette réaction. Après l'addition des fragments d'amination réductrice sur les résidus de lysine, ces modifications peuvent être détectées par spectrométrie de masse où on peut voir un signal qui correspond à chaque amination réductrice effectuée sur la même protéine ( $Mm + xn^+$ ; ou  $Mm$  = masse molaire de la protéine,  $x$  = masse molaire du fragment ajouté,  $n$  = nombre entier (0, 1, 2, 3)).

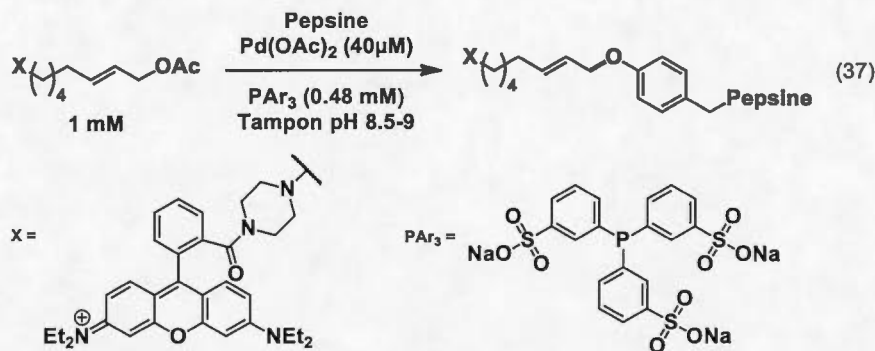


**Éq 36 :** Amination réductrice de la lysine du lysozyme en utilisant du formiate de sodium par catalyse à l'Ir(I)

#### 4.3.2. Modification de la tyrosine

En ce qui a trait au marquage de la tyrosine, le groupe de Francis a été en mesure de modifier ce résidu de façon chimiosélective par deux méthodes différentes, soit via une réaction de Tsuji-Trost avec un acétate allylique par catalyse au Pd(0) ou soit via une réaction de type Mannich en rajoutant une aniline et un aldéhyde (le phénol est considéré comme un énol).

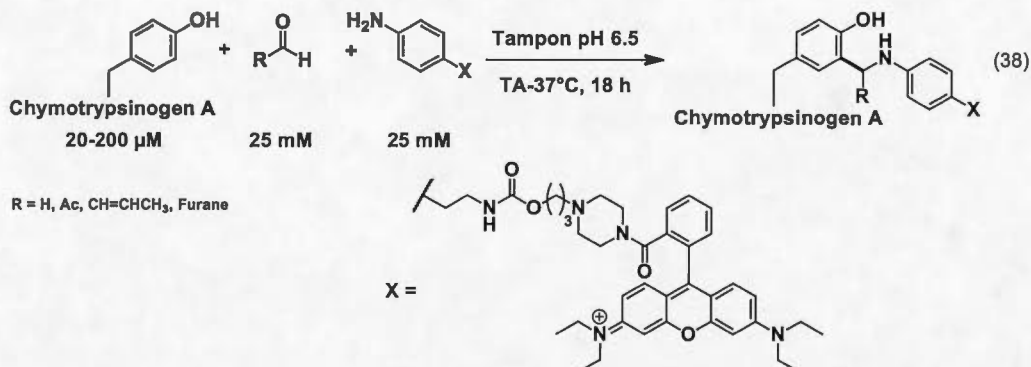
En ce qui concerne la réaction de Tsuji-Trost, cette réaction utilise du Pd(OAc)<sub>2</sub> comme source de palladium avec du triphenylphosphine tris-sulfonate (cette phosphine est soluble en milieu aqueux et elle génère du Pd(0) à partir du Pd(II)) dans un tampon phosphate (pH : 8.5-9). Cette réaction est très rapide et elle permet de transférer un acétate allylique comprenant une rhodamine, c'est-à-dire un fluorophore. En effet, ce groupe a été capable d'alkyler certains résidus de tyrosine sur la pepsine (plus particulièrement Y171). Le groupe de Francis a aussi été en mesure de prouver que l'alkylation a eu lieu sur la tyrosine en effectuant une digestion protéolytique de la pepsine alkylée (voir Éq 37).<sup>69</sup>



**Éq 37 :** Alkylation de type Tsuji-Trost d'un résidu de tyrosine sur une protéine avec un acétate allylique comprenant une rhodamine fluorescente

En ce qui touche la réaction de Mannich, cette réaction est effectuée dans un tampon phosphate de pH 6.5 (légèrement acide pour former l'iminium qui sera attaqué par le carbone en alpha du phénol). Cette méthode est surtout intéressante car elle est effectuée en milieu aqueux, elle ne nécessite pas de catalyseur métallique et elle est sélective pour la tyrosine. De plus, cette approche de marquage de la tyrosine permet aussi l'addition d'une sonde fluorescente telle que la rhodamine illustrée ci-dessous (voir **Éq 38**). Après la réaction, la protéine modifiée a été sujet à une digestion protéolytique qui a permis de déterminer que la réaction a bel et bien pris place sur le résidu de tyrosine, plus particulièrement Y146 et Y171 (les autres résidus de tyrosine sont significativement moins accessibles et il est présumé qu'ils ne réagissent pas), en effectuant un analyse MALDI-MS.<sup>70</sup>

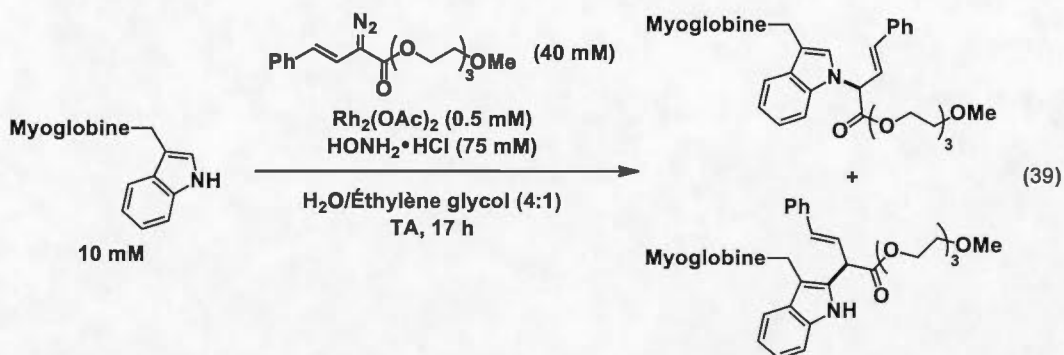




**Éq 38 :** Marquage de résidus de tyrosine avec une rhodamine sur la Chymotrypsinogen A par réaction de Mannich

#### 4.3.3. Modification du tryptophane

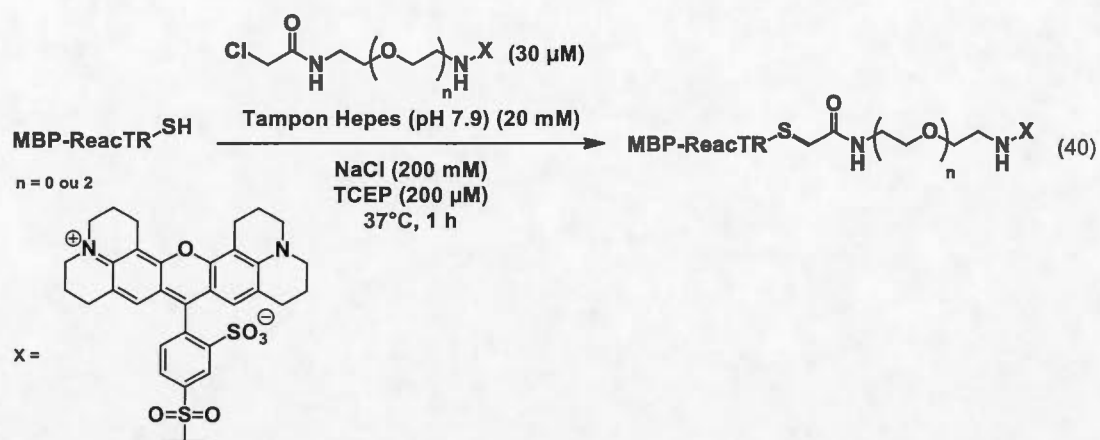
Toujours dans l'optique de la modification de protéines, le groupe de Francis a développé une méthode pour alkyler la tyrosine sur la myoglobine de façon sélective en utilisant des carbénoïdes de rhodium et de l'hydroxylamine en milieu aqueux. Il est à noter que l'alkylation peut livrer le produit de *N*-alkylation ou de *C*-alkylation en position C-2 du tryptophane (voir **Éq 39**). Ce phénomène a été élucidé en effectuant la réaction d'alkylation sur un substrat plus simple, soit le 3-méthylindole. Pour prouver que l'alkylation a pris place sur le tryptophane, une digestion protéolytique a été effectuée et il a été déterminé que la réaction d'alkylation a eu lieu sur les résidus de tryptophane W7 et W14 en analysant par spectrométrie de masse (ionisation par électronébuleur).<sup>71</sup>



**Éq 39 :** Marquage de résidus de tryptophane sur la myoglobine par addition de carbénoïdes de rhodium

#### 4.3.4. Modification de la cystéine

Depuis l'année 1935, les iodoacétamides ont été employés pour modifier et étudier les résidus de cystéine présents sur les protéines, notamment la kératine.<sup>72</sup> Or, il s'avère que la lysine était aussi capable de substituer l'iode de l'iodoacétamide. Conséquemment, les chloroacétamides sont plus fréquemment utilisés, car ils sont plus sélectifs pour l'alkylation de la cystéine.<sup>73</sup> À cet égard, le groupe de Jäschke a utilisé des chloroacétamides comportant une sonde fluorophore (TexasRed) afin de marquer la cystéine sur la protéine MBP-ReacTR (voir **Éq 40**).<sup>74</sup>



**Éq 40 :** Marquage de résidus cystéines sur la protéine MBP-ReacTR par addition d'un chloroacétamide contenant une sonde fluorophore (TexasRed)

## CONCLUSION

Pour conclure, nous avons été en mesure d'établir que l'utilisation des triarylbismuthines fonctionnalisés comme source d'aryle est très efficace pour effectuer des couplages-croisés catalysés au palladium ou au cuivre. Ces réactifs possèdent une faible toxicité et sont stables à l'air et à l'eau. De plus, les réactifs d'organobismuth sont capables de tolérer toutes sortes de groupes fonctionnels et de conditions (acide ou basique). Ces derniers peuvent également tolérer la présence de nucléophiles ainsi que des oxydants et des réducteurs. Afin d'obtenir des triarylbismuthines hautement fonctionnalisés, nous avons été capable d'effectuer la dérivatisation des groupes fonctionnels présents sur ces derniers.

L'obtention des structures rayons X de certains organobismuthines nous a permis de visualiser l'effet de la position et la nature des substituants sur la structure de ces derniers. Notamment, en présence d'atomes donneurs, tel que l'oxygène, sur des organobismuthines substitués en *ortho*, il peut y avoir une interaction entre l'atome donneur et le bismuth (tel qu'illustré par les structures rayons X que nous avons obtenues).

Les organobismuthines peuvent être utilisés pour aryle toutes sortes de substrats via des couplages-croisés catalysés au palladium (pour former des liens C-C) à partir d'halogénures d'aryle et au cuivre (pour former des liens C-N, C-O et C-S) à partir de nucléophiles de type N-H, O-H et S-H.

En ce qui concerne les couplages-croisés catalysés au palladium, nous avons déterminé que l'ajout de LiCl ou de CuI en tant qu'additif et l'emploi de bases telles que Cs<sub>2</sub>CO<sub>3</sub>, K<sub>3</sub>PO<sub>4</sub> et Rb<sub>2</sub>CO<sub>3</sub> était bénéfique pour le rendement de la réaction. De plus, les couplages-croisés catalysés au palladium ont l'avantage d'être capable de transférer leurs trois groupes organiques.



En ce qui concerne les couplages-croisés catalysés au cuivre, nous avons été en mesure d'aryler une grande variété de substrats, notamment des hétérocycles azotés, des phénols, des aminoalcools, etc.

Compte tenu de ceci, nous avons transposé nos méthodes à l'arylation d'acides aminés. Nous avons effectué l'arylation de la tyrosine, la cystéine, l'histidine, le tryptophane, la sérine, la lysine, la glutamine, la thréonine et l'acide glutamique. En somme, nous avons déterminé que notre méthode d'arylation démontrait une certaine sélectivité envers l'arylation de la tyrosine. Par la suite, nous avons démontré que nous étions capables de transférer une grande variété d'aryles sur un tripeptide contenant un résidu de tyrosine. En fait, nous avons pu aryle la tyrosine avec des aryles portant soit un atome de chlore ou de brome, ce qui pourrait être très utile pour marquer ce résidu dans des peptides afin de le suivre à travers de processus biochimiques en utilisant des techniques analytiques telle que la MSHR. Finalement, nous avons pu aryle la tyrosine de façon chimiosélective sur un dipeptide contenant à la fois la tyrosine et le tryptophane.

Malgré le fait qu'en catalyse au cuivre, les organobismuthines ne peuvent transférer qu'un aryle sur trois, cette méthode est très puissante et permet l'arylation d'une panoplie de substrats de type N-H, O-H et S-H. Ces types de couplages-croisés peuvent aussi être effectués en utilisant des acides boroniques comme source d'aryle par catalyse au cuivre (couplage de type Chan-Evans-Lam), mais il est important de considérer l'emploi d'organobismuthines comme une méthode alternative afin d'effectuer des couplages qui seraient difficiles en utilisant les acides boroniques.

## ANNEXE A

### CONTRIBUTIONS SCIENTIFIQUES

#### Article 1 : Synthesis of Highly Functionnalized Triarylbismuthines by Functional Group Manipulation and Use in Palladium- and Copper-Catalyzed Arylation Reactions

Hébert, M.; Petiot, P.; Benoit, E.; Dansereau, J.; Ahmad, T.; Le Roch, A.; Ottenwaelder, X.; Gagnon, A. *J. Org. Chem.* **2016**, *81*, 5401-5416.

#### Mise en Contexte

Au cours des dernières années, notre groupe a développé des méthodologies nous permettant de former des liaisons C–C, par catalyse au palladium, et des liaisons C–O et C–N, par catalyse au cuivre, en utilisant des triarylbismuthines fonctionnalisés. Étant donné que la synthèse des organobismuthines implique l'ajout d'une espèce organométallique (organomagnésien, organolithien, organozinznique, etc.) sur un sel de bismuth (typiquement BiCl<sub>3</sub>), certains groupes fonctionnels sensibles aux nucléophiles (aldéhyde, cétone, ester, etc.) ou aux bases fortes (protons acides de type N–H, O–H, S–H) ne peuvent pas être présents sur l'espèce organométallique. Dans cette optique, nous avons été en mesure d'effectuer une manipulation de groupes fonctionnels directement sur les organobismuthines afin d'obtenir des dérivés hautement fonctionnalisés.

Dans la littérature, lors des réactions de couplages-croisés catalysés au palladium, on emploie souvent des acides boroniques ou des organostannanes comme source organométallique. Cependant, en raison de leur toxicité, les organostannanes

sont de moins en moins utilisés. Par conséquent, les organobismuthines sont une excellente alternative aux acides boroniques pour les réactions de couplage difficiles.

En ce qui concerne les couplages-croisés catalysés au cuivre, encore une fois, les acides boroniques sont souvent utilisés dans des réactions de type Chan-Evans-Lam. Cependant, il s'avère que les organobismuthines donnent des résultats comparables ou supérieurs à ceux obtenus avec les acides boroniques. D'où l'intérêt d'investiguer leur efficacité à transférer des aryles hautement fonctionnalisés sur des substrats de type N-H et O-H.

### Contributions personnelles

La synthèse des organobismuthines hautement fonctionnalisés employés dans les réactions de couplage-croisé est une de mes contributions pour cet article. Notamment, j'ai effectué la synthèse de **1a**, **1d**, **1m**, **1n**, **1o** (Scheme 3) **1s**, **1t**, **21a**, **21b**, **21c** (eqs 1-5), **21d**, **21e**, **21f**, **21g** (Scheme 4) et **21h** (Scheme 5). De plus, j'ai cristallisé les organobismuthines **1s**, **1t** et **21e** afin d'obtenir les structures rayons X (Figure 1).

En ajout, j'ai effectué des couplages-croisés catalysés au cuivre avec l'emploi de triarylbismuthines trivalents et pentavalents. En effet, j'ai synthétisé les produits **17a**, **34a**, **34c**, **34d**, **35b** (Scheme 6), **37a**, **37b** (Scheme 7) et **37e** (Table 4).

Finalement, j'ai participé à la rédaction de l'article et j'ai préparé la partie expérimentale.

### Contributions des coauteurs

Pauline Petiot a également effectué la synthèse d'organobismuthines et elle a préparé la majorité des composés de la Table 1 et la moitié de la Table 2. Elle a aussi préparé le Scheme 9.

Emeline Benoit a complété la Table 1 et elle a fait un quart de la Table 2.

Julien Dansereau a complété un quart de la **Table 1**, la moitié du **Scheme 6** et a préparé la **Table 3**.

Tabinda Ahmad a préparé la majorité du Scheme 7 et elle a recristallisé les organobismuthines **1f**, **1i**, **1k**, **1n**, et **21c** afin d'avoir les structures rayons X.

Adrien Le Roch a effectué la repurification de certains produits impurs.

Xavier Ottenwaelder a effectué les analyses par rayons X.

Alexandre Gagnon était le chef d'équipe pour les expériences et les analyses effectuées. Il était également en charge de la recherche bibliographique et de la rédaction de l'article.



## **Manuscript 2: Chemoselective Copper-Catalyzed *O*-Arylation of Small Tyrosine-Containing Peptides using Functionnalized Organobismuth Reagents**

Hébert, M.; Le Roch, A.; Archambault, M.-J.; Pinsonneault, F.; Lachance, H.; Gagnon, A. *Org. Lett.* **2016**, (En préparation)

### **Mise en contexte**

Depuis 2014, le groupe du Pr. Gagnon a utilisé des triarylbismuthines fonctionnalisées pour effectuer de réactions de couplage-croisé catalysées au cuivre afin de former des liaisons C–O et C–N. Les indoles, les imidazoles, les phénols et les aminoalcools sont quelques exemples de substrats arylés en utilisant cette méthodologie. Étant donné que ces groupes fonctionnels se retrouvent sur les chaînes latérales des acides aminés, notamment le tryptophane, l’histidine, la tyrosine, la sérine et la thréonine, cette méthodologie a été transposée à l’arylation d’acides aminés et de peptides.

La modification des chaînes latérales d’acides aminés peut s’avérer très utile en chimie médicinale ou en biochimie. En effet, plusieurs médicaments possèdent des motifs d’acides aminés dans leur structure. En biochimie, il peut être utile de marquer certains acides aminés dans des peptides ou des protéines pour être en mesure de les suivre avec des techniques analytiques à travers de des processus biochimiques.

Dans la littérature, les méthodes d’arylation des chaînes latérales d’acides aminés qu’on retrouve sont surtout par catalyse au Cu(I) avec des halogénures d’aryles ou au Cu(II) utilisant des acides boroniques ou des trifluoroborates de potassium. Or, l’utilisation des organobismuthines pour l’arylation des chaînes latérales des acides aminés n’a toujours pas été rapportée dans la littérature (hormis l’arylation du tryptophane par le groupe de Gagnon).

### Contributions personnelles

La synthèse des organobismuthines hautement fonctionnalisées employés dans les réactions de couplage-croisé est une de mes contributions pour cet article. Notamment, j'ai effectué la synthèse des organobismuthines **6a**, **6b**, **6c**, **6d**, **6e**, **6f**, **6i**, **6j**, **6k**, **6l**, **6m**, **6n**, **6o**, **6p**, **6q**, **6r**, **6s** et **6t** (Scheme 5). Ensuite, j'ai effectué l'arylation des acides aminés **11a**, **11b**, **11d**, **11e**, **11f**, **11h** et **11i** (Scheme 3). Par la suite, j'ai synthétisé les tripeptides **12**, **13** et **15** (Scheme 4). Par après, j'ai préparé les tripeptides arylés **16a**, **17**, **18a**, **18b** et **19** (Scheme 4). J'ai effectué la table d'optimisation illustrée à la Table 1. De plus, j'ai effectué les arylations des tripeptides **16b**, **16c**, **16d**, **16e**, **16g**, **16h**, **16i**, **16j**, **16k**, **16l**, **16m**, **16n**, **16o**, **16p**, **16q** et **16r** (Scheme 5). En ajout, j'ai préparé le dipeptide **23** (Scheme 6).

Finalement, j'ai participé à la rédaction de l'article et j'ai préparé la partie expérimentale.

### Contributions des coauteurs

Adrien Le Roch a préparé le composé **11c**, il a fait un quart du Scheme 5 et il a fait le Scheme 6. Il a également effectué la synthèse des peptides **12** et **20**.

Marie-Jeanne Archambault a effectué la synthèse des composés **11f**, **11g** et elle a effectué la synthèse des peptides **14** et **15**.

Francis Pinsonneault a commencé le projet d'arylation des acides aminés en testant plusieurs conditions de réaction qui se retrouvent dans la table d'optimisation (Table 1).

Hugo Lachance est celui qui a eu l'idée pour le projet d'arylation des acides aminés.

Alexandre Gagnon était le chef d'équipe pour les expériences et les analyses effectuées. Il a aussi fait la recherche bibliographique et la rédaction de l'article.

## **ANNEXE B**

### **Article 1 : Synthesis of Highly Functionnalized Triarybismuthines by Functional Group Manipulation and Use in Palladium- and Copper-Catalyzed Arylation Reactions**



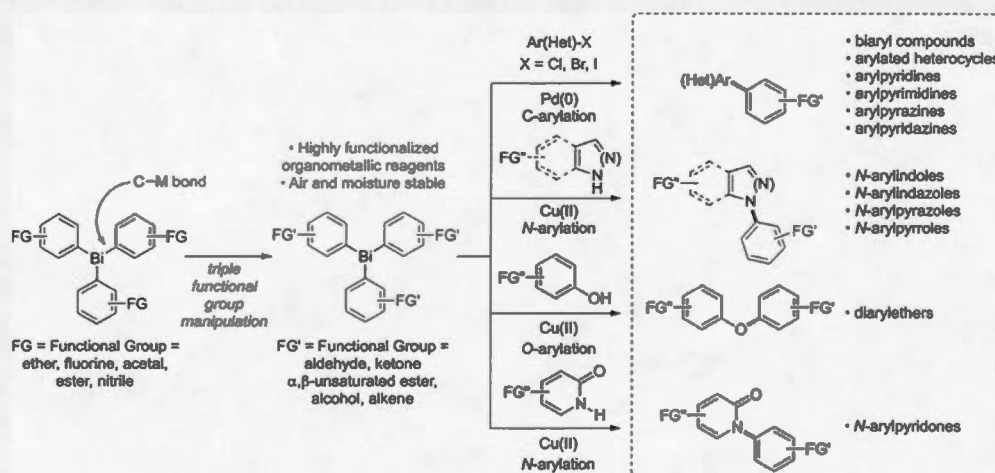
# Synthesis of Highly Functionalized Triarylbiomuthines by Functional Group Manipulation and Use in Palladium- and Copper-Catalyzed Arylation Reactions

Martin Hébert,<sup>†</sup> Pauline Petiot,<sup>†</sup> Emeline Benoit,<sup>†</sup> Julien Dansereau,<sup>†</sup> Tabinda Ahmad,<sup>†</sup> Adrien Le Roch,<sup>†</sup> Xavier Ottenwaelder,<sup>‡</sup> and Alexandre Gagnon<sup>\*,†</sup>

<sup>†</sup>Université du Québec à Montréal, Département de Chimie, C.P. 8888, Succursale Centre-Ville, Montréal, Québec, Canada, H3C 3P8

<sup>‡</sup>Concordia University, Department of Chemistry and Biochemistry, 7141 Sherbrooke Street West, Montréal, Québec, Canada, H4B 1R6

## Supporting Information



**ABSTRACT:** Organobismuthines are an attractive class of organometallic reagents that can be accessed from inexpensive and nontoxic bismuth salts. Triarylbiomuthines are particularly interesting due to their air and moisture stability and high functional group tolerance. We report herein a detailed study on the preparation of highly functionalized triarylbiomuth reagents by triple functional group manipulation and their use in palladium- and copper-catalyzed C-, N-, and O-arylation reactions.

## INTRODUCTION

Triarylbiomuthines (also known as triarylbiomuthanes) are a class of organometallic reagents that can be prepared from inexpensive and nontoxic bismuth salts.<sup>1,2</sup> These reagents are particularly attractive since they are air and moisture stable and can be purified by simple flash chromatography or crystallization. Moreover, organobismuth reagents are remarkably tolerant to numerous functional groups, making them highly suitable for methodology development.<sup>3</sup> They have also found applications in total synthesis,<sup>4</sup> in the preparation of transition metal complexes,<sup>5</sup> as catalysts for polymerization reactions,<sup>6</sup> and in medicinal chemistry.<sup>7</sup> Organobismuth reagents are divided into two main classes: trivalent and pentavalent organobismuthines, with bismuth in the +3 and +5 oxidation levels, respectively. Both classes have found applications in synthesis. For example, Barton and Finet reported in the 1980s a series of arylation reactions using triphenylbismuth and triphenylbismuth diacetate as arylating agents.<sup>8</sup> More recently,

Rao greatly contributed to expanding the use of this class of reagents in organic synthesis and more specifically in palladium-catalyzed reactions.<sup>9</sup>

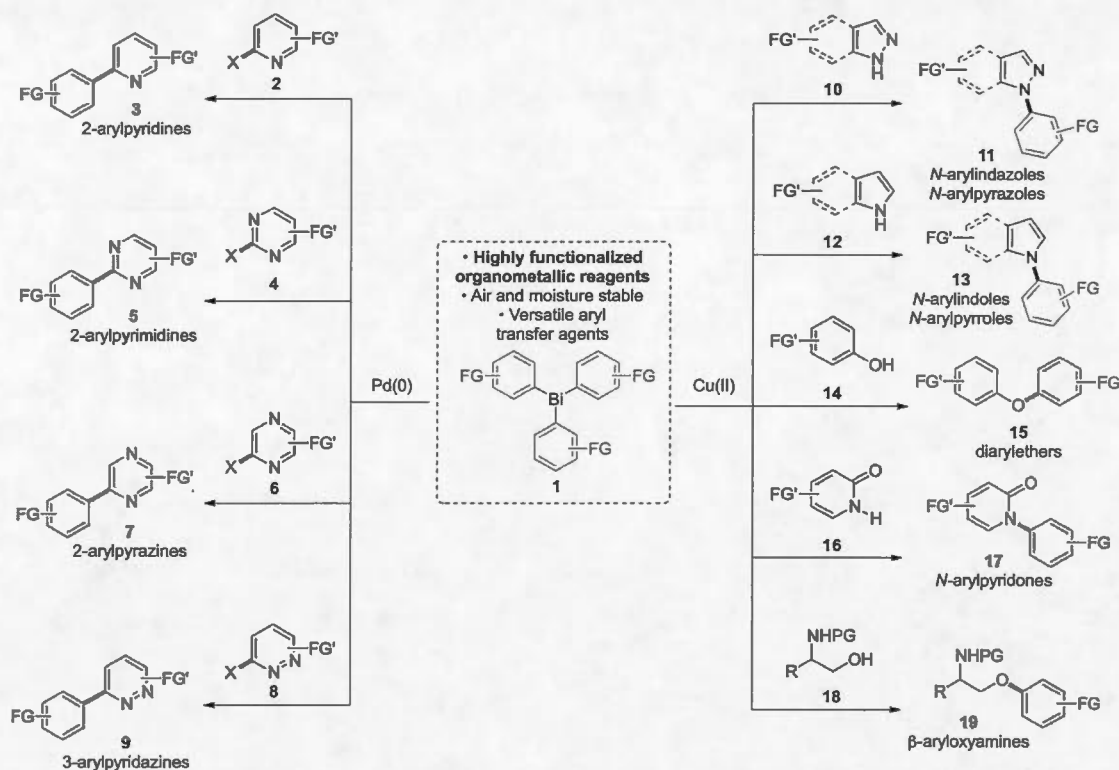
Our group has reported in recent years a portfolio of methods for the formation of C–C,<sup>10</sup> C–N,<sup>11</sup> and C–O<sup>12</sup> bonds using functionalized trivalent organobismuth reagents. These methods allow the transfer of functionalized aryl groups on scaffolds as diverse as pyridines 2, pyrimidines 4, pyrazines 6, pyridazines 8, indazoles and pyrazoles 10, indoles and pyrroles 12, phenols 14, pyridones 16, and 1,2-aminoalcohols 18, providing access to a range of medically relevant compounds (Scheme 1). These methodologies operate under mild conditions and tolerate a wide variety of functional groups on both coupling partners.

Received: April 7, 2016

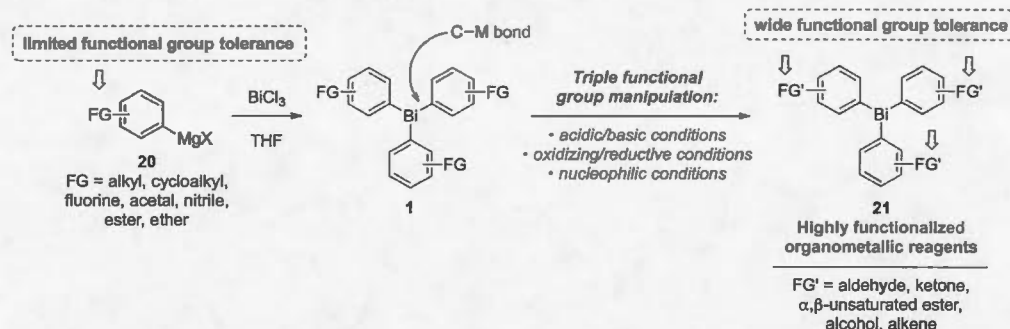
Published: May 27, 2016



Scheme 1. Triarylbiomuthines 1 in Arylation Reactions: Access to Highly Functionalized and Medicinally Relevant Compounds



Scheme 2. Synthesis of Highly Functionalized Organobismuthines 21 by Functional Group Manipulation Directly on Organometallic Species 1 (FG = Functional Group)



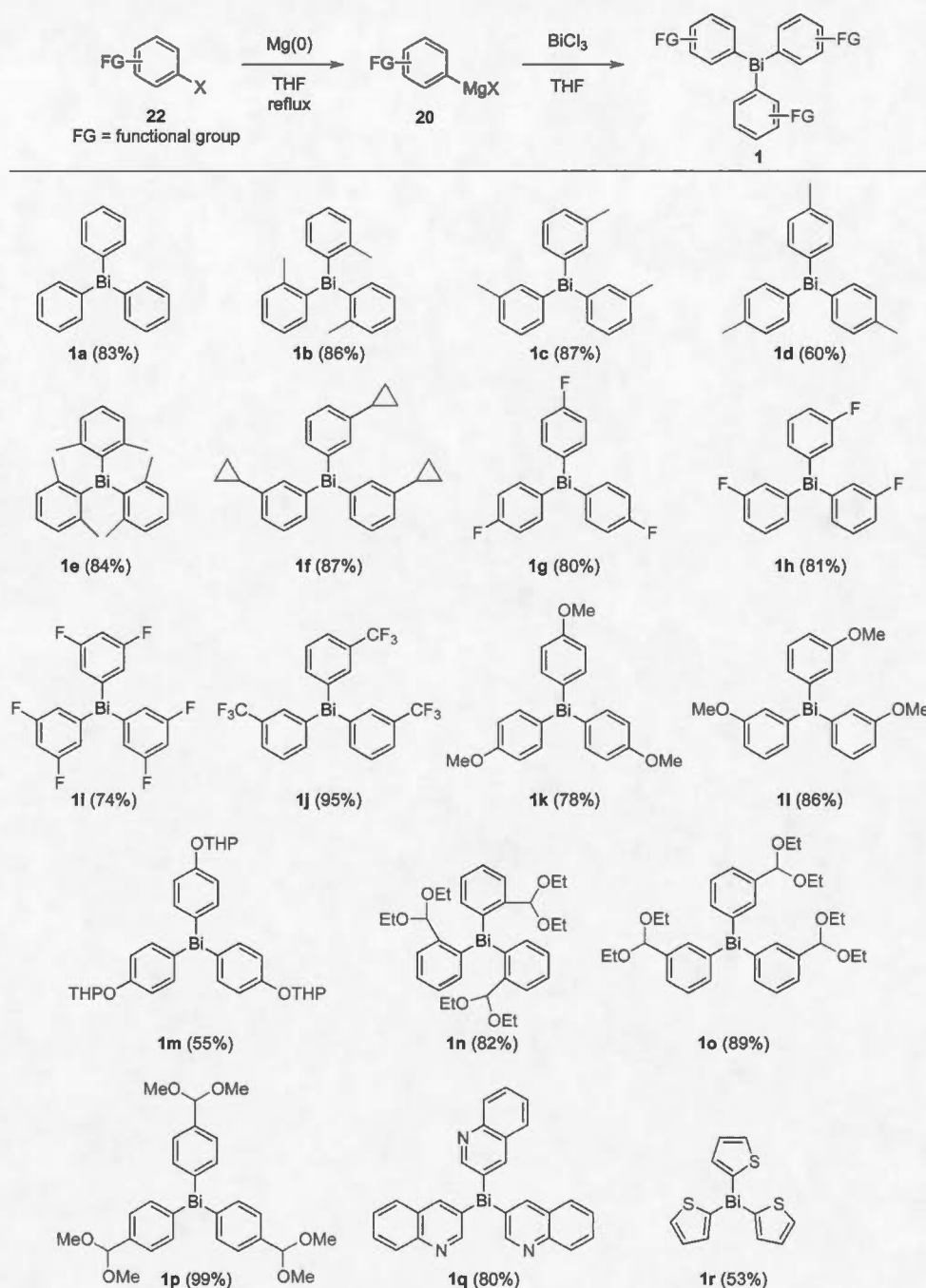
Triarylbiomuth reagents **1** can be easily accessed via addition of Grignard reagents **20** onto bismuth chloride (Scheme 2). However, due to the high reactivity of organomagnesium reagents, organobismuthines bearing electrophilic or acidic functional groups cannot be synthesized directly using this approach. Condon reported an elegant and powerful method to prepare functionalized organobismuth reagents by the addition of organozinc reagents obtained from a cobalt–zinc metal–halogen exchange reaction on aryl halides.<sup>13</sup> While this method is quite general, groups bearing acidic protons such as alcohols cannot be introduced using this methodology. Therefore, alternative methods are still desirable to access highly functionalized organobismuthines. The strategy that we explored consists of introducing the incompatible functional group by performing a functional group transformation directly on the organobismuth species. In the course of our studies, we found that the C–Bi bond in organobismuthines is remarkably resistant to acidic, reductive, and even oxidative conditions, thus enabling the transformation of the functional group FG in

**1** into a more elaborated functional group FG' in **21** (Scheme 2). This reaction actually corresponds to a triple group manipulation on a single substrate. To be efficient, it therefore requires a high level of control of the reaction conditions since, to a first approximation, the overall yield is given by the cube of the yield-per-function. We report herein an extensive study on the successful preparation of highly functionalized triarylbiomuth reagents using this approach and their use in palladium- and copper-catalyzed arylation reactions.

## RESULTS AND DISCUSSION

**a. Preparation of Triarylbiomuthines.** We began by synthesizing a set of substituted and unsubstituted organobismuthines **1a–r** bearing simple functional groups by adding organomagnesium reagents **20** to bismuth chloride (Scheme 3). The Grignard reagents **20** were prepared by reacting the corresponding aryl halides **22** with metallic magnesium at reflux of ether or THF. Using this approach, 18 different

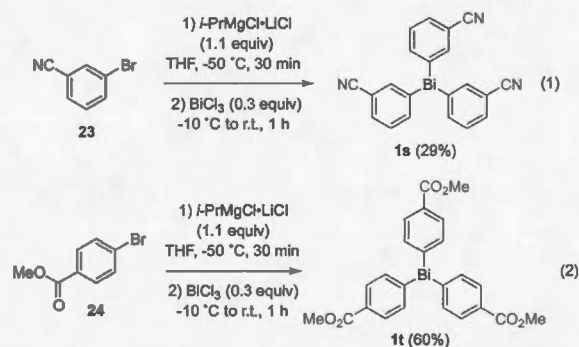
Scheme 3. Direct Preparation of Organobismuthines 1a–r by Addition of Organomagnesium Reagents 20 to Bismuth Chloride



unsubstituted and *ortho*, *meta*, and *para* substituted triaryl- and triheteroaryl-bismuthines were synthesized in 53% to 99% yields.

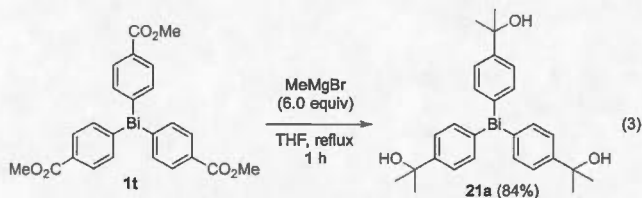
For the introduction of nitriles and esters, the required organomagnesium reagents were generated using Knochel's procedure.<sup>14</sup> Thus, addition of the isopropylmagnesium chloride lithium chloride complex onto 3-cyanobromobenzene 23 or methyl 4-bromobenzoate 24 at  $-50\text{ }^{\circ}\text{C}$  afforded the corresponding Grignard reagents, which were then reacted with bismuth chloride to provide 1s and 1t in moderate yields (eqs 1 and 2).

To begin our exploration of functional group manipulation on organobismuthines, we added 6.0 equiv of methylmagne-



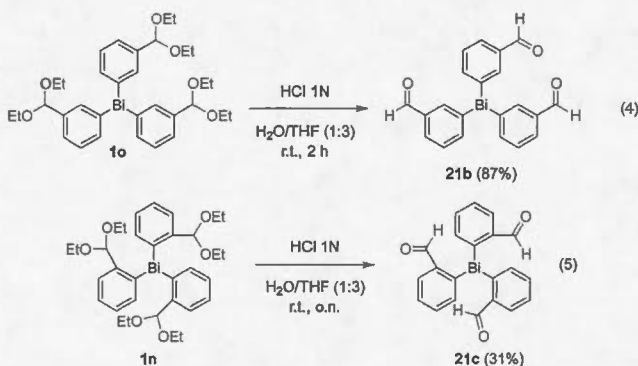


sium bromide on the ester derivative **1t**, affording **21a** in 84% yield (i.e., 94% yield per ester function) (eq 3). This simple



transformation demonstrates that the concept of functional group modification in the presence of a C–Bi bond is viable, giving us an impetus to explore other reactions directly on organobismuthines.

Due to their high electrophilicity, aldehydes cannot be present on organomagnesium reagents. Therefore, the preparation of organometallic reagents bearing aldehydes is only possible with less electropositive metals such as tin,<sup>15</sup> boron,<sup>16</sup> zinc,<sup>17</sup> or indium.<sup>18</sup> However, the toxic nature of tin greatly limits its use in synthesis. Although many organoboron acids bearing aldehydes are commercially available, their use in metal-catalyzed reactions often requires extensive optimization of the reaction conditions. Conversely, arylzinc reagents possessing aldehydes have proved to be very useful in palladium-catalyzed cross-coupling reactions, but strictly anhydrous conditions are mandatory due to their high sensitivity to moisture. Finally, while organoindiums have generated great interest in the synthetic community over the past decade, the cost of indium greatly limits their application in synthesis. In this context, organobismuthines bearing aldehydes can fill an important need in organic synthesis. Starting from tris((3-diethoxymethyl)phenyl)bismuthine **1o**, we prepared the tris-formyl derivative **21b** in excellent yield by hydrolysis of the acetal function under acidic conditions (eq 4).



Surprisingly, using the same conditions on tris((2-diethoxymethyl)phenyl)bismuthine **1n** led to a much lower yield of the *ortho* analogue **21c** (eq 5). While this overall yield is low, it still corresponds to a 67% yield per acetal function. This example illustrates the need for very efficient conditions to get a satisfactory triple functional group modification.

We then took advantage of the versatility of the aldehyde function in **21b** to introduce other functional groups on the organobismuth species (Scheme 4). For example, addition of methylmagnesium bromide on **21b** provided tris(3-(1-hydroxyethyl)phenyl)bismuthine **21d** in quantitative yield. Reduction of the aldehyde function in **21b** was also accomplished using sodium borohydride, affording tris(3-(hydroxymethyl)phenyl)bismuthine **21e** in 97% yield. An olefination reaction using

Wittig conditions was then performed on **21b**, leading to the corresponding cinnamyl ester and vinyl derivatives **21f** and **21g** in 80% and 82% yield, respectively. These examples show that organobismuthines are resistant to organometallic and reductive reagents in addition to phosphorus ylides.

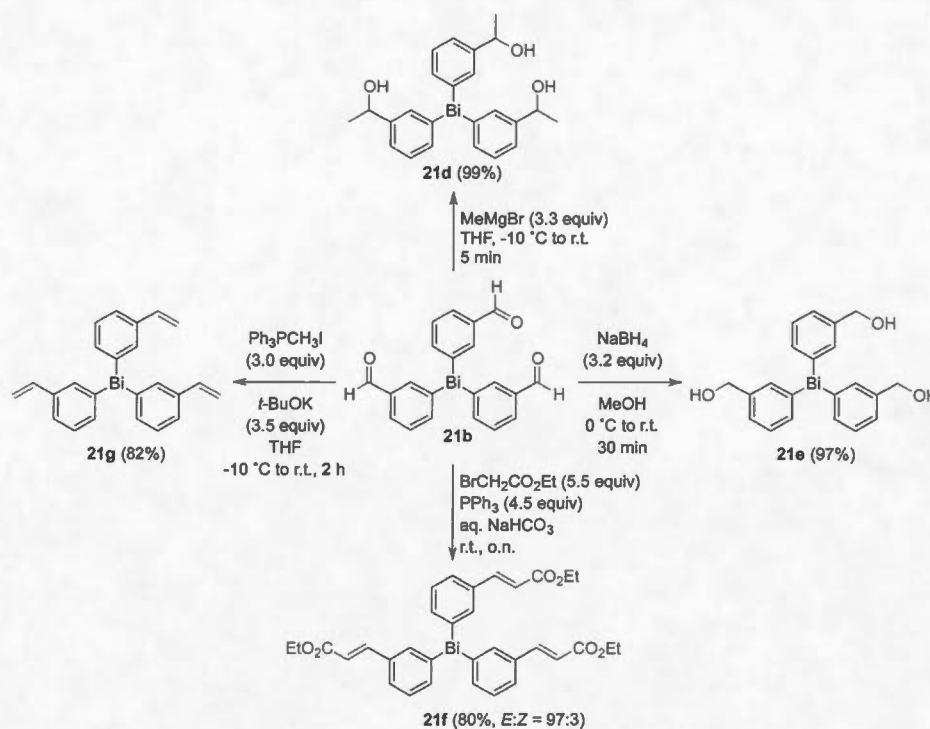
Next, we sought to introduce a ketone on the organobismuth reagent by oxidizing the secondary alcohol in **21d** (Scheme 5). Since trivalent organobismuthines can be oxidized into their pentavalent counterparts by oxidizing agents<sup>19</sup> and by hypervalent iodonium reagents,<sup>8c,20</sup> it was unclear if the bismuth(III) center would tolerate the oxidizing conditions required to oxidize the alcohol into the corresponding ketone. To our satisfaction, tris(methyl ketone) **21h** was obtained in acceptable yield using Dess–Martin periodinane **25**. Alternatively, compound **21h** could also be prepared in 63% yield via Swern oxidation of all three alcohol functions (86% per function).

**b. Structural Characterization of Selected Triarylbi-bismuthines.** To further gain insight into the structure of triarylbi-bismuthines,<sup>21</sup> we crystallized and analyzed by X-ray diffraction several derivatives bearing a cyclopropyl group at the *meta* position (**1f**), fluorine atoms at the 3 and 5 position (**1i**), a methoxy group at the *para* position (**1k**), an acetal at the *ortho* position (**1n**), a cyano group at the *meta* position (**1s**), a carbomethoxy function at the *para* position (**1t**), an aldehyde at the *meta* (**21b**) and *ortho* positions (**21c**), and an hydroxymethyl group at the *meta* position (**21e**).<sup>22</sup> The results show that the organobismuthines have a distorted trigonal pyramidal structure with Bi–C bond lengths ranging from 2.24 to 2.27 Å and C–Bi–C angles between 91° and 98° (Figure 1). These Bi–C bond lengths are consistent with known triarylbi-bismuthines crystal structures<sup>23</sup> and with the 2.256 Å value estimated in the gas phase by gas electron diffraction,<sup>24</sup> denoting innocuous crystal packing effects. The small C–Bi–C angles are due to known relativistic effects and fall within the trend observed when going down the pnictogen family ( $N > P > As > Sb > Bi$ ).

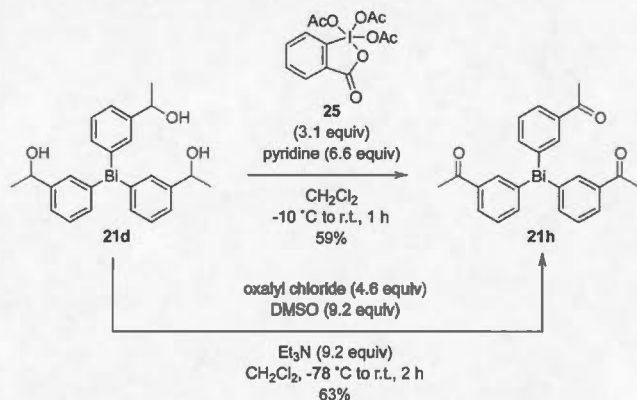
Interestingly, the nature and position of the substituent have little impact on the molecular structure of the organobismuthine, except in the case of the *ortho*-(diethylacetal) and *ortho*-formyl derivatives **1n** and **21c**. In these compounds with donor atoms in the *ortho* functional group, secondary intramolecular interactions are observed between the oxygen of the acetals or the carbonyls and the bismuth center (Bi...O from 3.085 to 3.255 Å in **1n** and from 2.911 to 2.991 Å in **21c**;  $r_{vdW}(\text{Bi}) = 2.07$  Å,  $r_{vdW}(\text{O}) = 1.52$  Å). This is not unprecedented, as bismuth is capable of acting as a weak acceptor toward other ligands or even metals<sup>25</sup> by accepting electrons in its C–Bi  $\sigma^*$  orbitals.<sup>5a,26</sup>

**c. Palladium-Catalyzed Cross-Coupling Reaction between Organobismuthines and Aryl or Heteroaryl Halides.** Multiple reports of palladium-catalyzed cross-coupling reactions involving organobismuth compounds have been disclosed in recent years.<sup>27,28</sup> One important aspect of triarylbi-bismuth reagents is their ability to deliver three aryl groups per equivalent of organometallic reagent in cross-coupling reactions, making them more atom-economical than other conventional Ar–M reagents. However, in most cases, a limited array of functional groups were present on the organobismuth partner. We recently reported a palladium-catalyzed reaction to cross-couple triarylbi-bismuthines with halogenated pyridines, pyrimidines, pyrazines, and pyridazines.<sup>10b</sup> Therefore, with our functionalized organobismuthines in hand, we next explored their reactivity in palladium-catalyzed

Scheme 4. Synthesis of Highly Functionalized Organobismuthines 21d–g by Derivatization of Tris(3-formylphenyl)bismuthine 21b



Scheme 5. Preparation of Ketone-Bearing Organobismuthine Derivative 21h by Dess-Martin and Swern Oxidation Starting from Tris-alcohol 21d



cross-coupling reactions with other electrophiles in order to further probe the scope and functional group tolerance of this transformation.

First, we optimized the reaction conditions using 1.0 equiv of 4-bromobenzaldehyde 26 and 0.4 equiv of organobismuthine 21d. Using our previously reported conditions, we obtained the desired cross-coupling product 27 in only 37% yield (Table 1, entry 1). The observed low yield shows that the coupling of highly functionalized organobismuthines represents a substantial challenge and that the reaction conditions had to be reoptimized. Suspecting that the high temperature was possibly responsible for the observed low yield, we performed the reaction at 80 °C but observed no improvement in yield (entry 2). Changing the catalyst for PdCl<sub>2</sub>(PPh<sub>3</sub>)<sub>2</sub> (entry 3), Pd(OAc)<sub>2</sub>/S-Phos (entry 4), Pd(OAc)<sub>2</sub>/PPh<sub>3</sub> (entry 5), or PEPPSI-*i*Pr<sup>29</sup> (entry 6) did not improve the yield either.

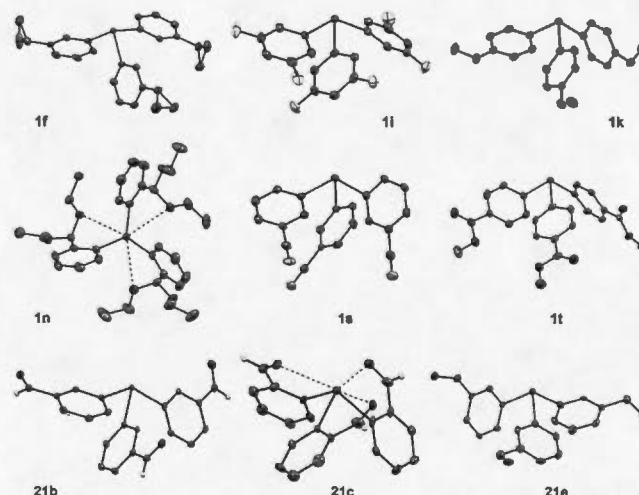
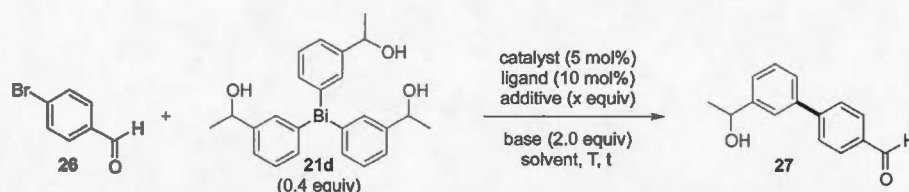


Figure 1. ORTEP view at 50% ellipsoid probability of 1f, 1i, 1k, 1n (one of three independent molecules), 1s, 1t, 21b,<sup>10b</sup> 21c and 21e. Hydrogen atoms are omitted for clarity, except for aldehyde functional groups. Dashed lines indicate weak intramolecular interactions between the bismuth atom and donor oxygen atoms at *ortho* positions.

Knowing the beneficial effect of lithium salts on cross-coupling reactions, as documented by Organ and others,<sup>30</sup> we found a substantial amelioration in yield when 2.0 equiv of lithium chloride were used as an additive (entry 7). Changing the solvent for THF (entry 8), toluene (entry 9), or a mixture of DMF and HMPA (entry 10) was detrimental to the reaction or at best inconsequential. The use of a stronger base such as potassium *tert*-butoxide led to a drastic drop in the yield of the reaction (entry 11). In addition, while potassium carbonate (entry 12) provided a similar yield as cesium carbonate (entry 7), we found that potassium phosphate (entry 13) and



**Table 1.** Optimization of Reaction Conditions for the Palladium-Catalyzed Cross-Coupling Reaction of 21d with 4-Bromobenzaldehyde 26

Entry	Catalyst	Ligand	Additive (x equiv)	Base	Solvent	T (°C)	t (h)	Yield (%) <sup>a</sup>
1	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	N.A.	Cs <sub>2</sub> CO <sub>3</sub>	DMF	130	18	37
2	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	N.A.	Cs <sub>2</sub> CO <sub>3</sub>	DMF	80	6	36
3	PdCl <sub>2</sub> (PPh <sub>3</sub> ) <sub>2</sub>	N.A.	N.A.	Cs <sub>2</sub> CO <sub>3</sub>	DMF	80	6	31
4	Pd(OAc) <sub>2</sub>	S-Phos	N.A.	Cs <sub>2</sub> CO <sub>3</sub>	DMF	80	6	7
5	Pd(OAc) <sub>2</sub>	PPh <sub>3</sub>	N.A.	Cs <sub>2</sub> CO <sub>3</sub>	DMF	80	6	31
6	PEPPSI- <i>i</i> Pr	N.A.	N.A.	Cs <sub>2</sub> CO <sub>3</sub>	DMF	80	6	23
7	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	Cs <sub>2</sub> CO <sub>3</sub>	DMF	80	6	64
8	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	Cs <sub>2</sub> CO <sub>3</sub>	THF	80	6	13
9	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	Cs <sub>2</sub> CO <sub>3</sub>	Toluene	80	6	25
10	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	Cs <sub>2</sub> CO <sub>3</sub>	DMF/HMPA <sup>b</sup>	80	6	59
11	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	<i>t</i> -BuOK	DMF	80	6	14
12	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	K <sub>2</sub> CO <sub>3</sub>	DMF	80	6	69
13	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	K <sub>3</sub> PO <sub>4</sub>	DMF	80	6	82
14	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	Rb <sub>2</sub> CO <sub>3</sub>	DMF	80	6	75
15	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (1.0 equiv)	K <sub>3</sub> PO <sub>4</sub>	DMF	80	6	57
16	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	K <sub>3</sub> PO <sub>4</sub> <sup>c</sup>	DMF	80	6	48
17	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	LiCl (2.0 equiv)	K <sub>3</sub> PO <sub>4</sub>	DMF/H <sub>2</sub> O <sup>d</sup>	80	6	73
18	Pd(PPh <sub>3</sub> ) <sub>4</sub>	N.A.	CuI (0.4 equiv)	Cs <sub>2</sub> CO <sub>3</sub>	DMF	80	6	83

<sup>a</sup>Isolated yield of pure product. <sup>b</sup>DMF/HMPA (4:1). <sup>c</sup>1.0 equiv of K<sub>3</sub>PO<sub>4</sub> was used. <sup>d</sup>DMF/H<sub>2</sub>O (5:1).

rubidium carbonate (entry 14) were much more efficient bases in this transformation, providing the desired cross-coupling product 27 in 82% and 75% isolated yield, respectively. Lowering the number of equivalents of lithium chloride (entry 15) or potassium phosphate (entry 16) proved disadvantageous. Interestingly, the yield remained satisfactory when the reaction was run in a 5:1 ratio of DMF/H<sub>2</sub>O, showing that strictly anhydrous conditions are not mandatory (entry 17). Rao recently reported the effect of copper salts in cross-coupling reactions involving organobismuthines.<sup>9a</sup> Simultaneously to Rao, we independently explored the effect of cuprous iodide as an additive and obtained a considerable improvement in the yield of the reaction (entry 18). Conditions from entries 13, 14, and 18 represent a substantial improvement over our previous protocol, as they allow the coupling reaction to be performed at a lower temperature, in a shorter time, and with a higher yield.

Having optimized the conditions for the cross-coupling of organobismuthine 21d, we next investigated the scope of the method using organobismuthines 1a–t and 21a–h with selected arylhalides 28, heteroarylhalides 29, 2-halopyridines 2, 2-halopyrimidines 4, 2-halopyrazines 6, and 3-halopyridazines 8 (Table 2). The choice of the electrophiles was motivated by the presence of functional groups that could demonstrate the applicability of our protocols in the preparation of highly functionalized compounds. In each case, our best conditions from entries 13, 14, and 18 in Table 1 (named Method A, B, and C, respectively, in Table 2) were tested in order to obtain the highest possible yield of each desired product. Thus, 4-(*N*-BOC-aminoethyl)bromobenzene 28a, methyl 4-iodobenzoate 28b, methyl 4-bromobenzoate 28c, 4-bromobenzaldehyde 28d, 2-iodo-4-furaldehyde 29a, 6-chloro-

pyridine-3-carboxaldehyde 2a, 2-acetyl-6-bromopyridine 2b, 2-chloro-4-methylpyrimidine 4a, 2-chloro-6-dimethylaminopyrazine 6a, and 3-chloro-6-phenylpyridazine 8a were engaged in cross-coupling reactions with organobismuthines 1a–t and 21a–h to afford the corresponding products in acceptable to excellent yields. The results demonstrate that the reaction works with aryl bromides (entries 1, 3, 5), aryl or heteroaryl iodides (entries 2, 4, 6), and 2-chloro and 2-bromo nitrogenated heterocycles (entries 7–12). However, entries 2 and 3 show that iodides are slightly more reactive than bromides. The identity of the optimal set of conditions (whether A, B, or C) depended greatly on the specific combination of Ar(Het)–X and Ar<sub>3</sub>Bi. Notwithstanding, these examples demonstrate that our protocols tolerate a wide diversity of functional groups on the electrophile and on the organobismuthine such as BOC-protected amines (30a),  $\alpha,\beta$ -unsaturated esters (30a), esters (30b,c), ethers (30b), nitriles (30c), aldehydes (30d, 31a, 3a), dialkylamines (7a), and acetals (3b, 9b). More striking is the ability of this method to transfer in acceptable to good yields aryl groups that possess functions that are susceptible to competitive arylation, elimination, or oxidation such as alcohols (31a, 7a), ketones (9a), and vinyl groups (30d, 5a).

**d. Copper-Catalyzed *N*-Arylation of Indoles, Pyrroles, Pyrazoles, and Pyridones.** We recently reported a protocol for the copper-catalyzed *N*-arylation of indoles, pyrroles, pyrazoles,<sup>11a</sup> and pyridones<sup>12b</sup> using organobismuthines with tolerance to a wide variety of functional groups. Other research groups have also reported various copper-catalyzed reactions based on trivalent or pentavalent reagents to *N*-arylate nitrogen-containing compounds.<sup>31</sup> To further expand the functional group diversity of our method, we tested some of the highly functionalized organobismuthines prepared in this

Table 2. Palladium-Catalyzed Cross-Coupling Reaction of Highly Functionalized Organobismuthines 1a–t and 21a–h with Arylhalides (28), Heteroarylhalides (29), 2-Halopyridines (2), 2-Halopyrimidines (4), 2-Halopyrazines (6), and 3-Halopyridazines (8)

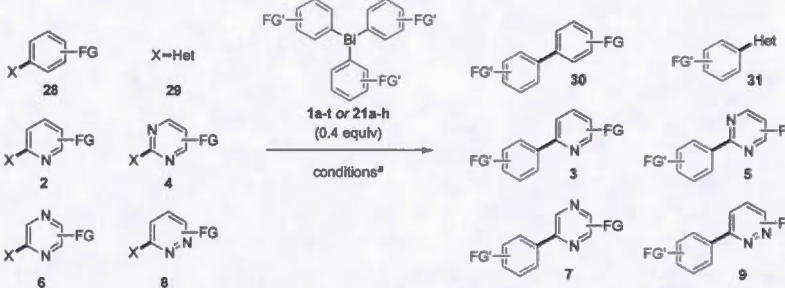
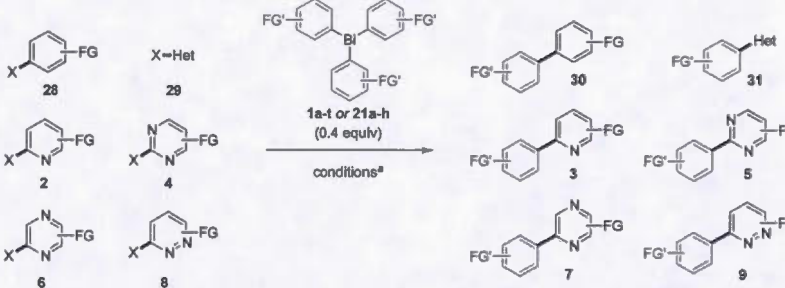
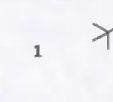
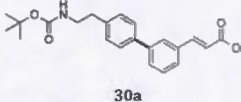

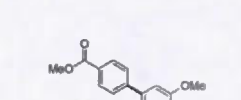

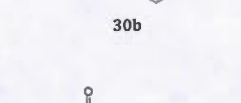

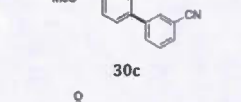
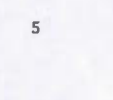
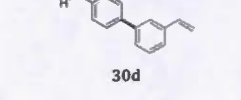

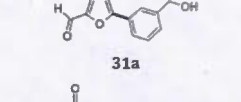

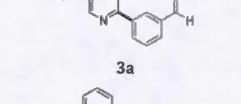

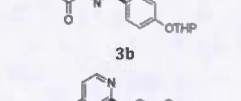
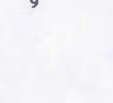
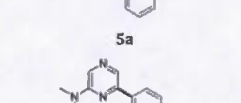
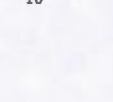
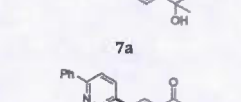
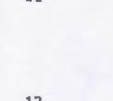
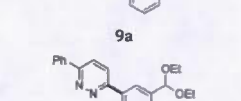

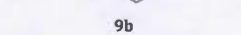
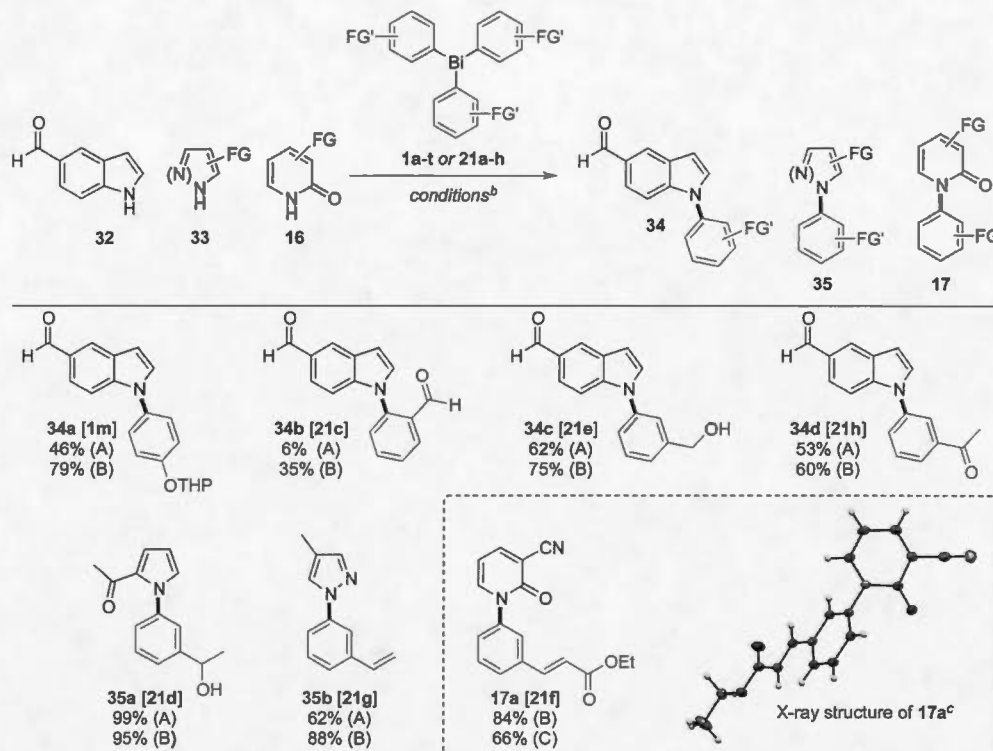
				
				
Entry	Ar(Het)-X	Ar <sub>3</sub> Bi	Product	Yield (%) <sup>b</sup>
1	 28a	21f	 30a	89 (A) 10 (B) 80 (C)
2	 28b	1l	 30b	91 (A) 95 (B) 98 (C)
3	 28c	1l	 30b	54 (A) 78 (B) 66 (C)
4	 28b	1s	 30c	84 (A) 87 (B) 41 (C)
5	 28d	21g	 30d	58 (A) 46 (B) 76 (C)
6	 29a	21d	 31a	82 (A) 65 (B) 29 (C)
7	 2a	21b	 3a	36 (A) 64 (B) 33 (C)
8	 2b	1m	 3b	0 (A) 80 (B) 31 (C)
9	 4a	21g	 5a	20 (A) 12 (B) 13 (C)
10	 6a	21a	 7a	41 (A) 12 (B) 10 (C)
11	 8a	21h	 9a	25 (A) 23 (B) 12 (C)
12	 8a	1o	 9b	0 (A) 45 (B) 0 (C)



Table 2. continued

<sup>a</sup>Method A: Ar<sub>3</sub>Bi (0.4 equiv), Pd(PPh<sub>3</sub>)<sub>4</sub> (5 mol %), LiCl (2.0 equiv), K<sub>3</sub>PO<sub>4</sub> (2.0 equiv), DMF, 80 °C, 6 h; Method B: Ar<sub>3</sub>Bi (0.4 equiv), Pd(PPh<sub>3</sub>)<sub>4</sub> (5 mol %), LiCl (2.0 equiv), Rb<sub>2</sub>CO<sub>3</sub> (2.0 equiv), DMF, 80 °C, 6 h; Method C: Ar<sub>3</sub>Bi (0.4 equiv), Pd(PPh<sub>3</sub>)<sub>4</sub> (5 mol %), CuI (0.4 equiv), Cs<sub>2</sub>CO<sub>3</sub> (2.0 equiv), DMF, 80 °C, 6 h. <sup>b</sup>Isolated yield of pure product.

Scheme 6. *N*-Arylation of Indole 32, Pyrrole and Pyrazole 33, and Pyridone 16 Using Highly Functionalized Organobismuthines 1a–t and 21a–h<sup>a</sup>



<sup>a</sup>The numbers in brackets indicate the organobismuthine used for the synthesis of each compound. <sup>b</sup>Conditions: Method A: Ar<sub>3</sub>Bi (1.0 equiv), Cu(OAc)<sub>2</sub> (0.1 equiv), pyridine (1.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, O<sub>2</sub>, 50 °C, o.n.; Method B: Ar<sub>3</sub>Bi (1.0 equiv), Cu(OAc)<sub>2</sub> (1.0 equiv), pyridine (3.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, O<sub>2</sub>, 50 °C, o.n.; Method C: Ar<sub>3</sub>Bi (1.0 equiv), Cu(OAc)<sub>2</sub> (1.0 equiv), pyridine (3.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, air, 50 °C, o.n. <sup>c</sup>ORTEP diagram at 50% ellipsoid probability of compound 17a.

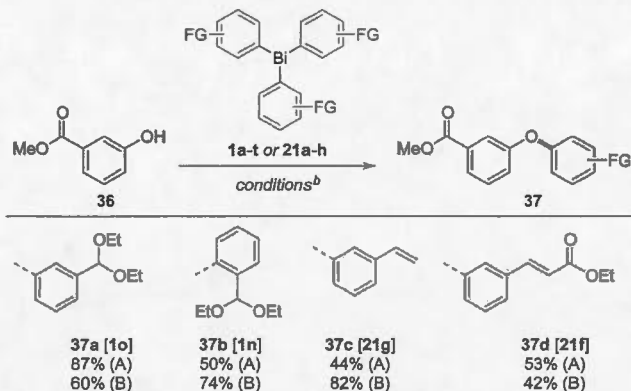
work in our copper-catalyzed *N*-arylation reaction. The arylations were performed at 50 °C under oxygen using 1.0 equiv of triarylbiarylborane, 1.0 equiv of pyridine, and 0.1 equiv of copper acetate (Scheme 6, Method A). Every substrate was also arylated under more “forcing” conditions, that is, using stoichiometric amounts of copper acetate and 3.0 equiv of pyridine (Scheme 6, Method B). Finally, pyridones were arylated using similar conditions, except that the reactions were also performed under air (Scheme 6, Method C). As illustrated in Scheme 6, indole 32, pyrrole and pyrazole 33, and pyridone 16 were *N*-arylated in up to 99% yield using these conditions, demonstrating that aryl groups possessing THP-protected phenols (34a), aldehydes (34b), benzylic alcohols (34c), methyl ketones (34d), secondary alcohols (35a), vinyl groups (35b), and  $\alpha,\beta$ -unsaturated esters (17a) can be efficiently transferred using our methods. It should be emphasized that groups that are susceptible to elimination (secondary alcohol in 35a), oxidation (benzylic and secondary alcohols in 34c and 35a respectively), enolization (methyl ketone in 34d), or arylation (alcohols in 34c and 35a) are also tolerated using our protocols. The main limitation of this method, as demonstrated previously,<sup>11a</sup> resides in the transfer of an *ortho*-substituted aryl group, as illustrated by compound 34b.

Mukaiyama reported a copper-free method to *N*-arylate pyridones using pentavalent organobismuth reagents.<sup>32</sup> We demonstrated previously,<sup>12b</sup> based on IR analysis, that the arylation of pyridones using our copper-catalyzed reaction involving trivalent organobismuthines also leads to the *N*-arylated product (as opposed to the *O*-arylated derivative). The analysis of compound 17a by X-ray diffraction further confirms the chemoselectivity of this arylation reaction toward nitrogen.

**e. Copper-Catalyzed *O*-Arylation of Phenols.** A few reports on the *O*-arylation of alcohols and phenols using organobismuth reagents have been disclosed in the literature.<sup>33</sup> However, most of these methods rely on pentavalent organobismuth compounds, which must be prepared from their corresponding trivalent analogues. Therefore, the development of *O*-arylation reactions directly from trivalent organobismuthines is of high interest, as it can reduce the number of steps required to obtain the desired *O*-arylated product. We recently reported an efficient and general method to prepare diarylethers via a copper-catalyzed *O*-arylation of phenols using triarylbiarylboranes.<sup>12b</sup> Using these protocols, we sought to evaluate the applicability of the highly functionalized organobismuthines in the *O*-arylation of methyl 3-hydroxybenzoate 36. Two sets of conditions were tested: in Method A, the

reaction was run at 50 °C for 3 h under air using 1.0 equiv of triarylbi-muthine, triethylamine as the base, and a stoichiometric amount of copper acetate; in Method B, the catalyst loading was lowered to 0.3 equiv and the reaction was performed under oxygen for 16 h. As shown in Scheme 7, these

**Scheme 7. O-Arylation of Methyl 3-Hydroxybenzoate 36 Using Highly Functionalized Organobismuthines 1a–t and 21a–h**



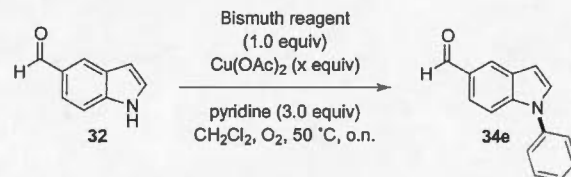
<sup>a</sup>The numbers in brackets indicate the organobismuthine used for the synthesis of each compound. <sup>b</sup>Conditions: Method A: Ar<sub>3</sub>Bi (1.0 equiv), Cu(OAc)<sub>2</sub> (1.0 equiv), Et<sub>3</sub>N (3.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 50 °C, air, 3 h; Method B: Ar<sub>3</sub>Bi (1.0 equiv), Cu(OAc)<sub>2</sub> (0.3 equiv), Et<sub>3</sub>N (3.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, 50 °C, O<sub>2</sub>, 16 h.

methods allow the transfer of aryl groups bearing a variety of groups such as acetals (37a,b), vinyl groups (37c), and  $\alpha,\beta$ -unsaturated esters (37d). Contrary to the *N*-arylation of indoles, pyrroles, and pyrazoles, the *O*-arylation of phenols even shows high tolerance for substitution in the *ortho* position, as shown by compound 37b.

**f. Mechanistic Investigations on the Copper-Catalyzed Arylation of Indoles and Phenols Using Trivalent and Pentavalent Organobismuth Reagents.** Pentavalent organobismuth reagents have been shown to be potent arylating species in copper-catalyzed<sup>8c,d,f,h,i</sup> and even metal-free reactions.<sup>8a</sup> Barton also proposed that, in copper-catalyzed arylation reactions using triphenylbismuth, triphenylbismuth diacetate is formed *in situ* and acts as the effective arylating agent. Therefore, to compare the arylating power of trivalent and pentavalent organobismuth species, we performed the arylation on indole 32 using triphenylbismuth (Ph<sub>3</sub>Bi, 1a) and triphenylbismuth diacetate (Ph<sub>3</sub>Bi(OAc)<sub>2</sub>, 38) in the presence and absence of copper acetate (Table 3). As expected, in the presence of copper acetate, triphenylbismuth was found to be an excellent arylating agent, providing the *N*-arylindole 34e in 97% yield (entry 1). By contrast, in the presence of copper acetate, triphenylbismuth diacetate proved to be a much less efficient arylating reagent, affording 34e in only 20% yield (entry 3). Both species were mostly inactive in the absence of copper acetate, showing that copper is essential for the *N*-arylation reaction (entries 2 and 4).

A comparative reactivity study between triphenylbismuth and triphenylbismuth diacetate was also performed on methyl 3-hydroxybenzoate 36 (Table 4). Contrary to the arylation of indoles, in the presence of copper acetate, triphenylbismuth diacetate was found to be more reactive than triphenylbismuth in the arylation of phenol 36 (entry 3 vs 1). However, similarly

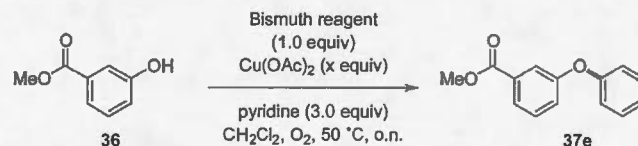
**Table 3. Comparison of the Reactivity of Triphenylbismuth 1a and Triphenylbismuth Diacetate 38 in the *N*-Arylation Reaction of Indole 32**



Entry	Bismuth reagent	Cu(OAc) <sub>2</sub> (x equiv)	Yield (%) <sup>a</sup>
1	Ph <sub>3</sub> Bi (1a)	1.0	97
2	Ph <sub>3</sub> Bi (1a)	0	2
3	Ph <sub>3</sub> Bi(OAc) <sub>2</sub> (38)	1.0	20
4	Ph <sub>3</sub> Bi(OAc) <sub>2</sub> (38)	0	1

<sup>a</sup>Yields are for isolated pure compound.

**Table 4. Comparison of the Reactivity of Triphenylbismuth 1a and Triphenylbismuth Diacetate 38 in the *O*-Arylation Reaction of Phenol 36**



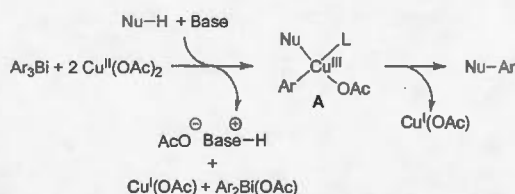
Entry	Bismuth reagent	Cu(OAc) <sub>2</sub> (x equiv)	Yield (%) <sup>a</sup>
1	Ph <sub>3</sub> Bi (1a)	1.0	70
2	Ph <sub>3</sub> Bi (1a)	0	0
3	Ph <sub>3</sub> Bi(OAc) <sub>2</sub> (38)	1.0	92
4	Ph <sub>3</sub> Bi(OAc) <sub>2</sub> (38)	0	4

<sup>a</sup>Yields are for isolated pure compound.

to the arylation of indole 32, in the absence of the catalyst, both reagents were unable to arylate phenol 36, showing once again that copper is essential for the *O*-arylation reaction (entries 2 and 4). The results from Table 3 and Table 4 suggest that copper acetate is necessary for both arylation reactions and that trivalent and pentavalent organobismuth species can both act as arylating agents, albeit with different efficiencies depending on the substrate.

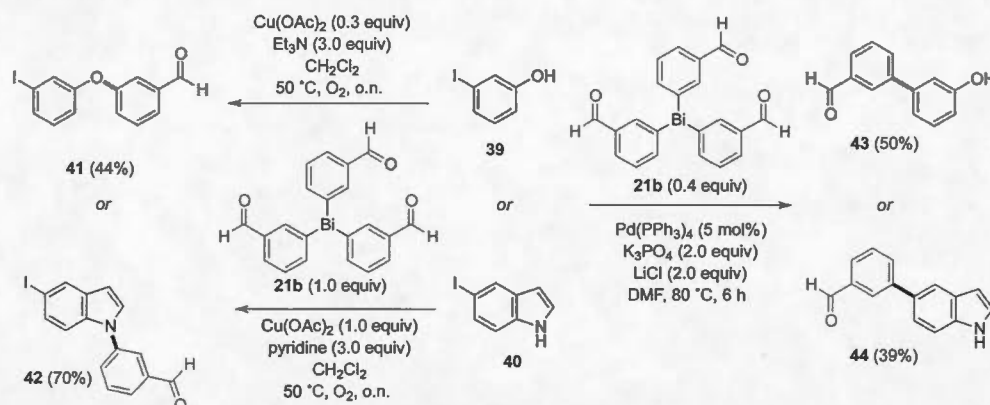
The mechanism that we propose for the copper-catalyzed *N*- and *O*-arylation of indoles, pyrroles, pyrazoles, pyridones, and phenols (summarized by Nu–H) using triarylbi-muthines involves the formation of an organocopper(III) species A where the deprotonated nucleophile, the aryl group, and one acetate are simultaneously ligated to the metal (Scheme 8).<sup>34</sup> This species would be formed by the reaction of triarylbi-muthine Ar<sub>3</sub>Bi and copper acetate, concomitantly with the

**Scheme 8. Proposed Mechanism for the Copper-Catalyzed *N*- and *O*-Arylation of Indoles, Pyrroles, Pyrazoles, Pyridones, and Phenols Using Triarylbi-muthines (Nu–H = Indole, Pyrrole, Pyrazole, Pyridone, or Phenol)**





Scheme 9. Orthogonal C-, N-Arylation and O-Arylation of 3-Iodophenol 39 and 5-Iodoindole 40 Using Tris(3-formylphenyl)bismuthine 21b



deprotonation of the nucleophile Nu–H by the base (either triethylamine or pyridine), leading to copper(III) intermediate A, copper(I) acetate, and diarylbismuth acetate ( $\text{Ar}_2\text{Bi}(\text{OAc})$ ). Reductive elimination from species A would then provide the arylated nucleophile Nu–Ar. This mechanism is similar to that proposed by Barton and Finet in the copper-catalyzed arylation of amines and indoles using triphenylbismuth diacetate.<sup>8g</sup> Moreover, intermediate A was also proposed by Evans in the arylation of phenols using arylboronic acids,<sup>35</sup> in the Ullmann–Goldberg copper-catalyzed N-arylation of amines using aryl halides<sup>36</sup> and in detailed mechanistic studies published by Stahl on the Chan–Evans–Lam reaction.<sup>37</sup>

**g. Orthogonal C-, N-, and O-Arylation Reactions on Iodo Phenols and Indoles.** To further test the scope of our copper- and palladium-catalyzed methods, we explored the orthogonal transfer of aryl groups on the halogenated phenol 39 and indole 40 (Scheme 9). Using copper acetate and organobismuthine 21b, the arylation could be directed chemoselectively to the oxygen of 39 or the nitrogen of 40, providing 41 and 42 in 44% and 70% yield, respectively. These products would be difficult to obtain using copper(I) or palladium(0) catalysis since reaction at the aryl–I bond would compete with the desired O- or N-arylation process.<sup>38</sup> In addition, the palladium-catalyzed cross-coupling reaction between 3-iodophenol 39 and 5-iodoindole 40 with triaryl bismuthine 21b delivered 43 and 44, respectively, in moderate yields. These cross-coupling reactions represent a substantial challenge since competitive dehalogenation can occur due to the presence of the phenol and N–H indole functions.

## CONCLUSION

In summary, we demonstrated that highly functionalized triaryl bismuthines can be prepared by triple functional group manipulation using acidic, nucleophilic, and reducing conditions or involving organometallic reagents or ylides. Using this approach, triaryl bismuthines bearing primary, secondary, and tertiary alcohols, aldehydes,  $\alpha,\beta$ -unsaturated esters, vinyl groups, and methyl ketones were prepared. These triaryl bismuthines represent a class of highly functionalized and versatile organometallic reagents that are in high demand. We then developed improved protocols for the palladium-catalyzed cross-coupling reaction between these highly functionalized reagents and aryl or heteroaryl halides. These modified procedures involve lithium chloride or copper iodide as an

additive and potassium phosphate or rubidium carbonate as a base and require shorter reaction times and lower temperatures. Using these protocols, highly functionalized C-arylated compounds were obtained in good to excellent yields. The highly functionalized organobismuthines prepared using the functional group manipulation approach were then engaged in a copper-catalyzed N-arylation of indoles, pyrroles, pyrazoles, and pyridones and O-arylation of phenols to afford highly functionalized arylated products. We then demonstrated that our palladium- and copper-catalyzed processes can be successfully used orthogonally on halogenated indoles and phenols. Preliminary mechanistic studies demonstrate that copper acetate is essential for the reaction to proceed and that the order of reactivity between the trivalent and pentavalent species depends on the nature of the substrate. Our palladium- and copper-catalyzed arylation reactions are simple to operate and show high functional group tolerance. The C-, N-, and O-arylation procedures developed in this work constitute an efficient and general portfolio of methods for the preparation of medicinally relevant scaffolds.

## EXPERIMENTAL SECTION

**General Information.** Unless otherwise indicated, all reactions were run under argon in nonflame-dried glassware. For reactions performed under oxygen, 99.6% extra dry oxygen was used. Unless otherwise stated, commercial reagents were used without further purification. Grignard reagents were prepared by conventional methods using metallic magnesium or via Knochel's procedure.<sup>14</sup> Anhydrous bismuth chloride >98% and triphenylbismuth diacetate 98% were purchased from Aldrich. Triaryl bismuthines 1b, 1d, 1g, 1l, 1m, 1q, 1r, 1s, and 1t were prepared according to a procedure that we previously reported.<sup>10b</sup> Triaryl bismuthines 1c, 1o, and 21b were prepared according to methods previously reported by us.<sup>12b</sup> Triaryl bismuthines 1e, 1f, 1h, 1i, 1j, 1k, 1p, 21a, 21d, and 21f were prepared according to a procedure that we previously reported.<sup>11a</sup> Anhydrous solvents were obtained using an encapsulated solvent purification system and were further dried over 4 Å molecular sieves. The evolution of reactions was monitored by analytical thin-layer chromatography using silica gel 60 F254 precoated plates. Flash chromatography was performed employing 230–400 mesh silica using the indicated solvent system according to standard techniques. For compounds 1n, 1o, 1p, 9b, 37a, and 37b, the silica gel was washed with 3 volumes of 0.5%  $\text{Et}_3\text{N}$ /hexanes prior to performing the flash chromatography to avoid hydrolysis of the acetal. Melting points are uncorrected. Nuclear magnetic resonance spectra ( $^1\text{H}$ ,  $^{13}\text{C}$ ) were recorded on a 300 or 600 MHz spectrometer. Chemical shifts for  $^1\text{H}$  NMR spectra are recorded in parts per million from tetramethylsilane with the solvent resonance as the internal standard (chloroform,  $\delta$  7.26



ppm; methanol,  $\delta$  3.31 ppm). Data are reported as follows: chemical shift, multiplicity (*s* = singlet, *d* = doublet, *t* = triplet, *q* = quartet, *qt* = quintuplet, *dd* = doublet of doublet, *dt* = doublet of triplet, *m* = multiplet), coupling constant *J* in Hz, and integration. Chemical shifts for  $^{13}\text{C}$  spectra are recorded in parts per million from tetramethylsilane using the central peak of deuteriochloroform ( $\delta$  77.16 ppm) or the central peak of tetradeuteromethanol ( $\delta$  49.00 ppm) as the internal standard. IR spectra were recorded on an FT-IR from thin films and are reported in reciprocal centimeters ( $\text{cm}^{-1}$ ). HRMS was performed on a TOF LCMS analyzer using the electrospray (ESI) mode.

#### General Procedure for the Synthesis of Triarylbiomuthines.

In a flask equipped with a magnetic stir bar and a condenser, bismuth chloride (500 mg, 1.6 mmol) was dissolved in anhydrous THF (23 mL) under argon and was cooled to  $-10^\circ\text{C}$  (ice/acetone bath). The organomagnesium reagent (5.23 mmol) was slowly added dropwise under argon. The reaction mixture was stirred at room temperature for 1 h and heated at  $65^\circ\text{C}$  for 30 min. After cooling to room temperature, the solution was diluted with sat. aq.  $\text{NaHCO}_3$  (100 mL) and extracted with EtOAc ( $2 \times 100$  mL). The combined organic phases were washed with sat. aq.  $\text{NaHCO}_3$  ( $2 \times 100$  mL), sat. aq.  $\text{NaCl}$  ( $2 \times 100$  mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated under reduced pressure. The crude product was purified by column chromatography using the indicated solvent system to afford the desired triarylbiomuthine.

**Triphenylbismuth (1a).** The general procedure was followed on a 2.27 mmol scale starting from bismuth chloride and phenylmagnesium bromide. The crude product was purified on silica gel (2% EtOAc/hexanes) to afford triphenylbismuth 1a as a white solid (831 mg, 83%): mp  $78-79^\circ\text{C}$ ;  $R_f$  0.33 (2% EtOAc/hexanes). Spectral data were identical to those of the literature compound.<sup>39</sup>  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.75 (d, *J* = 7.5 Hz, 6H), 7.42–7.30 (m, 9H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  155.3, 137.7, 130.6, 127.9; IR (neat) 3133, 3053, 3011, 2582, 1943, 1871, 1810, 1566, 1471, 1422, 1053, 992, 718, 692. Anal. Calcd for  $\text{C}_{18}\text{H}_{15}\text{Bi}$ : C, 49.10; H, 3.43. Found: C, 49.28; H, 3.39.

**Tris(2-(diethoxymethyl)phenyl)bismuthine (1n).** The general procedure was followed on a 5.33 mmol scale starting from bismuth chloride and 2-(benzaldehydediethylacetal)magnesium bromide. The crude product was purified on silica gel (5% EtOAc/hexanes) to afford tris(2-(diethoxymethyl)phenyl)bismuthine 1n as a white solid (3.3 g, 82%): mp  $78-79^\circ\text{C}$ ;  $R_f$  0.81 (20% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.70 (dd, *J* = 6.5, 1.4 Hz, 3H), 7.60 (dd, *J* = 7.3, 1.4 Hz, 3H), 7.30 (dt, *J* = 7.2, 1.4 Hz, 3H), 7.13 (dt, *J* = 7.3, 1.5 Hz, 3H), 5.53 (s, 3H), 3.55–3.34 (m, 12H), 1.01 (t, *J* = 7.0 Hz, 18H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  162.6, 144.5, 140.6, 131.0, 127.5, 126.8, 104.4, 61.4, 15.1; IR (neat) 3049, 2973, 2928, 2871, 1438, 1336, 1204, 1108, 1090, 1050, 757; HRMS (ESI) calcd for  $[\text{C}_{33}\text{H}_{45}\text{BiO}_6 + \text{Na}]^+$ : 769.2912, found 769.2868.

**Tris(2-formylphenyl)bismuthine (21c).**  $\text{H}_2\text{O}$  (10 mL) and HCl 12N (0.90 mL) were added at room temperature to a stirred solution of 1n (1.0 g, 1.3 mmol) in THF (30 mL). The reaction mixture was stirred overnight and then diluted with EtOAc (20 mL). The organic layer was washed with sat. aq.  $\text{NaHCO}_3$  (100 mL), sat. aq.  $\text{NaCl}$  ( $3 \times 20$  mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated under reduced pressure. The crude material was purified on silica gel (5% EtOAc/hexanes) to afford tris(2-formylphenyl)bismuthine 21c as a yellow solid (213 mg, 31%): mp  $179-180^\circ\text{C}$ ;  $R_f$  0.46 (20% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  10.23 (s, 3H), 8.02 (dd, *J* = 7.5, 1.4 Hz, 3H), 7.59–7.54 (m, 6H), 7.34 (td, *J* = 7.5, 1.5 Hz, 3H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  196.0, 171.2, 142.3, 140.9, 136.9, 136.4, 127.7; IR (neat) 3042, 2977, 2806, 2724, 1690, 1671, 1571, 1556, 1197, 835, 759; HRMS (ESI) calcd for  $[\text{C}_{21}\text{H}_{13}\text{BiO}_3 + \text{H}]^+$ : 525.0898, found 525.0896.

**Tris(3-(hydroxymethyl)phenyl)bismuthine (21e).** A solution of tris(3-formylphenyl)bismuthine 21b (400 mg, 0.76 mmol) in MeOH (10 mL) was cooled to  $0^\circ\text{C}$  (acetone/ice bath), and  $\text{NaBH}_4$  (90 mg, 2.4 mmol) was added. After 30 min, the reaction mixture was diluted with sat. aq.  $\text{NaHCO}_3$  (15 mL) and extracted with EtOAc (15 mL). The organic layer was washed with sat. aq.  $\text{NaHCO}_3$  (15 mL) and sat. aq.  $\text{NaCl}$  ( $3 \times 15$  mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated under reduced pressure. The crude material was purified on silica gel

(40% EtOAc/hexanes) to afford tris(3-(hydroxymethyl)phenyl)bismuthine 21e as a white solid (392 mg, 97%): mp  $107-108^\circ\text{C}$ ;  $R_f$  0.39 (80% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz, MeOD)  $\delta$  7.77 (s, 3H), 7.63 (d, *J* = 6.9 Hz, 3H), 7.37–7.28 (m, 6H), 4.53 (s, 6H);  $^{13}\text{C}$  NMR (75 MHz, MeOD)  $\delta$  156.7, 144.3, 137.6, 137.2, 131.4, 127.6, 65.3; IR (neat) 3314 (br), 3038, 2923, 2869, 1563, 1412, 1203, 1012, 776; HRMS (ESI) calcd for  $[\text{C}_{21}\text{H}_{21}\text{BiO}_3 + \text{Na}]^+$ : 553.1187, found 553.1177.

**Tris(3-vinylphenyl)bismuthine (21g).** To a solution of  $\text{Ph}_3\text{PCH}_2\text{I}$  (1.86 g, 4.58 mmol) in THF (8 mL) at  $-10^\circ\text{C}$ , *t*-BuOK was added (606 mg, 5.4 mmol), and the solution was stirred for 30 min. To the yellow solution, tris(3-formylphenyl)bismuthine 21b (800 mg, 1.53 mmol) was added, and the reaction was warmed up to room temperature and stirred for 2 h. The reaction mixture was diluted with sat. aq.  $\text{NaHCO}_3$  (50 mL) and extracted with EtOAc ( $2 \times 50$  mL). The combined organic layers were washed with sat. aq.  $\text{NaHCO}_3$  (50 mL), sat. aq.  $\text{NaCl}$  ( $3 \times 50$  mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated under reduced pressure. The crude material was purified on silica gel (5% EtOAc/hexanes) to afford tris(3-vinylphenyl)bismuthine 21g as a yellow oil (650 mg, 82%):  $R_f$  0.70 (20% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ ) 7.82 (s, 3H), 7.65–7.62 (m, 3H), 7.40–7.33 (m, 6H), 6.67 (dd, *J* = 17.6, 11.0 Hz, 3H), 5.66 (d, *J* = 17.6 Hz, 3H), 5.20 (d, *J* = 11.0 Hz, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  155.4, 139.4, 137.2, 137.0, 135.5, 130.7, 125.7, 114.1; IR (neat) 3156, 3084, 3063, 3040, 3006, 2984, 2927, 1939, 1821, 1629, 1580, 1554, 1467, 1401, 1380, 991, 906, 791, 710. Anal. Calcd for  $\text{C}_{24}\text{H}_{21}\text{Bi}$ : C, 55.60; H, 4.08. Found: C, 55.87; H, 4.01.

**Tris(3-acetylphenyl)bismuthine (21h).** *Preparation via Dess-Martin oxidation:* A solution of tris(3-(1-hydroxyethyl)phenyl)bismuthine 21d (50 mg, 0.09 mmol) in  $\text{CH}_2\text{Cl}_2$  (3 mL) was cooled to  $-10^\circ\text{C}$  (acetone/ice bath), and pyridine (46  $\mu\text{L}$ , 0.59 mmol) was added. After 5 min, Dess-Martin Periodinane (123 mg, 0.28 mmol) was added. After 1 h, the reaction mixture was diluted with EtOAc (15 mL). The organic layer was washed with sat. aq.  $\text{NaHCO}_3$  (15 mL), sat. aq.  $\text{NaCl}$  ( $3 \times 15$  mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated under reduced pressure. The crude material was purified on silica gel (20% EtOAc/hexanes) to afford tris(3-acetylphenyl)bismuthine 21h as a white solid (30 mg, 59%). *Preparation via Swern oxidation:* A solution of oxalyl chloride (67  $\mu\text{L}$ , 0.79 mmol) in anhydrous  $\text{CH}_2\text{Cl}_2$  (2.0 mL) was put under argon and cooled to  $-78^\circ\text{C}$  (acetone/ice bath). Anhydrous DMSO (112  $\mu\text{L}$ , 1.57 mmol) was added dropwise to the reaction mixture and stirred for 30 min at  $-78^\circ\text{C}$ . Tris(3-(1-hydroxyethyl)phenyl)bismuthine 21d (100 mg, 0.17 mmol) was dissolved in anhydrous  $\text{CH}_2\text{Cl}_2$  (10 mL) and added dropwise to the reaction mixture and stirred for 1 h at  $-78^\circ\text{C}$ .  $\text{Et}_3\text{N}$  (219  $\mu\text{L}$ , 1.57 mmol) was added, and the reaction mixture was warmed to room temperature and stirred for 30 min. The reaction mixture was diluted with sat. aq.  $\text{NaHCO}_3$  (50 mL) and extracted with  $\text{CH}_2\text{Cl}_2$  ( $2 \times 50$  mL). The combined organic phases were washed with sat. aq.  $\text{NaHCO}_3$  ( $2 \times 50$  mL), sat. aq.  $\text{NaCl}$  ( $2 \times 50$  mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated under reduced pressure. The crude material was purified on silica gel (20% EtOAc/hexanes) to afford tris(3-acetylphenyl)bismuthine 21h as a white solid (61 mg, 63%): mp  $85-86^\circ\text{C}$ ;  $R_f$  0.17 (20% EtOAc/hexanes). Spectral data were identical to those of the literature compound.<sup>40</sup>  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.36–8.35 (m, 3H), 7.90–7.88 (m, 6H), 7.49 (t, *J* = 7.5 Hz, 3H), 2.49 (s, 9H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  198.4, 156.0, 142.2, 138.8, 137.1, 131.1, 128.3, 26.8; IR (neat) 3344, 3051, 3008, 2923, 1678, 1578, 1559, 1404, 1356, 1255; HRMS (ESI) calcd for  $[\text{C}_{24}\text{H}_{21}\text{BiO}_3 + \text{H}]^+$ : 567.1367, found 567.1400.

**General Procedure for the Crystallization of Triarylbiomuthines 1f, 1i, 1k, 1n, 1s, 1t, 21b, 21c, 21e and Pyridone 17a.** Compounds 1f, 1i, 1k, 1n, 1s, 1t, 21b, 21c, 21e, and 17a were crystallized by the solvent diffusion technique according to the following procedure: 10 mg of the corresponding compound was dissolved in a minimal amount of dichloromethane in an open vial. The vial was then placed in a bigger vial filled with hexanes with a loosely tightened cap. The vials were kept at room temperature until crystals were obtained.



**General Procedures for the Palladium-Catalyzed Cross-Coupling Reactions.** Compounds 3a,b, 5a, 7a, 9a,b, 27, 30a–d, 31a, 43, and 44 were prepared according to the following procedures: **Method A** (Table 1, entry 13): In a sealed tube, the aryl or heteroaryl chloride, bromide, or iodide (1.0 equiv) was dissolved in *N,N*-dimethylformamide (4.0 mL). Potassium phosphate (2.0 equiv) was added, followed by tetrakis(triphenylphosphine)palladium (5 mol %), lithium chloride (2.0 equiv), and triarylbi-muth reagent (0.4 equiv). Argon was bubbled in the reaction mixture for 1 min. The tube was sealed and heated at 80 °C for 6 h. The reaction mixture was cooled to room temperature, diluted with sat. aq. NaHCO<sub>3</sub> (20 mL), and extracted with EtOAc (2 × 20 mL). The combined organic phases were washed with sat. aq. NaHCO<sub>3</sub> (2 × 20 mL), sat. aq. NaCl (2 × 20 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated under reduced pressure. The crude product was purified by flash column chromatography on silica gel using EtOAc/hexanes as the eluent to afford the corresponding product. **Method B** (Table 1, entry 14): Same as conditions A except that rubidium carbonate was used instead of potassium phosphate. **Method C** (Table 1, entry 18): Same as conditions A except that cesium carbonate was used instead of potassium phosphate and 0.4 equiv of cuprous iodide was used instead of lithium chloride.

**6-(3-Formylphenyl)nicotinaldehyde (3a).** Method B was followed on a 0.095 mmol scale starting from 6-chloronicotinaldehyde 2a and organobismuthine 21b. The crude material was purified on silica gel (15% EtOAc/hexanes) to afford 3a as a white solid (13 mg, 64%): mp 117–118 °C; *R*<sub>f</sub> 0.29 (20% EtOAc/hexanes). Spectral data were identical to those of the literature compound.<sup>41</sup> <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 10.15 (s, 1H), 10.12 (s, 1H), 9.15 (d, *J* = 1.4 Hz, 1H), 8.58 (s, 1H), 8.37 (dt, *J* = 7.7, 1.2 Hz, 1H), 8.26 (dd, *J* = 8.2, 2.1 Hz, 1H), 8.01–7.96 (m, 2H), 7.68 (t, *J* = 7.7 Hz, 1H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 192.0, 190.4, 160.6, 152.4, 139.0, 137.2, 137.1, 133.3, 131.3, 130.5, 129.9, 128.9, 120.8; IR (neat) 3053, 2852, 2745, 1689, 1587, 1359, 1212, 1183, 1166, 835, 795, 740; HRMS (ESI) calcd for [C<sub>13</sub>H<sub>9</sub>NO<sub>2</sub> + H]<sup>+</sup>: 212.0706, found 212.0708.

**1-(6-(4-(Tetrahydro-2H-pyran-2-yl)oxyphenyl)pyridine-2-yl)ethanone (3b).** Method B was followed on a 0.21 mmol scale starting from 2-acetyl-6-bromopyridine 2b and organobismuthine 1m. The crude material was purified on silica gel (10% EtOAc/hexanes) to afford 3b as a light pink solid (50 mg, 80%): mp 90–91 °C; *R*<sub>f</sub> 0.47 (20% EtOAc/hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.07–8.02 (m, 2H), 7.93–7.90 (m, 1H), 7.89–7.84 (m, 2H), 7.21–7.16 (m, 2H), 5.52 (t, *J* = 3.3 Hz, 1H), 3.97–3.89 (m, 1H), 3.67–3.60 (m, 1H), 2.82 (s, 3H), 2.10–2.00 (m, 1H), 1.93–1.89 (m, 2H), 1.88–1.60 (m, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 200.9, 158.5, 156.4, 153.4, 137.6, 132.0, 128.3, 122.9, 119.3, 116.8, 96.4, 62.2, 30.4, 25.9, 25.3, 18.8; IR (neat) 3060, 2943, 2872, 2840, 1696, 1606, 1513, 1449, 1355, 1239, 1177, 1036, 960, 919, 845, 806, 739, 613, 593; HRMS (ESI) calcd for [C<sub>18</sub>H<sub>19</sub>NO<sub>3</sub> + H]<sup>+</sup>: 298.1438, found 298.1426.

**4-Methyl-2-(3-vinylphenyl)pyrimidine (5a).** Method A was followed on a 0.40 mmol scale starting from 2-chloro-4-methylpyrimidine 4a and organobismuthine 21g. The crude material was purified on silica gel (10% EtOAc/hexanes) to afford 5a as a yellow oil (16 mg, 20%); *R*<sub>f</sub> 0.60 (20% EtOAc/hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.65 (d, *J* = 5.1 Hz, 1H), 8.48 (t, *J* = 1.9 Hz, 1H), 8.34 (dt, *J* = 7.7, 1.5 Hz, 1H), 7.54 (dt, *J* = 7.7, 1.6 Hz, 1H), 7.45 (t, *J* = 7.7 Hz, 1H), 7.05 (d, *J* = 5.0 Hz, 1H), 6.82 (dd, *J* = 17.6, 10.9 Hz, 1H), 5.87 (dd, *J* = 17.4, 0.9 Hz, 1H), 5.30 (dd, *J* = 10.8, 0.9 Hz, 1H), 2.60 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 167.5, 164.3, 156.9, 138.2, 138.0, 136.8, 128.9, 128.3, 127.8, 126.3, 118.8, 114.4, 24.6; IR (neat) 3061, 3038, 2954, 2928, 2867, 1692, 1572, 1555, 1431, 1384, 1364, 912, 789; HRMS (ESI) calcd for [C<sub>13</sub>H<sub>12</sub>N<sub>2</sub> + H]<sup>+</sup>: 197.1073, found 197.1080.

**2-(4-(6-(Dimethylamino)pyrazin-2-yl)phenyl)propan-2-ol (7a).** Method A was followed on a 0.32 mmol scale starting from 6-chloro-*N,N*-dimethylpyrazin-2-amine 6a and organobismuthine 21a. The crude material was purified on silica gel (40% EtOAc/hexanes) to afford 7a as a yellow solid (34 mg, 41%): mp 158–159 °C; *R*<sub>f</sub> 0.20 (40% EtOAc/hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.21 (s, 1H), 7.99–7.94 (m, 2H), 7.91 (s, 1H), 7.60–7.56 (m, 2H), 3.17 (s, 6H), 2.37 (s(br), 1H), 1.61 (s, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 154.4,

150.5, 149.2, 136.0, 128.4, 127.9, 126.8, 125.0, 72.6, 37.6, 31.9; IR (neat) 3371 (br), 3059, 2971, 2923, 2869, 1583, 1566, 1527, 1425, 1402, 1372, 1186, 1151, 996, 831; HRMS (ESI) calcd for [C<sub>15</sub>H<sub>19</sub>N<sub>3</sub>O + H]<sup>+</sup>: 258.1601, found 258.1603.

**1-(3-(6-Phenylpyridazin-3-yl)phenyl)ethanone (9a).** Method A was followed on a 0.26 mmol scale starting from 3-chloro-6-phenylpyridazine 8a and organobismuthine 21h. The crude material was purified on silica gel (20% EtOAc/hexanes) to afford 9a as a yellow solid (18 mg, 25%): mp 130–131 °C; *R*<sub>f</sub> 0.25 (20% EtOAc/hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.77 (t, *J* = 1.9 Hz, 1H), 8.41 (dt, *J* = 7.9, 1.4 Hz, 1H), 8.18 (dd, *J* = 8.1, 1.9 Hz, 2H), 8.11 (dt, *J* = 6.5, 1.5 Hz, 1H), 8.05–7.97 (m, 2H), 7.67 (t, *J* = 7.8 Hz, 1H), 7.60–7.52 (m, 3H), 2.72 (s, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 198.0, 158.1, 156.8, 137.9, 136.7, 135.9, 131.4, 130.4, 129.8, 129.5, 129.2, 127.1, 126.9, 124.5, 124.4, 27.0; IR (neat) 3061, 3000, 2932, 1683, 1601, 1585, 1432, 1398, 1358, 1236, 689; HRMS (ESI) calcd for [C<sub>18</sub>H<sub>14</sub>N<sub>2</sub>O + H]<sup>+</sup>: 275.1179, found 275.1191.

**3-(3-(Diethoxymethyl)phenyl)-6-phenylpyridazine (9b).** Method B was followed on a 0.10 mmol scale starting from 3-chloro-6-phenylpyridazine 8a and organobismuthine 1o. The crude material was purified on silica gel (20% EtOAc/hexanes) to afford 9b as a beige solid (15 mg, 45%): mp 88–89 °C; *R*<sub>f</sub> 0.36 (20% EtOAc/hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 8.25 (t, *J* = 1.8 Hz, 1H), 8.19–8.14 (m, 3H), 7.95 (dd, *J* = 11.3, 9.0 Hz, 2H), 7.63 (dt, *J* = 7.7, 1.5 Hz, 1H), 7.58–7.51 (m, 4H), 5.61 (s, 1H), 3.73–3.54 (m, 4H), 1.27 (t, *J* = 7.1 Hz, 6H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 157.8, 157.7, 140.3, 136.3, 130.2, 129.2, 129.1, 128.5, 127.1, 125.4, 124.5, 124.3, 101.5, 61.4, 15.4; IR (neat) 3052, 2974, 2917, 2848, 1704, 1450, 1397, 1328, 1095, 1050, 997, 780, 690; HRMS (ESI) calcd for [C<sub>21</sub>H<sub>22</sub>N<sub>2</sub>O<sub>2</sub> + H]<sup>+</sup>: 335.1754, found 335.1767.

**3'-(1-Hydroxyethyl)-[1,1'-biphenyl]-4-carbaldehyde (27).** Method C was followed on a 0.27 mmol scale starting from 4-bromobenzaldehyde 26 and organobismuthine 21d. The crude material was purified on silica gel (25% EtOAc/hexanes) to afford 27 as a yellow oil (51 mg, 83%): *R*<sub>f</sub> 0.22 (20% EtOAc/hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 10.05 (s, 1H), 7.97–7.93 (m, 2H), 7.78–7.74 (m, 2H), 7.67–7.66 (m, 1H), 7.55 (td, *J* = 7.1, 1.9 Hz, 1H), 7.49–7.41 (m, 2H), 4.99 (q, *J* = 6.5 Hz, 1H), 1.82 (s(br), 1H), 1.56 (d, *J* = 6.4 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 192.1, 147.2, 146.8, 139.9, 135.3, 130.3, 129.2, 127.8, 126.5, 125.6, 124.5, 70.3, 25.4; IR (neat) 3422 (br), 3059, 3032, 2971, 2923, 2836, 2731, 1689, 1603, 1567, 1170, 1077, 794, 703; HRMS (ESI) calcd for [C<sub>15</sub>H<sub>14</sub>O<sub>2</sub> + H]<sup>+</sup>: 227.1067, found 227.1068.

**(E)-Ethyl 3-(4'-(2-(tert-Butoxycarbonyl)amino)ethyl)-[1,1'-biphenyl]-3-yl)acrylate (30a).** Method A was followed on a 0.17 mmol scale starting from *tert*-butyl 4-bromophenethylcarbamate 28a and organobismuthine 21f. The crude material was purified on silica gel (15% EtOAc/hexanes) to afford 30a as a yellow solid (60 mg, 89%): mp 111–112 °C; *R*<sub>f</sub> 0.52 (20% EtOAc/hexanes); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ 7.75 (d, *J* = 16.0 Hz, 1H), 7.73 (s, 1H), 7.61–7.59 (m, 1H), 7.55–7.53 (m, 1H), 7.48 (d, *J* = 7.5 Hz, 1H), 7.45–7.40 (m, 2H), 7.06 (d, *J* = 8.2 Hz, 2H), 6.52 (d, *J* = 16.0 Hz, 1H), 4.53 (s(br), 1H), 4.28 (q, *J* = 7.1 Hz, 2H), 3.34 (q, *J* = 6.4 Hz, 2H), 2.75 (t, *J* = 6.9 Hz, 2H), 1.43 (s, 9H), 1.35 (t, *J* = 7.1 Hz, 3H); <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 170.0, 155.9, 144.3, 141.3, 138.1, 135.2, 131.7, 130.6, 129.6, 129.0, 127.3, 126.9, 120.3, 119.0, 79.4, 60.7, 41.7, 35.8, 28.5, 14.4; IR (neat) 3437, 3363, 2977, 2931, 2866, 1703, 1637, 1508, 1488, 1365, 1307, 1267, 1248, 1163, 1036, 1011, 789. Anal. Calcd for C<sub>24</sub>H<sub>29</sub>NO<sub>4</sub>: C, 72.89; H, 7.39; N, 3.54. Found: C, 72.90; H, 7.41; N, 3.52.

**Methyl 3'-Methoxy-[1,1'-biphenyl]-4-carboxylate (30b).** Starting from methyl 4-iodobenzoate 28b: Method C was followed on a 0.19 mmol scale starting from methyl 4-iodobenzoate 28b and organobismuthine 1l. The crude material was purified on silica gel (10% EtOAc/hexanes) to afford 30b as a white solid (45 mg, 98%). Starting from methyl 4-bromobenzoate 28c: Method B was followed on a 0.30 mmol scale starting from methyl 4-bromobenzoate 28c and organobismuthine 1l. The crude material was purified on silica gel (10% EtOAc/hexanes) to afford 30b as a white solid (57 mg, 78%): mp 55–56 °C; *R*<sub>f</sub> 0.50 (10% EtOAc/hexanes). Spectral data were identical to those of the literature compound.<sup>42</sup> <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ



8.10 (d,  $J = 4.2$  Hz, 2H), 7.65 (d,  $J = 4.2$  Hz, 2H), 7.38 (t,  $J = 3.9$  Hz, 1H), 7.20 (d,  $J = 3.8$  Hz, 1H), 7.16–7.15 (m, 1H), 6.94 (dd,  $J = 4.1$ , 1.0 Hz, 1H) 3.94 (s, 3H), 3.87 (s, 3H);  $^{13}\text{C}$  NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  167.1, 160.1, 145.6, 141.6, 130.2, 130.1, 129.1, 127.2, 119.9, 113.6, 113.1, 55.5, 52.2; IR (neat) 3064, 2999, 2950, 2835, 1932, 1716, 1605, 1434, 1270, 1103, 1017, 849, 764, 694; HRMS (ESI): calcd for  $[\text{C}_{15}\text{H}_{14}\text{O}_3 + \text{H}]^+$ : 243.1016, found 243.1016.

**Methyl 3'-Cyano-[1,1'-biphenyl]-4-carboxylate (30c).** Method B was followed on a 0.30 mmol scale starting from methyl 4-iodobenzoate 28b and organobismuthine 1s. The crude material was purified on silica gel (10% EtOAc/hexanes) to afford 30c as a white solid (62 mg, 87%): mp 137–138 °C;  $R_f$  0.60 (20% EtOAc/hexanes). Spectral data were identical to those of the literature compound:<sup>43</sup>  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.14–8.10 (m, 2H), 7.88–7.87 (m, 1H), 7.85–7.81 (m, 1H), 7.68–7.54 (m, 4H) 3.94 (s, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  166.7, 143.2, 141.3, 131.7, 131.6, 130.9, 130.5, 130.1, 129.9, 127.2, 118.6, 113.3, 52.4; IR (neat) 3411, 3068, 2954, 2836, 2232, 1608, 1429, 1281, 1186, 1102, 764, 684; HRMS (ESI): calcd for  $[\text{C}_{15}\text{H}_{11}\text{NO}_2 + \text{H}]^+$ : 238.0863, found 238.0870.

**3'-Vinyl-[1,1'-biphenyl]-4-carbaldehyde (30d).** Method C was followed on a 0.17 mmol scale starting from 4-bromobenzaldehyde 28d and organobismuthine 21g. The crude material was purified on silica gel (5% EtOAc/hexanes) to afford 30d as a colorless oil (27 mg, 76%):  $R_f$  0.30 (5% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  10.07 (s, 1H), 7.98–7.94 (m, 2H), 7.79–7.74 (m, 2H), 7.66–7.65 (m, 1H), 7.55–7.51 (m, 1H), 7.49–7.42 (m, 2H), 6.80 (dd,  $J = 17.6$ , 10.8 Hz, 1H), 5.84 (d,  $J = 17.6$  Hz, 1H), 5.33 (d,  $J = 10.9$  Hz, 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  192.0, 147.2, 140.2, 138.5, 136.6, 135.4, 130.4, 129.3, 127.9, 126.9, 126.3, 125.5, 114.9; IR (neat) 3407 (br), 3057, 2916, 2844, 2734, 1692, 1601, 1384, 1209, 1167, 988, 836, 791, 699; HRMS (ESI): calcd for  $[\text{C}_{15}\text{H}_{12}\text{O} + \text{H}]^+$ : 209.0961, found 209.0965.

**5-(3-(1-Hydroxyethyl)phenyl)furan-2-carbaldehyde (31a).** Method A was followed on a 0.23 mmol scale starting from 5-iodofuran-2-carbaldehyde 29a and organobismuthine 21d. The crude material was purified on silica gel (30% EtOAc/hexanes) to afford 31a as a yellow oil (41 mg, 82%):  $R_f$  0.25 (40% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  9.65 (s, 1H), 7.85 (s, 1H), 7.74–7.71 (m, 1H), 7.43–7.41 (m, 2H), 7.32 (d,  $J = 3.7$  Hz, 1H), 6.86 (d,  $J = 3.7$  Hz, 1H), 4.97 (q,  $J = 6.5$  Hz, 1H), 1.96 (s(br), 1H), 1.53 (d,  $J = 6.5$  Hz, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  177.4, 159.5, 151.9, 146.9, 129.2, 129.1, 126.8, 124.4, 123.8, 122.3, 107.9, 70.1, 25.4; IR (neat) 3416 (br), 3103, 2972, 2916, 2871, 2821, 1663, 1516, 1468, 1426, 1265, 1071, 1028, 792; HRMS (ESI) calcd for  $[\text{C}_{13}\text{H}_{12}\text{O}_3 + \text{H}]^+$ : 217.0859, found 217.0867.

**3'-Hydroxy-[1,1'-biphenyl]-3-carbaldehyde (43).** Method A was followed on a 0.23 mmol scale starting from 3-iodophenol 39 and organobismuthine 21b. The crude material was purified on silica gel (20% EtOAc/hexanes) to afford 43 as a white solid (23 mg, 50%): mp 88–89 °C;  $R_f$  0.38 (20% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  10.09 (s, 1H), 8.08 (t,  $J = 1.6$  Hz, 1H), 7.86 (dt,  $J = 7.4$ , 1.4 Hz, 2H), 7.61 (t,  $J = 7.7$  Hz, 1H), 7.34 (t,  $J = 7.9$  Hz, 1H), 7.20 (dt,  $J = 7.6$ , 1.4 Hz, 1H), 7.11 (t,  $J = 2.2$  Hz, 1H), 6.87 (ddd,  $J = 8.1$ , 2.5, 0.8 Hz, 1H), 4.97 (s(br), 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  192.7, 156.3, 141.9, 141.5, 137.0, 133.3, 130.4, 129.7, 129.1, 128.3, 119.8, 115.2, 114.3; IR (neat) 3363 (br), 3062, 2927, 2831, 2734, 1686, 1599, 1588, 1458, 1308, 1212, 1169, 779, 690; HRMS (ESI) calcd for  $[\text{C}_{13}\text{H}_{10}\text{O}_2 + \text{H}]^+$ : 199.0754, found 199.0750.

**3-(1H-Indol-5-yl)benzaldehyde (44).** Method A was followed on 0.21 mmol scale starting from 5-iodo-1H-indole 40 and organobismuthine 21b. The crude material was purified on silica gel (20% EtOAc/hexanes) to afford 44 as a yellow oil (18 mg, 39%):  $R_f$  0.43 (20% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  10.11 (s, 1H), 8.26 (s(br), 1H), 8.17 (t,  $J = 1.8$  Hz, 1H), 7.95–7.91 (m, 1H), 7.91 (s, 1H), 7.82 (dt,  $J = 7.6$ , 1.4 Hz, 1H), 7.60 (t,  $J = 7.6$  Hz, 1H), 7.49 (s, 2H), 7.28 (t,  $J = 2.9$  Hz, 1H), 6.64 (dd,  $J = 3.1$ , 1.9 Hz, 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  192.8, 143.6, 137.0, 135.8, 133.4, 132.0, 129.5, 128.6, 127.8, 125.3, 121.8, 119.6, 111.6, 103.1; IR (neat) 3414 (br), 3057, 2825, 2738, 1689, 1599, 1578, 1466, 1424, 1316, 1197, 792; HRMS (ESI) calcd for  $[\text{C}_{15}\text{H}_{11}\text{NO} + \text{H}]^+$ : 222.0913, found 222.0924.

**General Procedure for the N-Arylation of Indoles, Pyrroles, Pyrazoles, and Pyridones.** Compounds 17a, 34a–e, 35a,b, and 42 were prepared according to the following procedures: **Method A:** In a sealed tube, the triarylbi-muthine (1.0 equiv) was added, followed by copper(II) acetate (0.1 equiv) and the indole, pyrrole, pyrazole, or pyridone (1.0 equiv). The reagents were dissolved in anhydrous dichloromethane (4 mL), and pyridine (1.0 equiv) was added to the mixture. The reaction tube was purged with dry oxygen for 30 s, sealed, and heated at 50 °C overnight. The reaction mixture was cooled to room temperature, transferred, and rinsed with EtOAc in a round-bottom flask. Silica gel was added, and the mixture was concentrated under reduced pressure. The crude product was purified by flash column chromatography on silica gel using the indicated solvent system as the eluent to give the corresponding product. **Method B:** Same as method A except for copper(II) acetate (1.0 equiv instead of 0.1 equiv) and pyridine (3.0 equiv instead of 1.0 equiv). **Method C:** Same as method A except that 1.0 equiv of copper(II) acetate and 3.0 equiv of pyridine were used and the reaction was performed at 50 °C under air in a sealed tube overnight.

**(E)-Ethyl 3-(3-(3-Cyano-2-oxopyridin-1(2H)-yl)phenyl)acrylate (17a).** Method B was followed on a 0.21 mmol scale starting from 2-oxo-1,2-dihydropyridine-3-carbonitrile and organobismuthine 21f. The crude product was purified on silica gel (50% EtOAc/hexanes) to afford 17a as a white solid (52 mg, 84%): mp 173–174 °C;  $R_f$  0.37 (60% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.91 (dd,  $J = 7.1$ , 2.1 Hz, 1H), 7.66 (d,  $J = 16.1$  Hz, 1H), 7.63–7.59 (m, 2H), 7.54 (d,  $J = 7.7$  Hz, 1H), 7.52–7.51 (m, 1H), 7.38 (dt,  $J = 7.9$ , 1.6 Hz, 1H), 6.45 (d,  $J = 16.1$  Hz, 1H), 6.38 (t,  $J = 7.0$  Hz, 1H), 4.26 (q,  $J = 7.1$  Hz, 2H), 1.33 (t,  $J = 7.1$  Hz, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  166.5, 159.3, 148.0, 142.9, 142.6, 140.1, 136.4, 130.3, 129.0, 127.7, 125.6, 120.5, 115.3, 107.0, 105.8, 60.9, 14.4; IR (neat) 3080, 2977, 2924, 2905, 2228, 1707, 1662, 1640, 1542, 1269, 1181; HRMS (ESI) calcd for  $[\text{C}_{17}\text{H}_{14}\text{N}_2\text{O}_3 + \text{H}]^+$ : 295.1077, found 295.1099.

**1-(4-(Tetrahydro-2H-pyran-2-yl)oxy)phenyl)-1H-indole-5-carbaldehyde (34a).** Method B was followed on a 0.17 mmol scale starting from 1H-indole-5-carbaldehyde 32 and organobismuthine 1m. The crude product was purified on silica gel (15% EtOAc/hexanes) to afford 34a as a yellow solid (43 mg, 79%): mp 105–106 °C;  $R_f$  0.52 (20% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  10.04 (s, 1H), 8.20 (s, 1H), 7.76 (d,  $J = 8.6$  Hz, 1H), 7.53–7.46 (m, 1H), 7.41–7.30 (m, 3H), 7.26–7.20 (m, 2H), 6.80 (d,  $J = 3.2$  Hz, 1H), 5.49 (t,  $J = 3.3$  Hz, 1H), 3.99–3.91 (m, 1H), 3.70–3.63 (m, 1H), 2.08–1.99 (m, 1H), 1.94–1.89 (m, 2H), 1.78–1.62 (m, 3H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  192.5, 156.5, 139.7, 132.7, 130.5, 130.0, 128.8, 126.5, 126.2, 122.6, 117.6, 111.2, 104.8, 96.7, 62.3, 30.4, 25.3, 18.8; IR (neat) 3101, 3039, 2943, 2873, 2850, 2715, 1683, 1615, 1509, 1330, 1237, 1221, 1201, 1102, 1044, 921, 731; HRMS (ESI) calcd for  $[\text{C}_{20}\text{H}_{19}\text{NO}_3 + \text{H}]^+$ : 322.1438, found 322.1430.

**1-(2-Formylphenyl)-1H-indole-5-carbaldehyde (34b).** Method B was followed on a 0.17 mmol scale starting from 1H-indole-5-carbaldehyde 32 and organobismuthine 21c. The crude product was purified on silica gel (20% EtOAc/hexanes) to afford 34b as a yellow oil (15 mg, 35%):  $R_f$  0.44 (20% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  10.07 (s, 1H), 9.63 (s, 1H), 8.25 (d,  $J = 0.9$  Hz, 1H), 8.13 (dd,  $J = 7.7$ , 1.5 Hz, 1H), 7.83–7.77 (m, 2H), 7.65 (t,  $J = 7.6$  Hz, 1H), 7.51 (dd,  $J = 7.9$ , 0.8 Hz, 1H), 7.39 (d,  $J = 3.3$  Hz, 1H), 7.24 (d,  $J = 8.2$  Hz, 1H), 6.91 (dd,  $J = 3.3$ , 0.9 Hz, 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  192.3, 189.1, 141.5, 141.0, 135.4, 132.3, 131.9, 131.7, 130.7, 129.2, 128.7, 128.4, 126.4, 123.6, 110.9, 106.0; IR (neat) 3357, 3106, 3065, 2977, 2920, 2867, 2745, 1687, 1597, 1490, 1331, 1195; HRMS (ESI) calcd for  $[\text{C}_{16}\text{H}_{11}\text{NO}_2 + \text{H}]^+$ : 250.0868, found 250.0873.

**1-(3-(Hydroxymethyl)phenyl)-1H-indole-5-carbaldehyde (34c).** Method B was followed on a 0.09 mmol scale starting from 1H-indole-5-carbaldehyde 32 and organobismuthine 21e. The crude product was purified on silica gel (30% EtOAc/hexanes) to afford 34c as a yellow oil (17 mg, 75%):  $R_f$  0.15 (20% EtOAc/hexanes);  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  10.06 (s, 1H), 8.22 (d,  $J = 1.1$  Hz, 1H), 7.78 (dd,  $J = 8.7$ , 1.5 Hz, 1H), 7.59 (d,  $J = 8.8$  Hz, 1H), 7.55–7.53 (m, 2H), 7.44–7.41 (m, 3H), 6.84 (d,  $J = 3.3$  Hz, 1H), 4.83 (d,  $J = 4.0$  Hz, 2H), 1.84 (t,  $J = 5.0$  Hz, 1H);  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  192.5, 143.2,



139.3, 130.2, 130.1, 130.0, 129.2, 126.5, 125.8, 123.8, 123.1, 122.8, 111.2, 105.4, 105.4, 64.8; IR (neat) 3411 (br), 3105, 3055, 2920, 2858, 2815, 2727, 1682, 1601, 1589, 1565, 1493, 1449, 1331, 1223, 1103, 1032, 894, 726, 698; HRMS (ESI) calcd for  $[C_{16}H_{13}NO_2 + H]^+$ : 252.1019, found 252.1012.

**1-(3-Acetylphenyl)-1H-indole-5-carbaldehyde (34d).** Method B was followed on a 0.17 mmol scale starting from 1H-indole-5-carbaldehyde 32 and organobismuthine 21h. The crude product was purified on silica gel (30% EtOAc/hexanes) to afford 34d as a yellow oil (27 mg, 60%);  $R_f$  0.34 (20% EtOAc/hexanes);  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  10.07 (s, 1H), 8.23 (d,  $J$  = 1.5 Hz, 1H), 8.09 (t,  $J$  = 1.8 Hz, 1H), 8.00 (dt,  $J$  = 7.4, 1.6 Hz, 1H), 7.81 (dd,  $J$  = 8.7, 1.6 Hz, 1H), 7.73 (dt,  $J$  = 8.0, 1.7 Hz, 1H), 7.70–7.65 (m, 1H), 7.57 (d,  $J$  = 8.7 Hz, 1H), 7.45 (d,  $J$  = 3.3 Hz, 1H), 6.87 (d,  $J$  = 3.3 Hz, 1H), 2.68 (s, 3H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  197.1, 192.3, 139.6, 139.2, 139.0, 130.5, 130.4, 129.9, 129.3, 129.1, 127.3, 126.5, 124.3, 123.2, 111.0, 106.0, 26.9; IR (neat) 3357, 3103, 3057, 2924, 2825, 2741, 1682, 1599, 1586, 1491, 1445, 1330, 1236, 1105, 769; HRMS (ESI) calcd for  $[C_{17}H_{13}NO_2 + H]^+$ : 264.1019, found 264.1012.

**1-Phenyl-1H-indole-5-carbaldehyde (34e).** Method B was followed on a 0.29 mmol scale starting from 1H-indole-5-carbaldehyde 32 and organobismuthine 1a. The crude material was purified on silica gel (20% EtOAc/hexanes) to afford 34e as a yellow oil (62 mg, 97%);  $R_f$  0.14 (10% EtOAc/hexanes). Spectral data were identical to those of the literature compound.<sup>44</sup>  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  10.06 (s, 1H), 8.22 (s, 1H), 7.57 (d,  $J$  = 8.7 Hz, 1H), 7.61–7.54 (m, 3H), 7.52–7.49 (m, 2H), 7.46–7.41 (m, 2H), 6.84 (d,  $J$  = 3.2 Hz, 1H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  192.4, 139.4, 139.1, 130.3, 130.2, 130.0, 129.2, 127.6, 126.5, 124.9, 122.8, 111.2, 105.4; IR (neat) 3060, 2962, 2921, 2815, 2778, 2709, 1683, 1593, 1498, 1448, 1338, 1222, 1103, 886, 755, 695; HRMS (ESI) calcd for  $[C_{15}H_{11}NO + H]^+$ : 222.0913, found 222.0919.

**1-(1-(3-(1-Hydroxyethyl)phenyl)-1H-pyrrol-2-yl)ethanone (35a).** Method A was followed on a 0.13 mmol scale starting from 1-(1H-pyrrol-2-yl)ethanone and organobismuthine 21d. The crude product was purified on silica gel (35% EtOAc/hexanes) to afford 35a as a yellow oil (30 mg, 99%);  $R_f$  0.19 (20% EtOAc/hexanes);  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.34–7.32 (m, 2H), 7.26 (s, 1H), 7.13–7.08 (m, 2H), 6.94 (dd,  $J$  = 2.4, 1.8 Hz, 1H), 6.28 (dd,  $J$  = 4.1, 2.6 Hz, 1H), 4.84 (q,  $J$  = 6.4 Hz, 1H), 3.51 (s(br), 1H), 2.38 (s, 3H), 1.45 (d,  $J$  = 6.5 Hz, 3H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  187.5, 147.1, 140.8, 131.5, 131.4, 128.6, 124.8, 124.7, 123.4, 120.9, 109.3, 69.6, 27.2, 25.0; IR (neat) 3409 (br), 3114, 3057, 2973, 2920, 2867, 1644, 1489, 1405, 1366, 1348, 1110, 1084, 1050, 940, 793, 738, 654; HRMS (ESI) calcd for  $[C_{14}H_{15}NO_2 + H]^+$ : 230.1176, found 230.1178.

**4-Methyl-1-(3-vinylphenyl)-1H-pyrazole (35b).** Method B was followed on a 0.43 mmol scale starting from 4-methyl-1H-pyrazole and organobismuthine 21g. The crude product was purified on silica gel (20% EtOAc/hexanes) to afford 35b as a yellow oil (70 mg, 88%);  $R_f$  0.53 (20% EtOAc/hexanes);  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.72 (t,  $J$  = 2.0 Hz, 1H), 7.70–7.69 (m, 1H), 7.53 (s, 1H), 7.51–7.48 (m, 1H), 7.35 (t,  $J$  = 7.7 Hz, 1H), 7.27 (dt,  $J$  = 7.7, 1.5 Hz, 1H), 6.73 (dd,  $J$  = 17.6, 10.8 Hz, 1H), 5.82 (d,  $J$  = 17.6 Hz, 1H), 5.31 (d,  $J$  = 10.9 Hz, 1H), 2.14 (s, 3H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  141.9, 140.6, 139.1, 136.3, 129.6, 125.5, 123.9, 118.4, 118.0, 116.6, 115.1, 9.1; IR (neat) 3141, 3118, 3065, 2926, 2863, 1695, 1606, 1585, 1533, 1489, 1454, 1399, 1362, 1047, 791, 755, 697; HRMS (ESI) calcd for  $[C_{12}H_{12}N_2 + H]^+$ : 185.1073, found 185.1078.

**3-(5-Iodo-1H-indol-1-yl)benzaldehyde (42).** Method B was followed on 0.12 mmol scale starting from 5-iodo-1H-indole 40 and organobismuthine 21b. The crude material was purified on silica gel (20% EtOAc/hexanes) to afford 42 as a yellow oil (29 mg, 70%);  $R_f$  0.54 (20% EtOAc/hexanes);  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  10.10 (s, 1H), 8.03 (d,  $J$  = 1.5 Hz, 1H), 7.98–7.97 (m, 1H), 7.87 (dt,  $J$  = 7.0, 1.6 Hz, 1H), 7.76–7.68 (m, 2H), 7.49 (dd,  $J$  = 8.7, 1.6 Hz, 1H), 7.33 (s, 1H), 7.31 (d,  $J$  = 5.2 Hz, 1H), 6.65 (d,  $J$  = 3.3 Hz, 1H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  191.4, 140.3, 138.0, 134.9, 132.1, 131.2, 130.7, 130.3, 129.9, 128.5, 128.4, 124.3, 112.3, 103.8, 84.4; IR (neat) 3106, 3068, 2958, 2920, 2829, 2734, 1698, 1588, 1513, 1487, 1459, 1237, 793; HRMS (ESI) calcd for  $[C_{15}H_{10}INO + H]^+$ : 347.9880, found 347.9896.

**General Procedure for the O-Arylation of Phenols.** Compounds 37a–e and 41 were prepared according to the following procedures: **Method A:** In a sealed tube, the phenol (1.0 equiv) was dissolved in nonanhydrous solvent grade dichloromethane (3 mL). The organobismuthine (1.0 equiv) was added followed by copper(II) acetate (1.0 equiv) and  $Et_3N$  (3.0 equiv). The tube was sealed and heated at 50 °C for 3 h under air. The reaction mixture was cooled to room temperature, and silica gel was added. The mixture was concentrated under reduced pressure, and the crude product was purified by flash column chromatography using the indicated solvent system to afford the corresponding product. **Method B:** Same as method A except that 0.3 equiv of  $Cu(OAc)_2$  was used and the reaction was performed under  $O_2$  for 16 h.

**Methyl 3-(3-(Diethoxymethyl)phenoxy)benzoate (37a).** Method A was followed on a 0.26 mmol scale starting from methyl 3-hydroxybenzoate 36 and organobismuthine 1o. The crude material was purified on silica gel (15% EtOAc/hexanes) to afford 37a as a colorless oil (75 mg, 87%);  $R_f$  0.62 (20% EtOAc/hexanes);  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.77 (dt,  $J$  = 7.7, 1.3 Hz, 1H), 7.65 (dd,  $J$  = 2.6, 1.5 Hz, 1H), 7.42–7.31 (m, 2H), 7.27–7.24 (m, 1H), 7.20 (ddd,  $J$  = 8.2, 2.6, 1.1 Hz, 1H), 7.16 (t,  $J$  = 2.1 Hz, 1H), 6.97 (ddd,  $J$  = 7.9, 2.6, 1.2 Hz, 1H), 5.48 (s, 1H), 3.88 (s, 3H), 3.67–3.48 (m, 4H), 1.22 (t,  $J$  = 7.0 Hz, 6H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  166.6, 157.5, 156.8, 141.6, 132.0, 129.9, 129.8, 124.4, 123.4, 122.2, 119.6, 119.0, 117.6, 101.1, 61.2, 52.4, 15.3; IR (neat) 3068, 2973, 2920, 2878, 1726, 1684, 1582, 1483, 1441, 1357, 1270, 1049, 901, 794, 753, 696; HRMS (ESI) calcd for  $[C_{19}H_{22}O_5 + Na]^+$ : 353.1359, found 353.1369.

**Methyl 3-(2-(Diethoxymethyl)phenoxy)benzoate (37b).** Method B was followed on a 0.16 mmol scale starting from methyl 3-hydroxybenzoate 36 and organobismuthine 1n. The crude material was purified on silica gel (15% EtOAc/hexanes) to afford 37b as a colorless oil (39 mg, 74%);  $R_f$  0.73 (20% EtOAc/hexanes);  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.78–7.73 (m, 1H), 7.70 (dd,  $J$  = 7.6, 1.8 Hz, 1H), 7.64 (dd,  $J$  = 2.6, 1.5 Hz, 1H), 7.37 (t,  $J$  = 7.9 Hz, 1H), 7.28 (dt,  $J$  = 7.4, 1.8 Hz, 1H), 7.20 (td,  $J$  = 7.5, 1.3 Hz, 1H), 7.14 (ddd,  $J$  = 8.3, 2.6, 1.1 Hz, 1H), 6.88 (dd,  $J$  = 8.2, 1.5 Hz, 1H), 5.73 (s, 1H), 3.89 (s, 3H), 3.70–3.60 (m, 2H), 3.56–3.49 (m, 2H), 1.16 (t,  $J$  = 7.1 Hz, 6H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  166.6, 158.2, 153.8, 131.9, 131.2, 129.9, 129.7, 128.0, 124.4, 124.0, 122.6, 119.7, 119.0, 97.6, 62.4, 52.3, 15.2; IR (neat) 3066, 2975, 2877, 1724, 1580, 1482, 1444, 1271, 1230, 1202, 1052, 994, 905, 754; HRMS (ESI) calcd for  $[C_{19}H_{22}O_5 + Na]^+$ : 353.1359, found 353.1363.

**Methyl 3-(3-Vinylphenoxy)benzoate (37c).** Method B was followed on a 0.13 mmol scale starting from methyl 3-hydroxybenzoate 36 and organobismuthine 21g. The crude material was purified on silica gel (10% EtOAc/hexanes) to afford 37c as a yellow oil (27 mg, 82%);  $R_f$  0.74 (20% EtOAc/hexanes);  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.78 (dt,  $J$  = 7.7, 1.3 Hz, 1H), 7.66 (dd,  $J$  = 2.6, 1.6 Hz, 1H), 7.41 (t,  $J$  = 8.0 Hz, 1H), 7.31 (t,  $J$  = 7.8 Hz, 1H), 7.23–7.17 (m, 2H), 7.07 (t,  $J$  = 2.0 Hz, 1H), 6.90 (ddd,  $J$  = 8.1, 2.4, 1.0 Hz, 1H), 6.67 (dd,  $J$  = 17.6, 10.9 Hz, 1H), 5.70 (dd,  $J$  = 17.6, 0.8 Hz, 1H), 5.27 (d,  $J$  = 10.9 Hz, 1H), 3.90 (s, 3H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  166.6, 157.5, 157.1, 139.8, 136.3, 132.1, 130.1, 129.9, 124.5, 123.4, 122.0, 119.7, 118.6, 116.8, 115.0, 52.4; IR (neat) 3076, 3008, 2952, 2928, 2844, 1724, 1573, 1485, 1442, 1275, 1250, 1208, 1143, 1098, 991; HRMS (ESI) calcd for  $[C_{16}H_{14}O_3 + H]^+$ : 255.1016, found 255.1020.

**(E)-Methyl 3-(3-(3-Ethoxy-3-oxoprop-1-en-1-yl)phenoxy)benzoate (37d).** Method A was followed on a 0.25 mmol scale starting from methyl 3-hydroxybenzoate 36 and organobismuthine 21f. The crude material was purified on silica gel (15% EtOAc/hexanes) to afford 37d as a yellow oil (43 mg, 53%);  $R_f$  0.61 (20% EtOAc/hexanes);  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.81 (td,  $J$  = 7.8, 1.3 Hz, 1H), 7.67–7.66 (m, 1H), 7.62 (d,  $J$  = 16.0 Hz, 1H), 7.46–7.40 (m, 1H), 7.39–7.34 (m, 1H), 7.30–7.27 (m, 1H), 7.22 (ddd,  $J$  = 8.2, 2.6, 1.1 Hz, 1H), 7.14 (t,  $J$  = 2.1 Hz, 1H), 7.02 (ddd,  $J$  = 8.0, 1.9, 1.2 Hz, 1H), 6.37 (d,  $J$  = 16.0 Hz, 1H), 4.25 (q,  $J$  = 7.1 Hz, 2H), 3.90 (s, 3H), 1.32 (t,  $J$  = 7.1 Hz, 3H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  166.9, 166.5, 157.5, 157.0, 143.8, 136.6, 132.2, 130.5, 130.1, 124.9, 123.7, 123.6, 120.7, 120.0, 119.3, 118.0, 60.7, 52.4, 14.4; IR (neat) 3065,



2980, 2953, 2901, 1709, 1639, 1575, 1484, 1442, 1269, 1230, 1177, 1097, 1036, 983, 756; HRMS (ESI) calcd for  $[C_{19}H_{18}O_5 + H]^+$ : 327.1227, found 327.1224.

**Methyl 3-Phenoxybenzoate (37e).** A modified method B using 1.0 equiv of  $Cu(OAc)_2$  instead of 0.3 equiv and pyridine instead of triethylamine was followed on a 0.33 mmol scale starting from methyl 3-hydroxybenzoate 36 and organobismuthine 1a. The crude material was purified on silica gel (5% EtOAc/hexanes) to afford 37e as a pale yellow oil (53 mg, 70%);  $R_f$  0.43 (10% EtOAc/hexanes). Spectral data were identical to those of the literature compound:<sup>45</sup>  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.79 (td,  $J$  = 7.7, 1.3 Hz, 1H), 7.68 (t,  $J$  = 2.1 Hz, 1H), 7.43–7.32 (m, 3H), 7.21 (dd,  $J$  = 8.2, 2.5 Hz, 1H), 7.14 (t,  $J$  = 7.4 Hz, 1H), 7.02 (d,  $J$  = 8.2 Hz, 2H), 3.89 (s, 3H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  166.5, 157.5, 156.7, 131.9, 129.9, 129.8, 124.3, 123.8, 123.3, 119.6, 119.1, 52.2; IR (neat) 3068, 2947, 2840, 1722, 1582, 1479, 1437, 1266, 1228, 1091, 988, 904, 749, 688; HRMS (ESI) calcd for  $[C_{14}H_{12}O_3 + H]^+$ : 229.0859, found 229.0863.

**3-(3-Iodophenoxy)benzaldehyde (41).** Method B was followed on a 0.23 mmol scale starting from 3-iodophenol 39 and organobismuthine 21b. The crude material was purified on silica gel (15% EtOAc/hexanes) to afford 41 as a yellow oil (33 mg, 44%);  $R_f$  0.69 (20% EtOAc/hexanes);  $^1H$  NMR (300 MHz,  $CDCl_3$ )  $\delta$  9.97 (s, 1H), 7.64 (dt,  $J$  = 7.4, 1.1 Hz, 1H), 7.52–7.46 (m, 3H), 7.37 (t,  $J$  = 2.0 Hz, 1H), 7.28 (ddd,  $J$  = 8.1, 2.5, 1.0 Hz, 1H), 7.09 (t,  $J$  = 8.1 Hz, 1H), 6.99 (ddd,  $J$  = 8.2, 2.3, 0.9 Hz, 1H);  $^{13}C$  NMR (75 MHz,  $CDCl_3$ )  $\delta$  191.5, 157.7, 157.1, 138.3, 133.3, 131.4, 130.8, 128.4, 125.5, 125.0, 118.7, 118.6, 94.5; IR (neat) 3059, 2923, 2827, 2734, 1697, 1572, 1465, 1450, 1243, 1208, 846, 781; HRMS (ESI) calcd for  $[C_{13}H_9IO_2 + H]^+$ : 324.9720, found 324.9714.

## ■ ASSOCIATED CONTENT

### Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.6b00767.

Crystallographic data for 1f, 1i, 1k, 1n, 1s, 1t, 17a, 21c, and 21e (ZIP)

Copies of  $^1H$ ,  $^{13}C$ , and IR spectra (PDF)

## ■ AUTHOR INFORMATION

### Corresponding Author

\*E-mail: gagnon.alexandre@uqam.ca.

### Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

This work was supported by the Natural Sciences and Engineering Research Council of Canada (NSERC), the Université du Québec à Montréal (UQÀM) and the Centre in Green Chemistry and Catalysis. M.H. thanks NSERC and FRQNT for graduate scholarships. P.P. and J.D. thank UQÀM for a FARE scholarship.

## ■ REFERENCES

- (1) For a study on the toxicity of bismuth salts and its elimination from human cells by conjugation with glutathione, see: Hong, Y.; Lai, Y.-T.; Chan, G. C.-F.; Sun, H. *Proc. Natl. Acad. Sci. U. S. A.* **2015**, *112*, 3211–3216.
- (2) For reviews on organobismuth reagents, see: (a) Condon, S.; Pichon, C.; Davi, M. *Org. Prep. Proced. Int.* **2014**, *46*, 89–131. (b) Elliott, G. I.; Konopelski, J. P. *Tetrahedron* **2001**, *57*, S683–S705. (c) Suzuki, H.; Ikegami, T.; Matano, Y. *Synthesis* **1997**, *1997*, 249–267. (d) Postel, M.; Dunach, E. *Coord. Chem. Rev.* **1996**, *155*, 127–144. (e) Finet, J.-P. *Chem. Rev.* **1989**, *89*, 1487–1501. (f) Barton, D. H. R.; Finet, J.-P. *Pure Appl. Chem.* **1987**, *59*, 937–946. (g) Freedman, L. D.; Doak, G. O. *Chem. Rev.* **1982**, *82*, 15–57. (h) Gilman, H.; Yale, H. L. *Chem. Rev.* **1942**, *30*, 281–320.

- (3) For selected examples of applications of organobismuth reagents in methodology development, see: (a) Kobiki, Y.; Kawaguchi, S.-i.; Ogawa, A. *Org. Lett.* **2015**, *17*, 3490–3493. (b) Wang, T.; Sang, S.; Liu, L.; Qiao, H.; Gao, Y.; Zhao, Y. *J. Org. Chem.* **2014**, *79*, 608–617. (c) Sueda, T.; Oshima, A.; Teno, N. *Org. Lett.* **2011**, *13*, 3996–3999. (d) Sato, I.; Toyota, Y.; Asakura, N. *Eur. J. Org. Chem.* **2007**, *2007*, 2608–2610. (e) Ikegai, K.; Fukumoto, K.; Mukaiyama, T. *Chem. Lett.* **2006**, *35*, 612–613. (f) Mukaiyama, T.; Sakurai, N.; Ikegai, K. *Chem. Lett.* **2006**, *35*, 1140–1141. (g) Koech, P. K.; Krische, M. J. *J. Am. Chem. Soc.* **2004**, *126*, 5350–5351. (h) Ooi, T.; Goto, R.; Maruoka, K. *J. Am. Chem. Soc.* **2003**, *125*, 10494–10495. (i) Cho, C. S.; Yoshimori, Y.; Uemura, S. *Bull. Chem. Soc. Jpn.* **1995**, *68*, 950–957.

- (4) For examples of applications of organobismuth reagents in total synthesis, see: (a) Nicolaou, K. C.; Sarlah, D.; Wu, T. R.; Zhan, W. *Angew. Chem., Int. Ed.* **2009**, *48*, 6870–6874. (b) Krawczuk, P. J.; Schöne, N.; Baran, P. S. *Org. Lett.* **2009**, *11*, 4774–4776.

- (5) For reviews on organobismuth compounds as ligands in transition metal complexes, see: (a) Benjamin, S. L.; Reid, G. *Coord. Chem. Rev.* **2015**, *297–298*, 168–180. (b) Braunschweig, H.; Cogswell, P.; Schwab, K. *Coord. Chem. Rev.* **2011**, *255*, 101–117.

- (6) For reviews on polymerization reactions using organobismuth reagents, see: Yamago, S. *Chem. Rev.* **2009**, *109*, 5051–5068 and references cited therein.

- (7) For reviews on the application of organobismuth compounds in medicinal chemistry, see: (a) Yang, Y.; Ouyang, R.; Xu, L.; Guo, N.; Li, W.; Feng, K.; Ouyang, L.; Yang, Z.; Zhou, S.; Miao, Y. *J. Coord. Chem.* **2015**, *68*, 379–397. (b) Briand, G. G.; Burford, N. *Chem. Rev.* **1999**, *99*, 2601–2657.

- (8) (a) Combes, S.; Finet, J.-P. *Tetrahedron* **1999**, *55*, 3377–3386. (b) Fedorov, A. Y.; Finet, J.-P. *Tetrahedron Lett.* **1999**, *40*, 2747–2748. (c) Combes, S.; Finet, J.-P. *Tetrahedron* **1998**, *54*, 4313–4318. (d) Arnauld, T.; Barton, D. H. R.; Doris, E. *Tetrahedron* **1997**, *53*, 4137–4144. (e) Abramovitch, R. A.; Barton, D. H. R.; Finet, J.-P. *Tetrahedron* **1988**, *44*, 3039–3071. (f) Barton, D. H. R.; Finet, J.-P.; Khamsi, J. *Tetrahedron Lett.* **1988**, *29*, 1115–1118. (g) Barton, D. H. R.; Finet, J.-P.; Khamsi, J. *Tetrahedron Lett.* **1987**, *28*, 887–890. (h) Barton, D. H. R.; Finet, J. P.; Khamsi, J.; Pichon, C. *Tetrahedron Lett.* **1986**, *27*, 3619–3622. (i) Barton, D. H. R.; Finet, J.-P.; Khamsi, J. *Tetrahedron Lett.* **1986**, *27*, 3615–3618.

- (9) For selected references, see: (a) Rao, M. L. N.; Dhanorkar, R. J. *RSC Adv.* **2016**, *6*, 1012–1017. (b) Rao, M. L. N.; Dhanorkar, R. J. *Eur. J. Org. Chem.* **2014**, *2014*, 5214–5228. (c) Rao, M. L. N.; Dhanorkar, R. J. *RSC Adv.* **2014**, *4*, 13134–13144. (d) Rao, M. L. N.; Dhanorkar, R. J. *Tetrahedron* **2014**, *70*, 8067–8078. (e) Rao, M. L. N.; Jadhav, D. N.; Dasgupta, P. *Eur. J. Org. Chem.* **2013**, *2013*, 781–788. (f) Rao, M. L. N.; Banerjee, D.; Giri, S. J. *Organomet. Chem.* **2010**, *695*, 1518–1525. (g) Rao, M. L. N.; Jadhav, D. N.; Dasgupta, P. *Org. Lett.* **2010**, *12*, 2048–2051. (h) Rao, M. L. N.; Venkatesh, V.; Dasgupta, P. *Tetrahedron Lett.* **2010**, *51*, 4975–4980.

- (10) (a) Petiot, P.; Gagnon, A. *Heterocycles* **2014**, *88*, 1615–1624. (b) Petiot, P.; Gagnon, A. *Eur. J. Org. Chem.* **2013**, *2013*, 5282–5289. (c) Gagnon, A.; Albert, V.; Duplessis, M. *Synlett* **2010**, *2010*, 2936–2940. (d) Gagnon, A.; Duplessis, M.; Fader, L. *Org. Prep. Proced. Int.* **2010**, *42*, 1–69. (e) Gagnon, A.; Duplessis, M.; Alsabeh, P.; Barabé, F. *J. Org. Chem.* **2008**, *73*, 3604–3607.

- (11) (a) Petiot, P.; Dansereau, J.; Gagnon, A. *RSC Adv.* **2014**, *4*, 22255–22259. (b) Gagnon, A.; St-Onge, M.; Little, K.; Duplessis, M.; Barabé, F. *J. Am. Chem. Soc.* **2007**, *129*, 44–45.

- (12) (a) Petiot, P.; Dansereau, J.; Hébert, M.; Khene, I.; Ahmad, T.; Samaali, S.; Leroy, M.; Pinsonneault, F.; Legault, C. Y.; Gagnon, A. *Org. Biomol. Chem.* **2015**, *13*, 1322–1327. (b) Crifar, C.; Petiot, P.; Ahmad, T.; Gagnon, A. *Chem. - Eur. J.* **2014**, *20*, 2755–2760.

- (13) Urgin, K.; Aubé, C.; Pichon, C.; Pipelier, M.; Blot, V.; Thobie-Gautier, C.; Léonel, E.; Dubreuil, D.; Condon, S. *Tetrahedron Lett.* **2012**, *53*, 1894–1896.

- (14) Dohle, W.; Lindsay, D. M.; Knochel, P. *Org. Lett.* **2001**, *3*, 2871–2873.

- (15) (a) Flöistrup, E.; Goede, P.; Strömberg, R.; Malm, J. *Tetrahedron Lett.* **2011**, *52*, 209–211. (b) Chang, J. H.; Kang, H.-U.; Jung, I.-H.;



- Cho, C.-G. *Org. Lett.* 2010, 12, 2016–2018. (c) de Frutos, O.; Atienza, C.; Echavarren, A. M. *Eur. J. Org. Chem.* 2001, 2001, 163–171.
- (16) (a) Mamane, V.; Louërât, F.; Iehl, J.; Abboud, M.; Fort, Y. *Tetrahedron* 2008, 64, 10699–10705. (b) Wang, N.; Xiang, J.; Quan, J.; Chen, J.; Yang, Z. *J. Comb. Chem.* 2008, 10, 825–834. (c) Kuhnert, N.; Patel, C.; Fatemeh, J. *Tetrahedron Lett.* 2005, 46, 7575–7579. (d) Xiong, Z.; Wang, N.; Dai, M.; Li, A.; Chen, J.; Yang, Z. *Org. Lett.* 2004, 6, 3337–3340. (e) Massa, M. A.; Spangler, D. P.; Durley, R. C.; Hickory, B. S.; Connolly, D. T.; Witherbee, B. J.; Smith, M. E.; Sikorski, J. A. *Bioorg. Med. Chem. Lett.* 2001, 11, 1625–1628. (f) Yamamoto, Y.; Seko, T.; Rong, F. G.; Nemoto, H. *Tetrahedron Lett.* 1989, 30, 7191–7194.
- (17) Bolliger, J. L.; Oberholzer, M.; Frech, C. M. *Adv. Synth. Catal.* 2011, 353, 945–954.
- (18) Adak, L.; Yoshikai, N. *J. Org. Chem.* 2011, 76, 7563–7568.
- (19) (a) Moiseev, D. V.; Malysheva, Y. B.; Shavryin, A. S.; Kurskii, Y. A.; Gushchin, A. V. *J. Organomet. Chem.* 2005, 690, 3652–3663. (b) Malysheva, Y. B.; Moiseev, D. V.; Gushchin, A. V.; Dodonov, V. A. *Russ. J. Gen. Chem.* 2005, 75, 1766–1770. (c) Sharutin, V. V.; Sharutina, O. K.; Senchurin, V. S. *Russ. J. Inorg. Chem.* 2013, 58, 1470–1474. (d) Ong, Y. C.; Blair, V. L.; Kedzierski, L.; Andrews, P. C. *Dalton Trans.* 2014, 43, 12904–12916. (e) Ong, Y. C.; Blair, V. L.; Kedzierski, L.; Tuck, K. L.; Andrews, P. C. *Dalton Trans.* 2015, 44, 18215–18226.
- (20) Rasmussen, L. K.; Begtrup, M.; Ruhland, T. *J. Org. Chem.* 2004, 69, 6890–6893.
- (21) For a review on the structure of organobismuth compounds, see: Silvestru, C.; Breunig, H. J.; Althaus, H. *Chem. Rev.* 1999, 99, 3277–3327.
- (22) Compound 21d was also crystallographically characterized, but positional disorder amidst the 8 possible stereoisomers led to an unsatisfactory structural model.
- (23) (a) Hawley, D. M.; Ferguson, G. J. *Chem. Soc. A* 1968, 2059–2063. (b) Stavila, V.; Thurston, J. H.; Prieto-Centurió, D.; Whitmire, K. H. *Organometallics* 2007, 26, 6864–6866. (c) Schuster, O.; Schier, A.; Schmidbaur, H. *Organometallics* 2003, 22, 4079–4083. (d) Ogawa, T.; Ikegami, T.; Hikasa, T.; Ono, N.; Suzuki, H. *J. Chem. Soc., Perkin Trans. I* 1994, 3479–3483. (e) Kamepalli, S.; Carmalt, C. J.; Culp, R. D.; Cowley, A. H.; Jones, R. A. *Inorg. Chem.* 1996, 35, 6179–6183. (f) Schulz, A.; Villinger, A. *Organometallics* 2011, 30, 284–289. (g) Hassan, A.; Breeze, S. R.; Courtenay, S.; Deslippe, C.; Wang, S. *Organometallics* 1996, 15, 5613–5621. (h) Li, X.-W.; Lorberth, J.; Massa, W.; Wocadlo, S. *J. Organomet. Chem.* 1995, 485, 141–147.
- (24) Berger, R. J. F.; Rettenwander, D.; Spirk, S.; Wolf, C.; Patzschke, M.; Ertl, M.; Monkowius, U.; Mitzel, N. W. *Phys. Chem. Chem. Phys.* 2012, 14, 15520–15524.
- (25) Lin, T.-P.; Ke, I.-S.; Gabbai, F. P. *Angew. Chem., Int. Ed.* 2012, 51, 4985–4988.
- (26) (a) Urbanová, I.; Jambor, R.; Ruzicka, A.; Jirásko, R.; Dostál, L. *Dalton Trans.* 2014, 43, 505–512. (b) Rat, C. I.; Silvestru, C.; Breunig, H. J. *Coord. Chem. Rev.* 2013, 257, 818–879. (c) Simon, P.; Jambor, R.; Ruzicka, A.; Dostál, L. *Organometallics* 2013, 32, 239–248. (d) Benjamin, S. L.; Karagiannidis, L.; Levason, W.; Reid, G.; Rogers, M. C. *Organometallics* 2011, 30, 895–904. (e) Soran, A.; Breunig, H. J.; Lippolis, V.; Arca, M.; Silvestru, C. *J. Organomet. Chem.* 2010, 695, 850–862.
- (27) (a) Urgin, K.; Aubé, C.; Pipelier, M.; Blot, V.; Thobie-Gautier, C.; Sengmany, S.; Lebreton, J.; Léonel, E.; Dubreuil, D.; Condon, S. *Eur. J. Org. Chem.* 2013, 2013, 117–124. (b) Monguchi, Y.; Hattori, T.; Miyamoto, Y.; Yanase, T.; Sawama, Y.; Sajiki, H. *Adv. Synth. Catal.* 2012, 354, 2561–2567. (c) Chaudhari, K. R.; Wadawale, A. P.; Jain, V. K. *J. Organomet. Chem.* 2012, 698, 15–21. (d) Rasmussen, L. K.; Begtrup, M.; Ruhland, T. *J. Org. Chem.* 2004, 69, 6890–6893. (e) Shimada, S.; Yamazaki, O.; Tanaka, T.; Rao, M. L. N.; Suzuki, Y.; Tanaka, M. *Angew. Chem., Int. Ed.* 2003, 42, 1845–1848. (f) Rao, M. L. N.; Yamazaki, O.; Shimada, S.; Tanaka, T.; Suzuki, Y.; Tanaka, M. *Org. Lett.* 2001, 3, 4103–4105.
- (28) For a review on transition-metal catalyzed C–C bond formation using organobismuth compounds, see: Shimada, S.; Rao, M. L. N. *Top. Curr. Chem.* 2011, 311, 199–228.
- (29) Organ, M. G.; Avola, S.; Dubovyk, I.; Hadei, N.; Kantchev, E. A. B.; O'Brien, C. J.; Valente, C. *Chem. - Eur. J.* 2006, 12, 4749–4755.
- (30) (a) McCann, L. C.; Organ, M. G. *Angew. Chem., Int. Ed.* 2014, 53, 4386–4389 and references cited therein. (b) McCann, L. C.; Hunter, H. N.; Clyburne, J. A. C.; Organ, M. G. *Angew. Chem., Int. Ed.* 2012, 51, 7024–7027. (c) Achonduh, G. T.; Hadei, N.; Valente, C.; Avola, S.; O'Brien, C. J.; Organ, M. G. *Chem. Commun.* 2010, 46, 4109–4111. (d) Böck, K.; Feil, J. E.; Karahiosoff, K.; Koszinowski, K. *Chem. - Eur. J.* 2015, 21, 5548–5560. (e) Fagnou, K.; Lautens, M. *Angew. Chem., Int. Ed.* 2002, 41, 26–47. (f) Amatore, C.; Jutand, A. *Acc. Chem. Res.* 2000, 33, 314–321.
- (31) (a) Sorenson, R. J. *J. Org. Chem.* 2000, 65, 7747–7749. (b) Chan, D. M. T. *Tetrahedron Lett.* 1996, 37, 9013–9016. (c) Banfi, A.; Bartoletti, M.; Bellora, E.; Bignotti, M.; Turconi, M. *Synthesis* 1994, 1994, 775–776.
- (32) (a) Ikegai, K.; Nagata, Y.; Mukaiyama, T. *Bull. Chem. Soc. Jpn.* 2006, 79, 761–767. (b) Ikegai, K.; Mukaiyama, T. *Chem. Lett.* 2005, 34, 1496–1497.
- (33) (a) Harada, S.; Hayashi, D.; Sato, I.; Hiram, M. *Synlett* 2012, 23, 405–408. (b) Ikegai, K.; Fukumoto, K.; Mukaiyama, T. *Chem. Lett.* 2006, 35, 612–613. (c) Mukaiyama, T.; Sakurai, N.; Ikegai, K. *Chem. Lett.* 2006, 35, 1140–1141. (d) Coles, S. J.; Costello, J. F.; Hursthouse, M. B.; Smith, S. J. *Organomet. Chem.* 2002, 662, 98–104. (e) Sheppard, G. S. *Synlett* 1999, 1999, 1207–1210. (f) Dodonov, V. A.; Starostina, T. I.; Kuznetsova, Y. L.; Gushchin, A. V. *Russ. Chem. Bull.* 1995, 44, 151–152. (g) Brunner, H.; Obermann, U.; Wimmer, P. *Organometallics* 1989, 8, 821–826. (h) David, S.; Thieffry, A. *J. Org. Chem.* 1983, 48, 441–447.
- (34) For an excellent review on aerobic copper-catalyzed organic reactions, see: Allen, S. E.; Walvoord, R. R.; Padilla-Salinas, R.; Kozlowski, M. C. *Chem. Rev.* 2013, 113, 6234–6458.
- (35) Evans, D. A.; Katz, J. L.; West, T. R. *Tetrahedron Lett.* 1998, 39, 2937–2940.
- (36) Sperotto, E.; van Klink, G. P. M.; van Koten, G.; de Vries, J. G. *Dalton Trans.* 2010, 39, 10338–10351.
- (37) (a) King, A. E.; Brunold, T. C.; Stahl, S. S. *J. Am. Chem. Soc.* 2009, 131, 5044–5045. (b) McCann, S. D.; Stahl, S. S. *Acc. Chem. Res.* 2015, 48, 1756–1766.
- (38) For an excellent review on palladium- and copper-catalyzed C–N cross-coupling reactions, see: Beletskaya, I. P.; Cheprakov, A. V. *Organometallics* 2012, 31, 7753–7808.
- (39) Robertson, A. P. M.; Burford, N.; McDonald, R.; Ferguson, M. J. *Angew. Chem., Int. Ed.* 2014, 53, 3480–3483.
- (40) Urgin, K.; Aubé, C.; Pichon, C.; Pipelier, M.; Blot, V.; Thobie-Gautier, C.; Léonel, E.; Dubreuil, D.; Condon, S. *Tetrahedron Lett.* 2012, 53, 1894–1896.
- (41) Wong, M. Y.; Xie, G.; Tourbillon, C.; Sandroni, M.; Cordes, D. B.; Slawin, A. M. Z.; Samuel, I. D. W.; Zysman-Colman, E. *Dalton Trans.* 2015, 44, 8419–8432.
- (42) Leowanawat, P.; Zhang, N.; Safi, M.; Hoffman, D. J.; Fryberger, M. C.; George, A.; Percec, V. *J. Org. Chem.* 2012, 77, 2885–2892.
- (43) Zhu, L.; Duquette, J.; Zhang, M. *J. Org. Chem.* 2003, 68, 3729–3732.
- (44) Lai, M.-J.; Huang, H.-L.; Pan, S.-L.; Liu, Y.-M.; Peng, C.-Y.; Lee, H.-Y.; Yeh, T.-K.; Huang, P.-H.; Teng, C.-M.; Chen, C.-S.; Chuang, H.-Y.; Liou, J.-P. *J. Med. Chem.* 2012, 55, 3777–3791.
- (45) Zhu, Y.; Yan, H.; Lu, L.; Liu, D.; Rong, G.; Mao, J. *J. Org. Chem.* 2013, 78, 9898–9905.

## Supporting Information

### Synthesis of Highly Functionalized Triarylbiomuthines by Functional Group Manipulation and Use in Palladium- and Copper-Catalyzed Arylation Reactions

Martin Hébert,<sup>a</sup> Pauline Petiot,<sup>a</sup> Emeline Benoit,<sup>a</sup> Julien Dansereau,<sup>a</sup>  
Tabinda Ahmad,<sup>a</sup> Adrien Le Roch,<sup>a</sup> Xavier Ottenwaelder<sup>b</sup> and Alexandre Gagnon<sup>a\*</sup>

<sup>a</sup> Université du Québec à Montréal, Département de chimie, C.P. 8888, Succursale Centre-Ville,  
Montréal, Québec, Canada, H3C 3P8. E-mail: gagnon.alexandre@uqam.ca

<sup>b</sup> Concordia University, Department of Chemistry and Biochemistry, 7141 Sherbrooke St. West,  
Montréal, Québec, Canada, H4B 1R6

### Table of Contents

#### Spectral data:

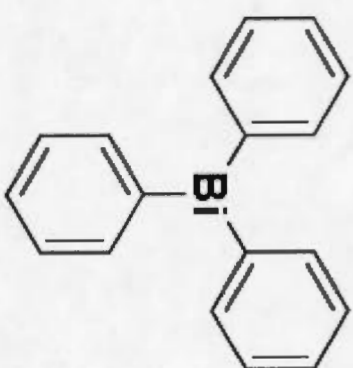
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>1a</b> .....	S1
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>1n</b> .....	S4
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>3a</b> .....	S7
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>3b</b> .....	S10
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>5a</b> .....	S13
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>7a</b> .....	S16
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>9a</b> .....	S19
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>9b</b> .....	S22
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>17a</b> .....	S25
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>21c</b> .....	S28
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>21e</b> .....	S31
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>21g</b> .....	S34
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>21h</b> .....	S37
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>27</b> .....	S40
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>30a</b> .....	S43
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>30b</b> .....	S46



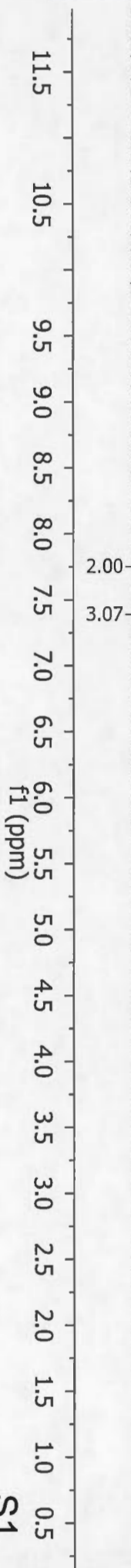
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>30c</b> .....	S49
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>30d</b> .....	S52
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>31a</b> .....	S55
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>34a</b> .....	S58
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>34b</b> .....	S61
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>34c</b> .....	S64
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>34d</b> .....	S67
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>34e</b> .....	S70
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>35a</b> .....	S73
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>35b</b> .....	S76
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>37a</b> .....	S79
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>37b</b> .....	S82
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>37c</b> .....	S85
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>37d</b> .....	S88
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>37e</b> .....	S91
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>41</b> .....	S94
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>42</b> .....	S97
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>43</b> .....	S100
<sup>1</sup> H, <sup>13</sup> C NMR and IR spectra of <b>44</b> .....	S103
<b>Crystallographic analysis:</b>	
Single-crystal X-ray diffraction data of <b>1f</b> .....	S106
Single-crystal X-ray diffraction data of <b>1i</b> .....	S107
Single-crystal X-ray diffraction data of <b>1k</b> .....	S108
Single-crystal X-ray diffraction data of <b>1n</b> .....	S109

Single-crystal X-ray diffraction data of <b>1s</b> .....	S110
Single-crystal X-ray diffraction data of <b>1t</b> .....	S111
Single-crystal X-ray diffraction data of <b>21c</b> .....	S112
Single-crystal X-ray diffraction data of <b>21e</b> .....	S113
Single-crystal X-ray diffraction data of <b>17a</b> .....	S114
<b>Table S1:</b> Crystal and solution data for <b>1f, 1i, 1k, 1n, 1s, 1t, 21c, 21e</b> and <b>17a</b> .....	S115
<b>Table S2:</b> Bi-C bond lengths and C-Bi-C angles in <b>1f, 1i, 1k, 1n, 1s, 1t, 21c,</b> and <b>21e</b> .....	S117

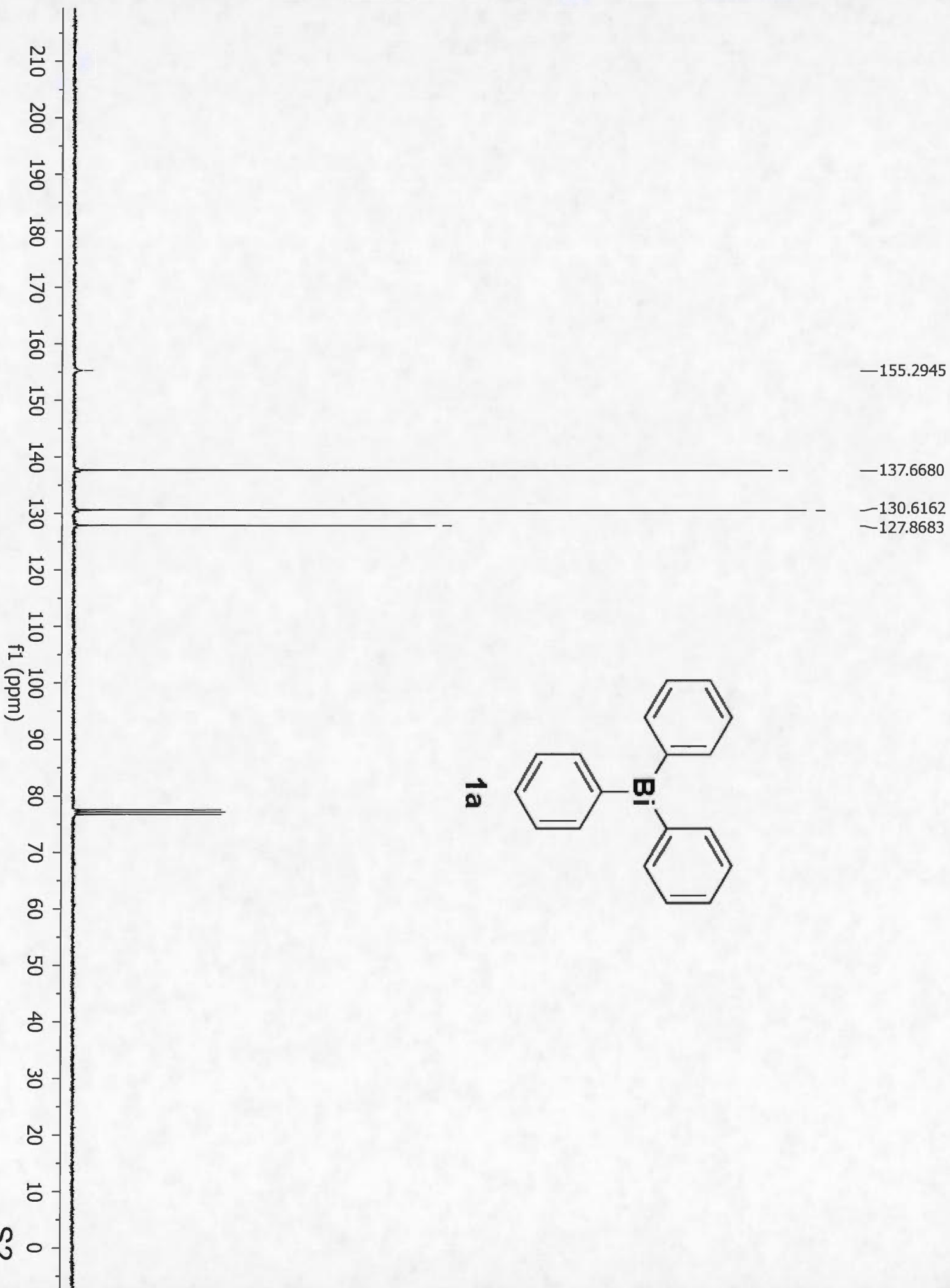
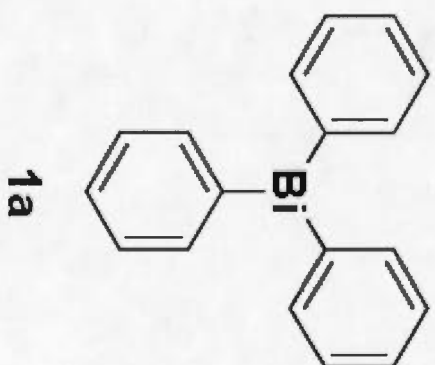
7.7630  
7.7380  
7.4161  
7.4104  
7.3900  
7.3668  
7.3466  
7.3413  
7.3203  
7.2961

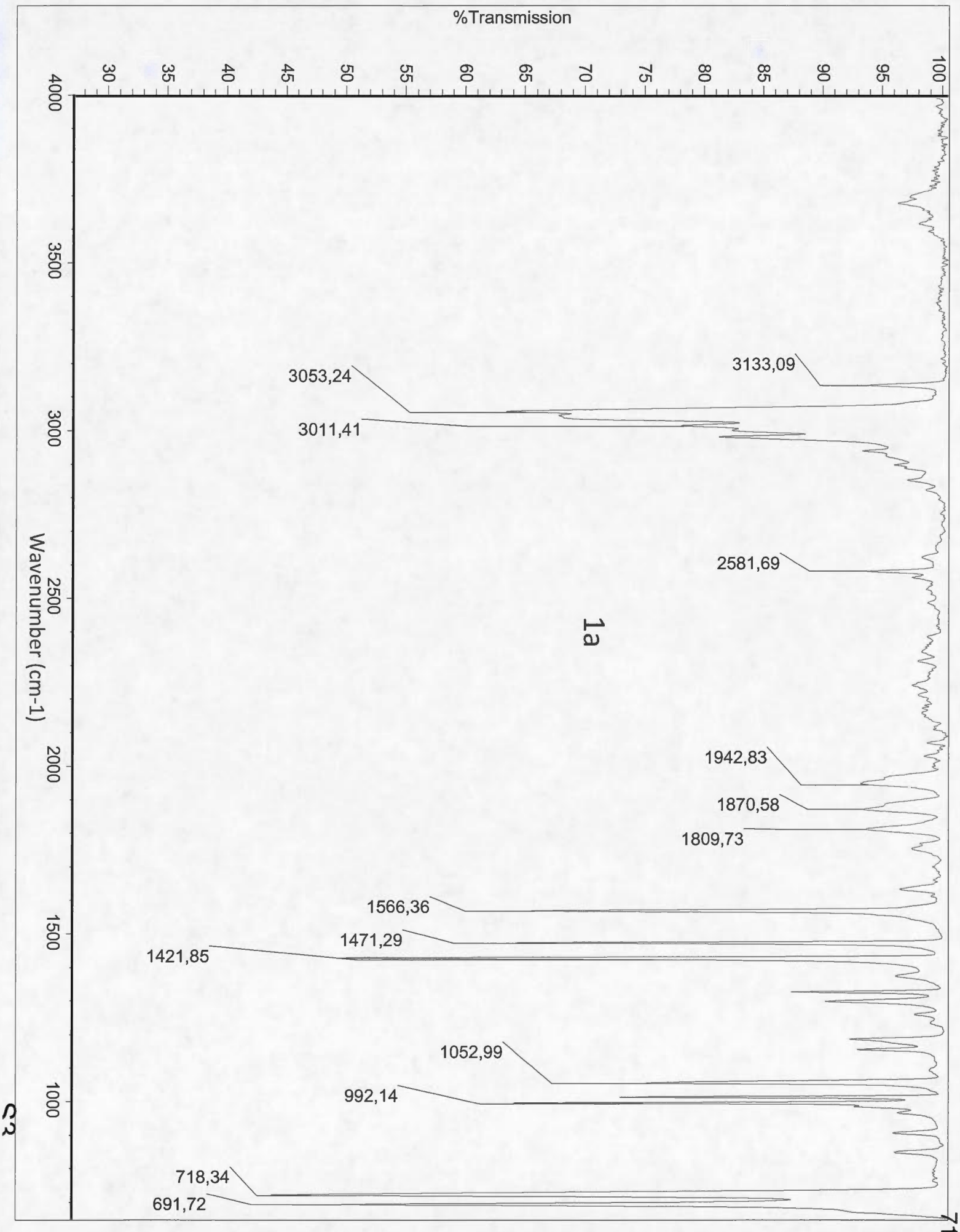


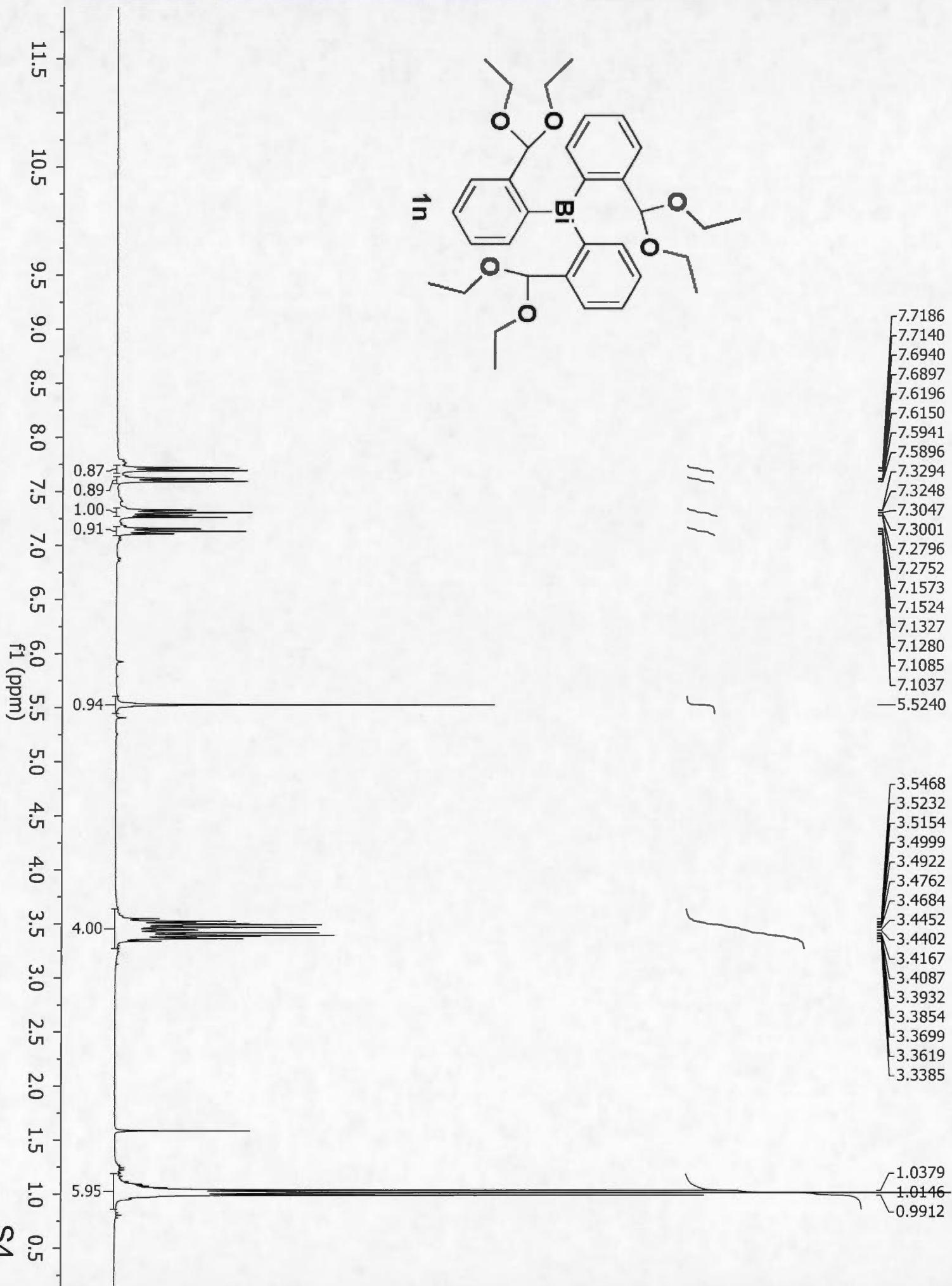
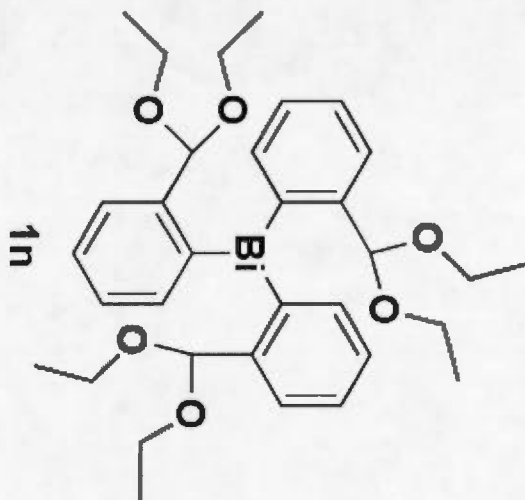
1a



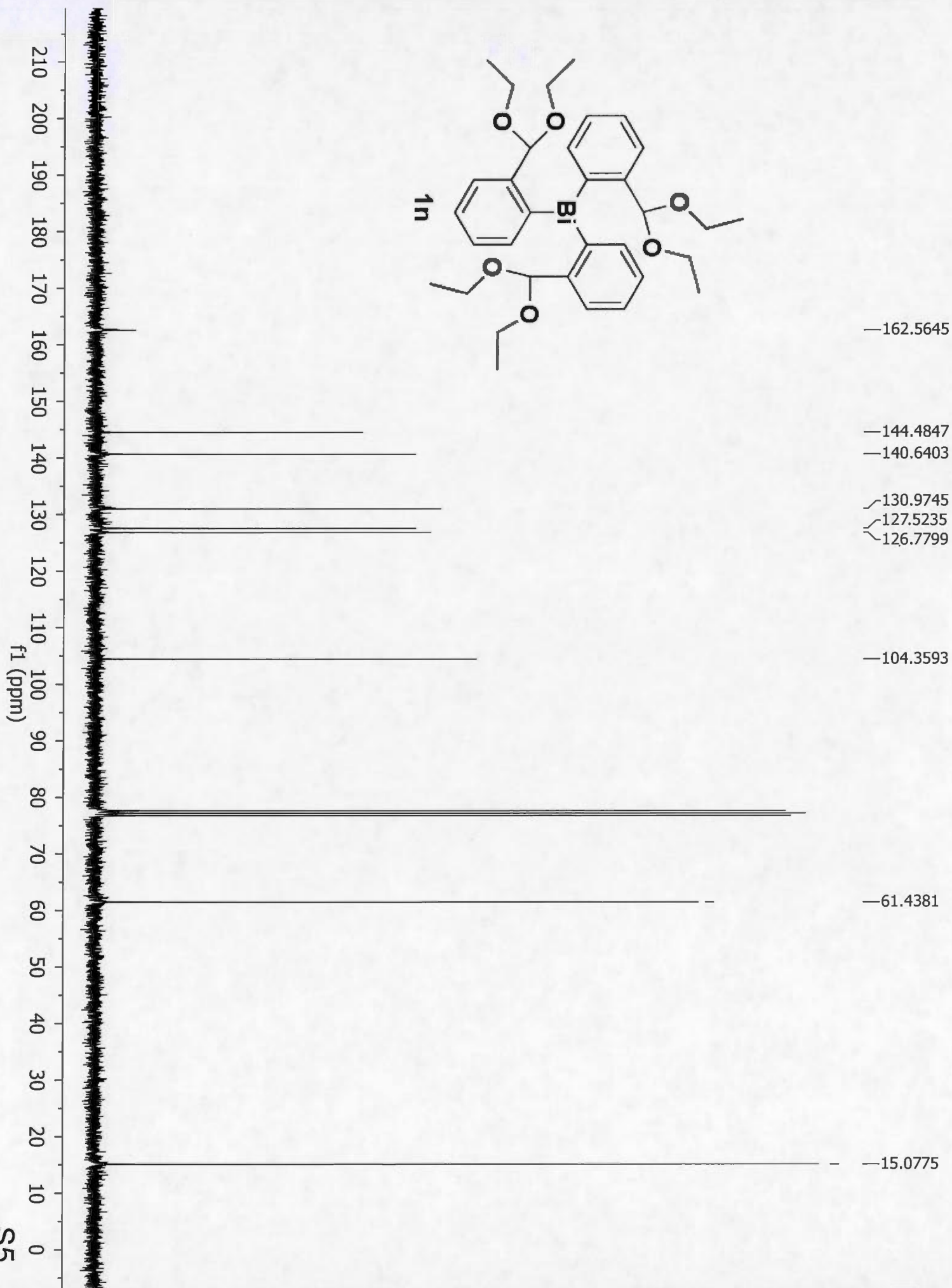
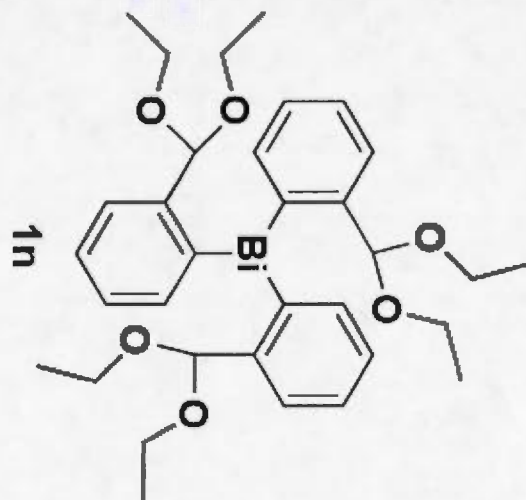


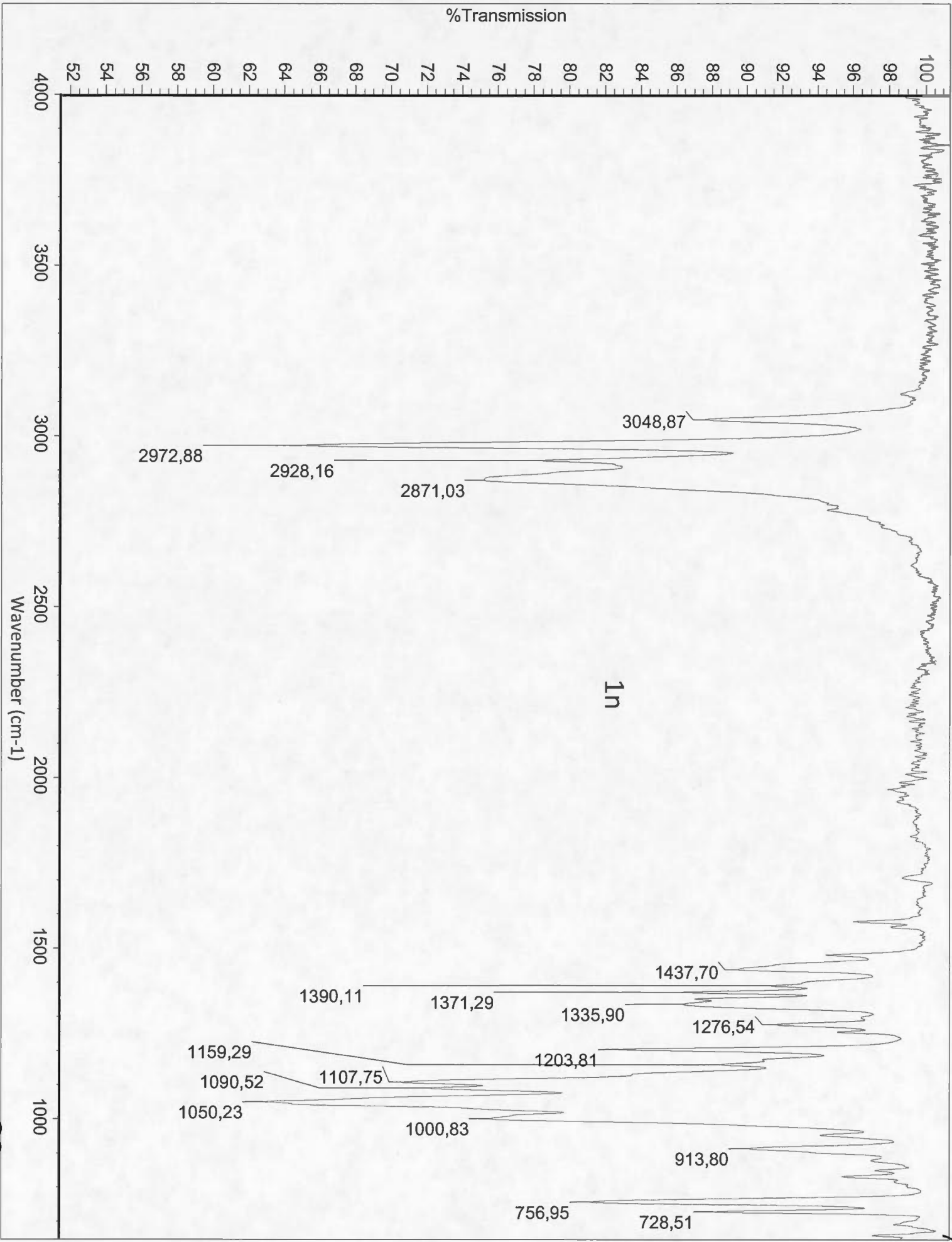


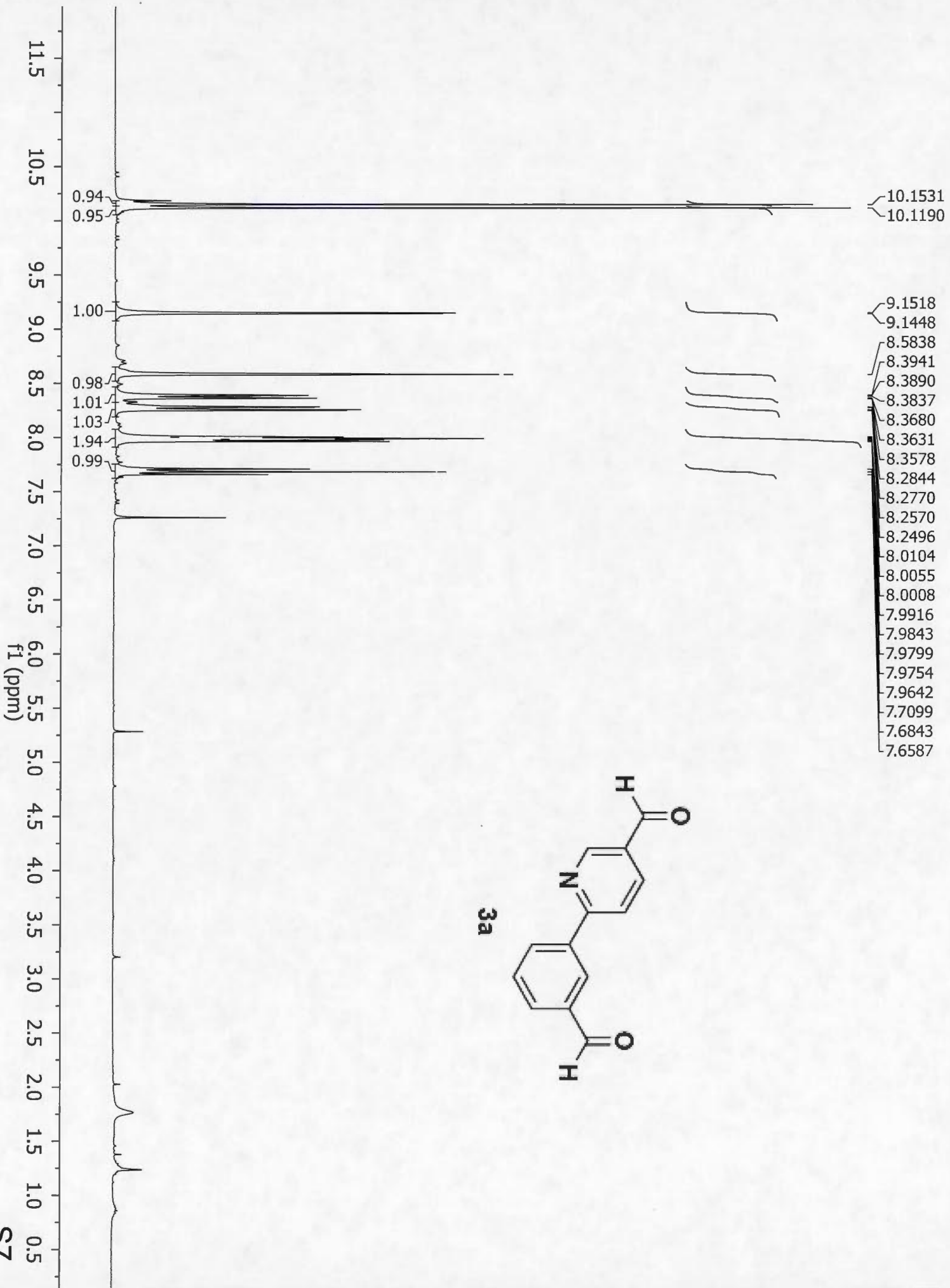




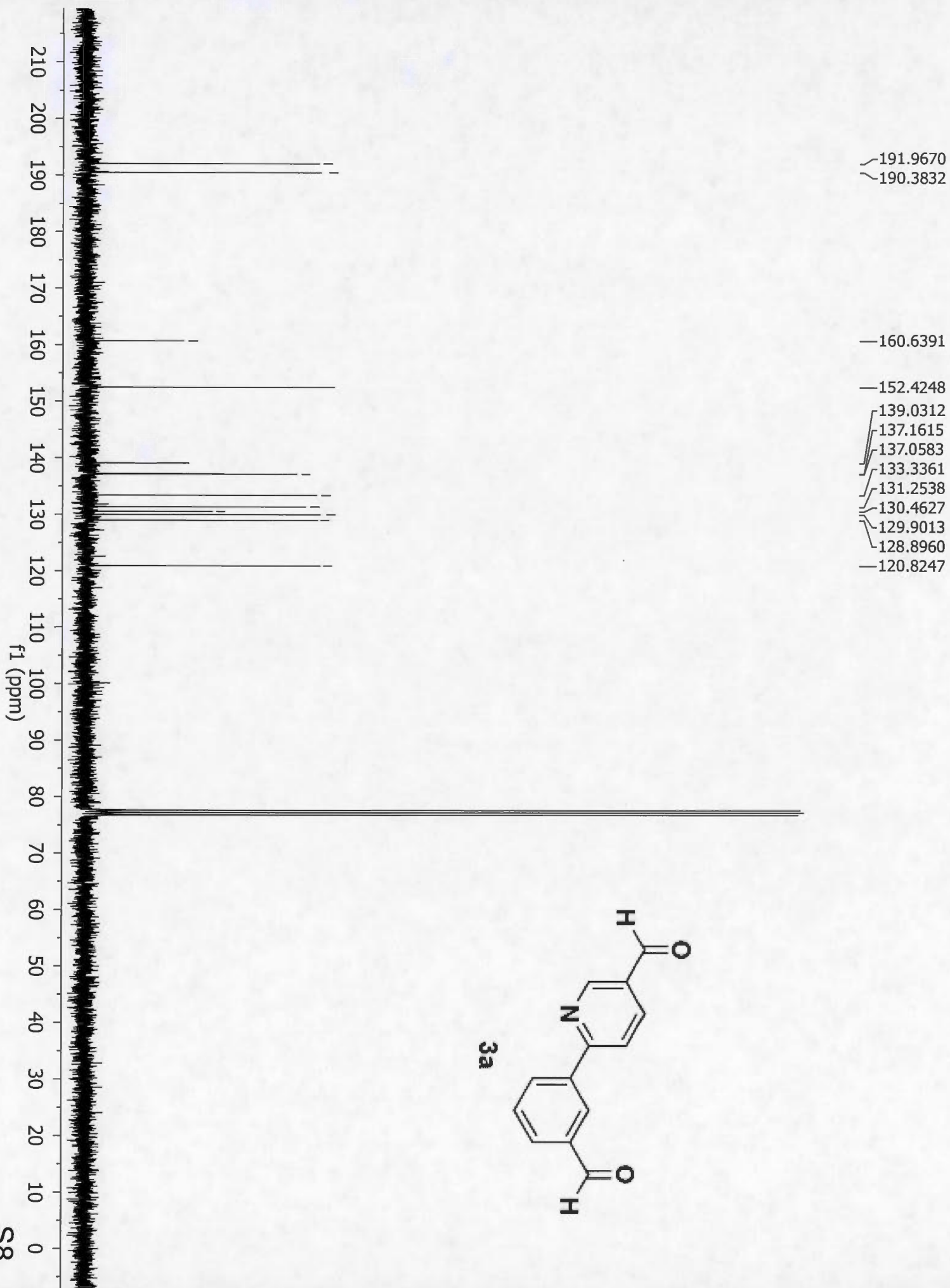
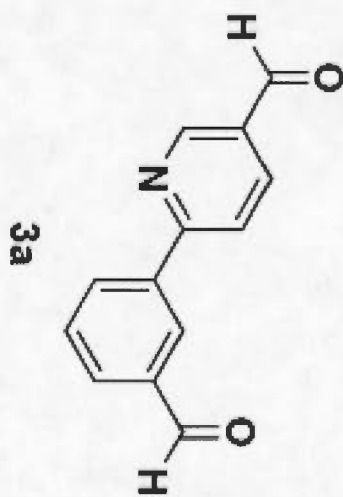


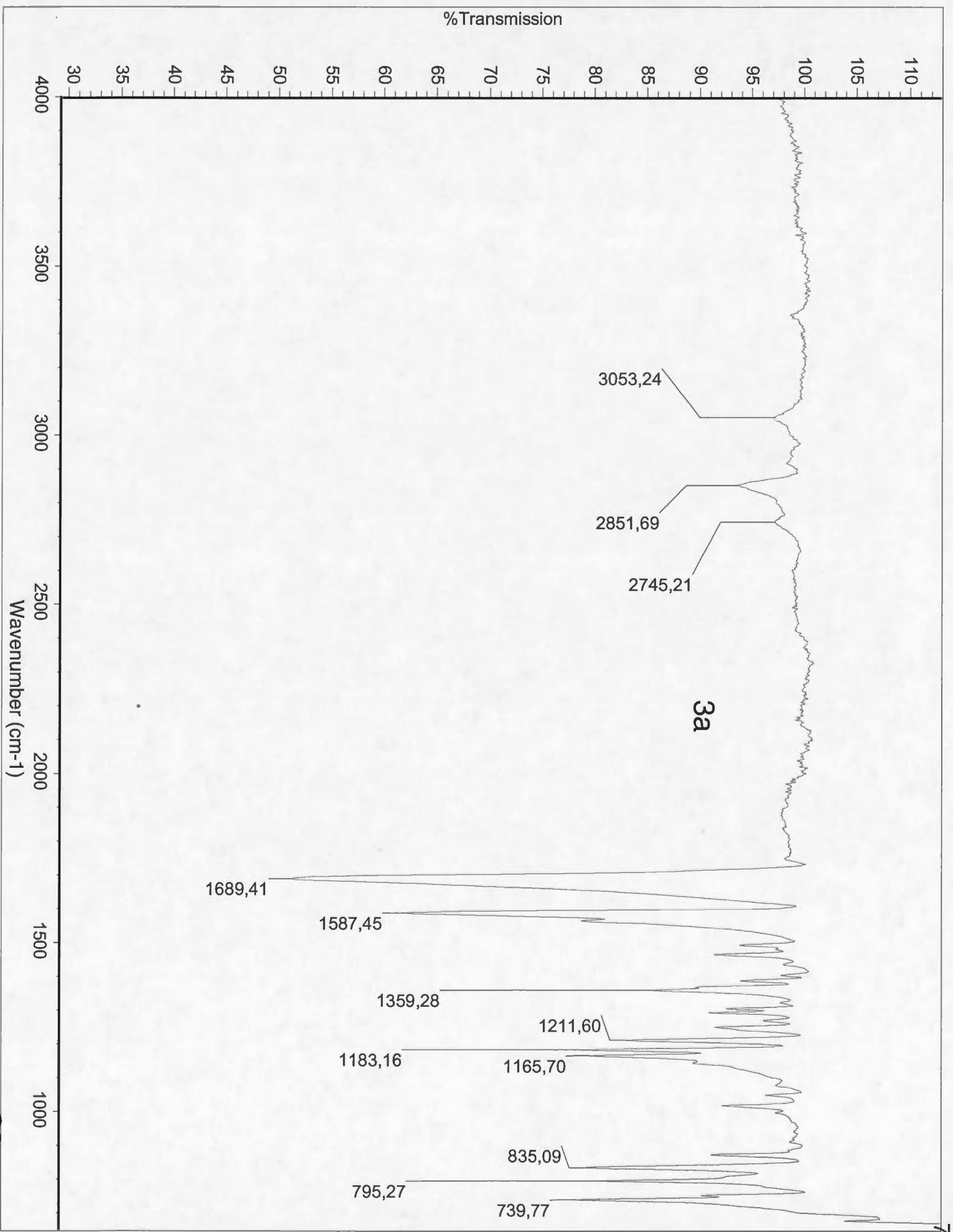




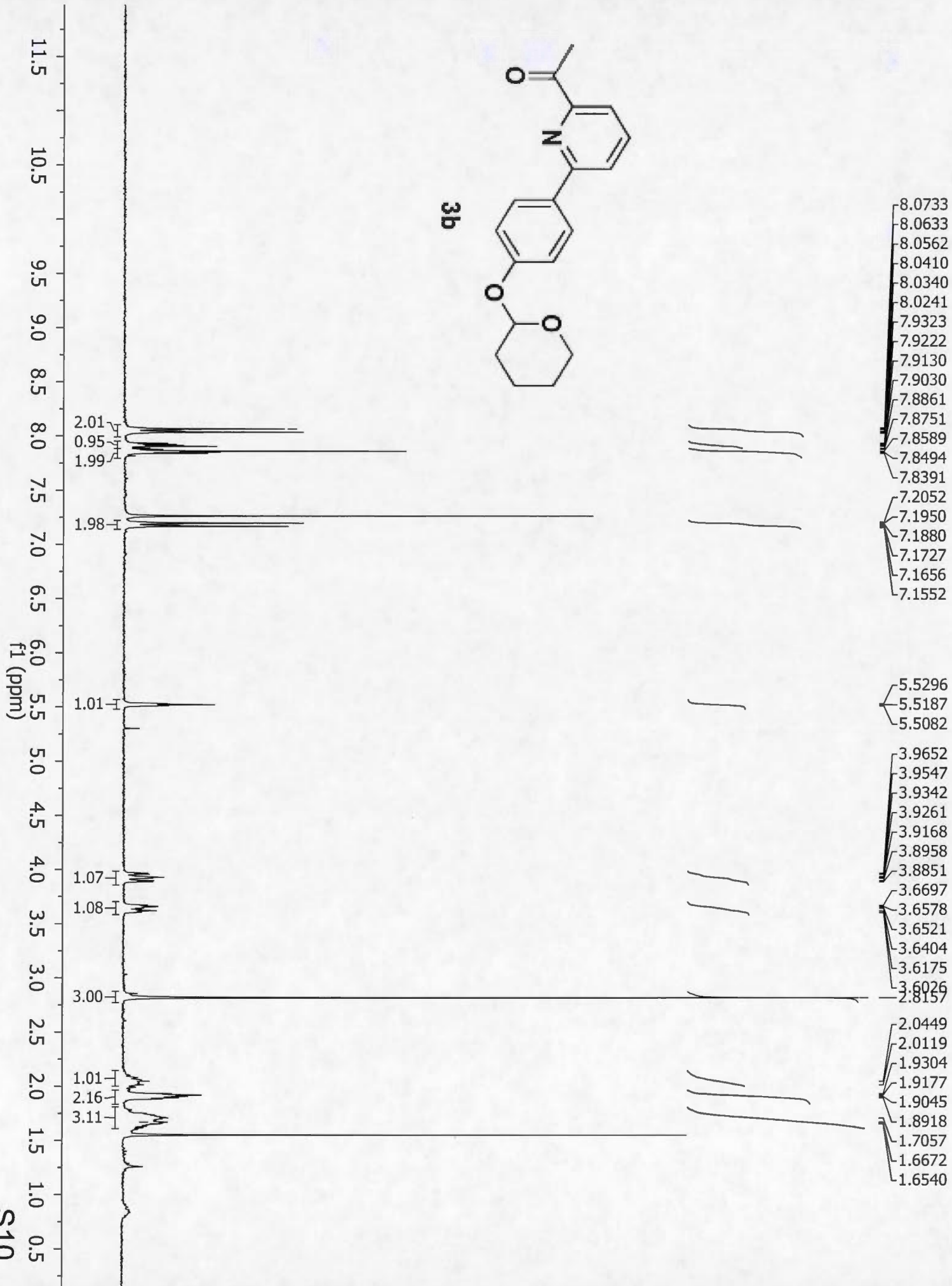
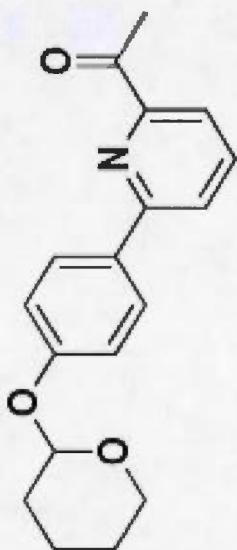






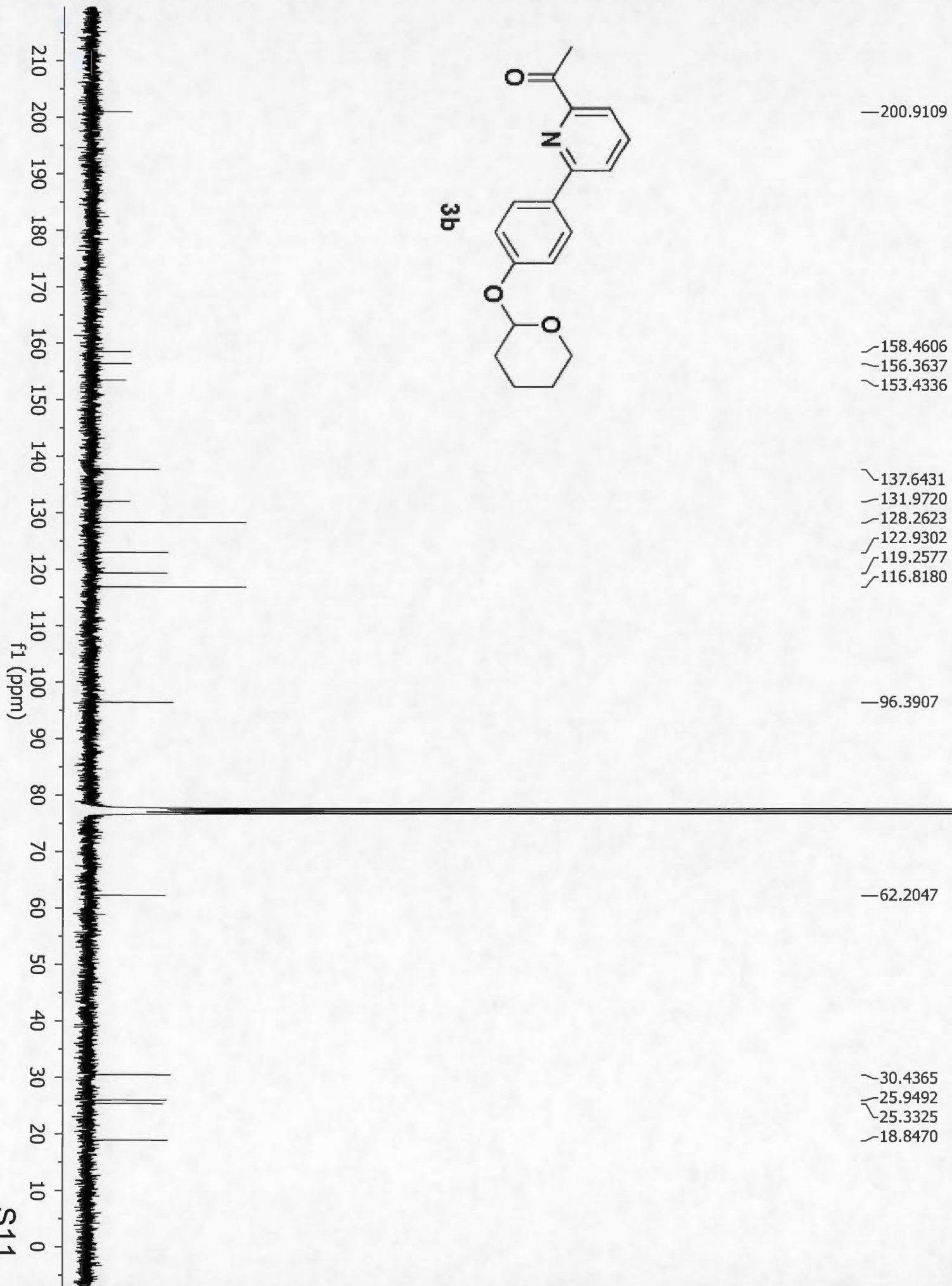
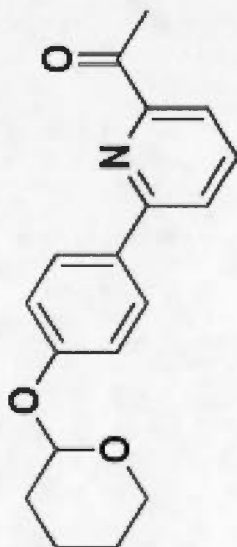


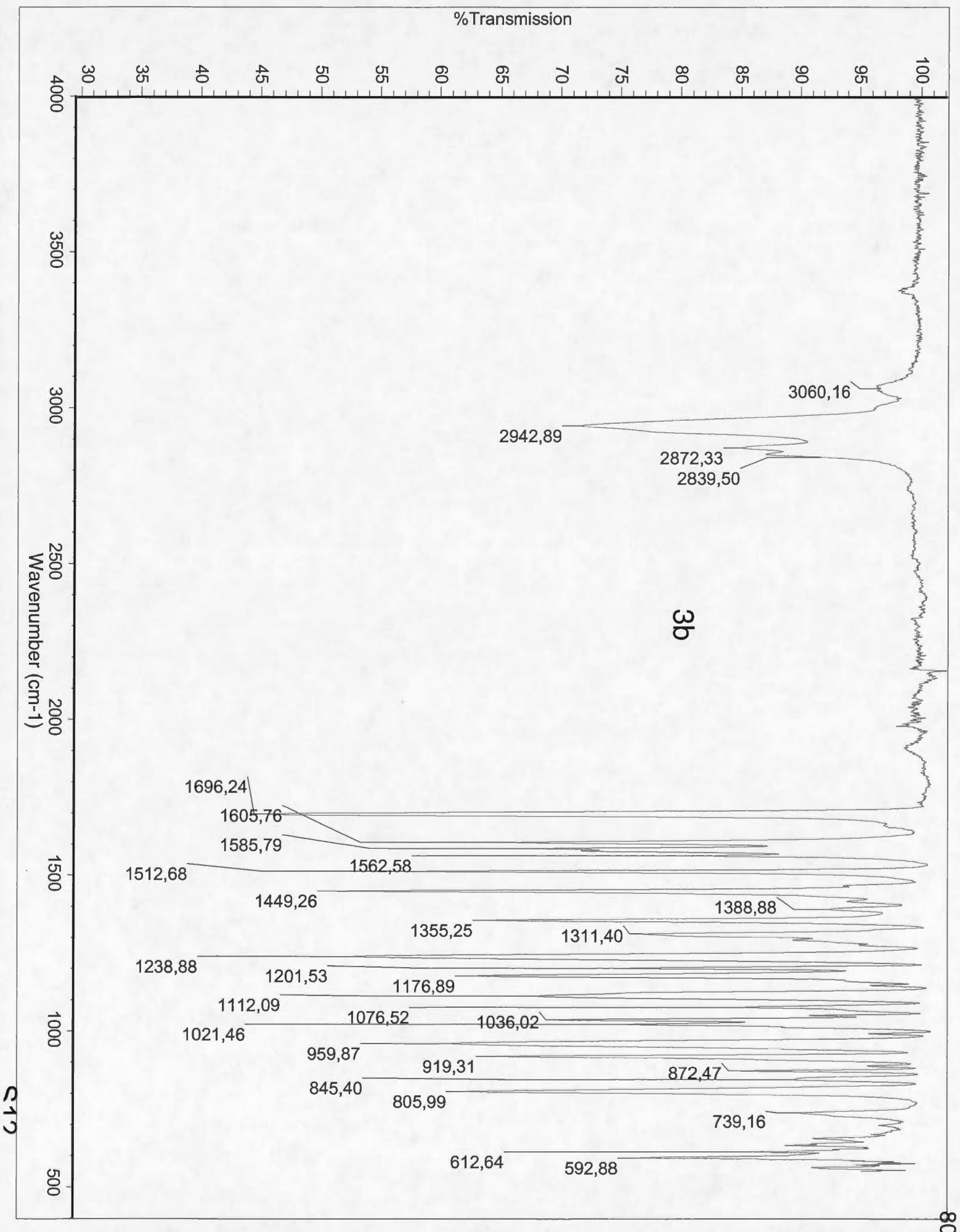
3b



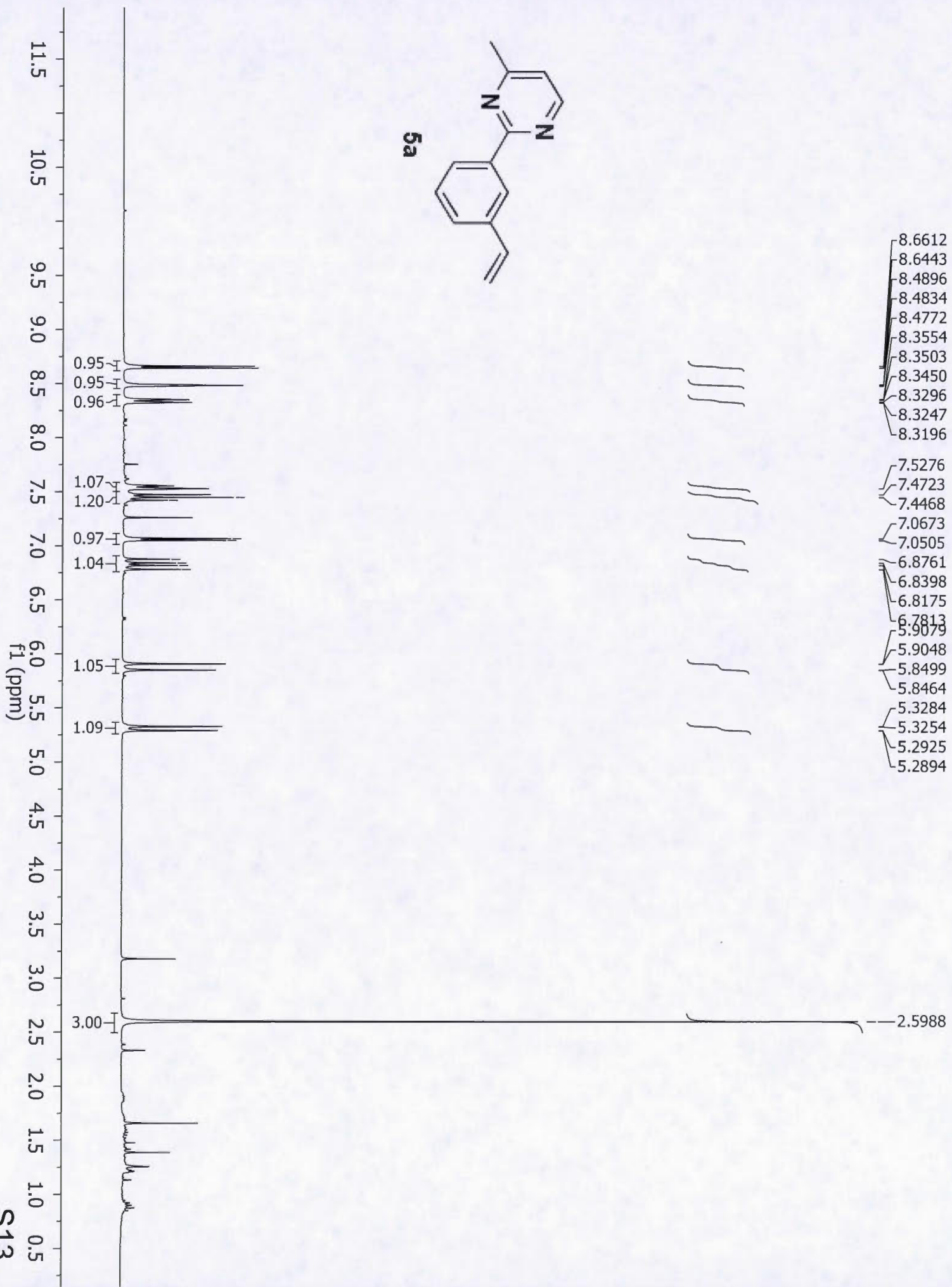
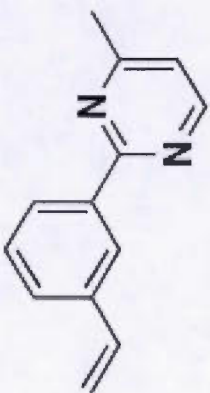


3b

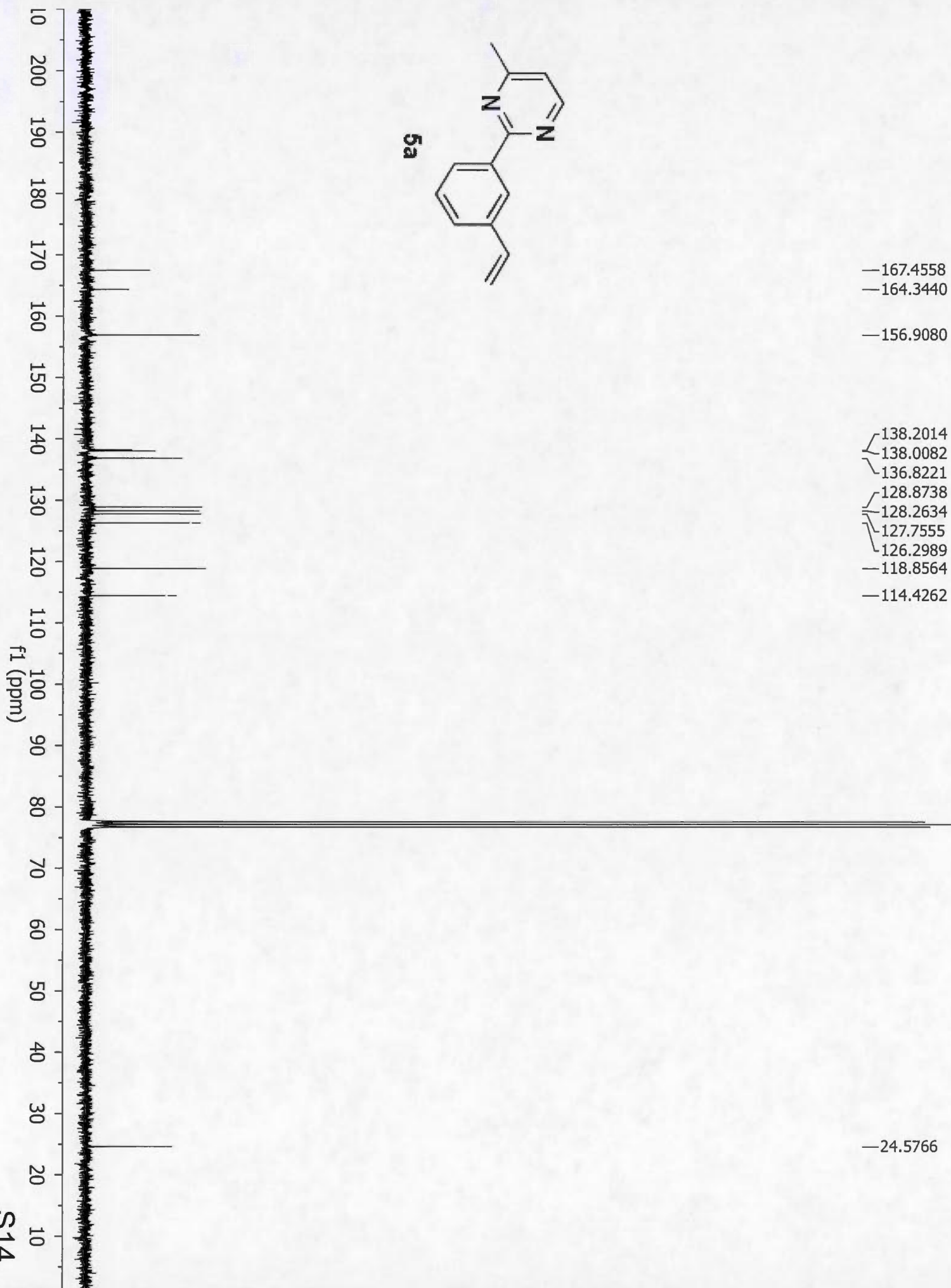
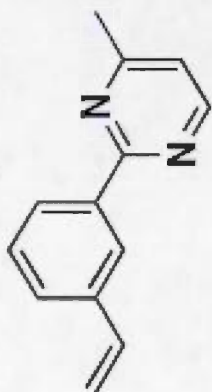




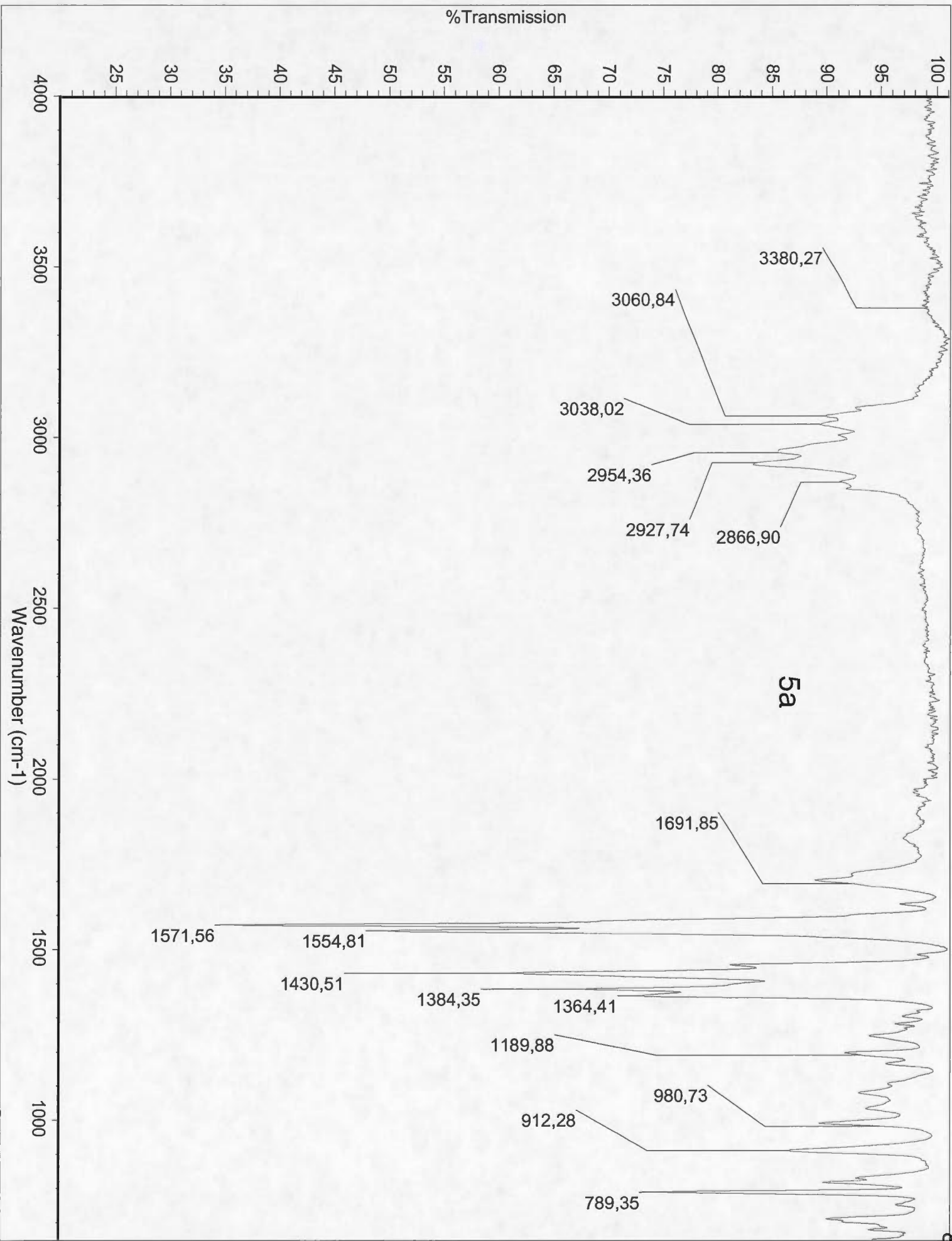
5a

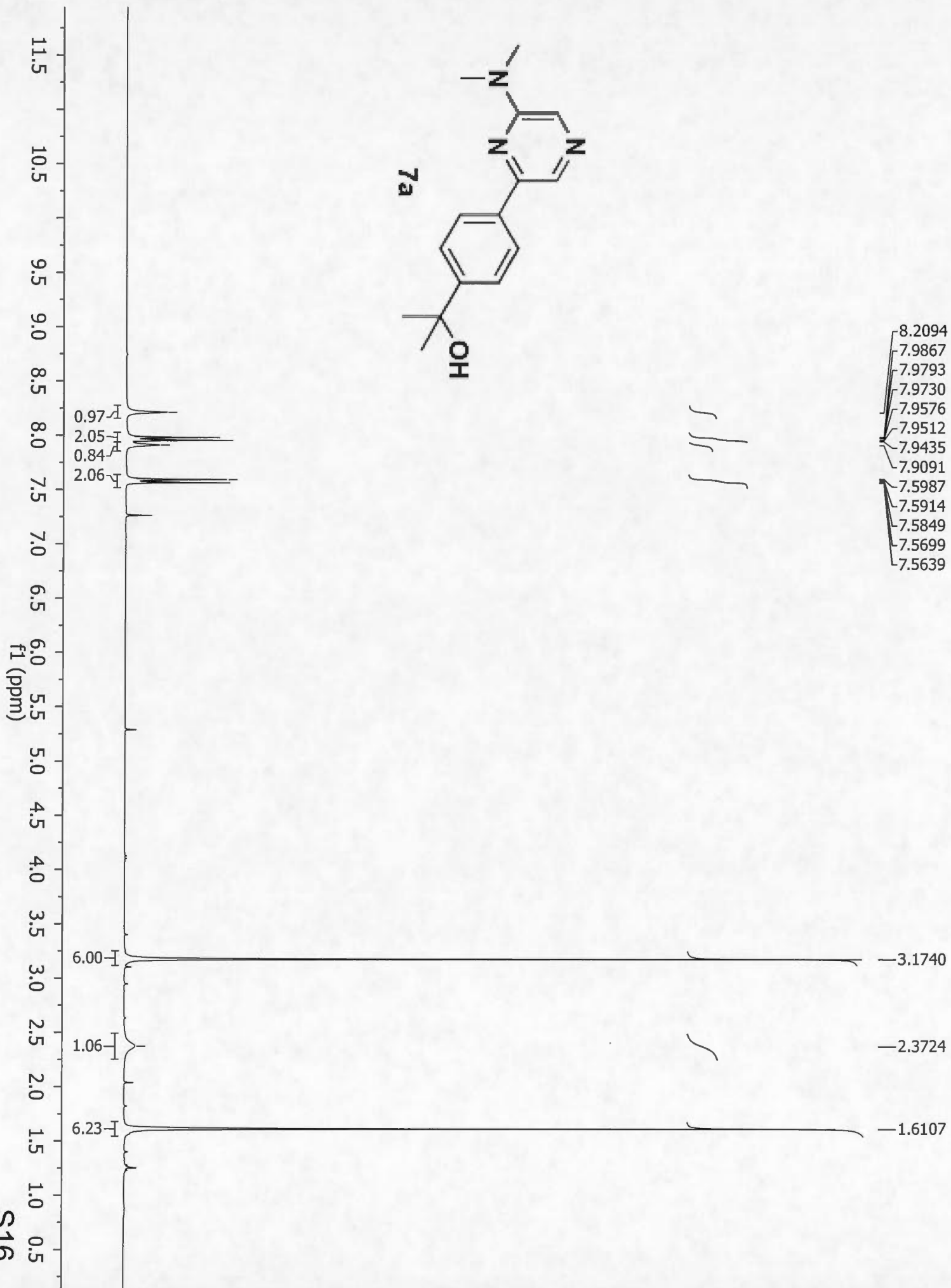
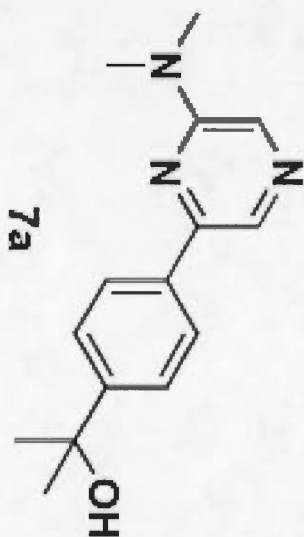


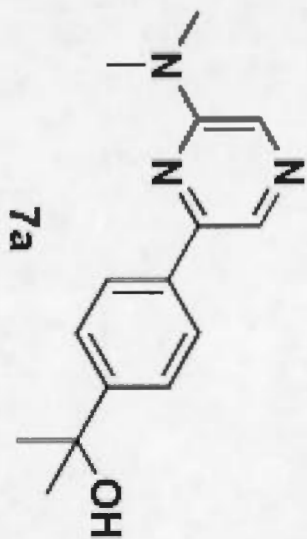
5a



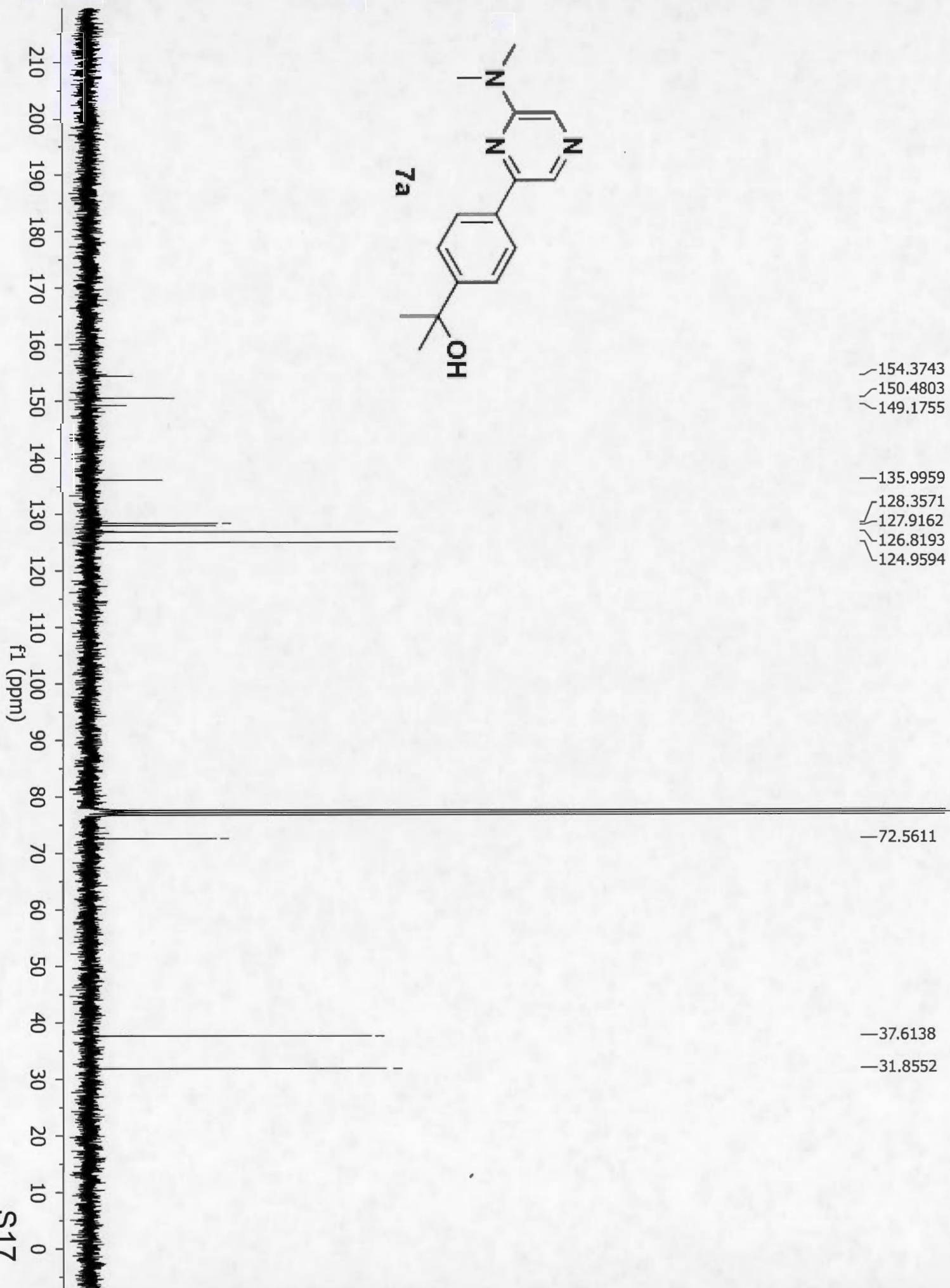


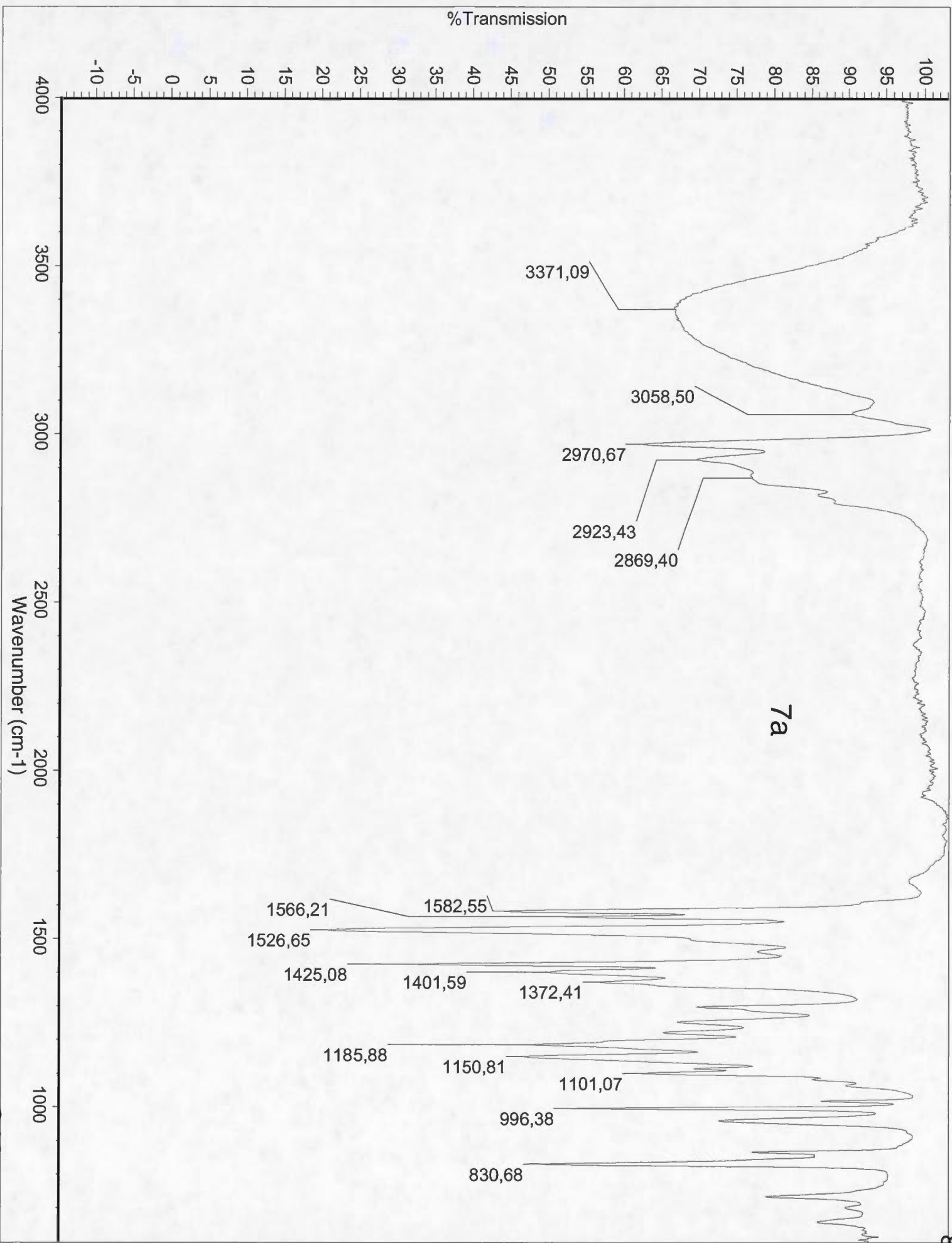






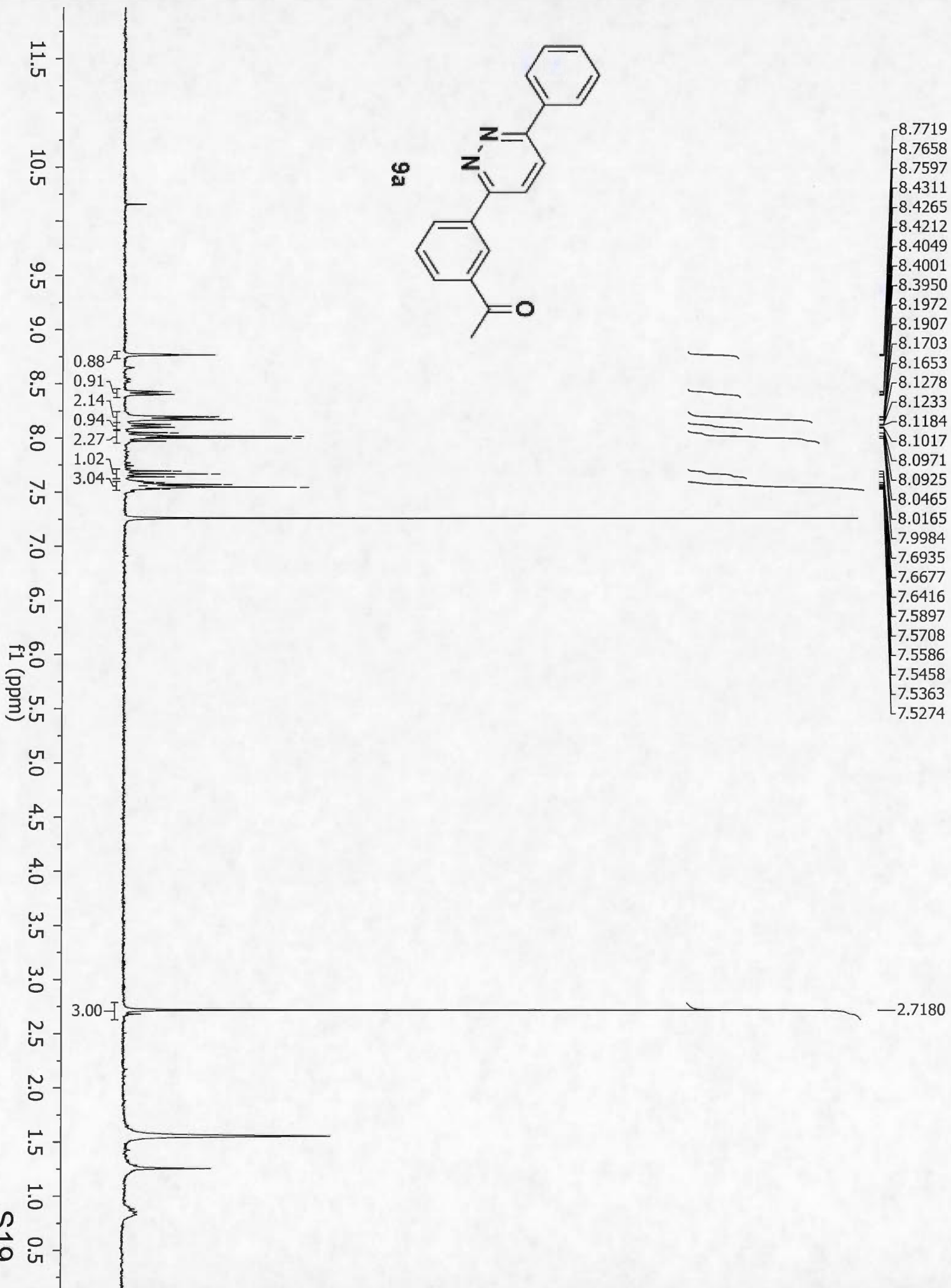
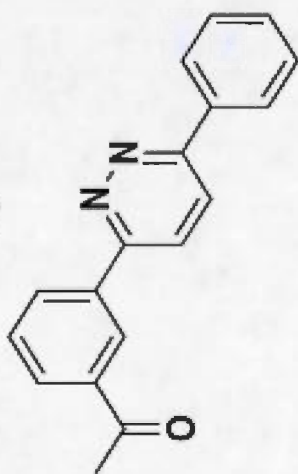
7a

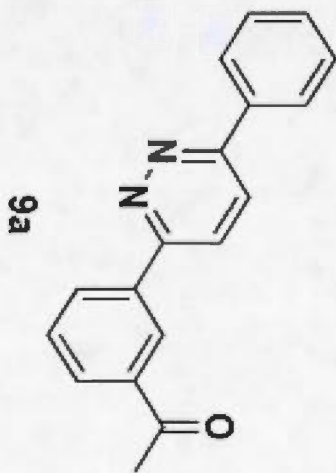




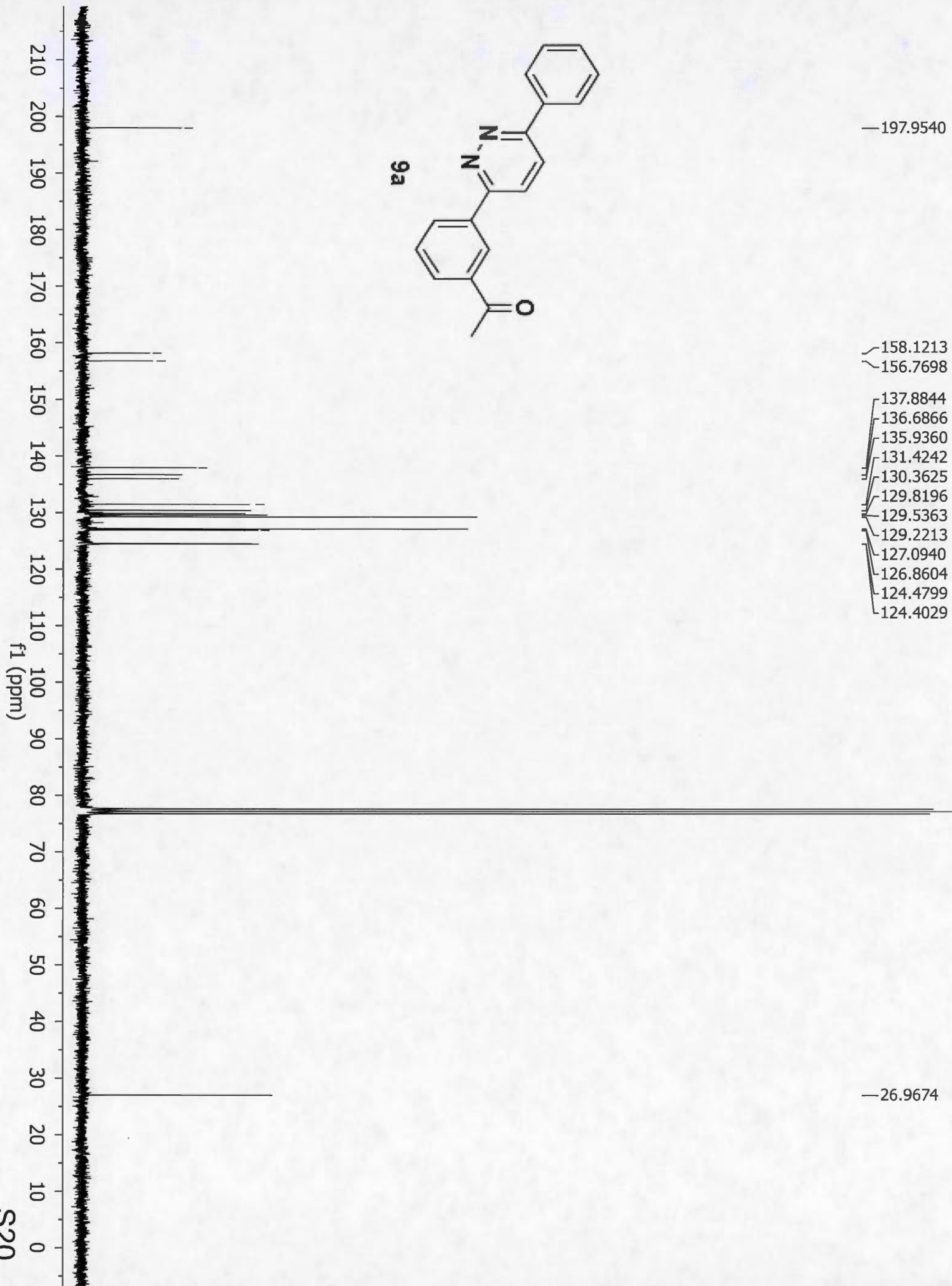


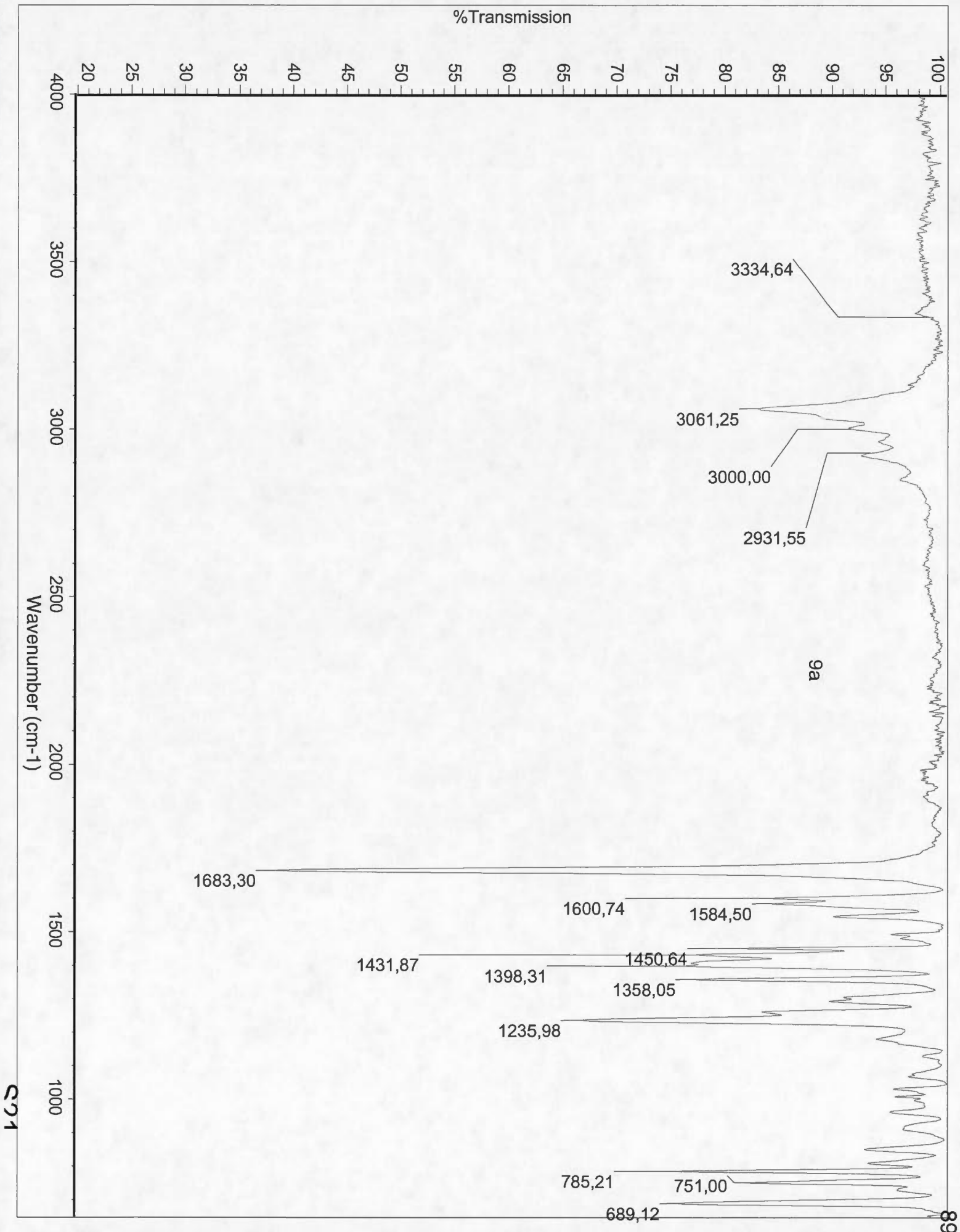
9a

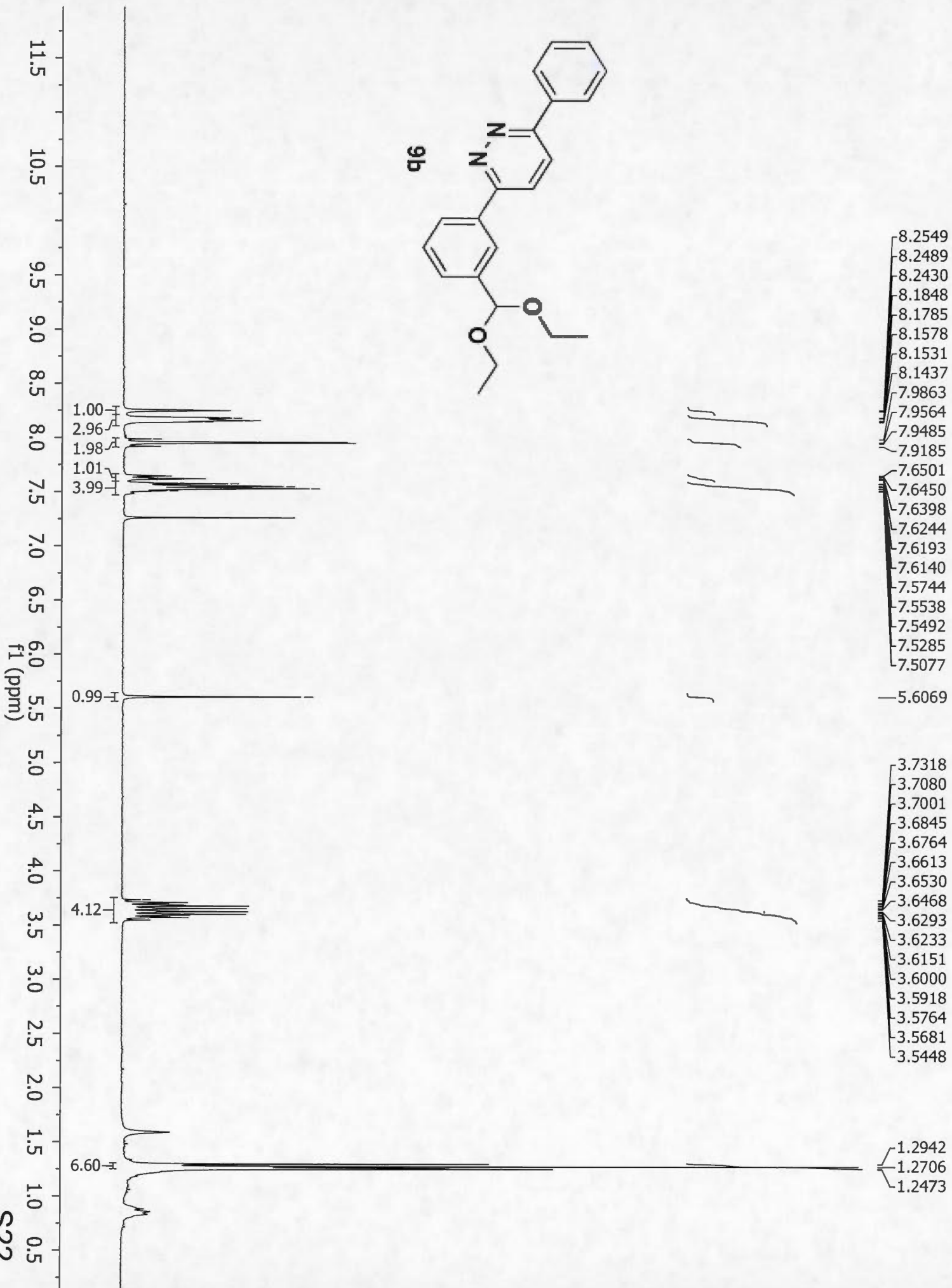
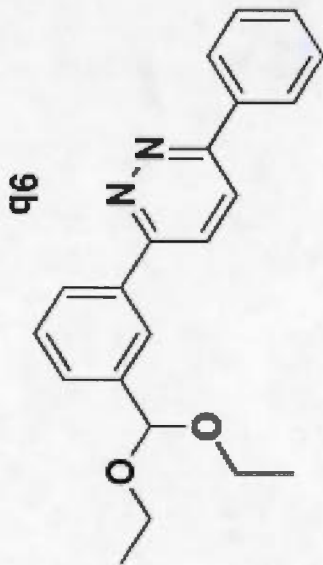




9a

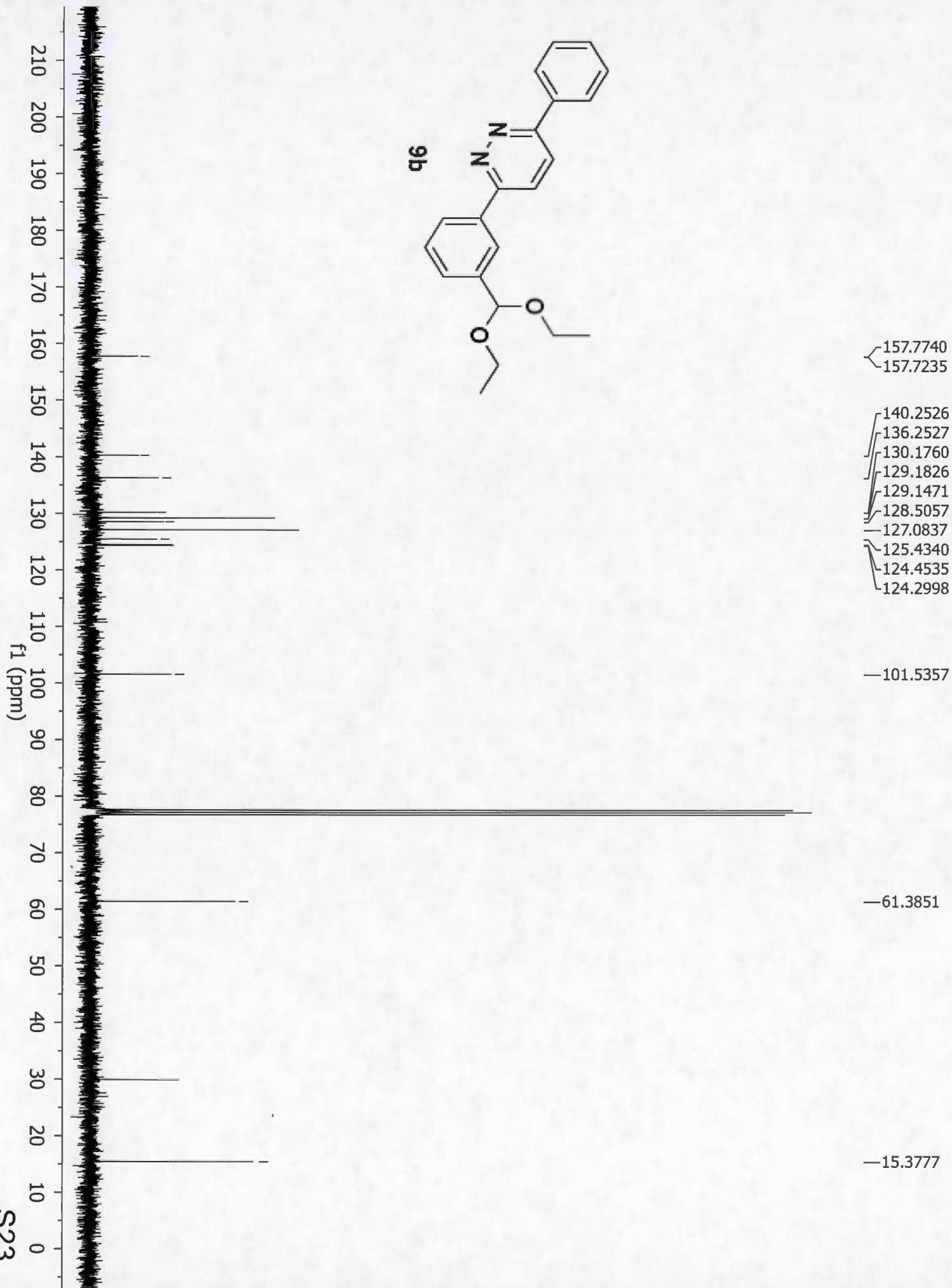
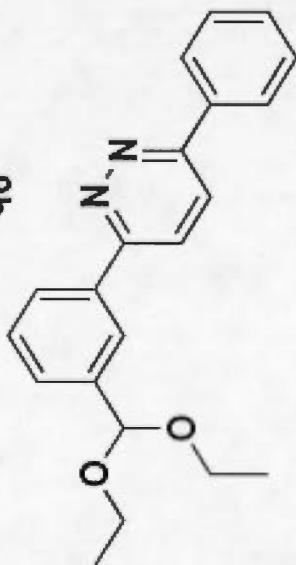


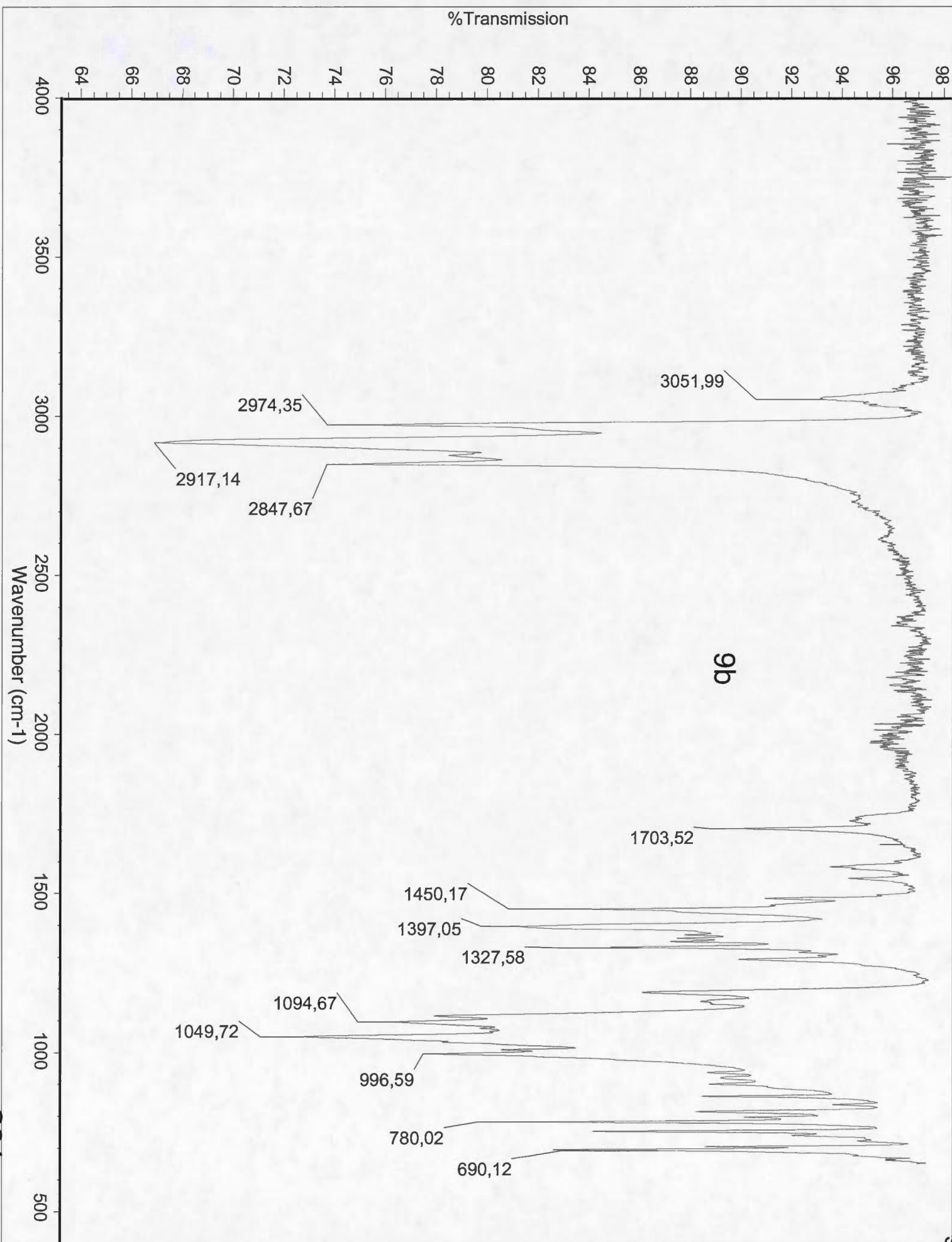




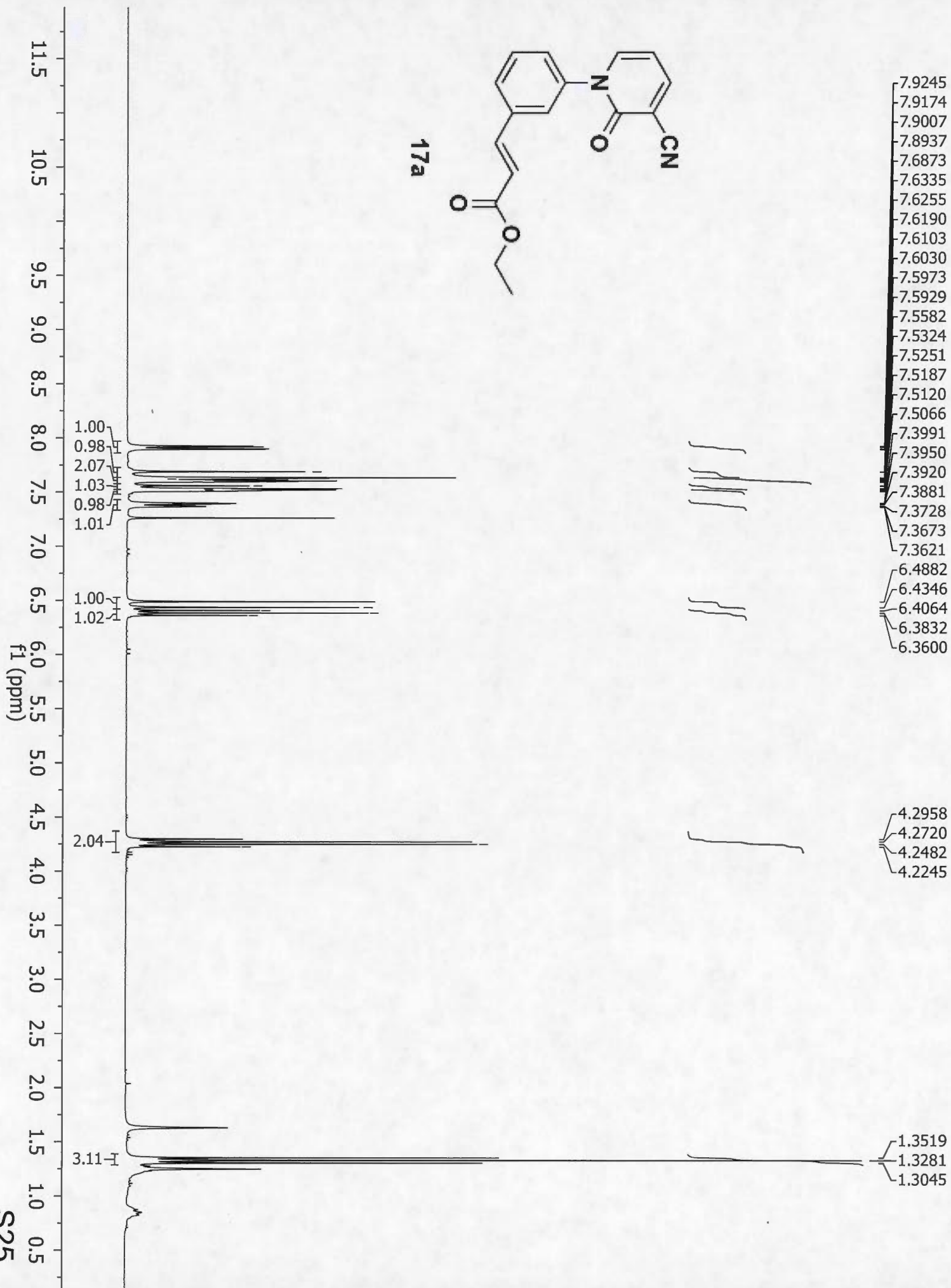
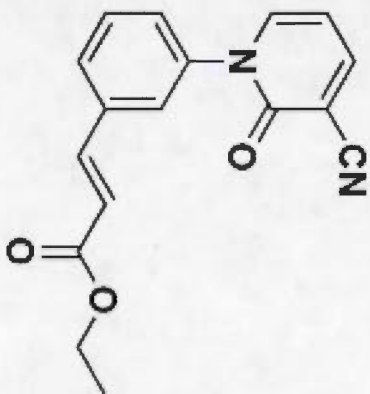


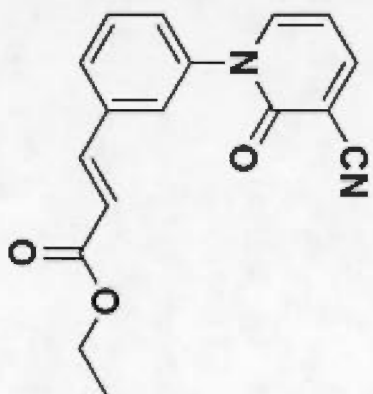
9b



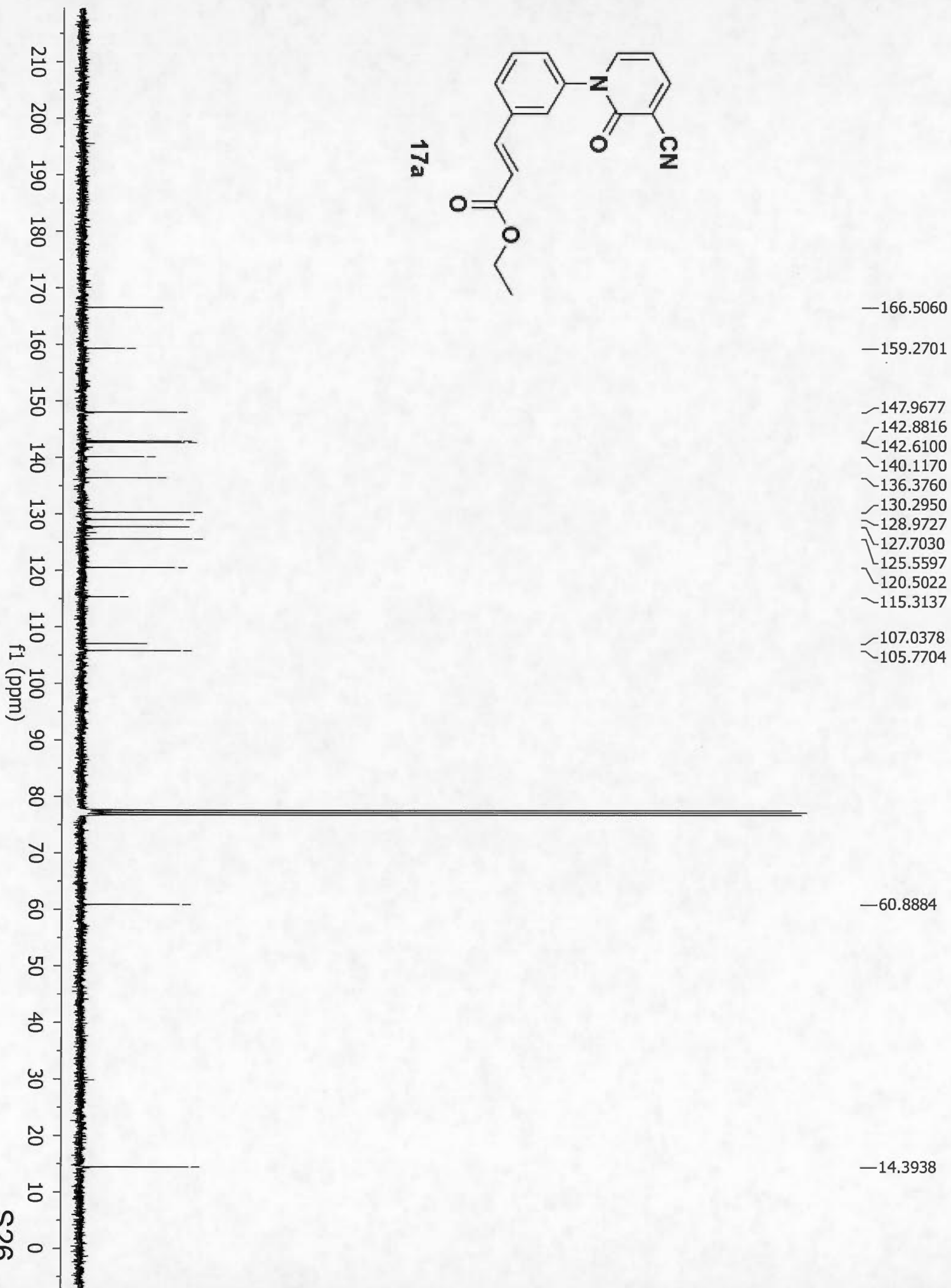


17a

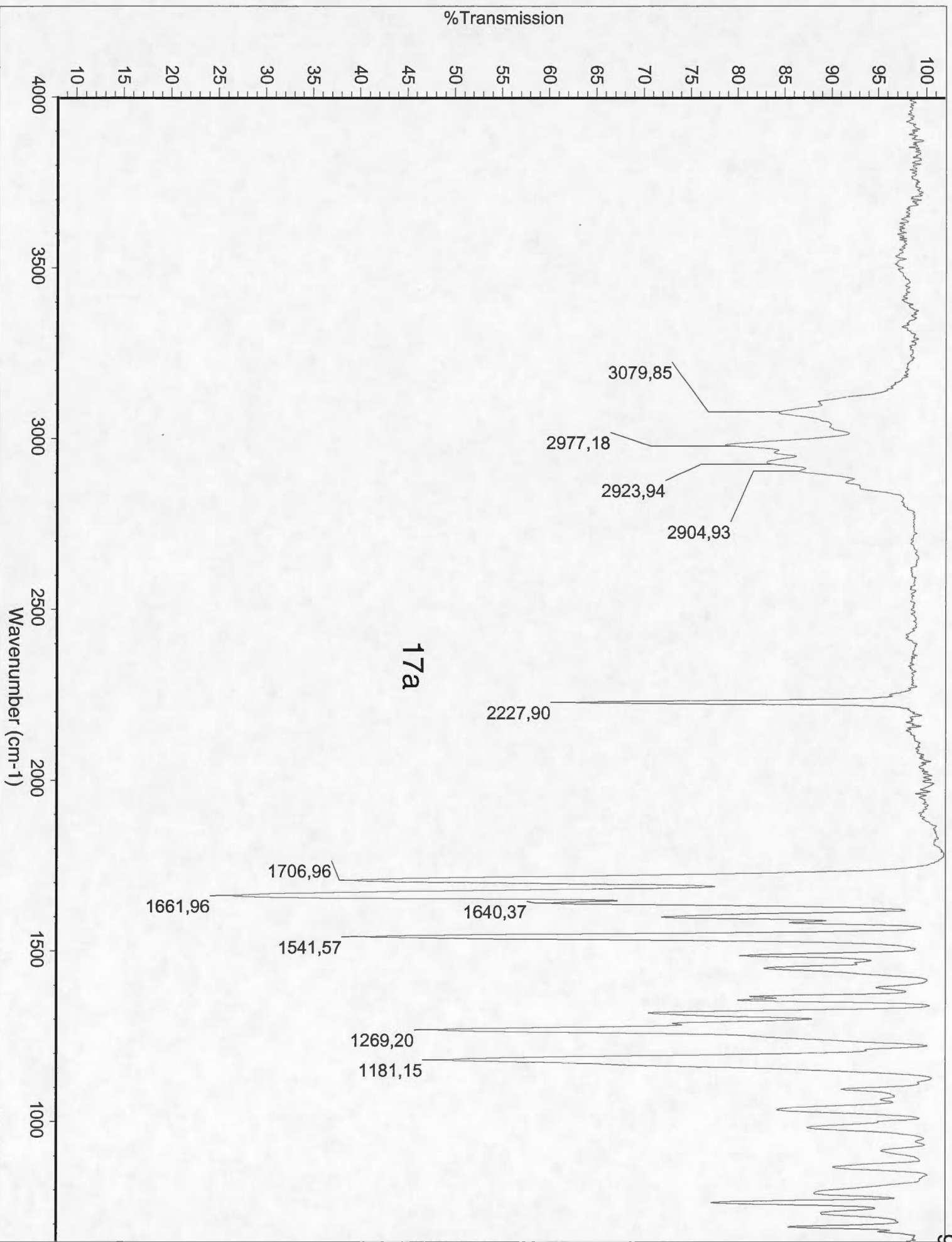


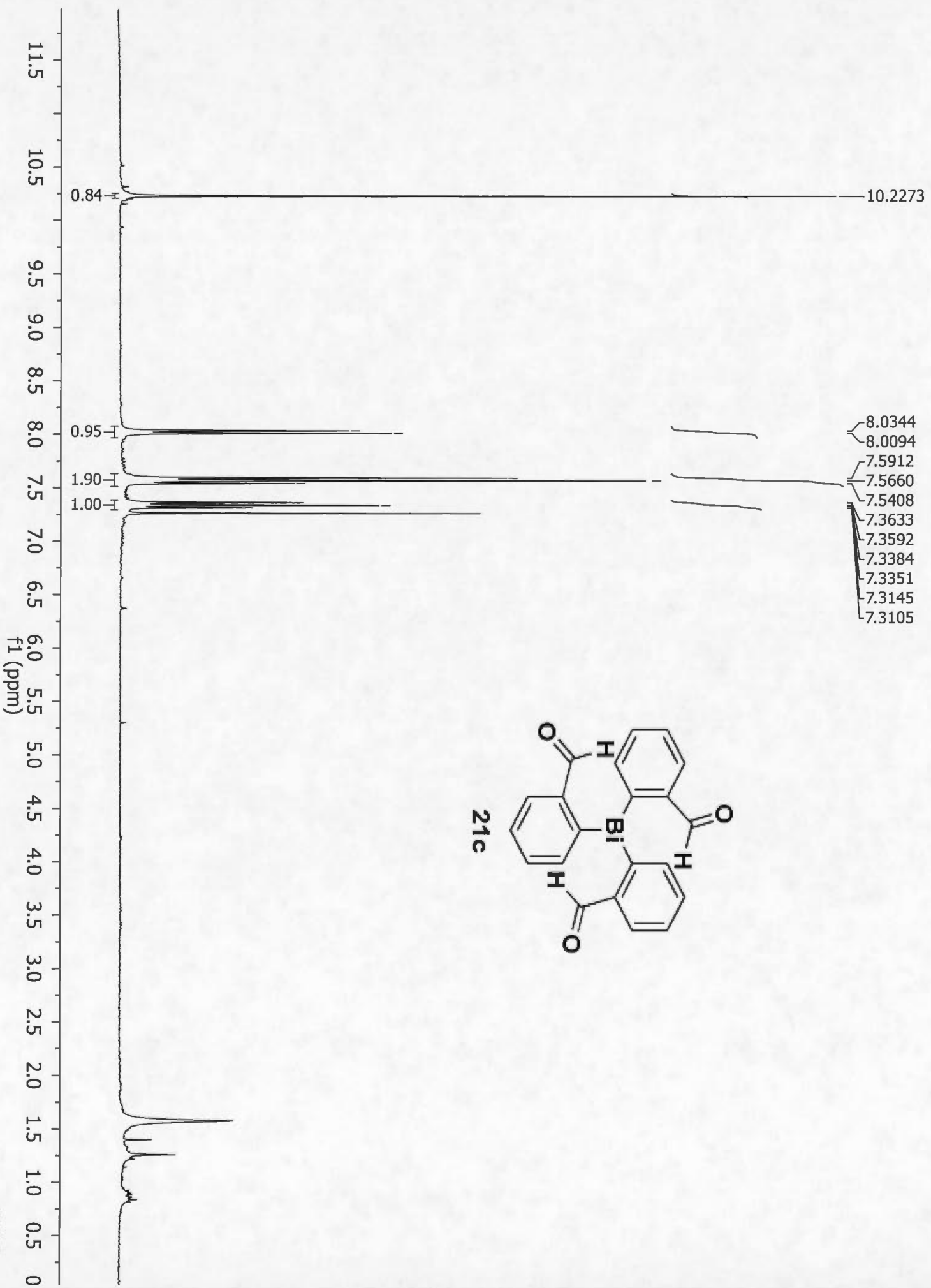


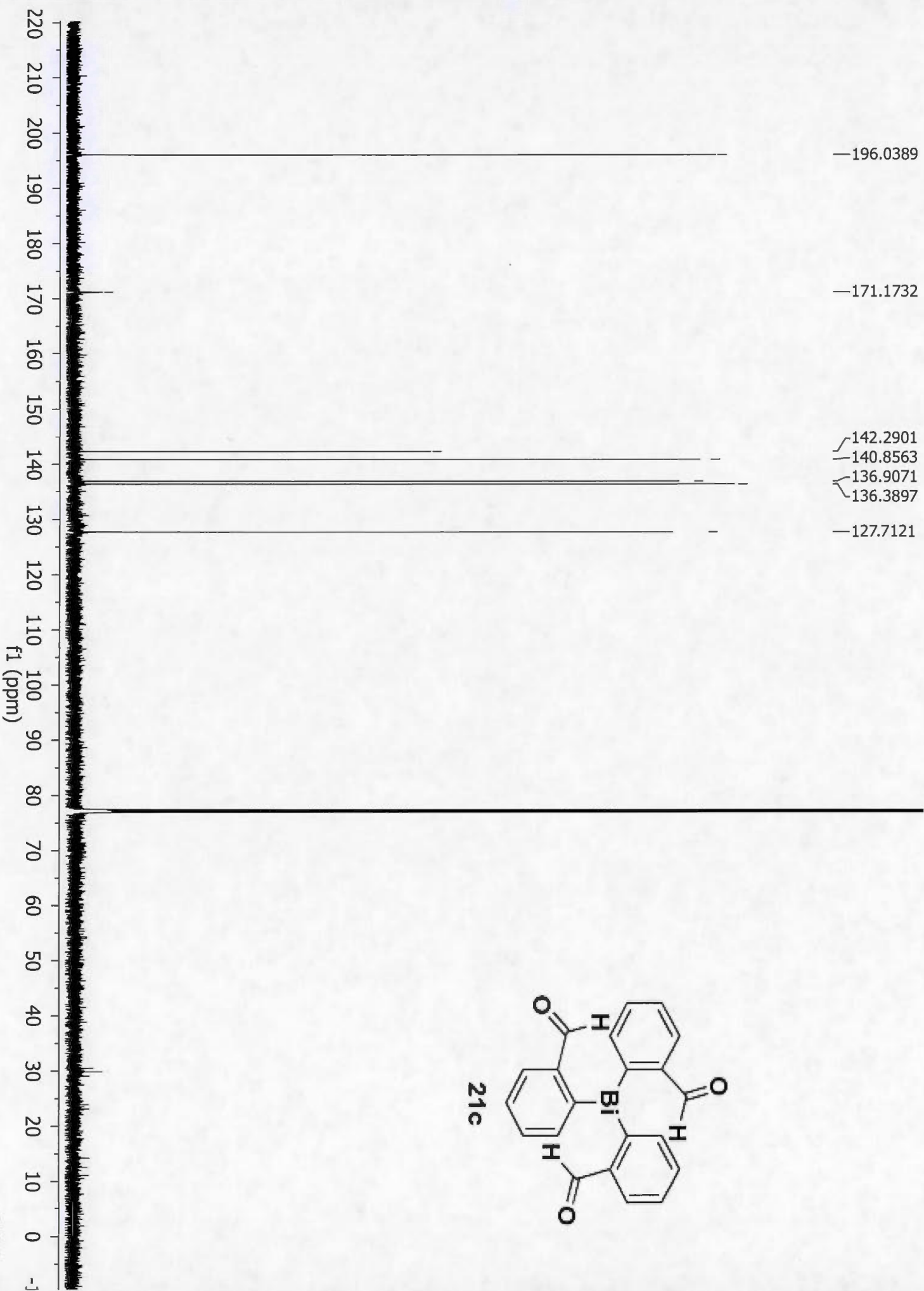
17a

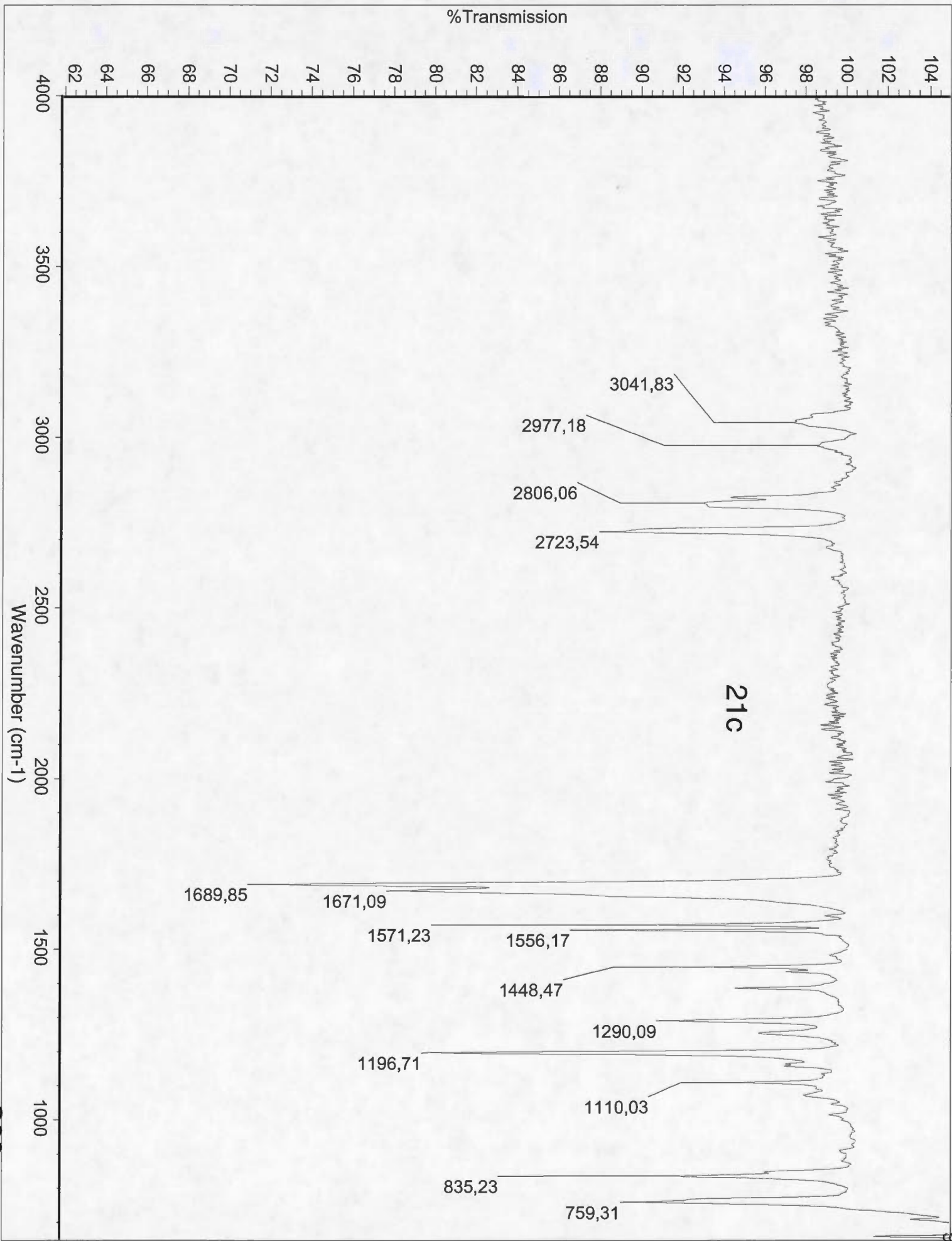




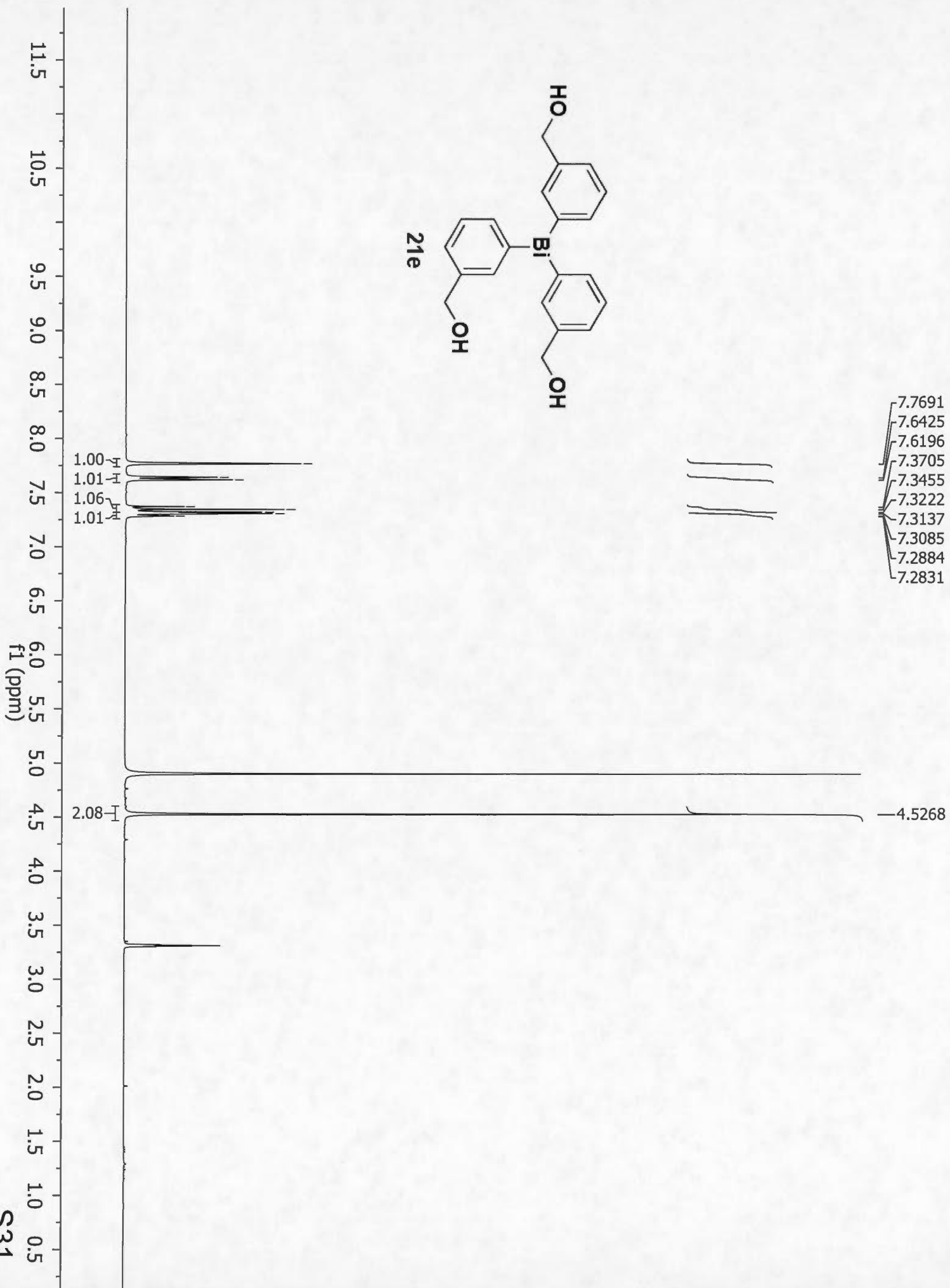
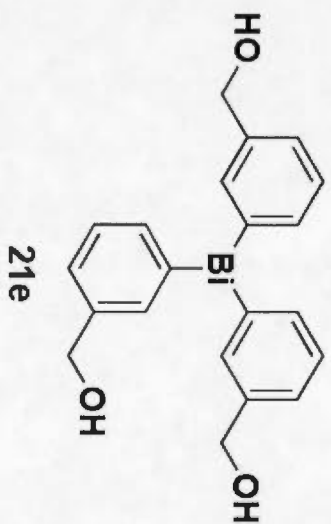


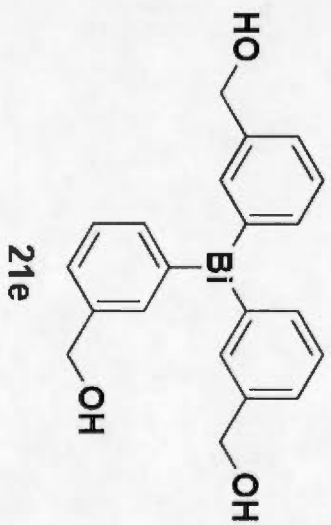




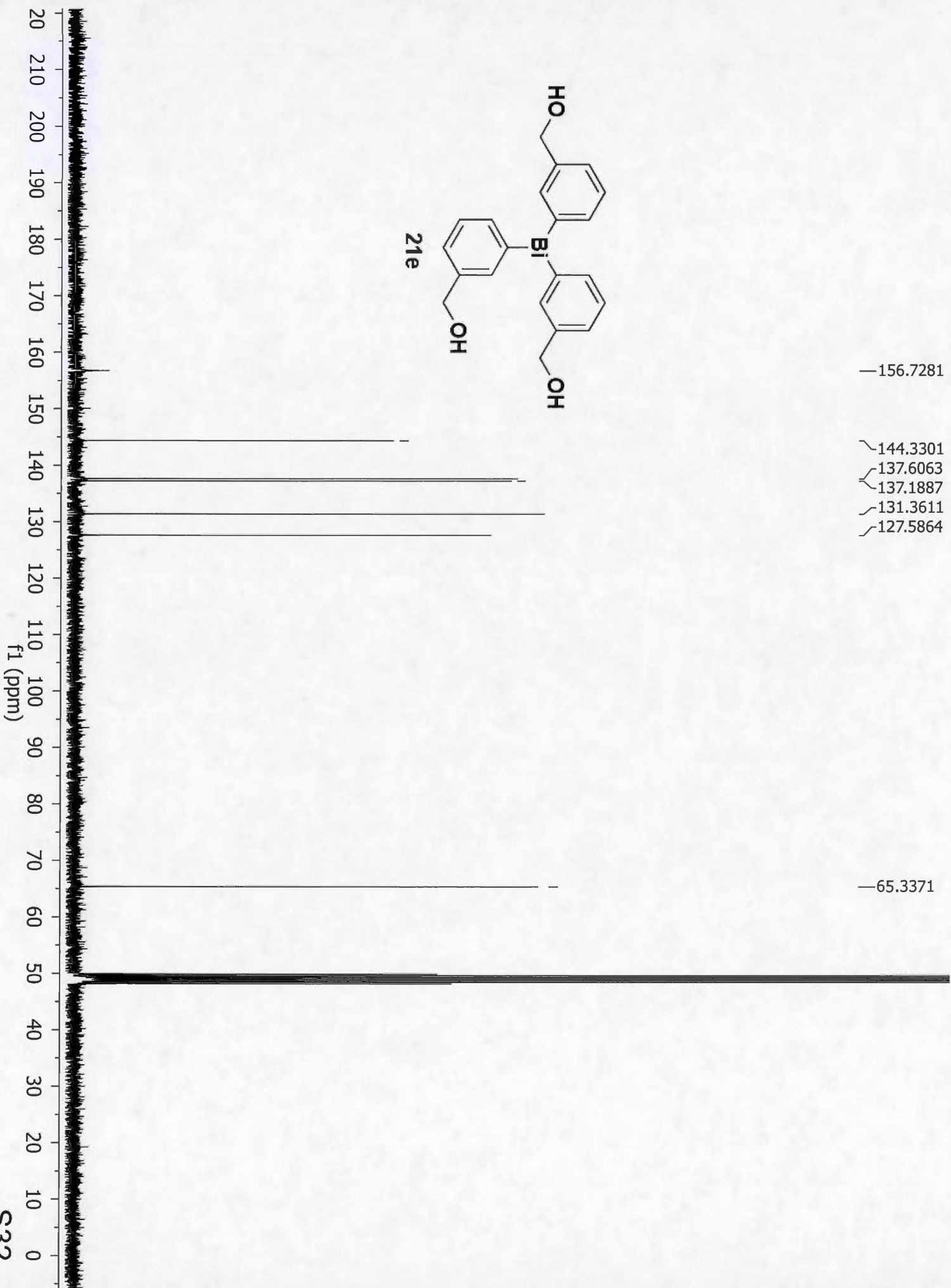


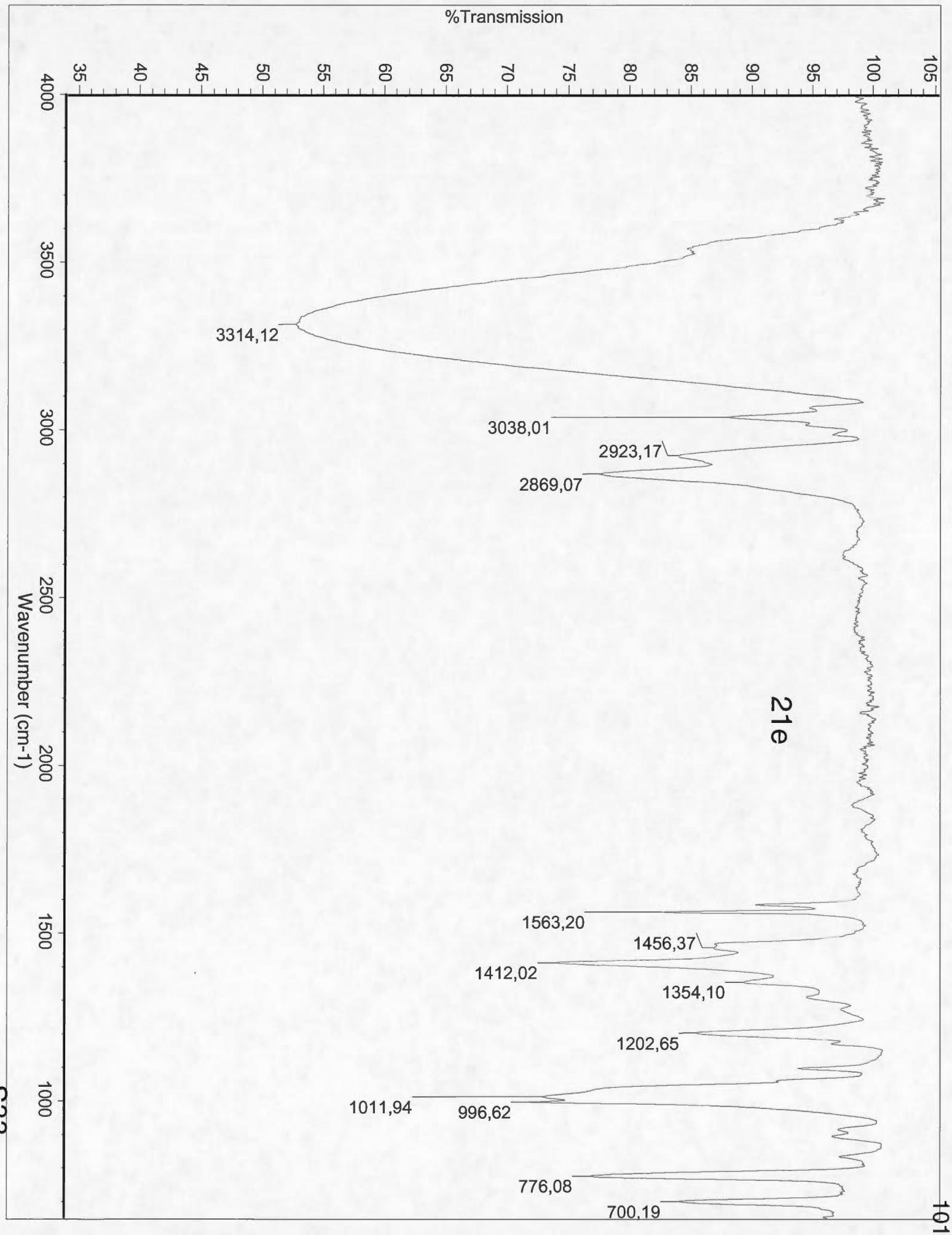


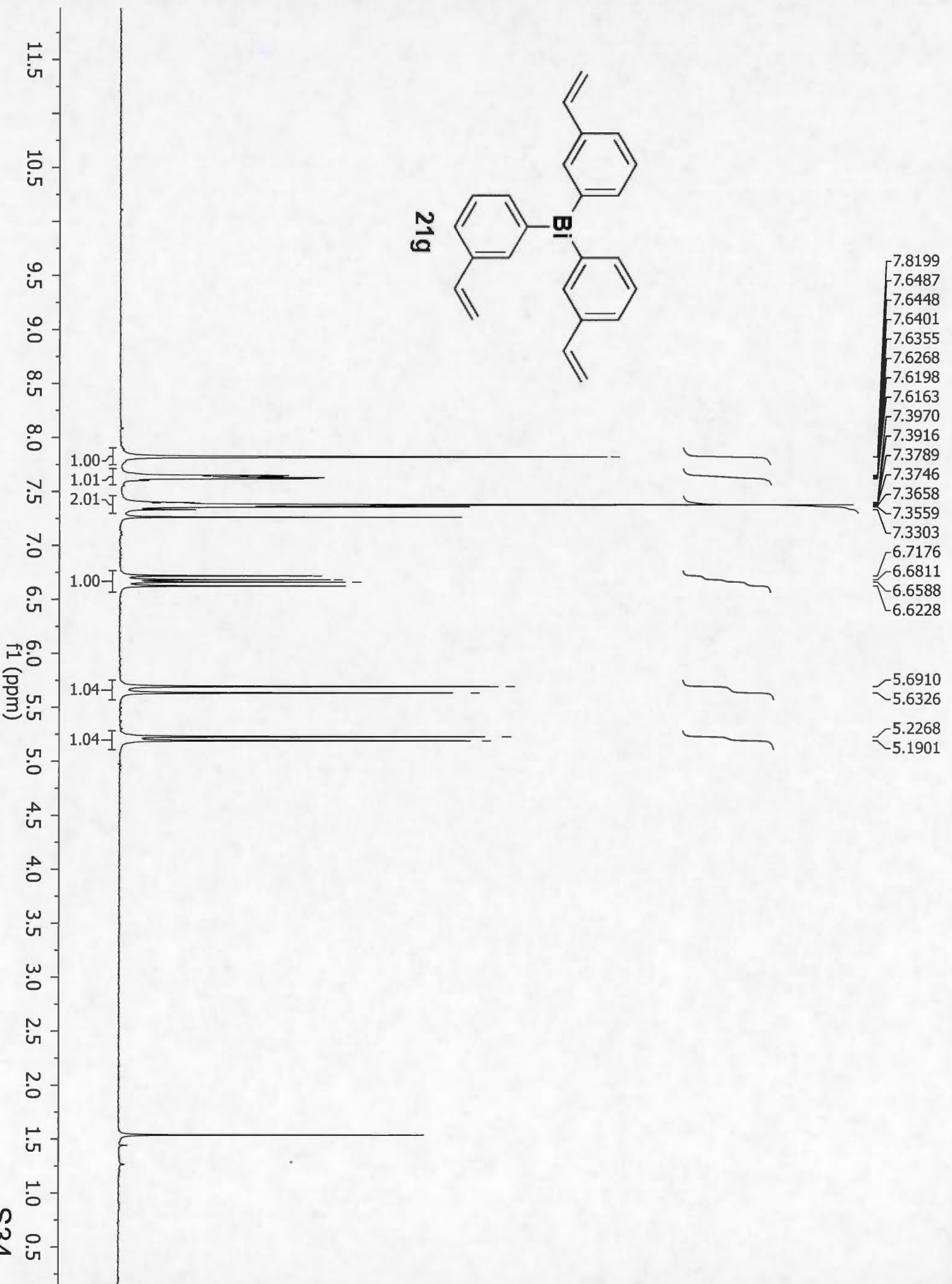
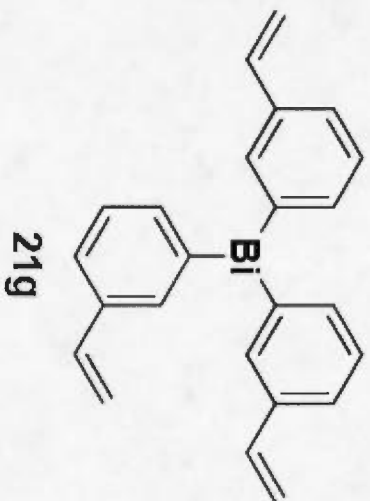




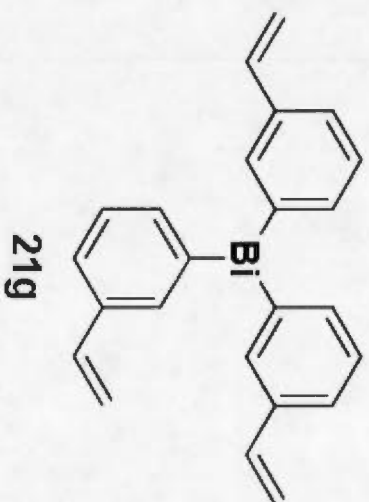
21e



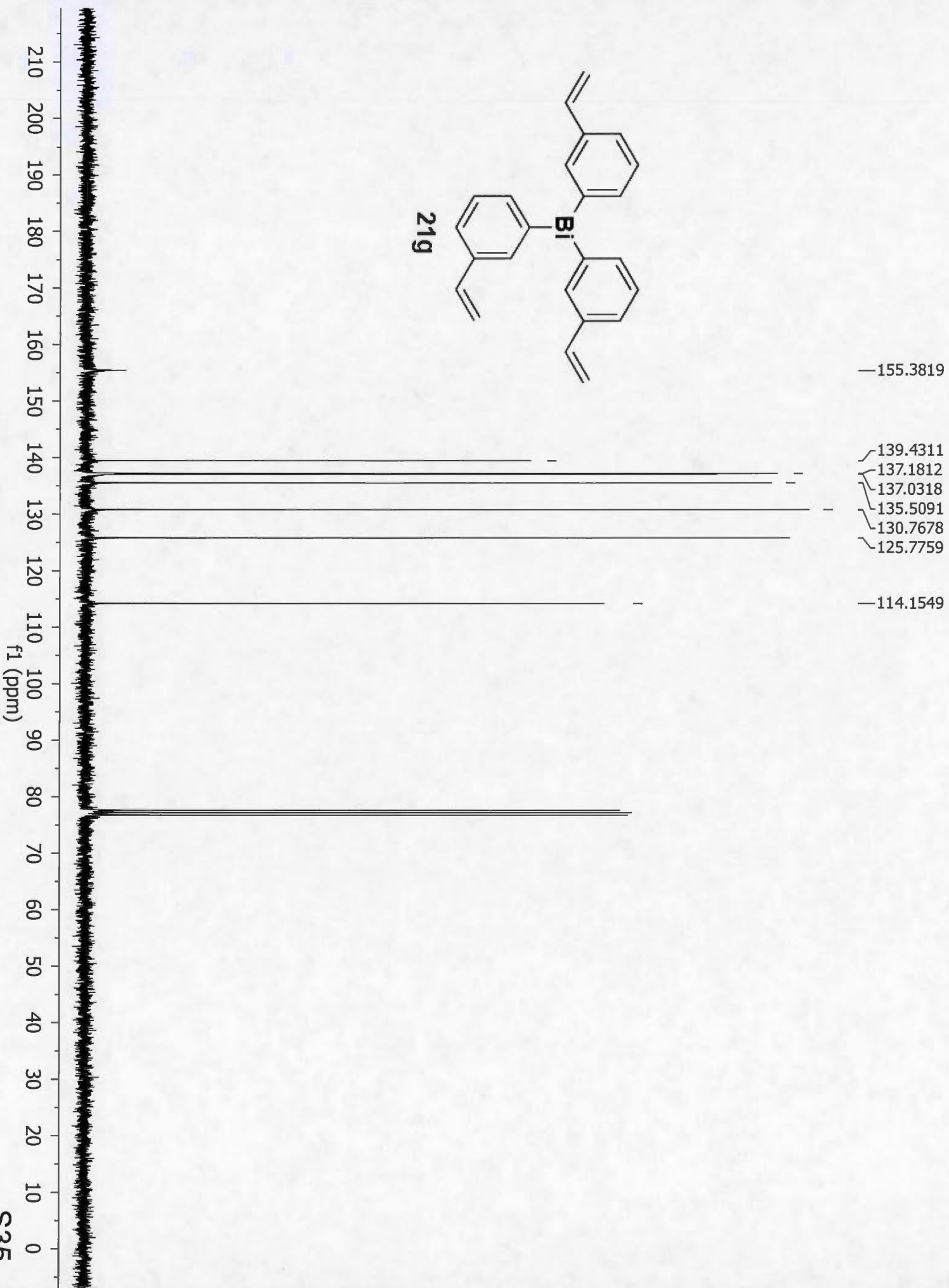


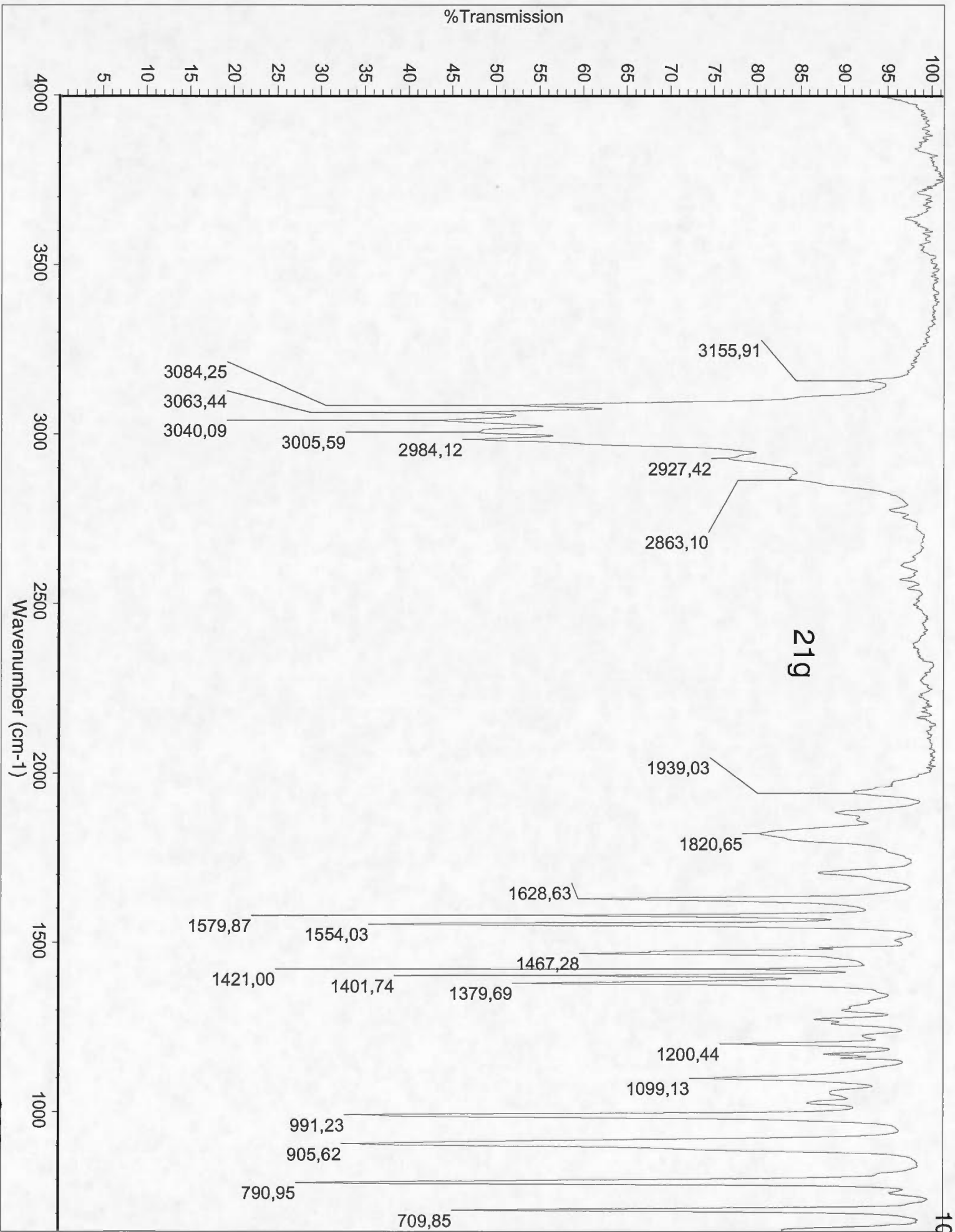


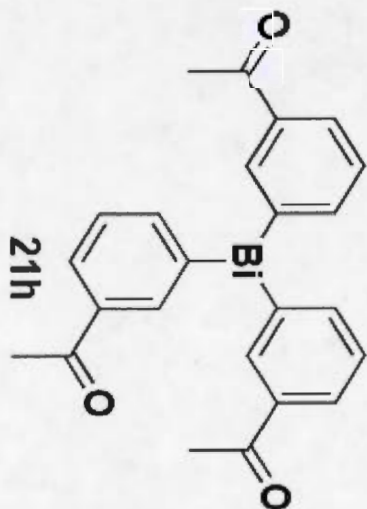




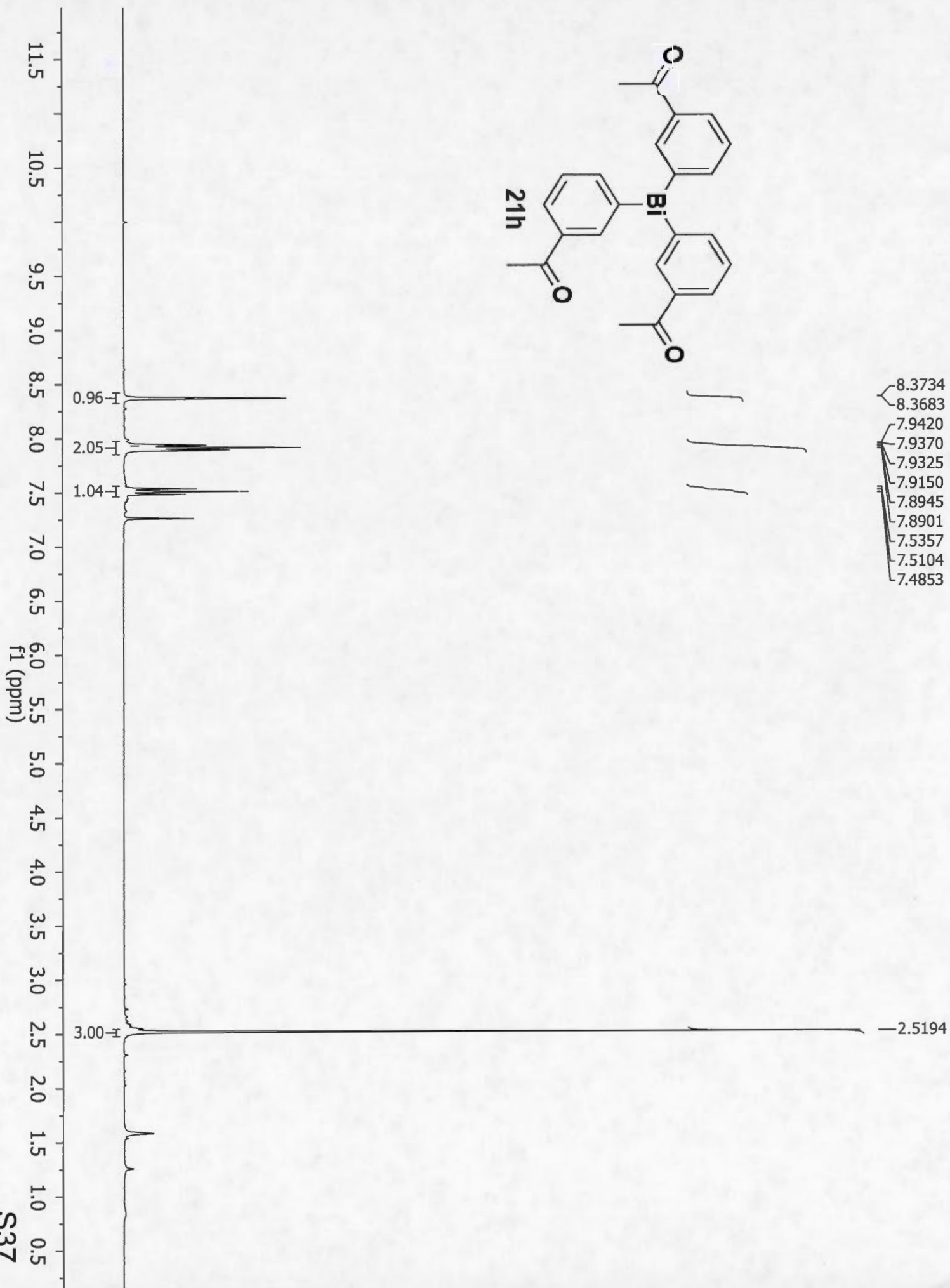
21g

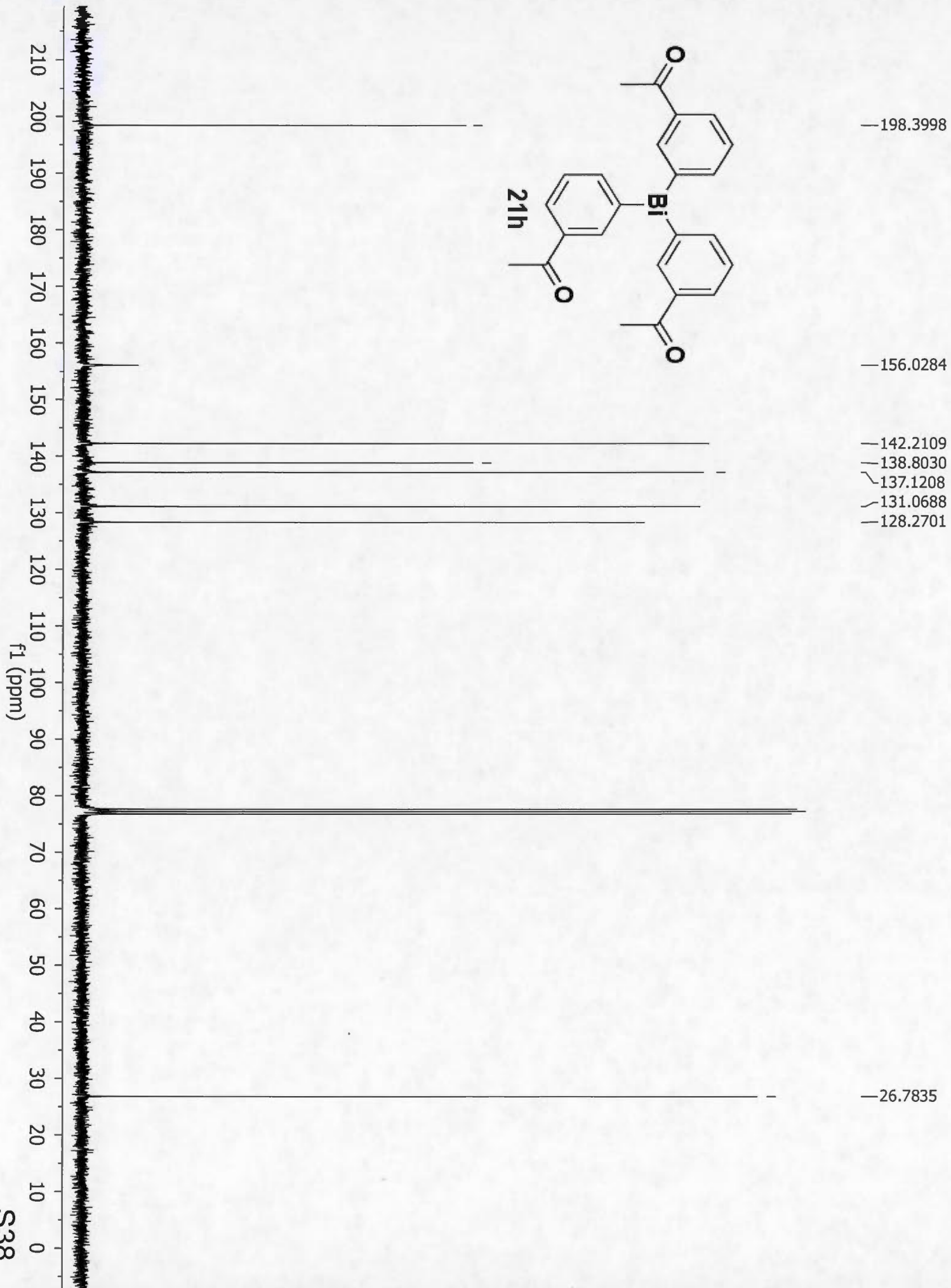
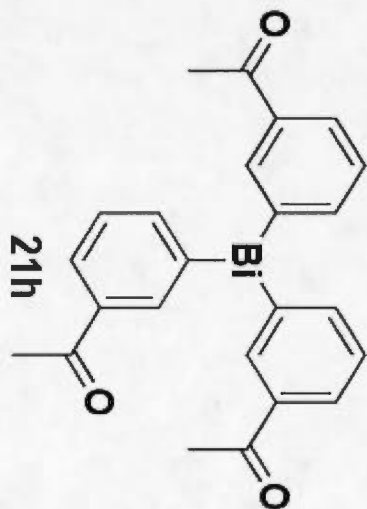




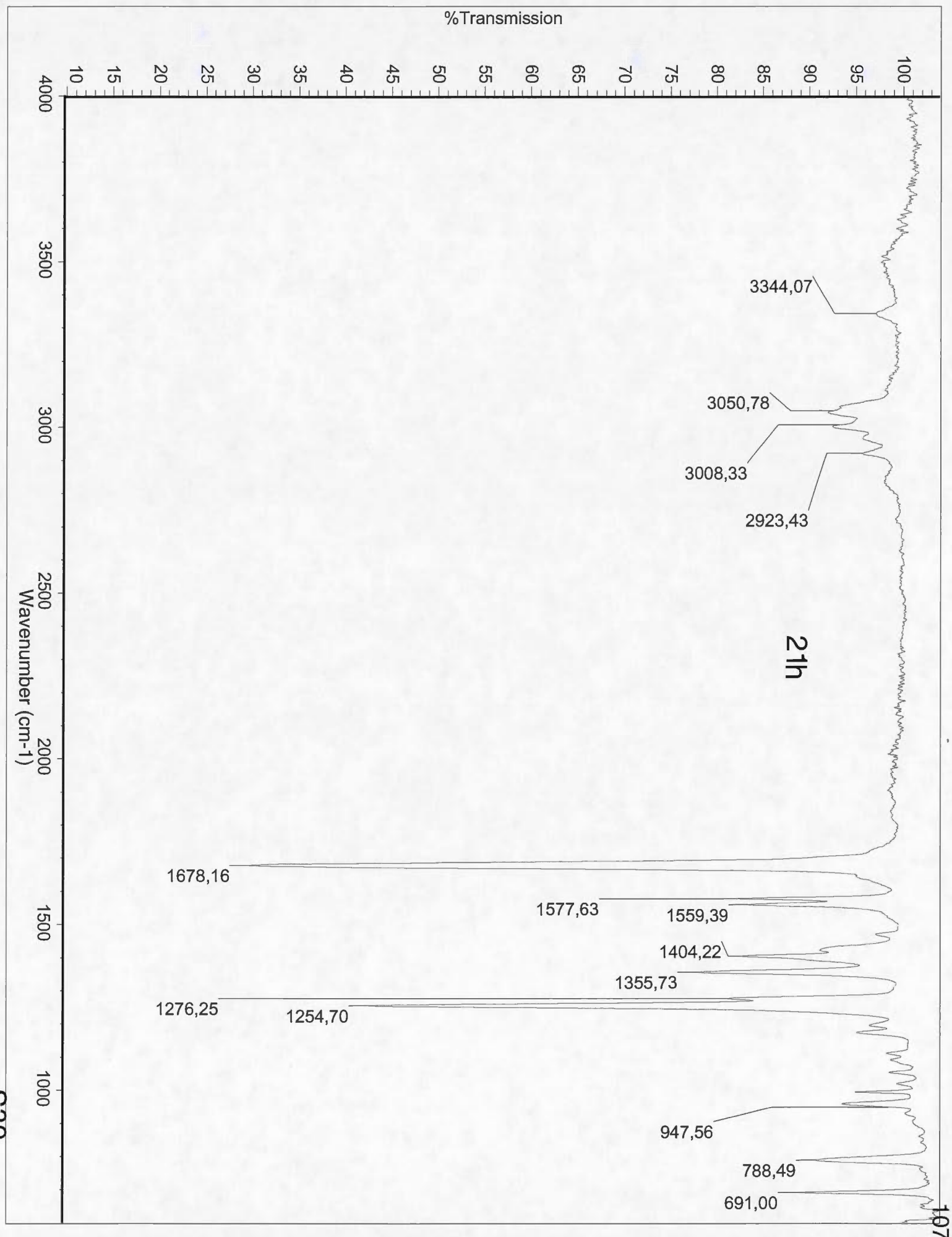


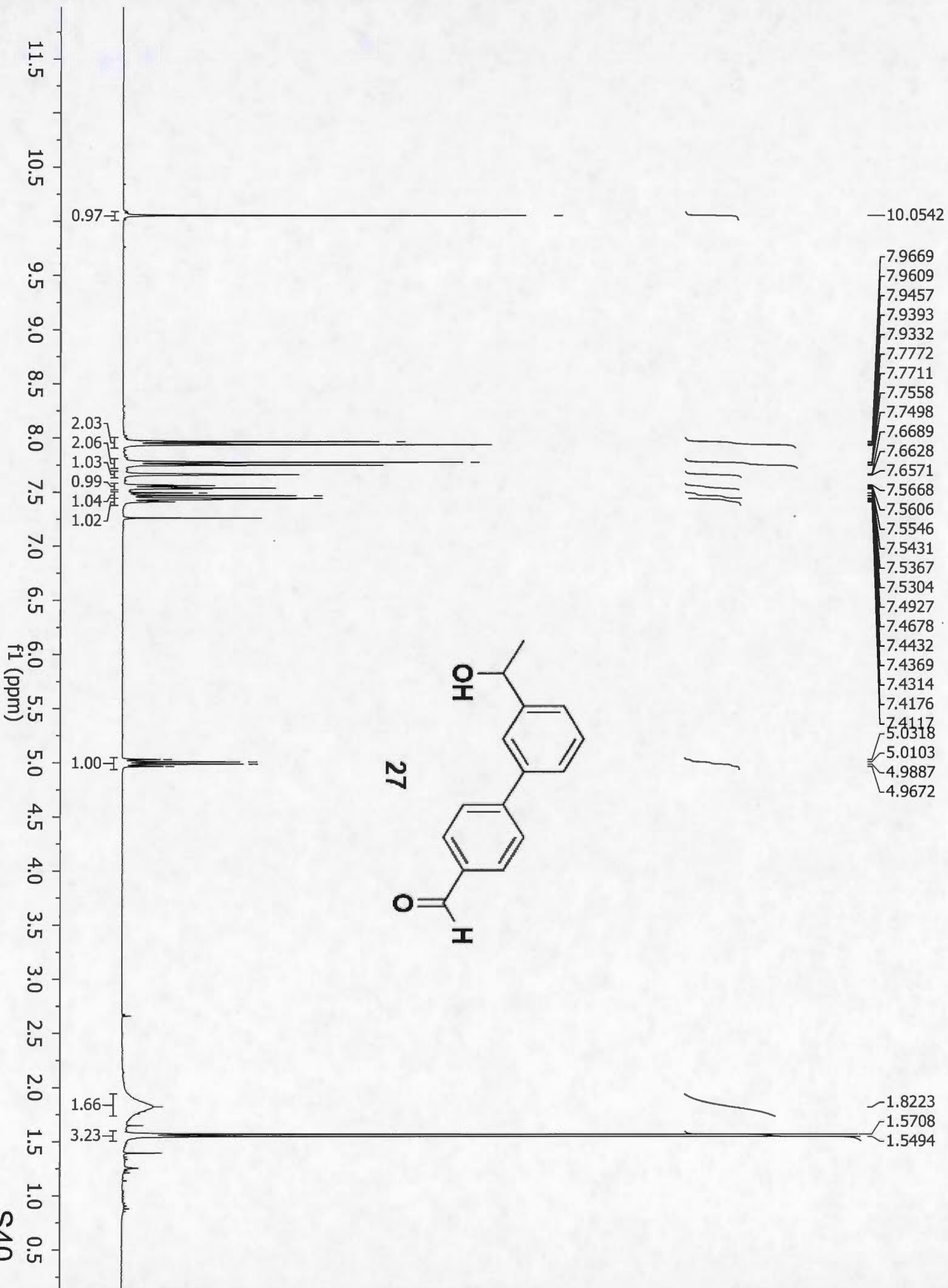
21h

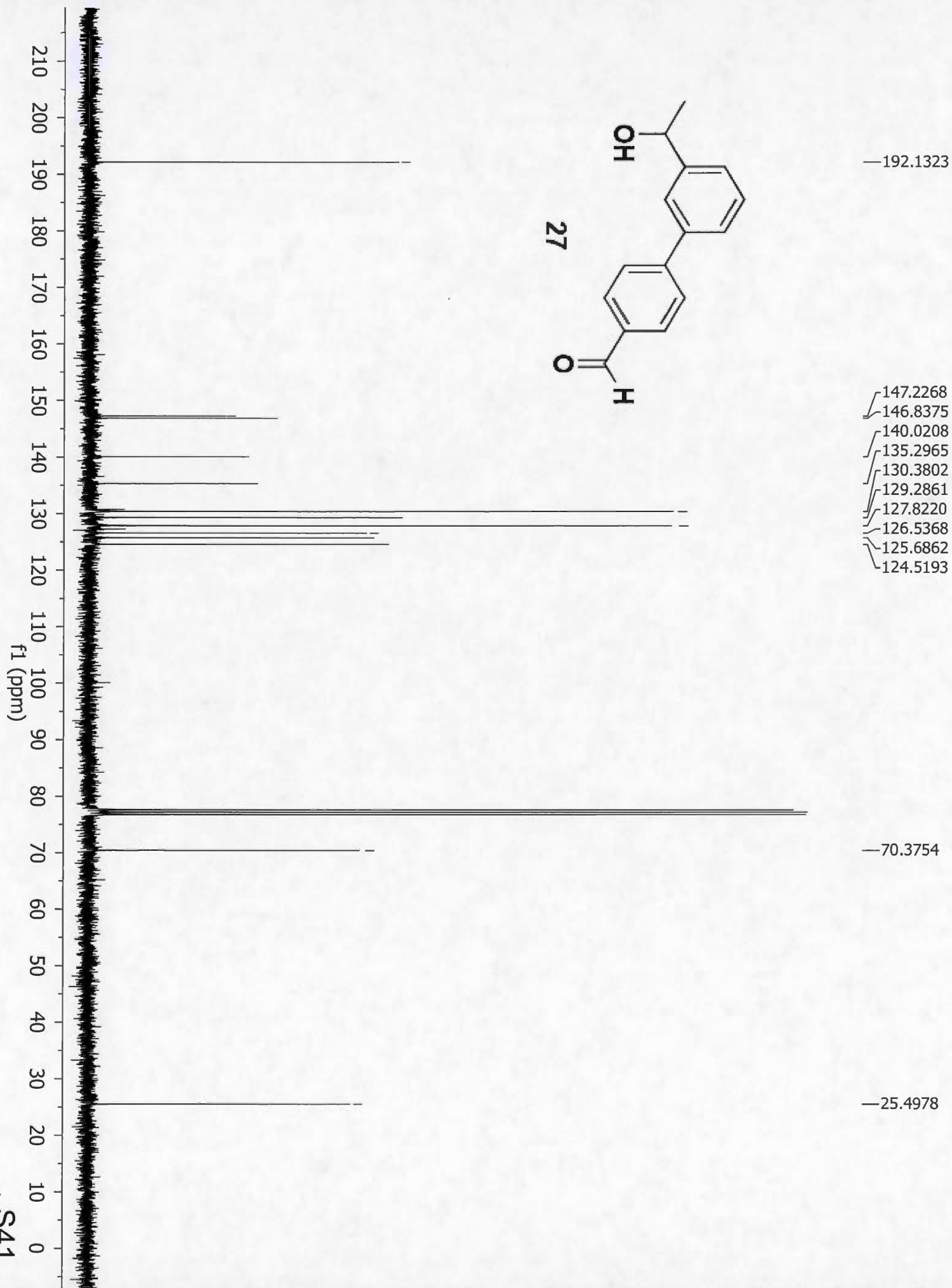
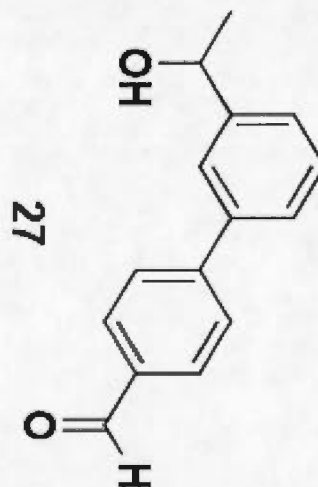


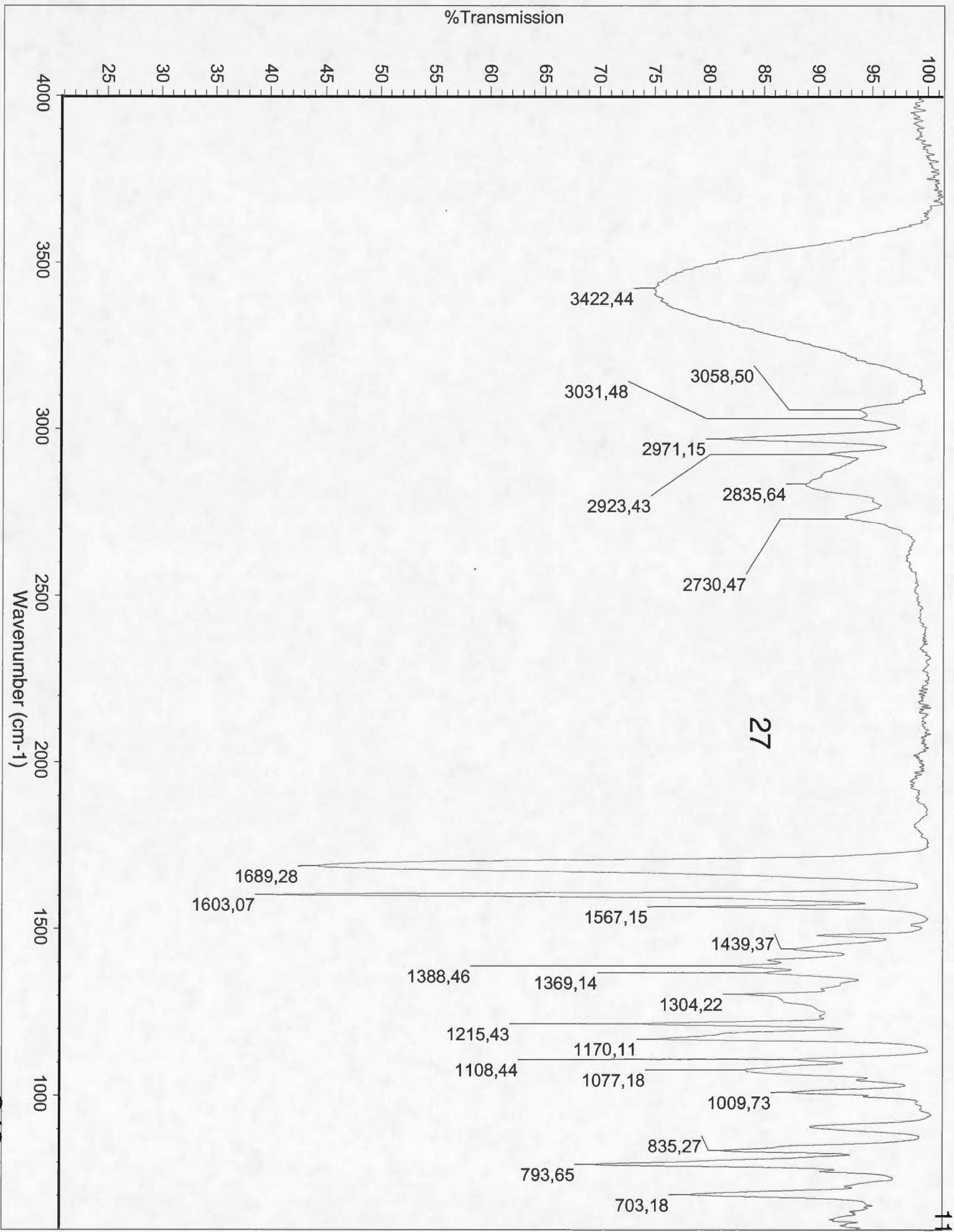






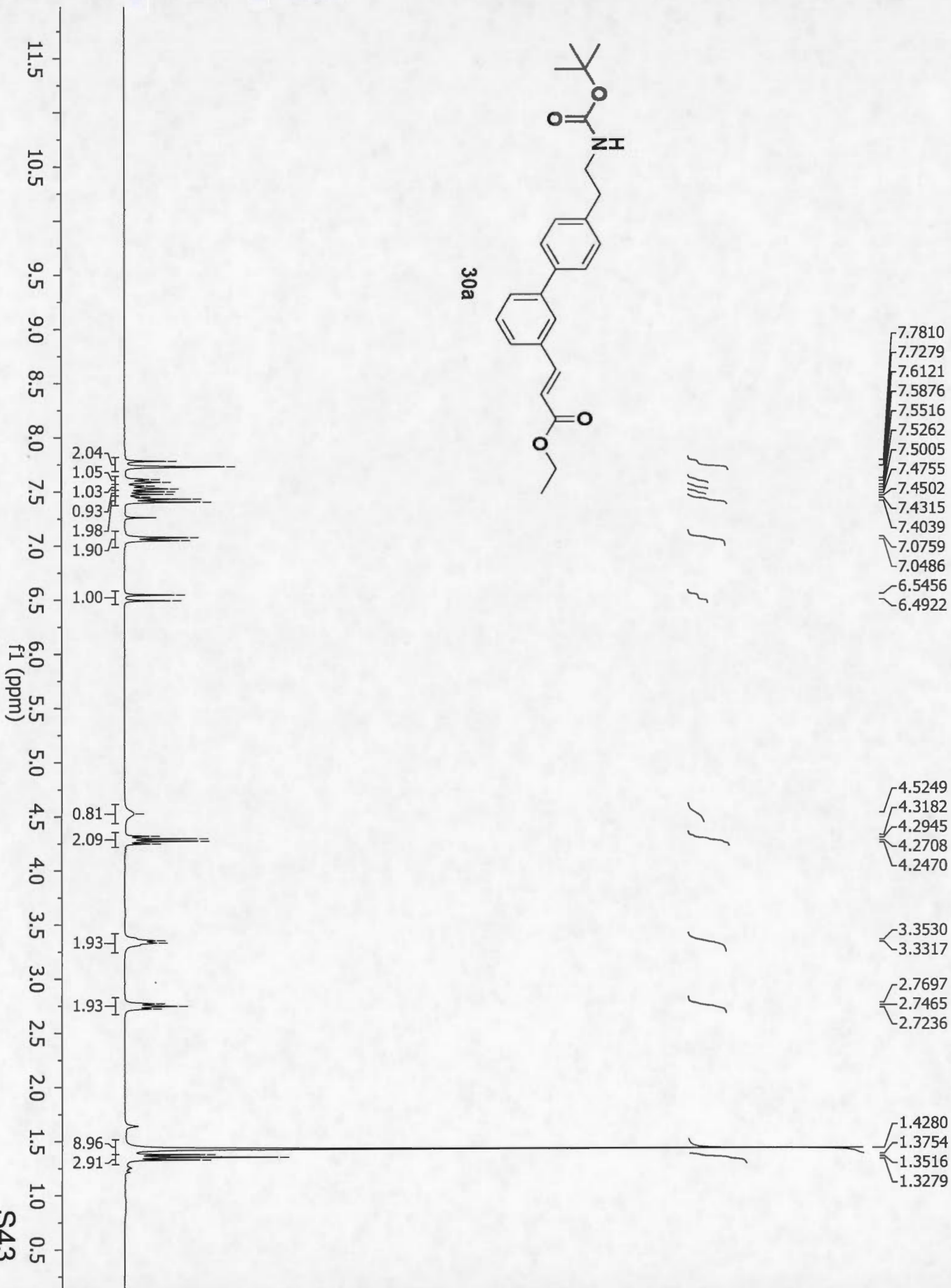
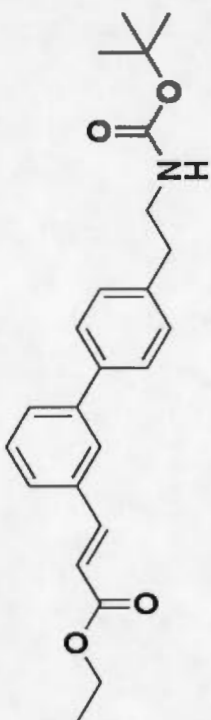


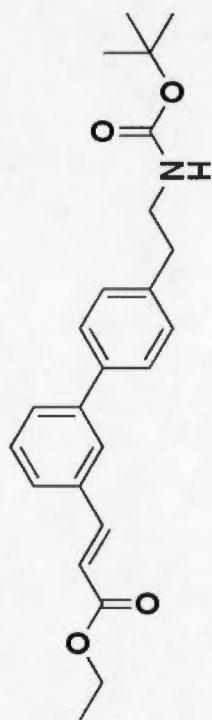




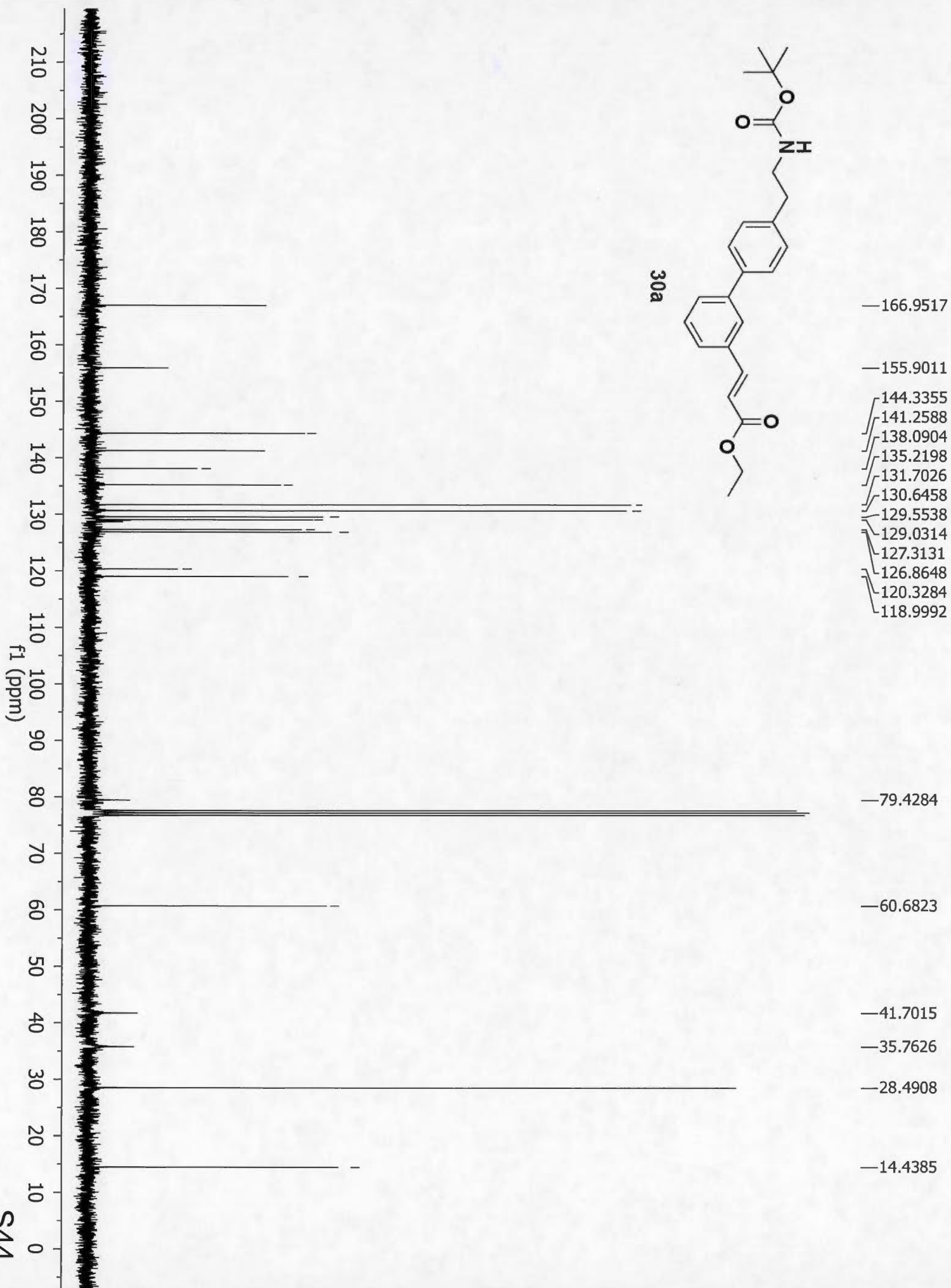


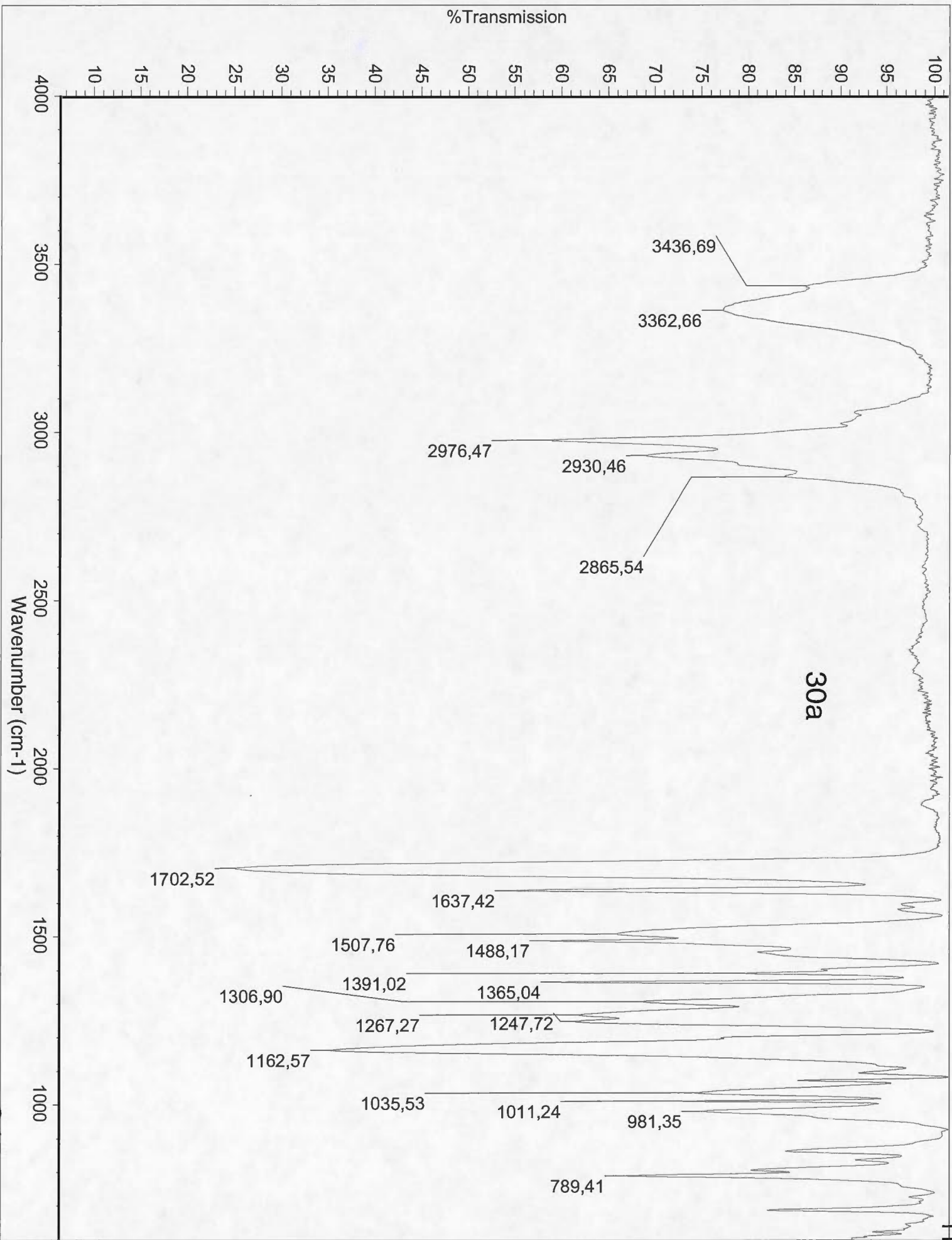
30a

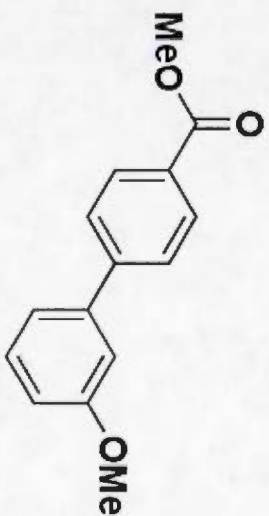




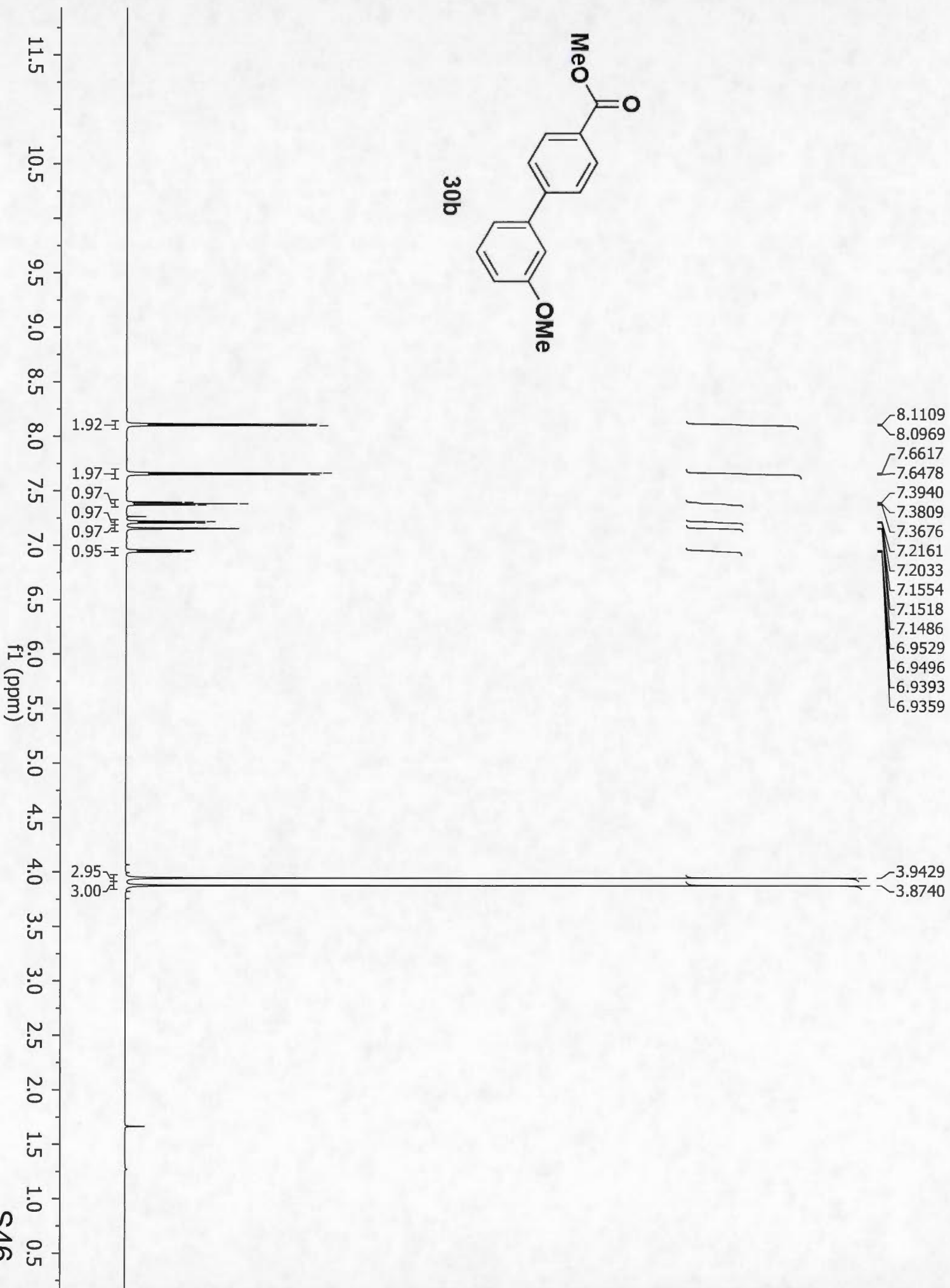
30a



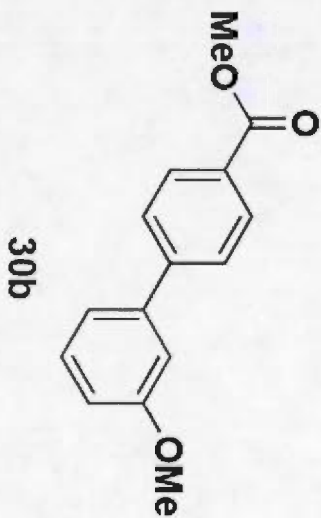




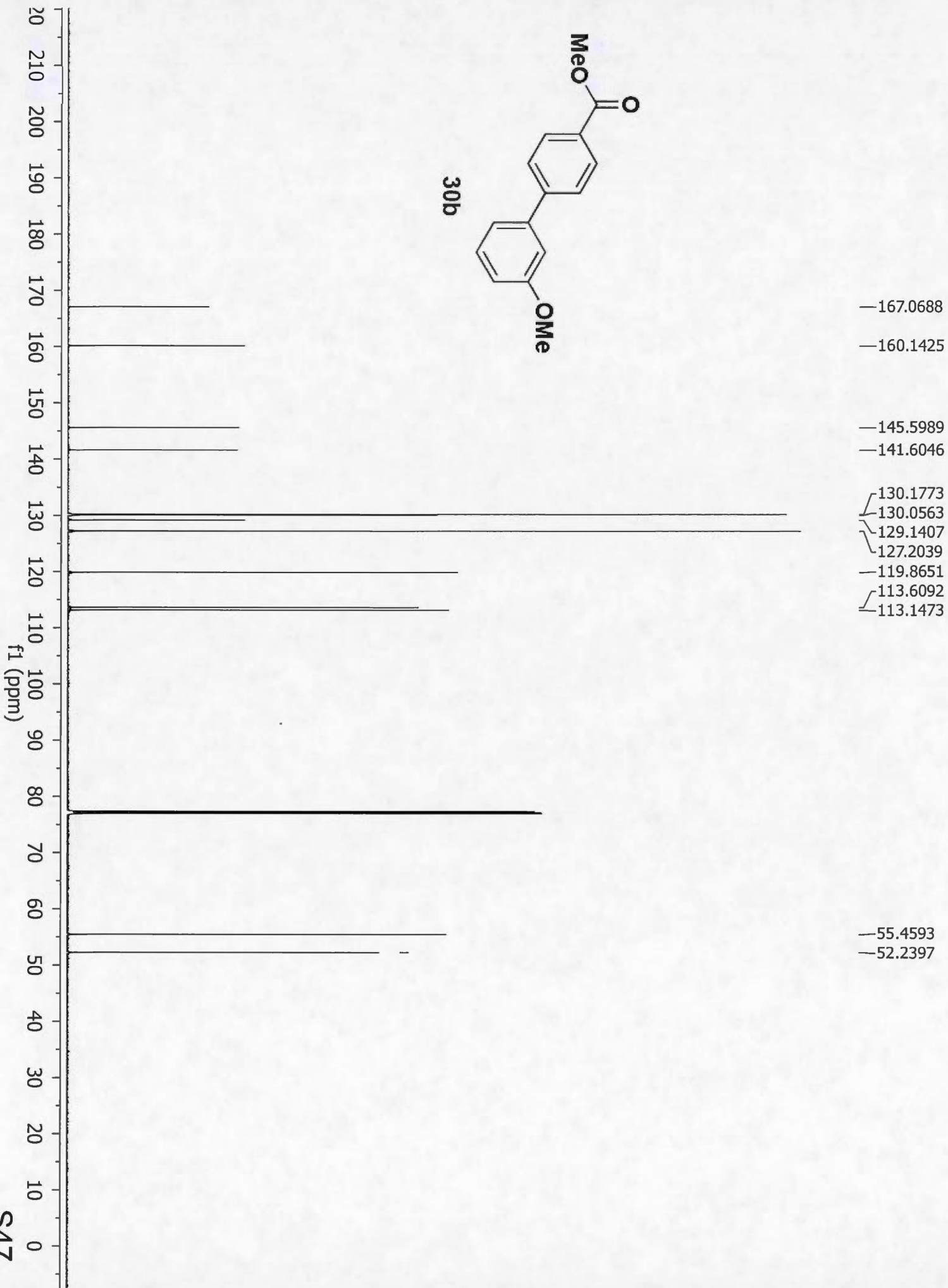
30b

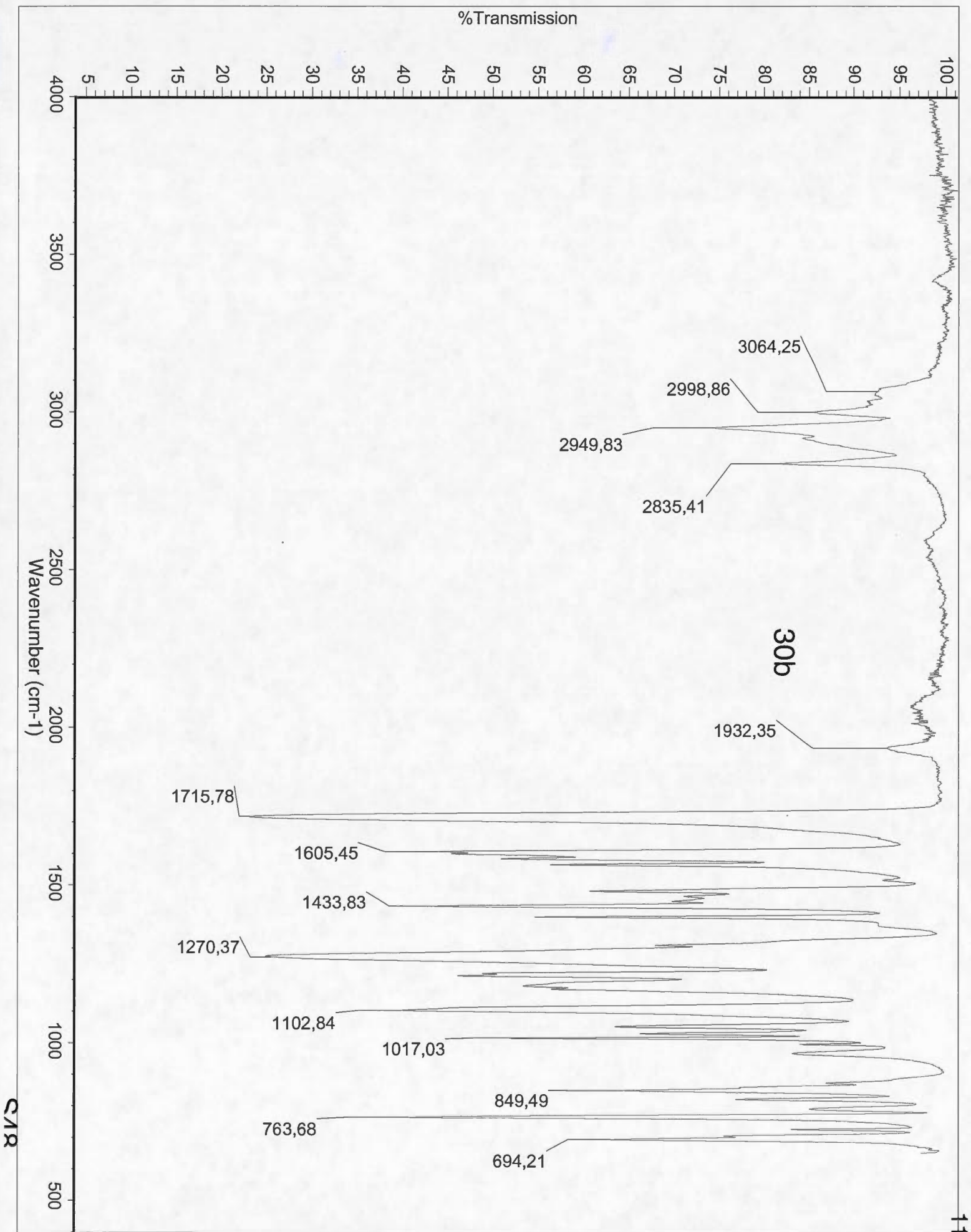


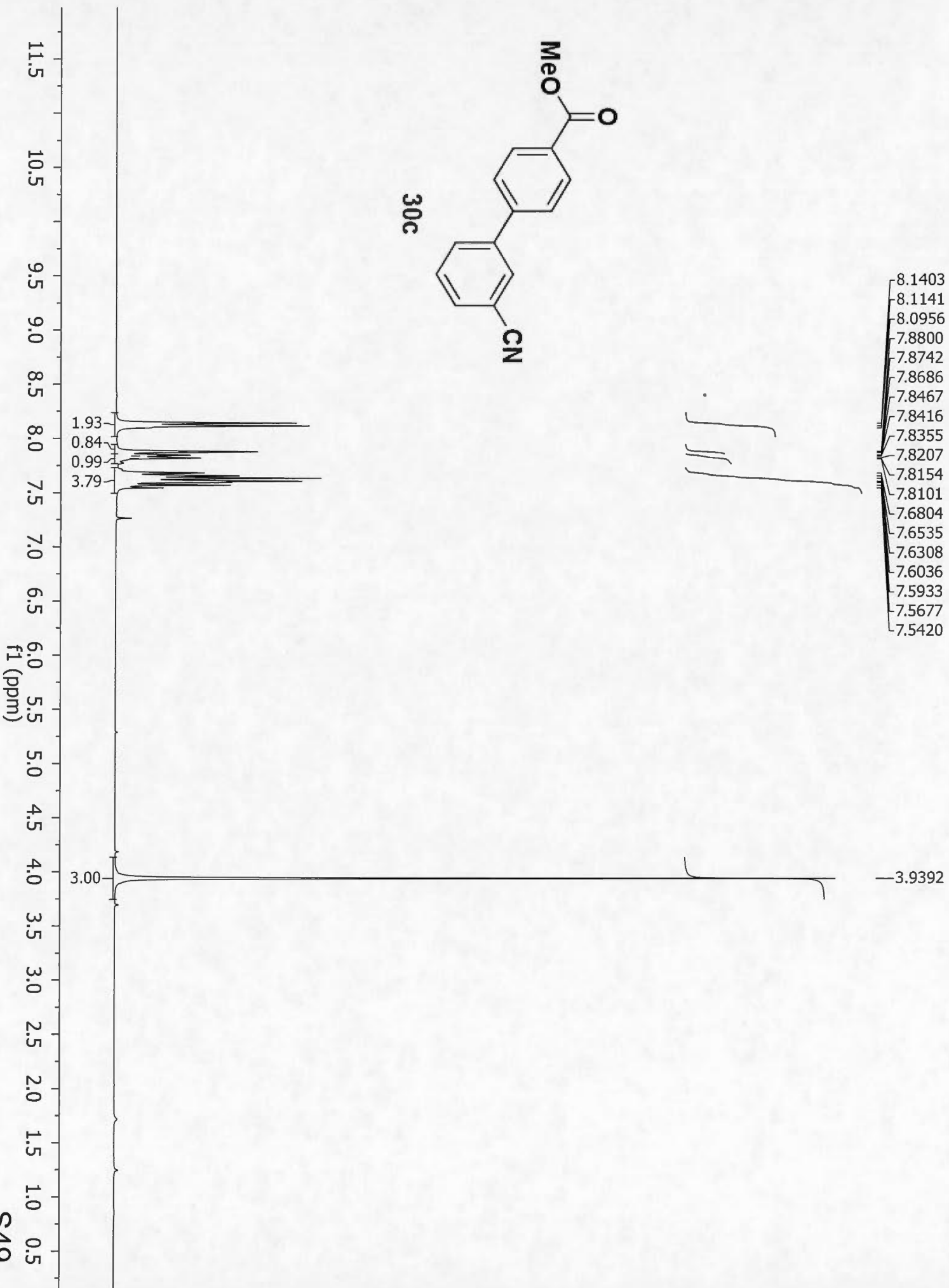


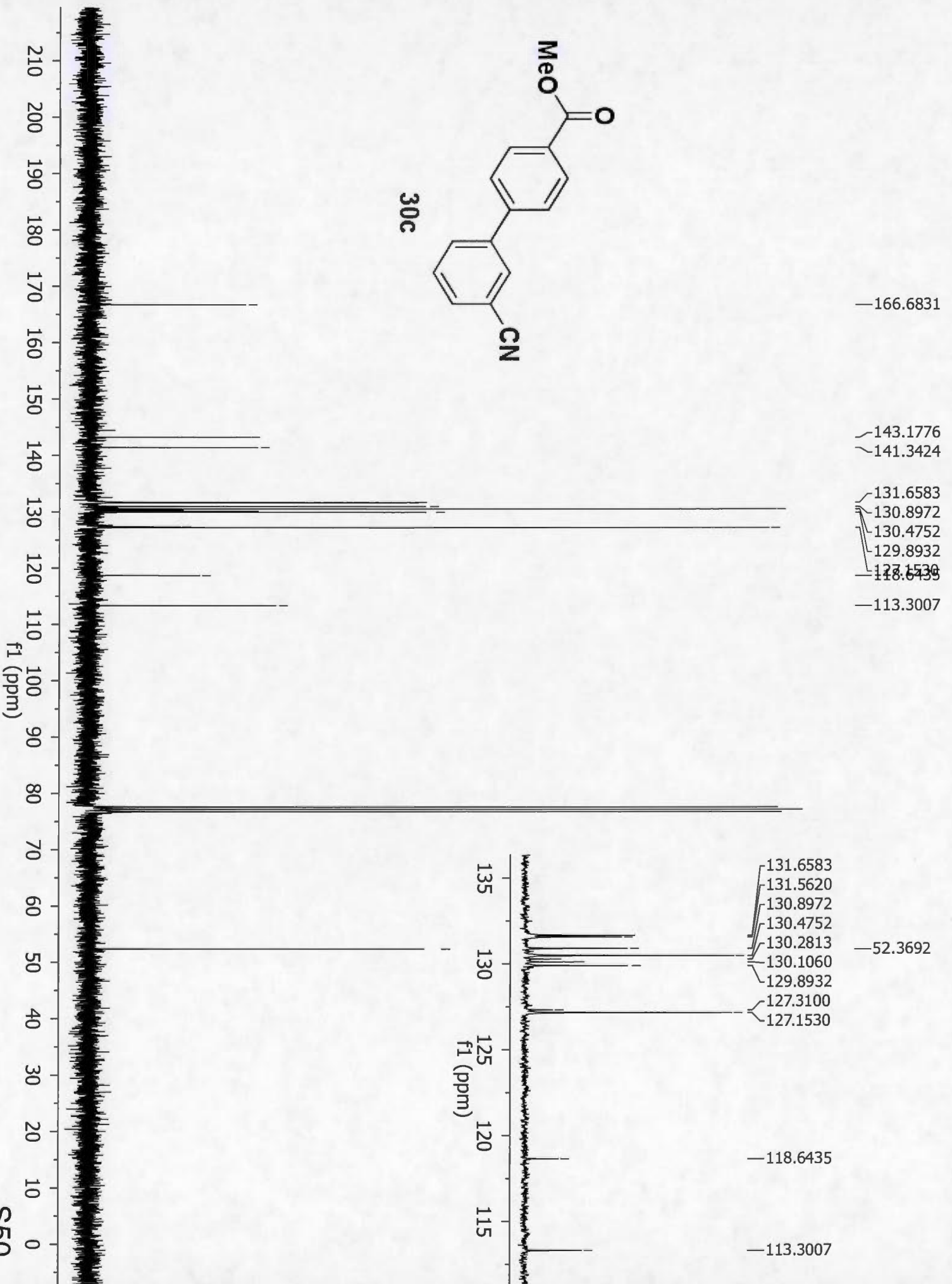


30b

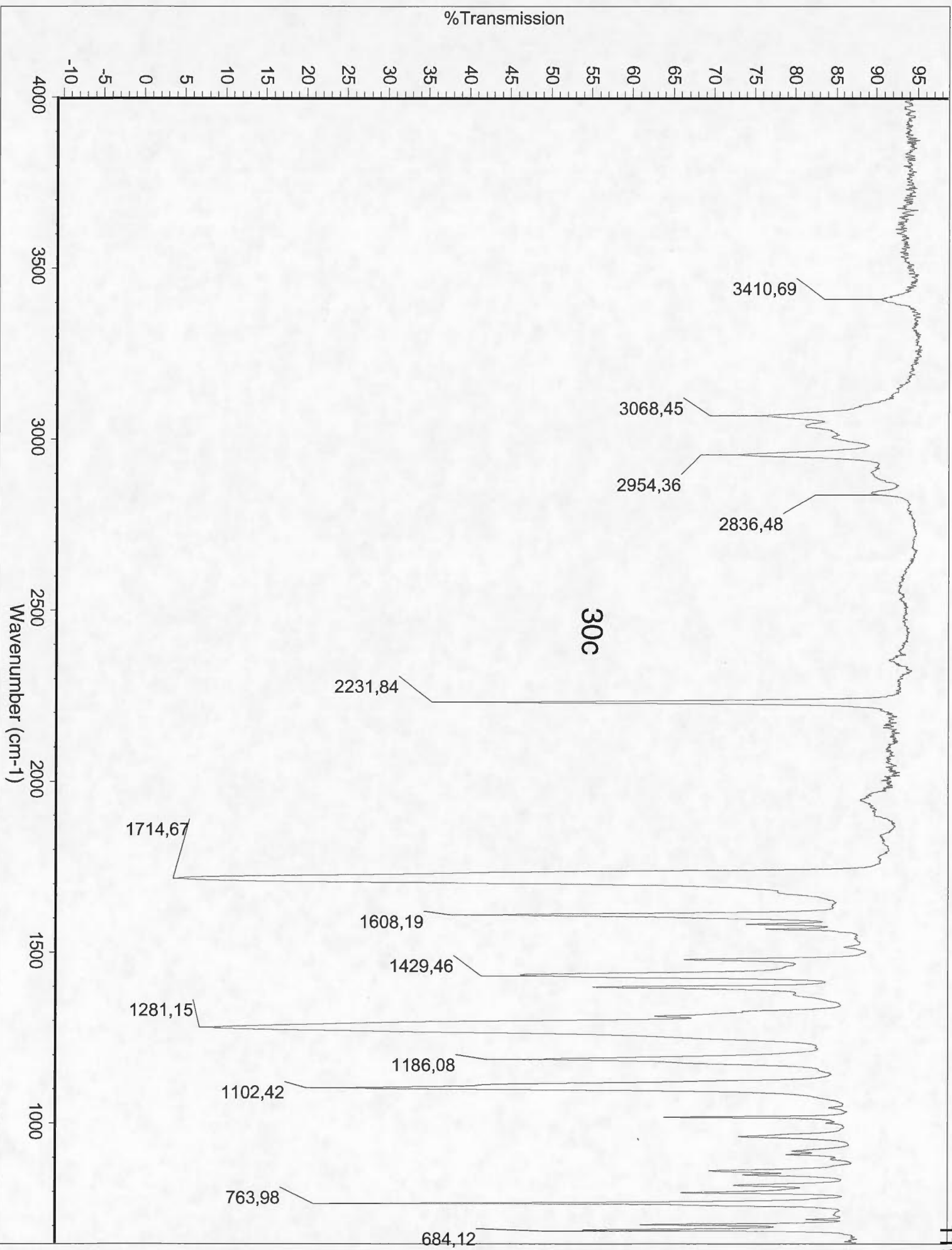


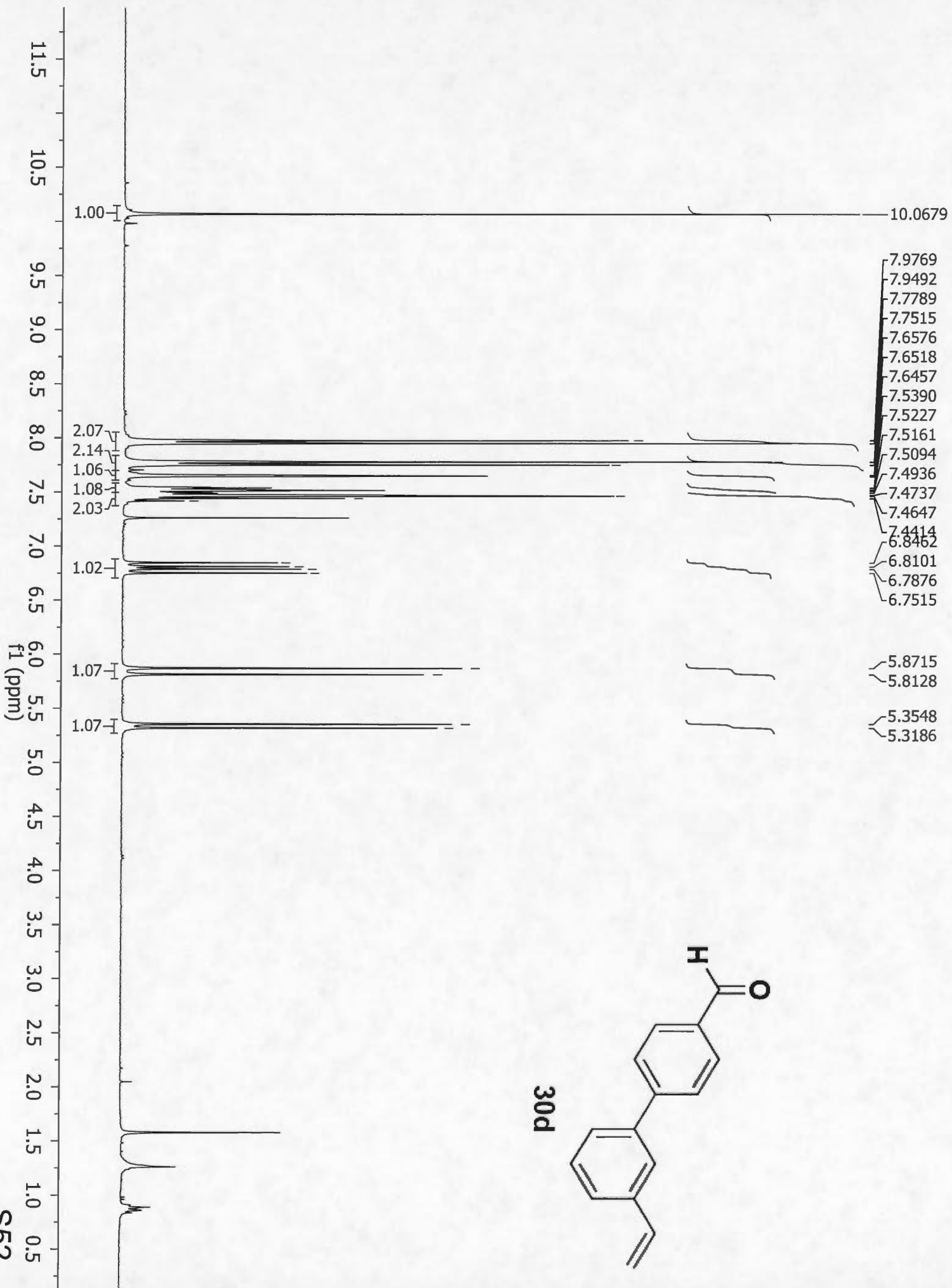
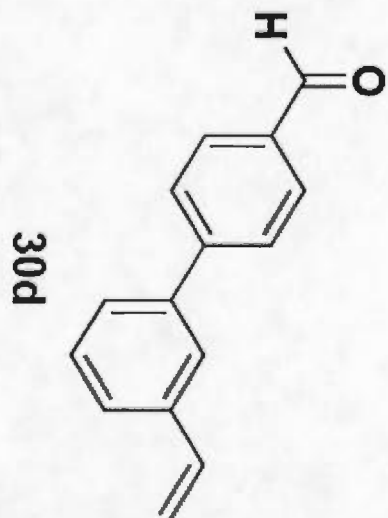


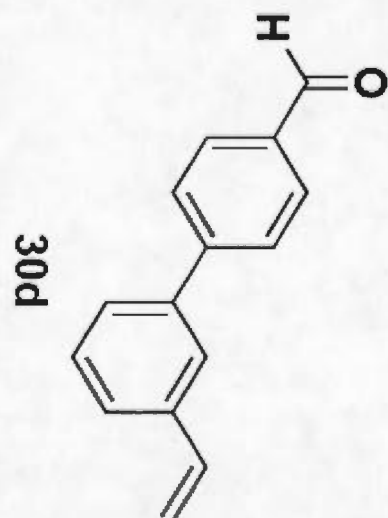




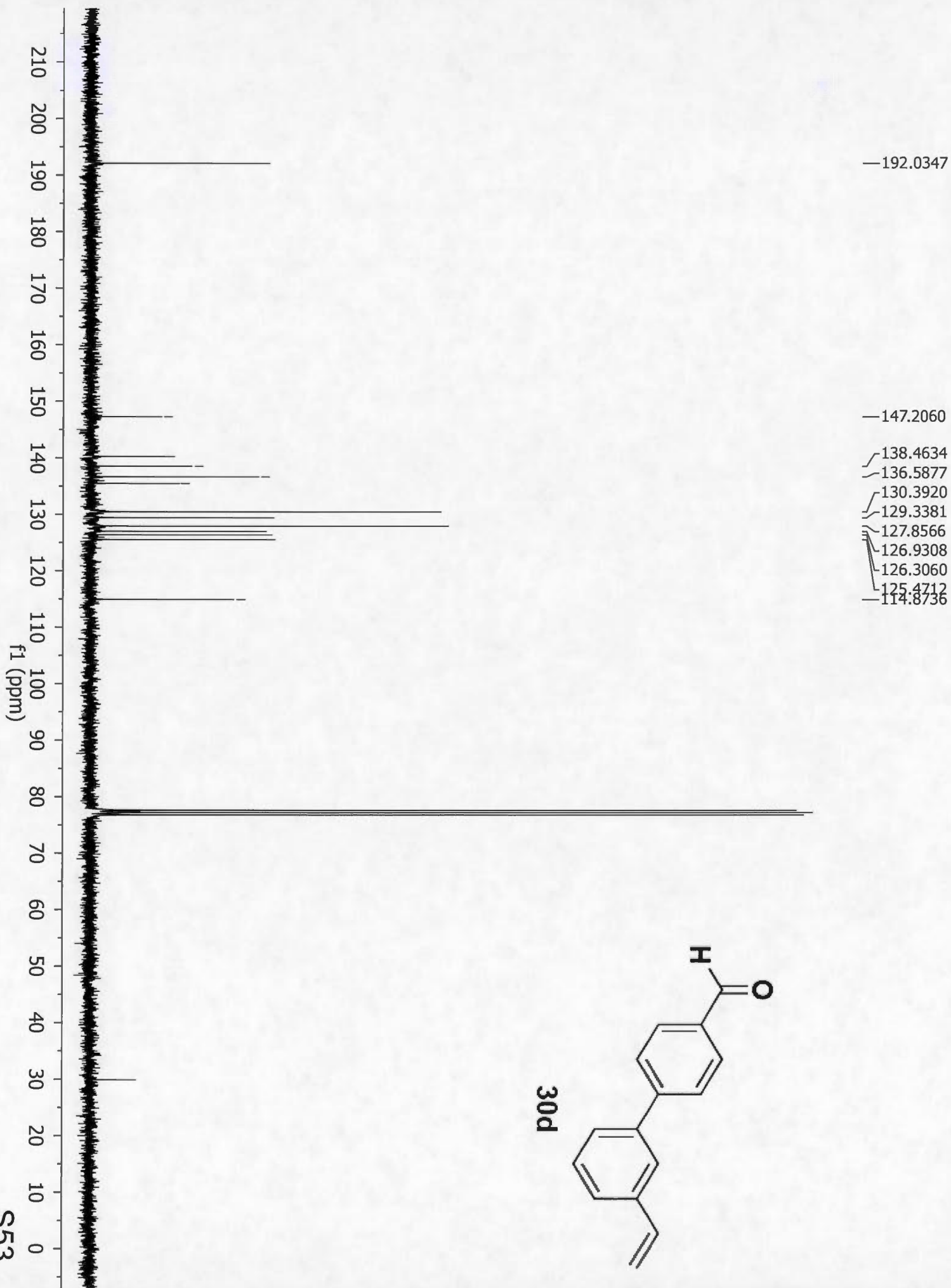




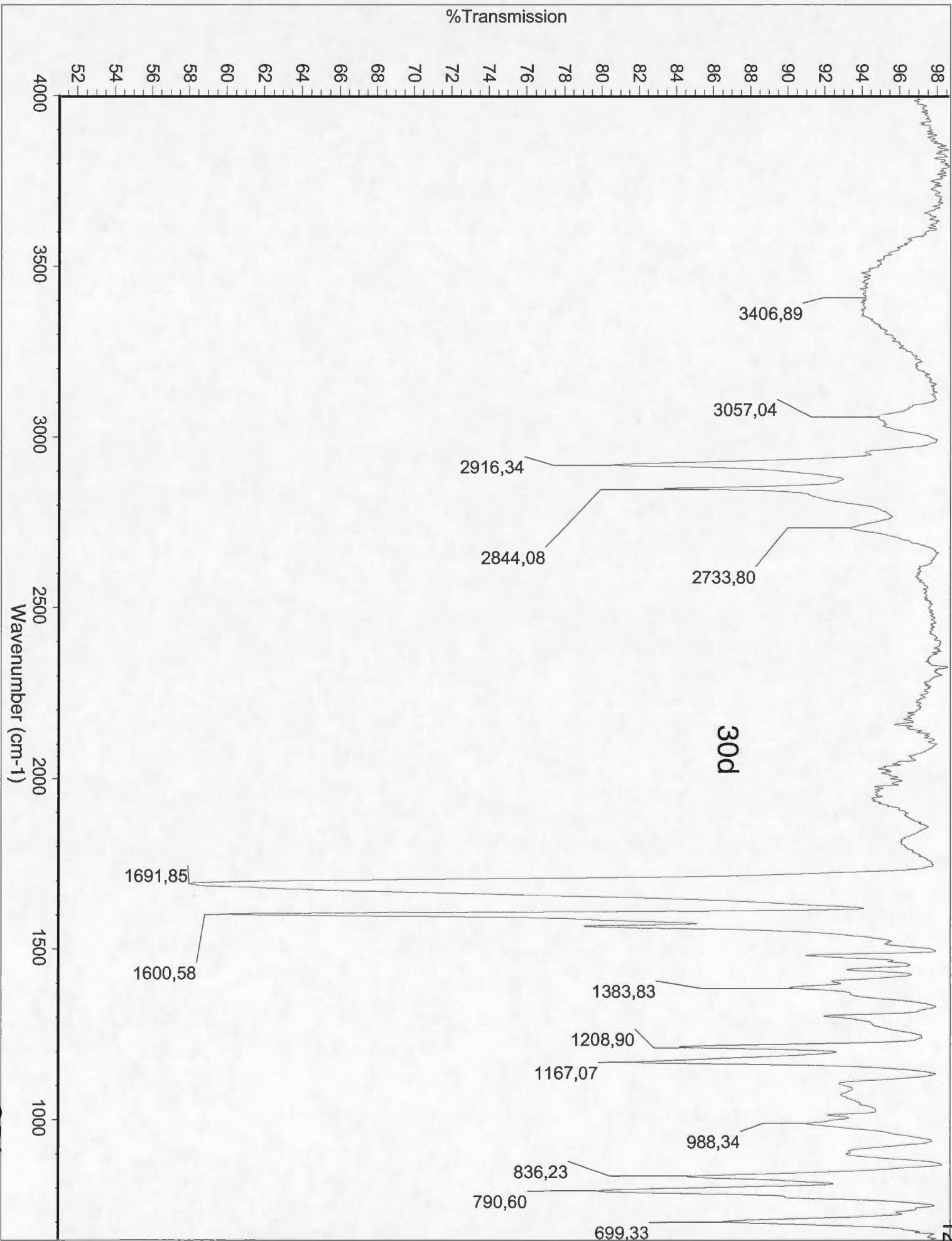




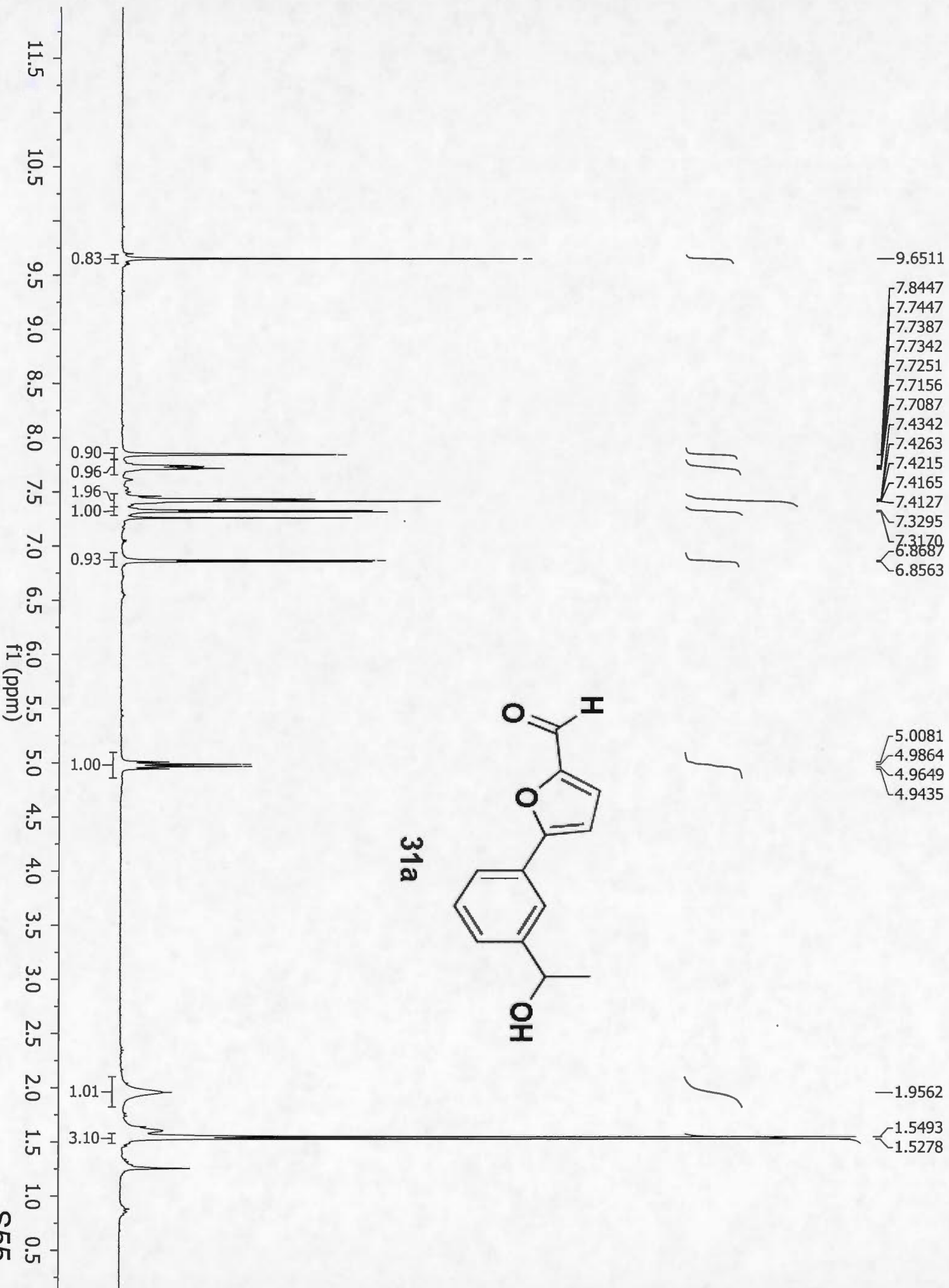
30d

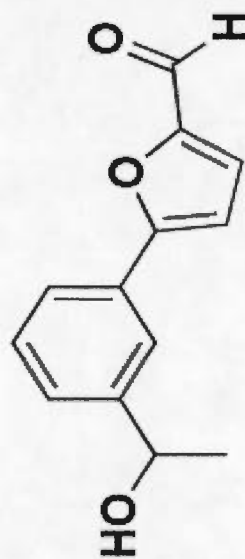


S53

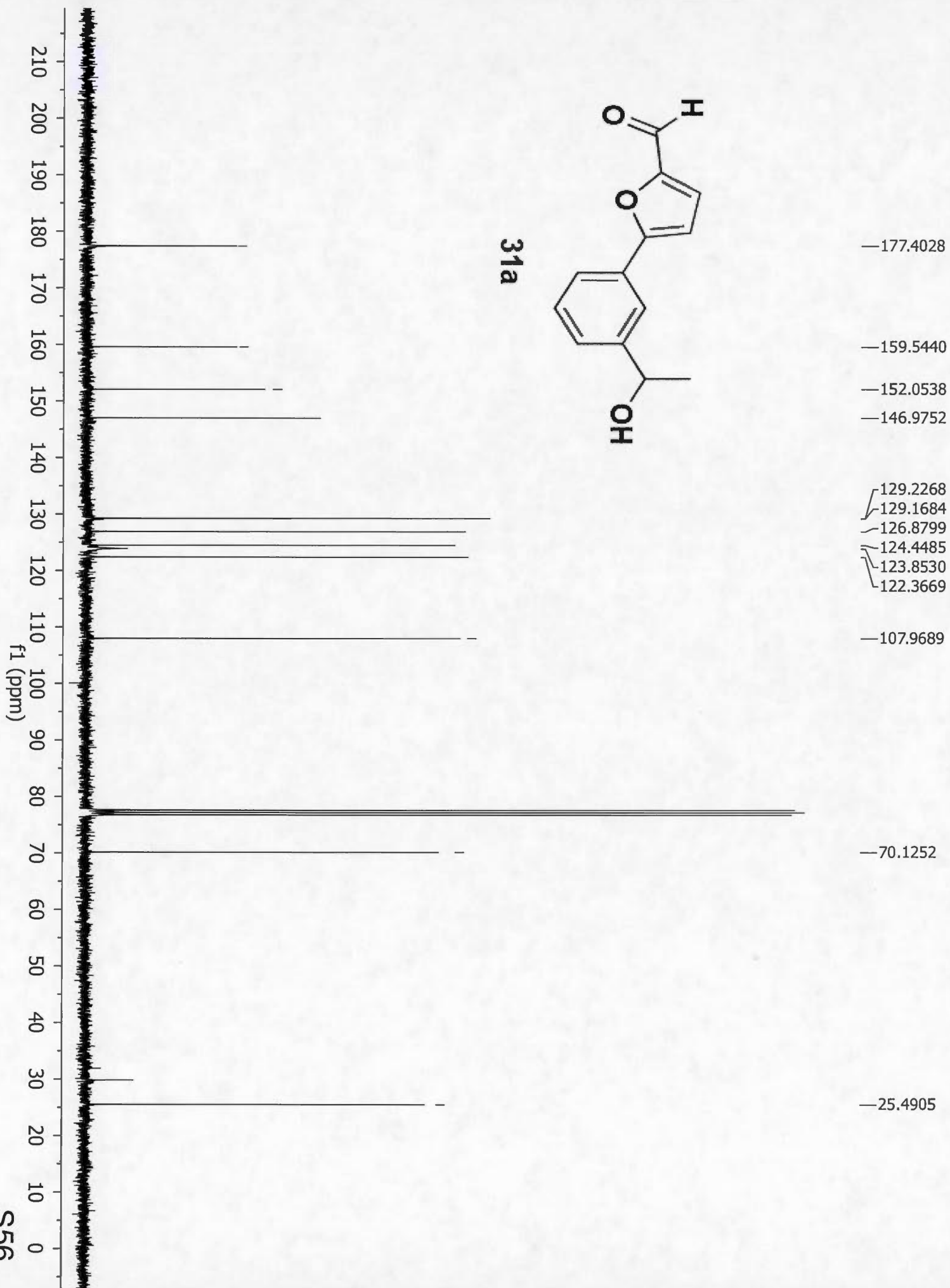


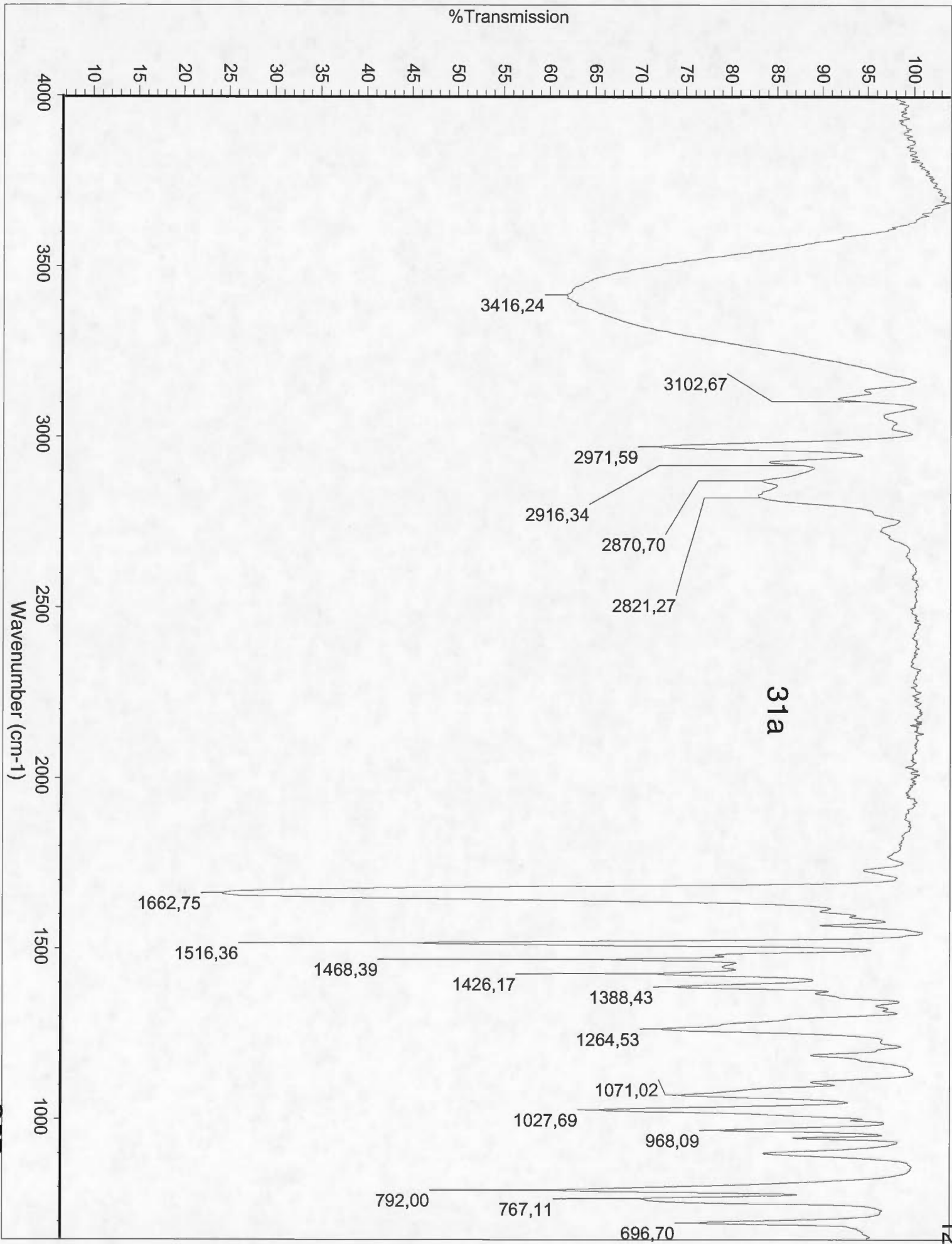


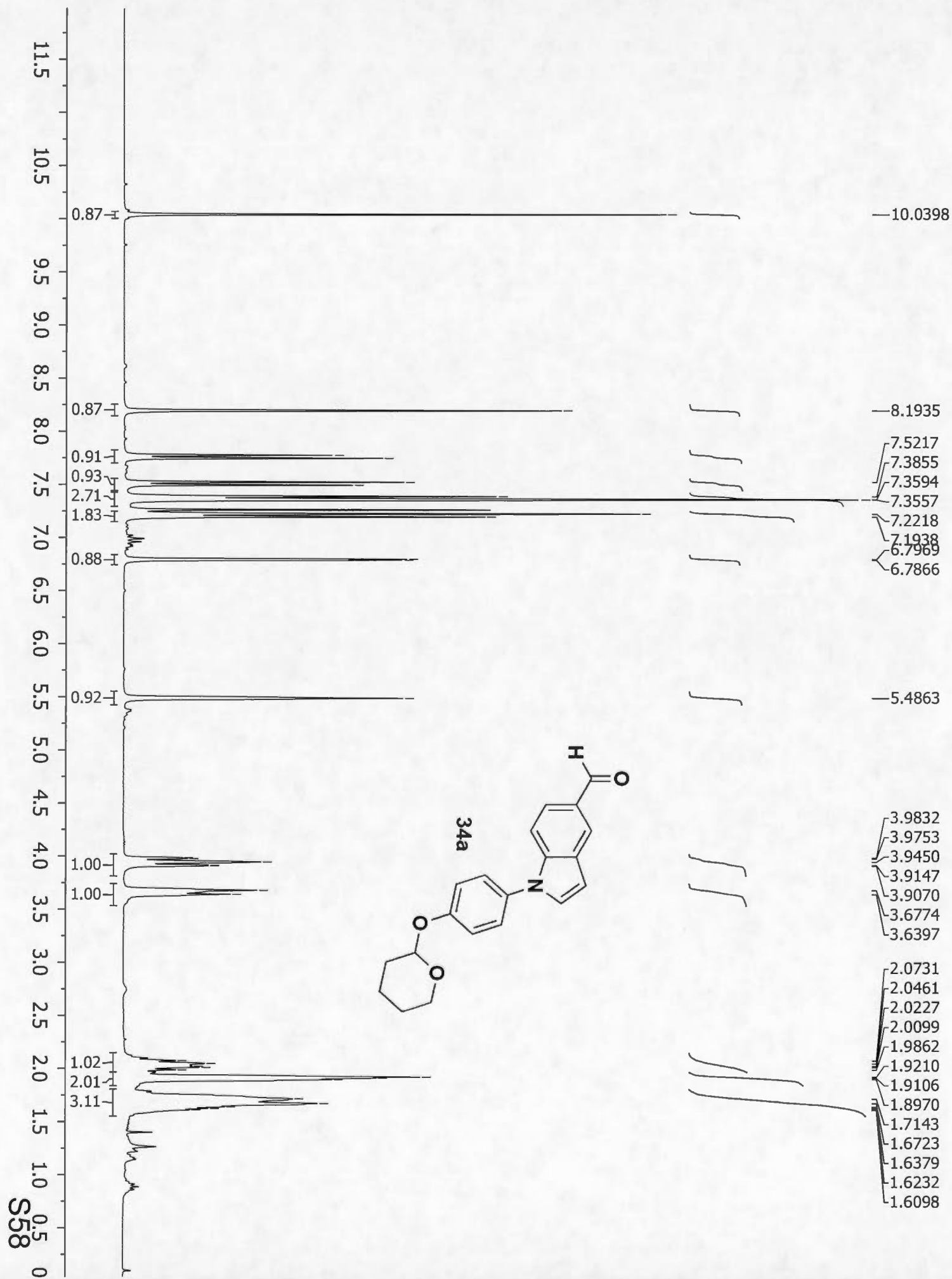




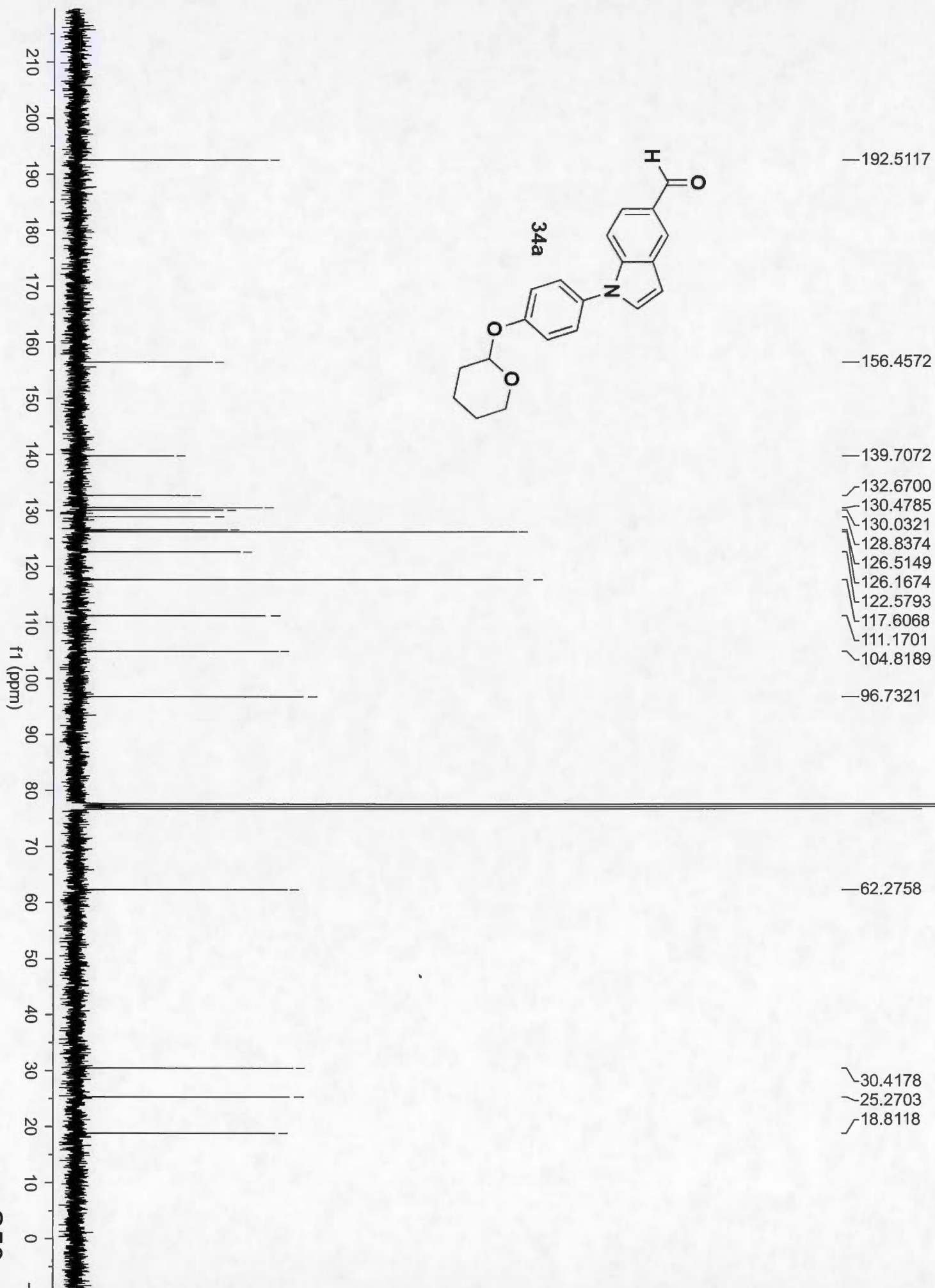
**31a**

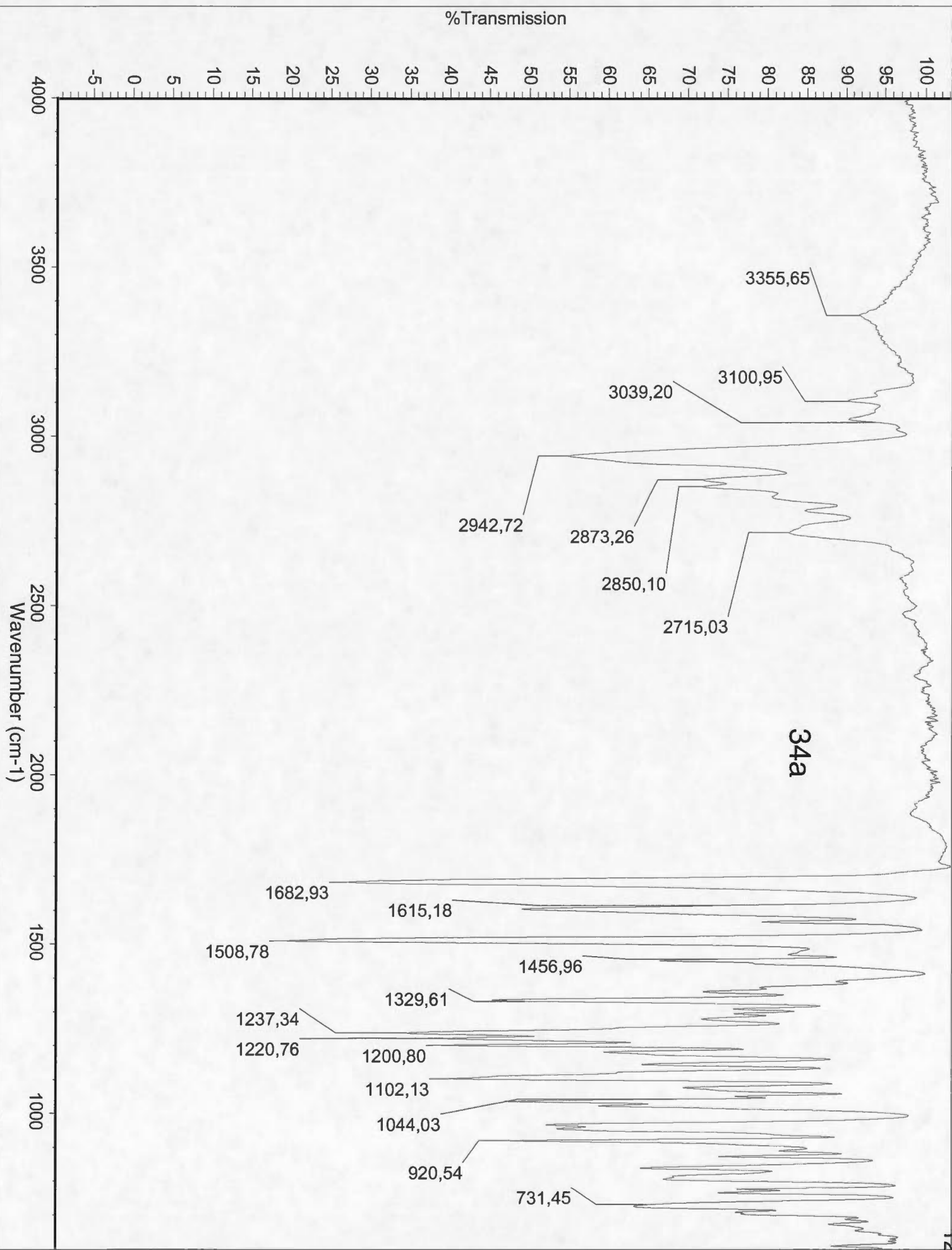


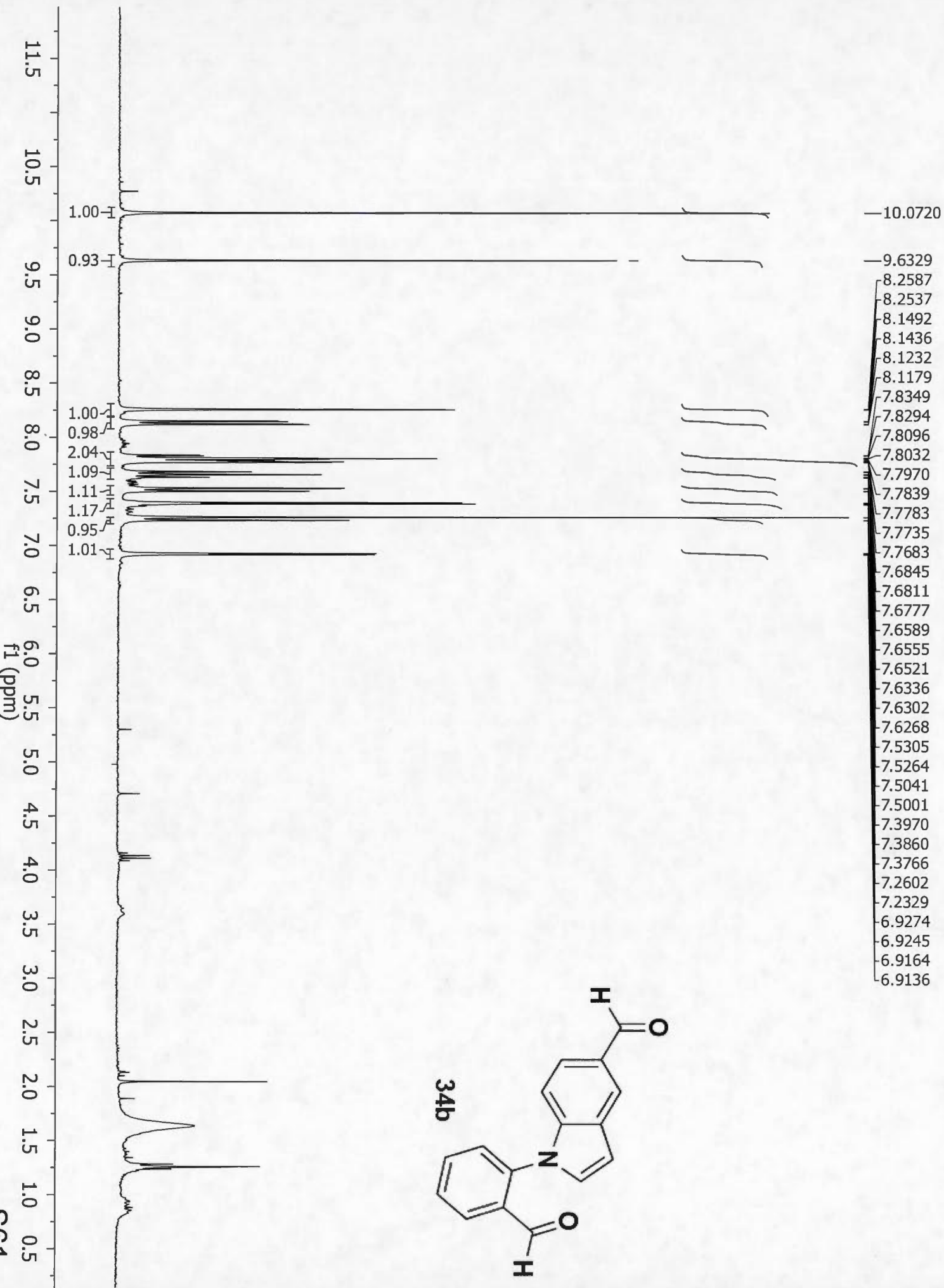


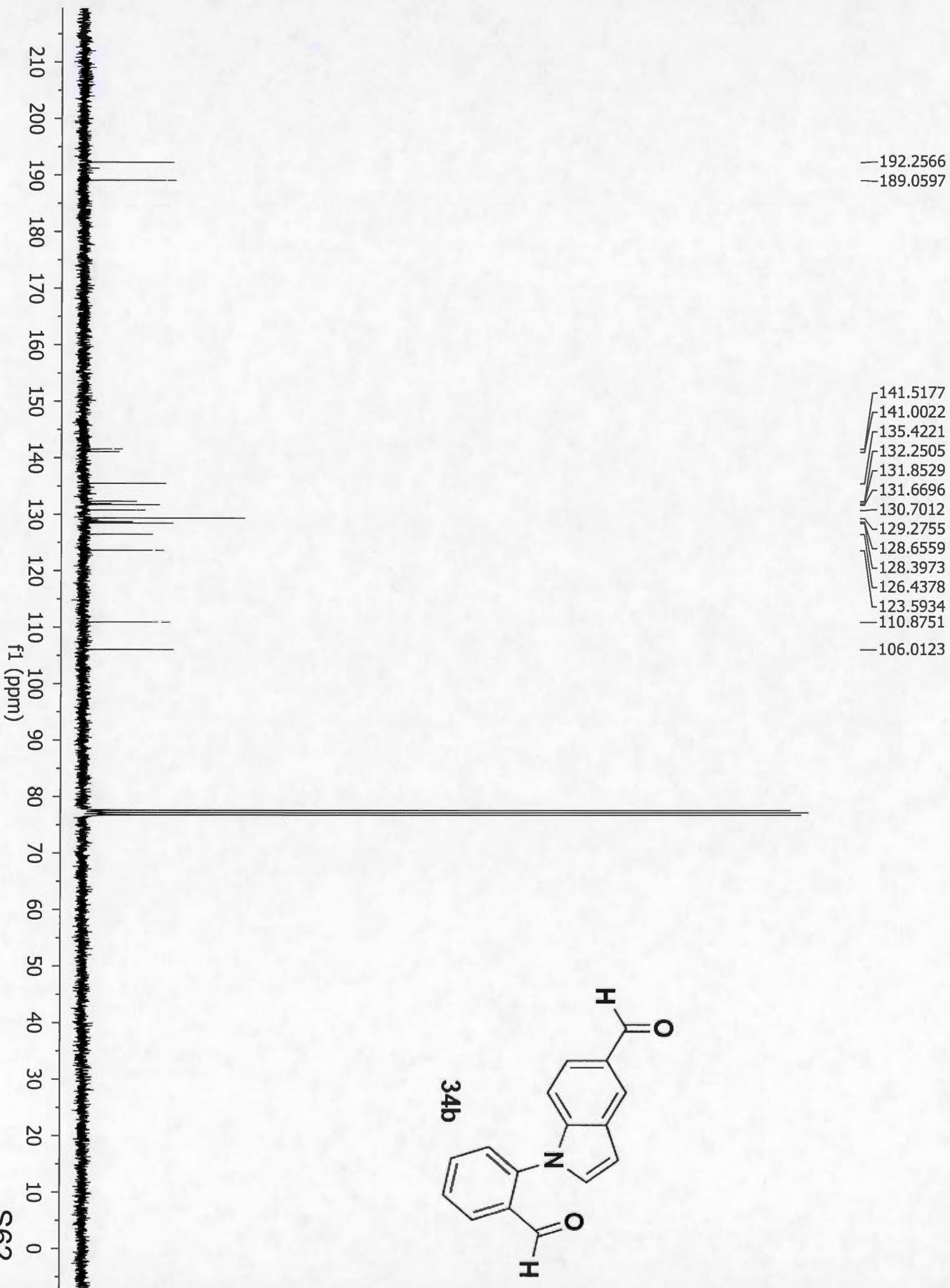
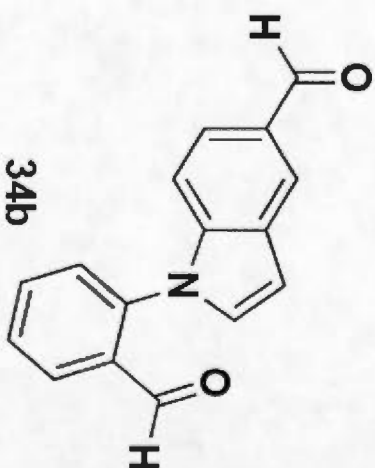




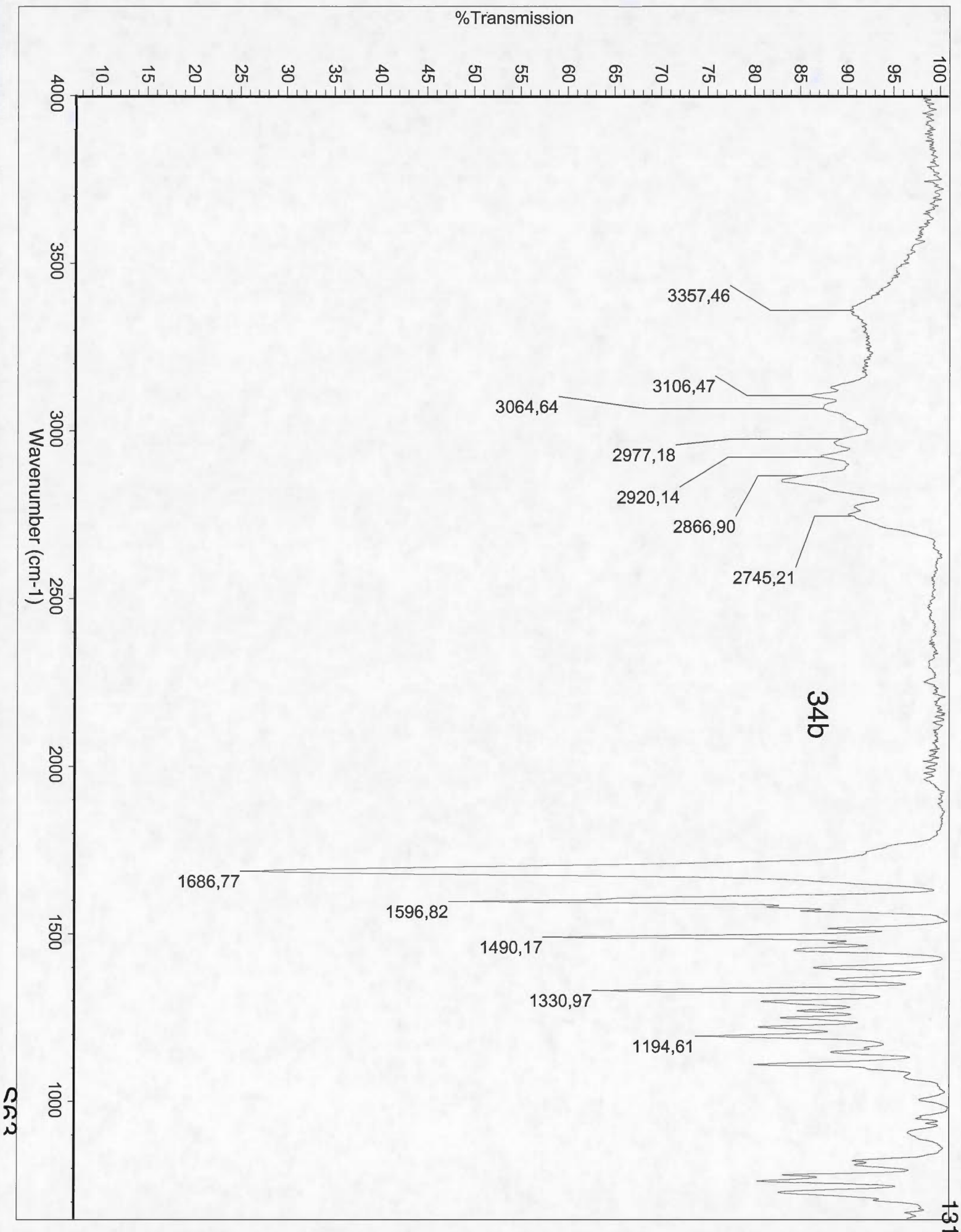


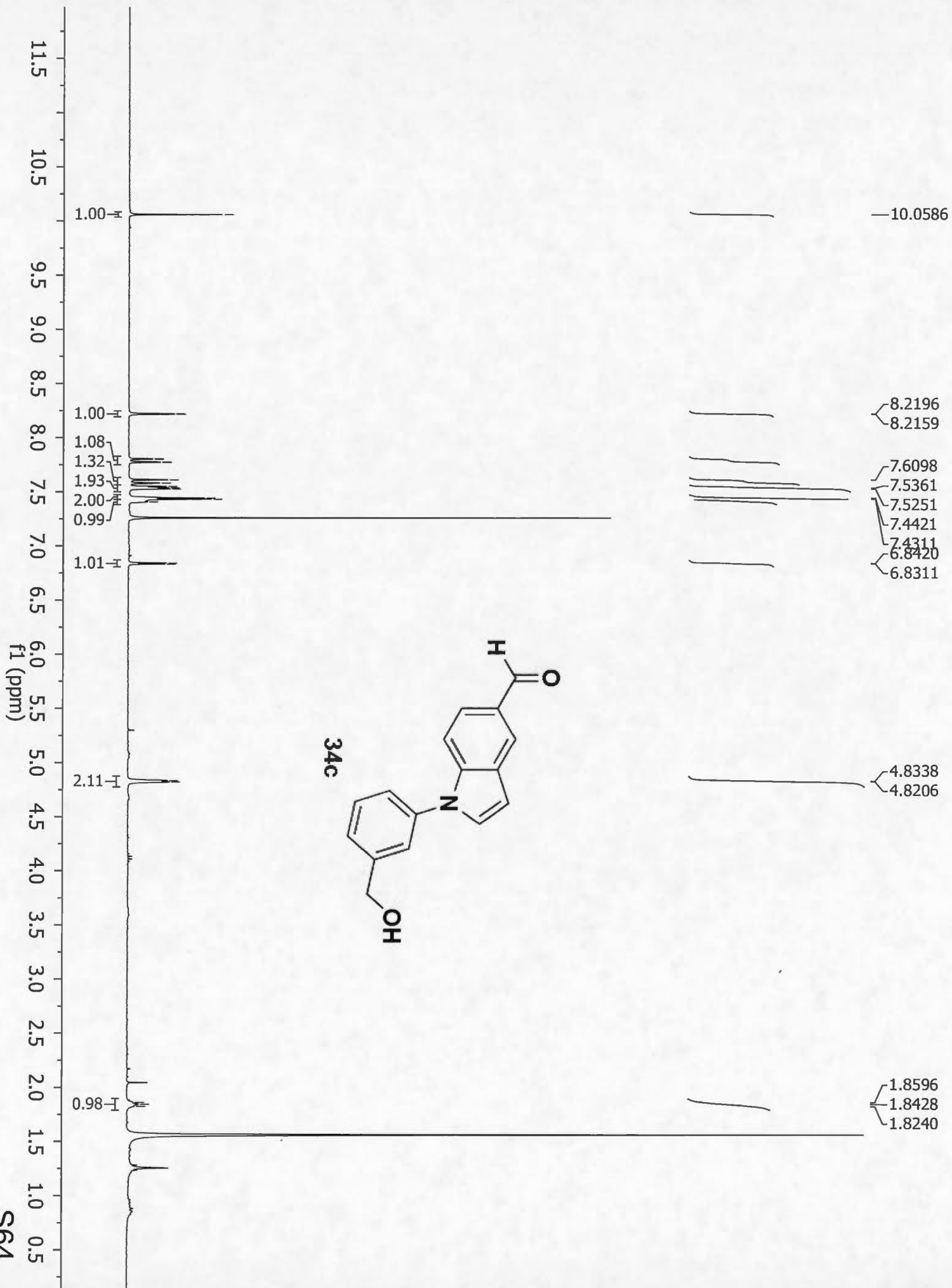


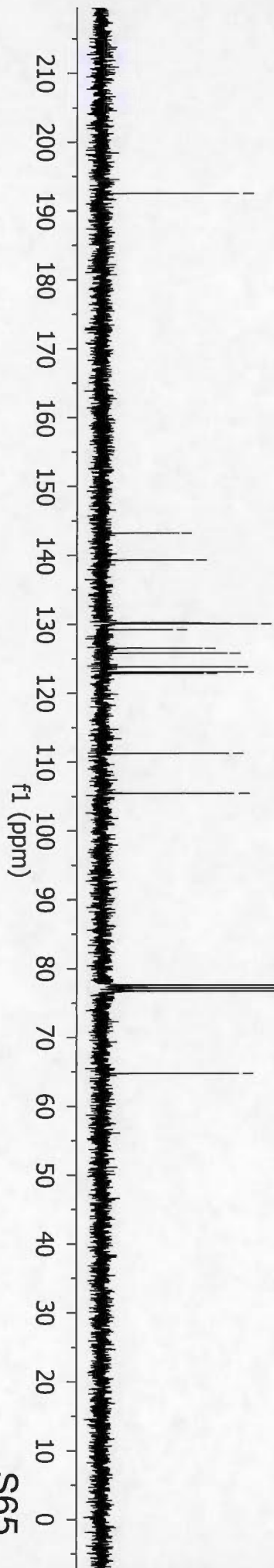
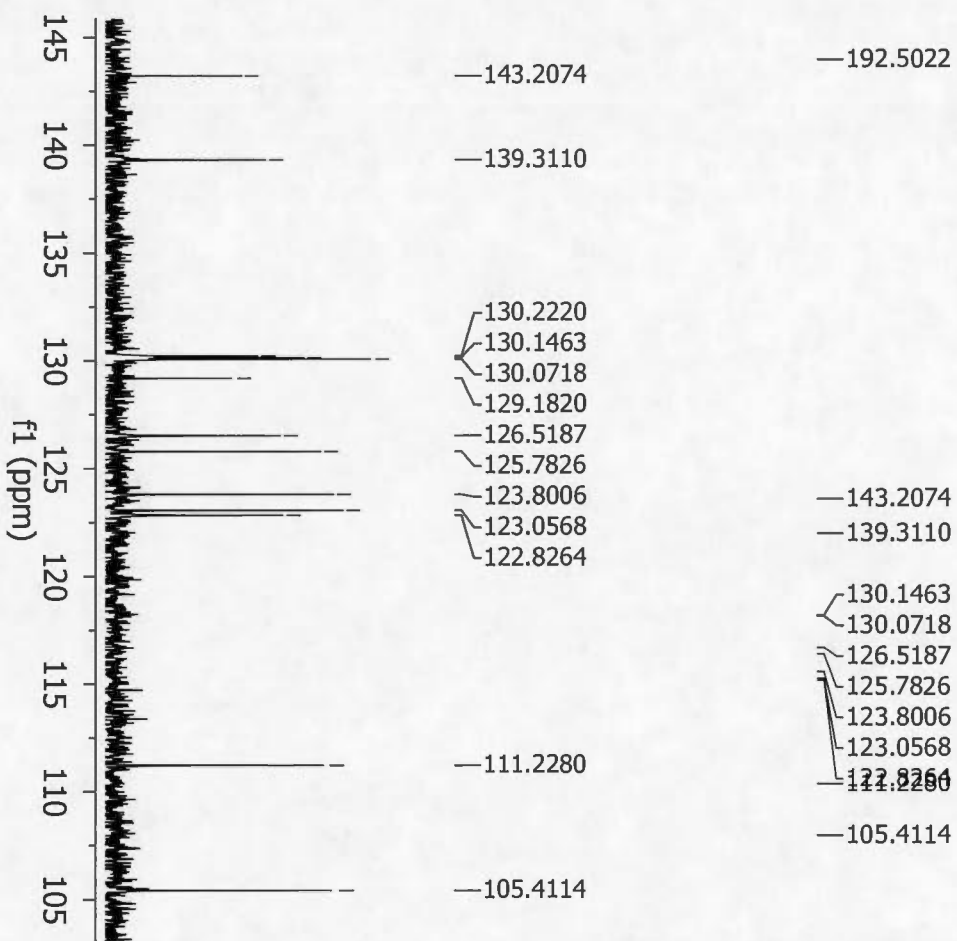
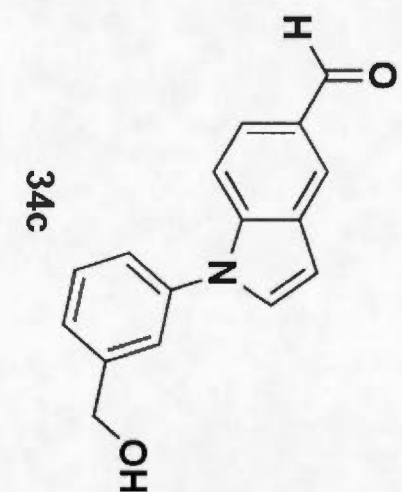


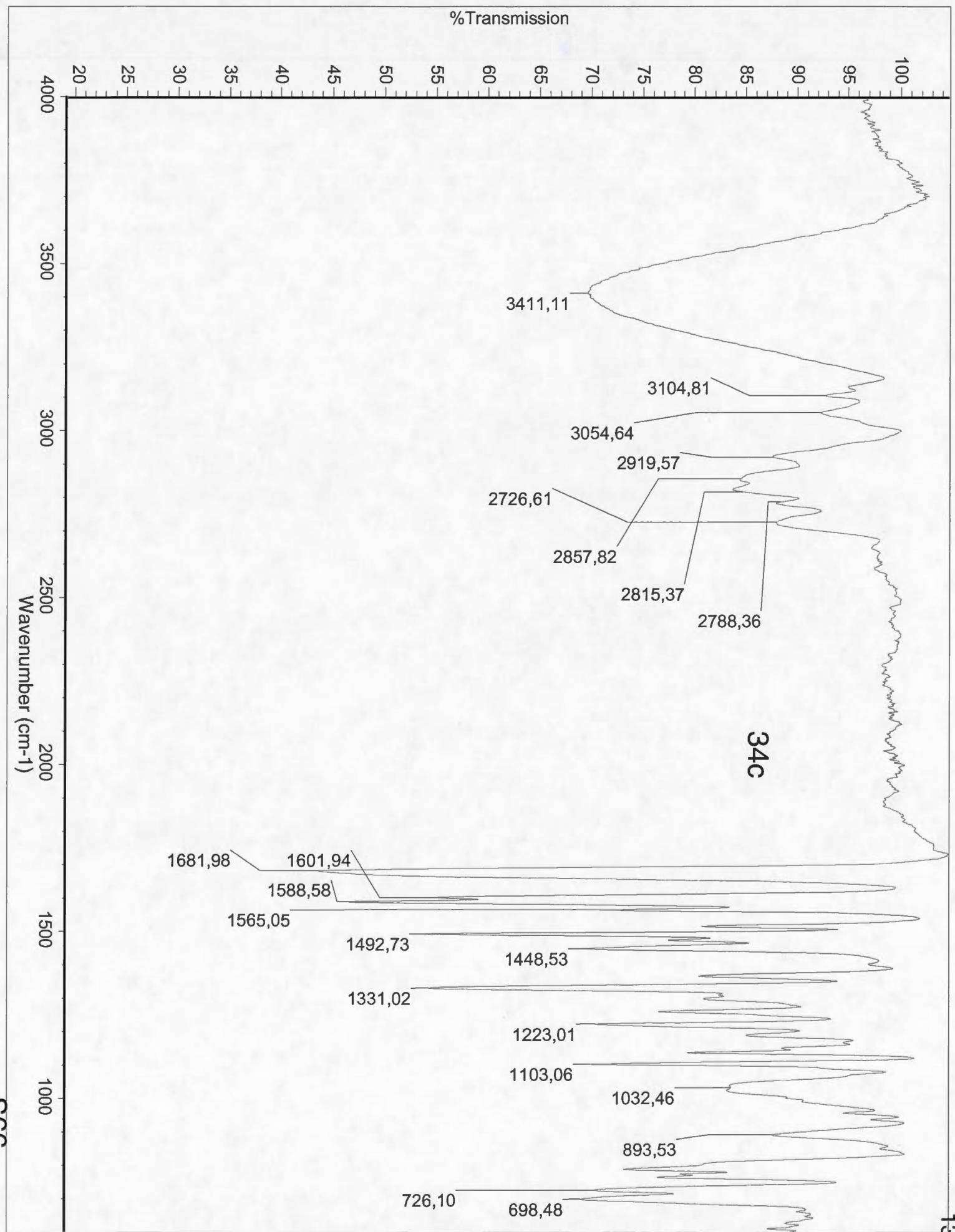




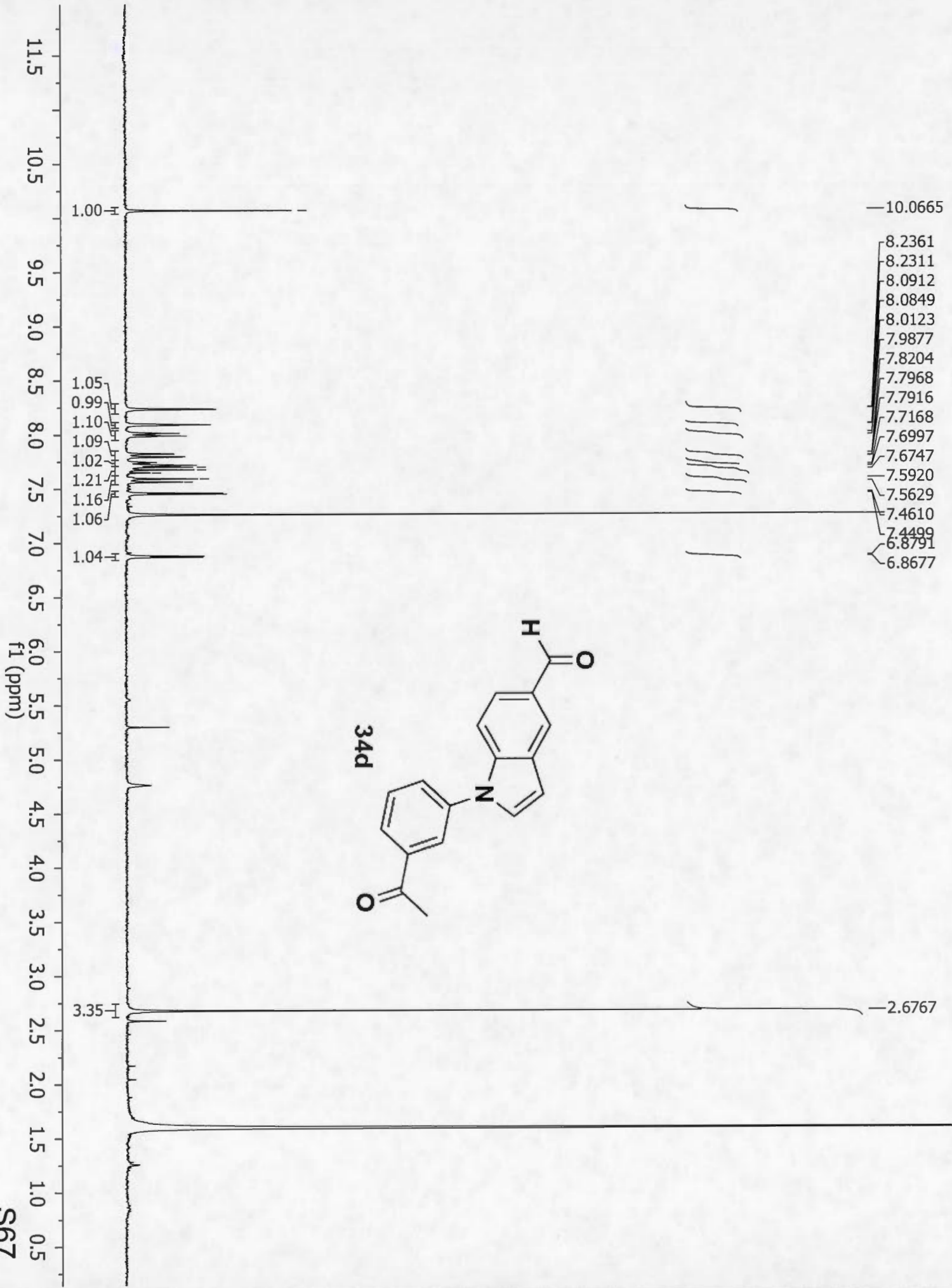


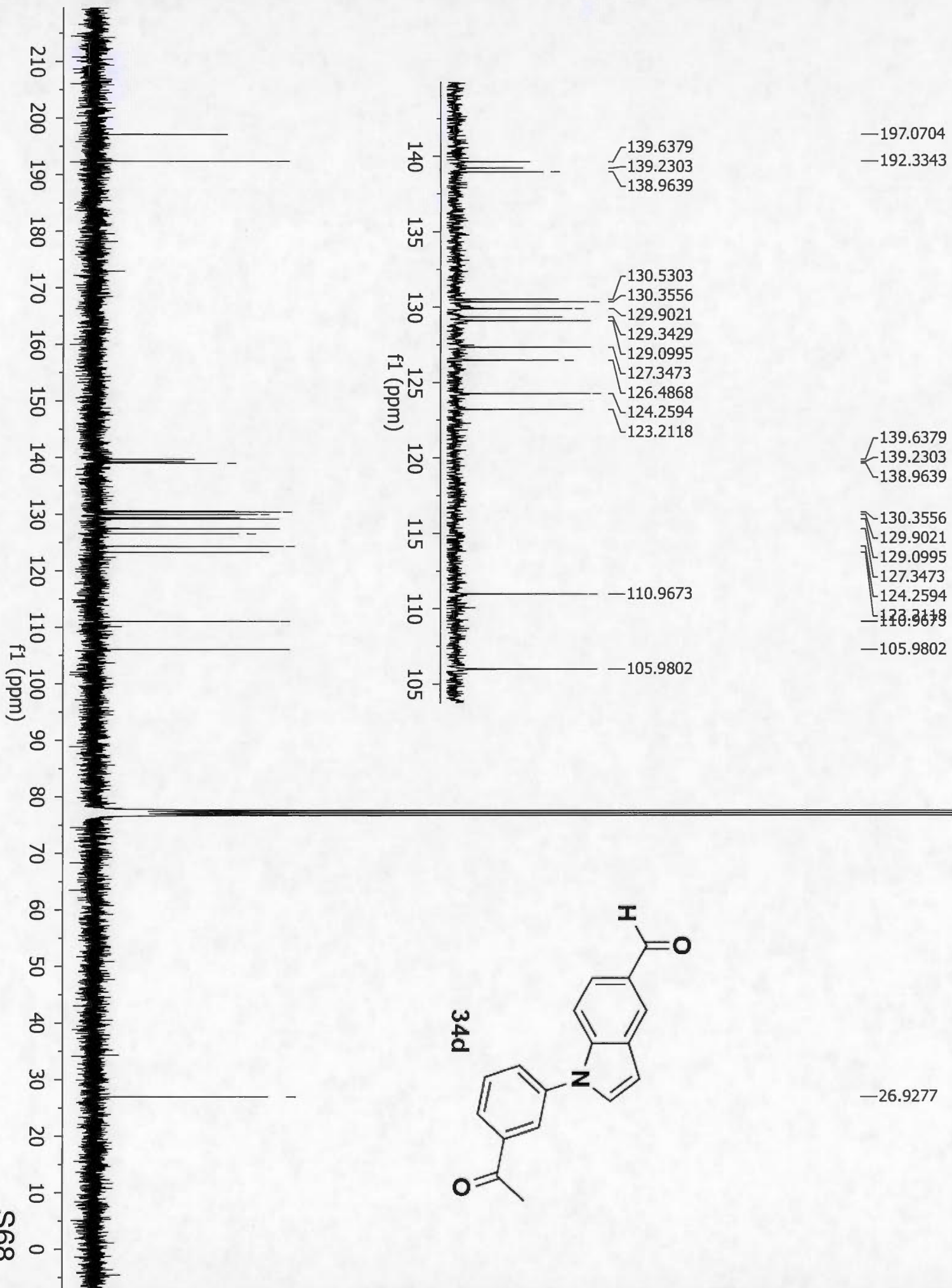


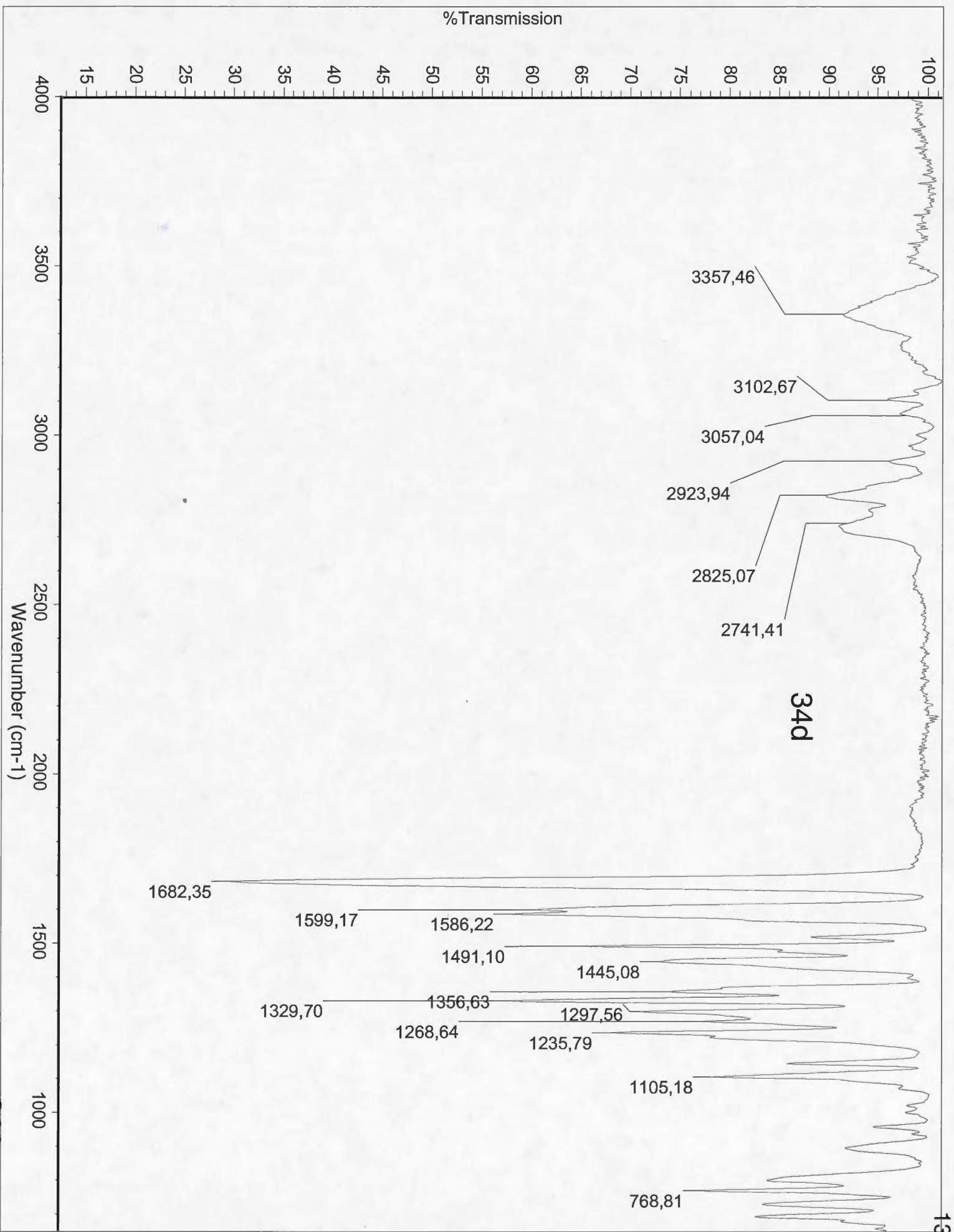


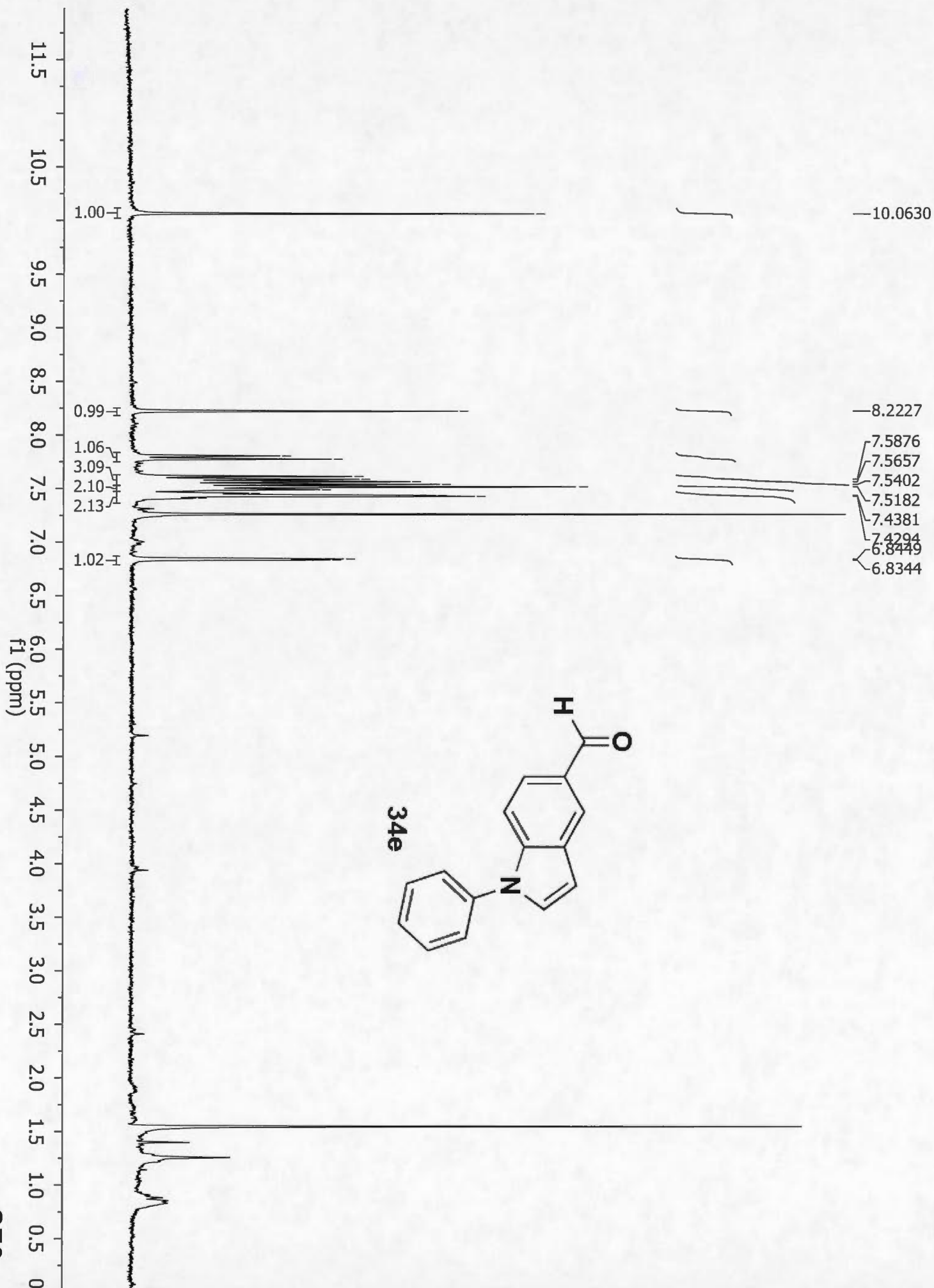




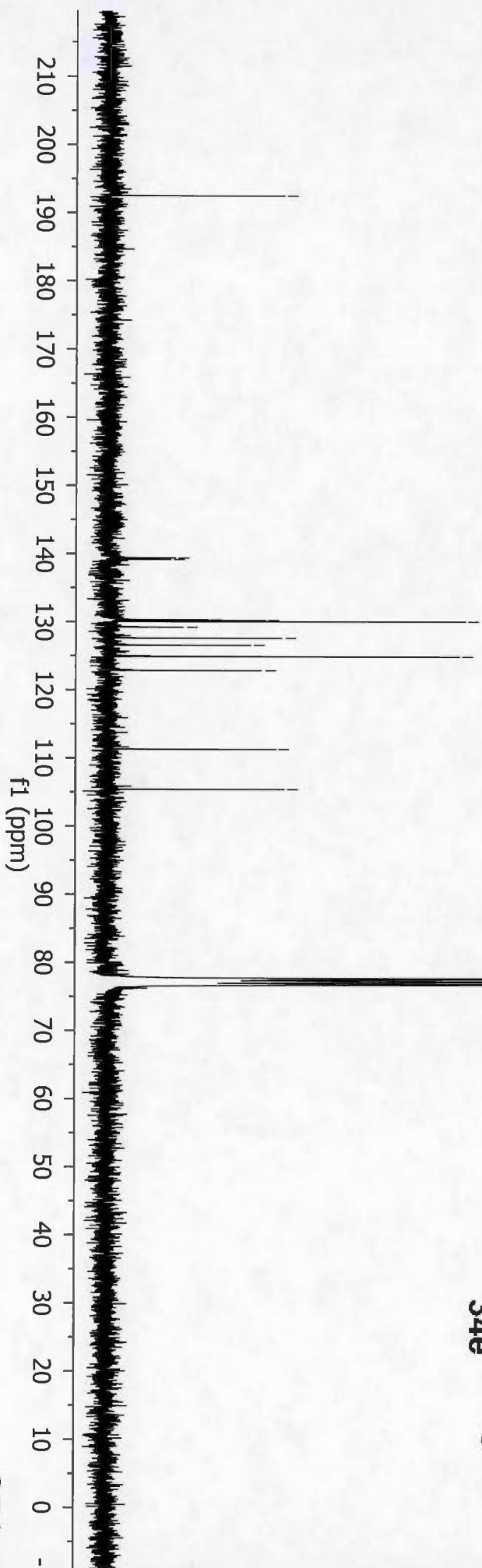
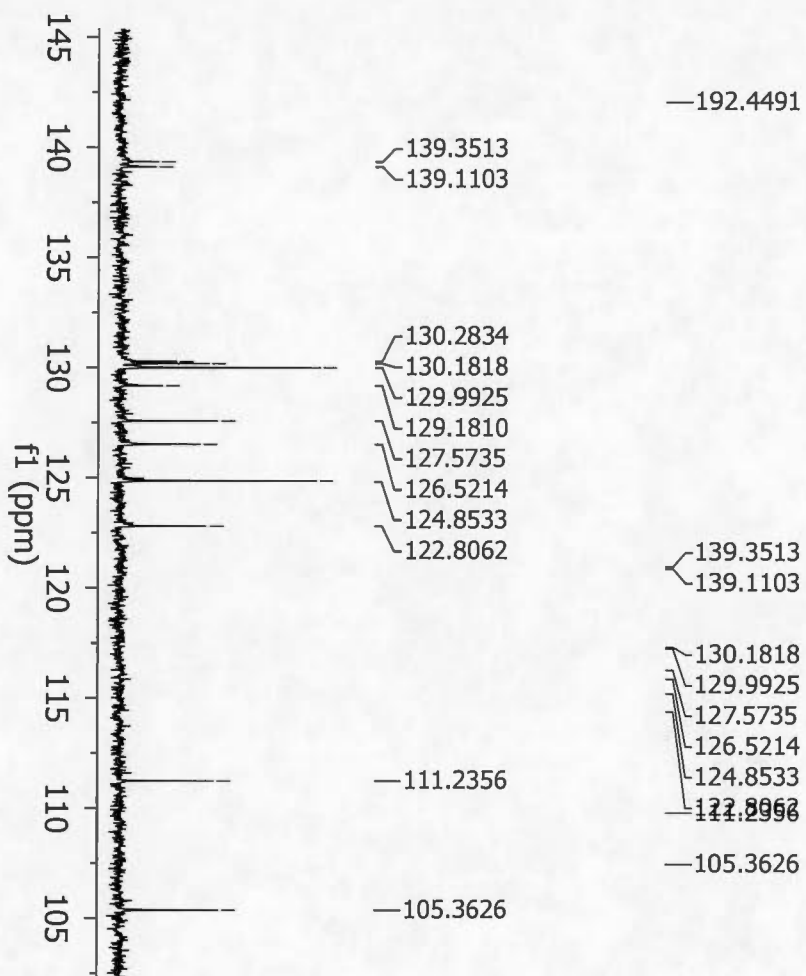
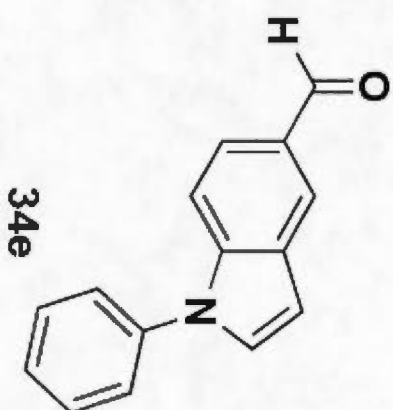


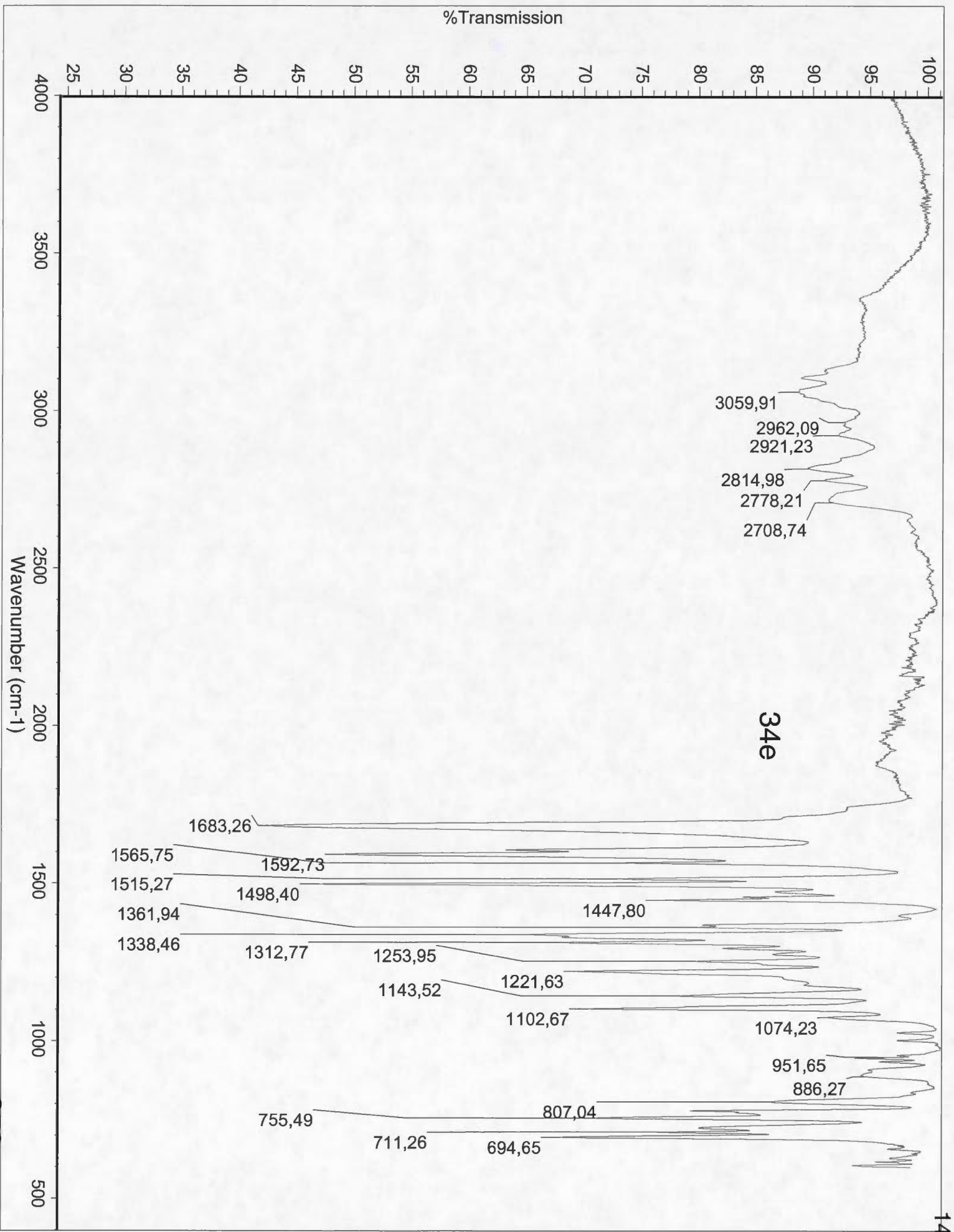


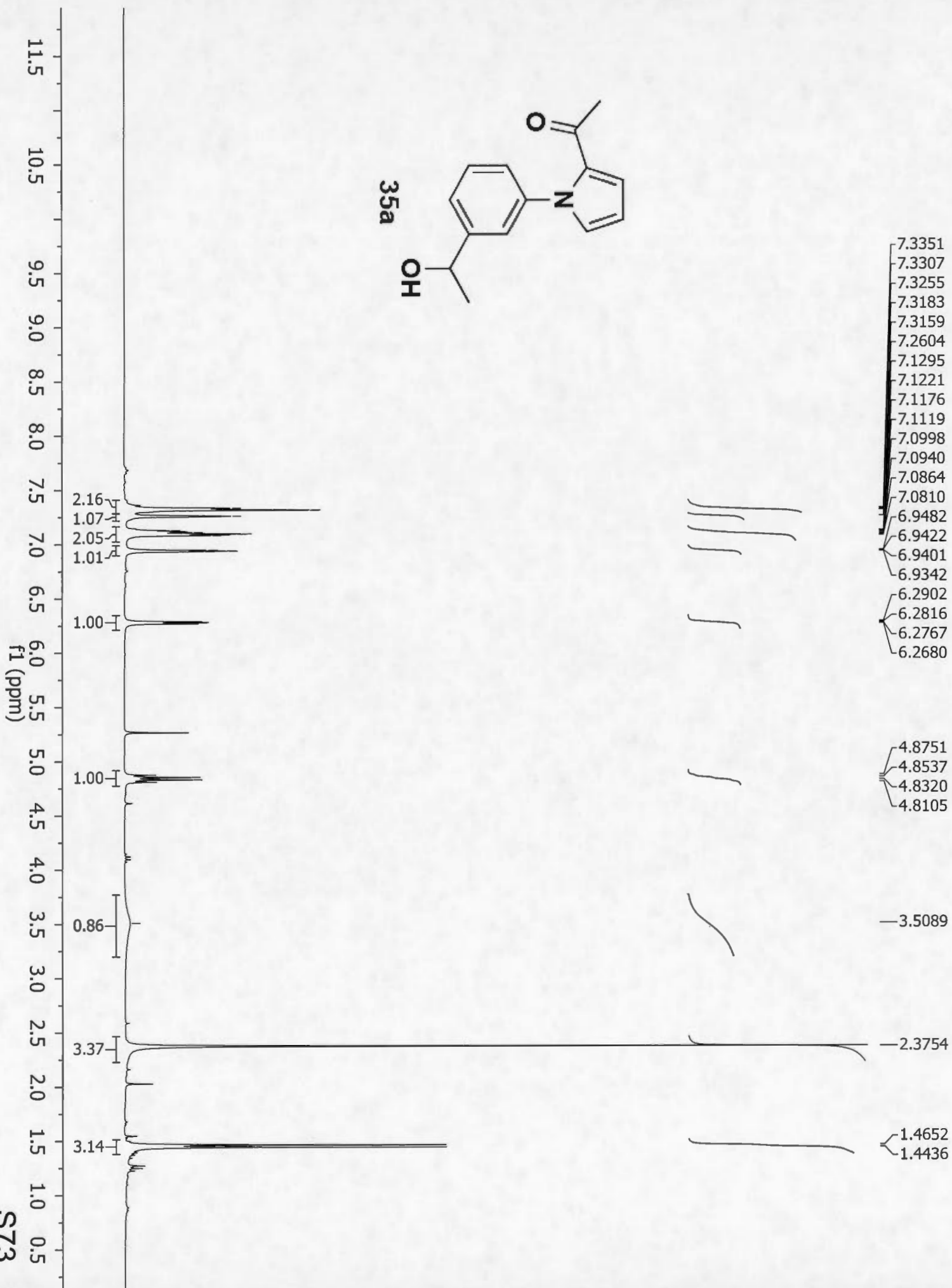
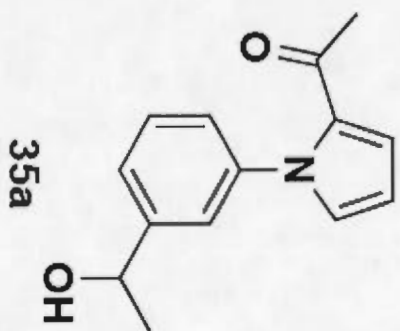


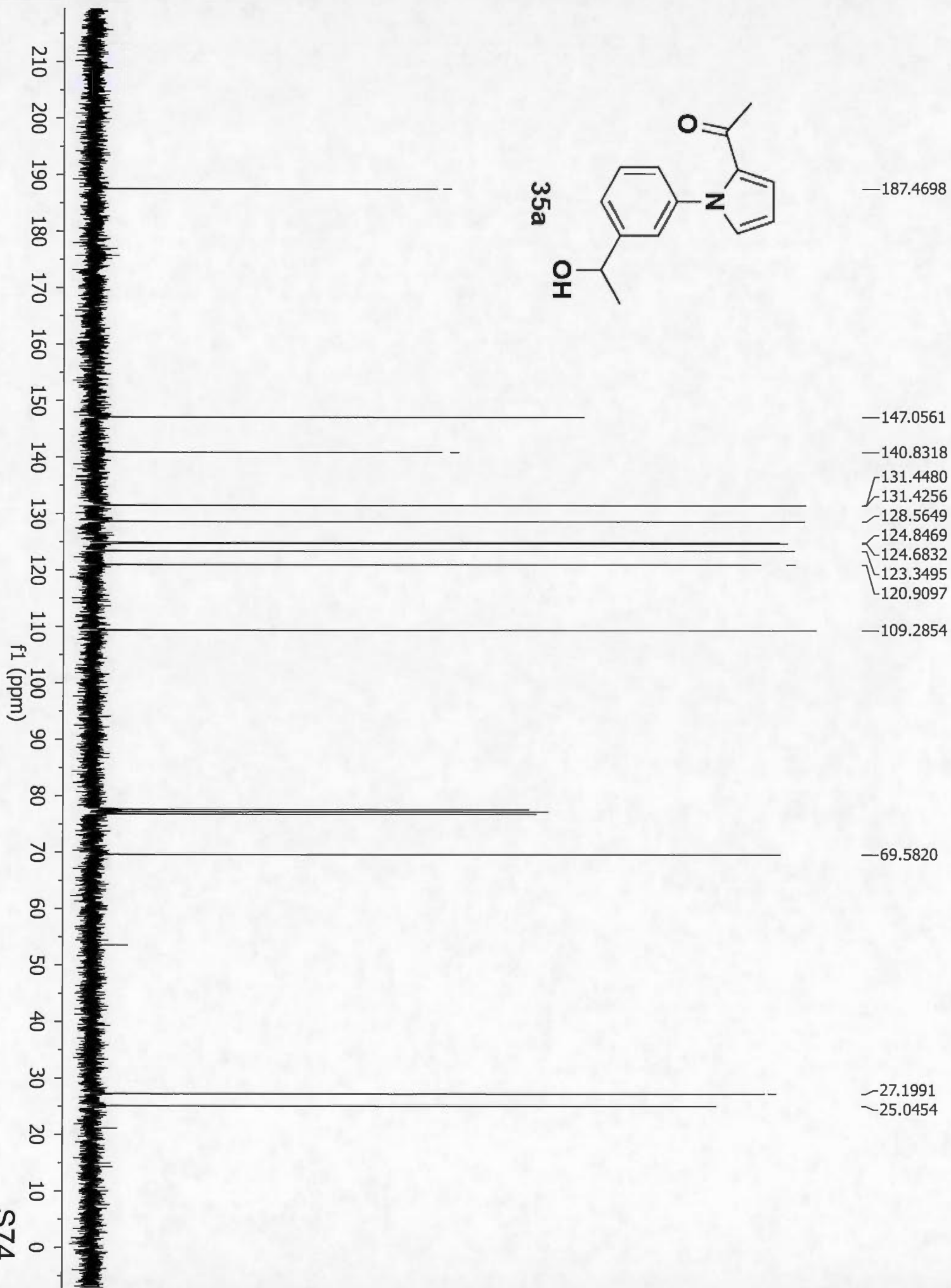
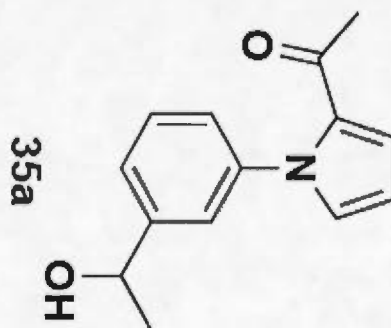




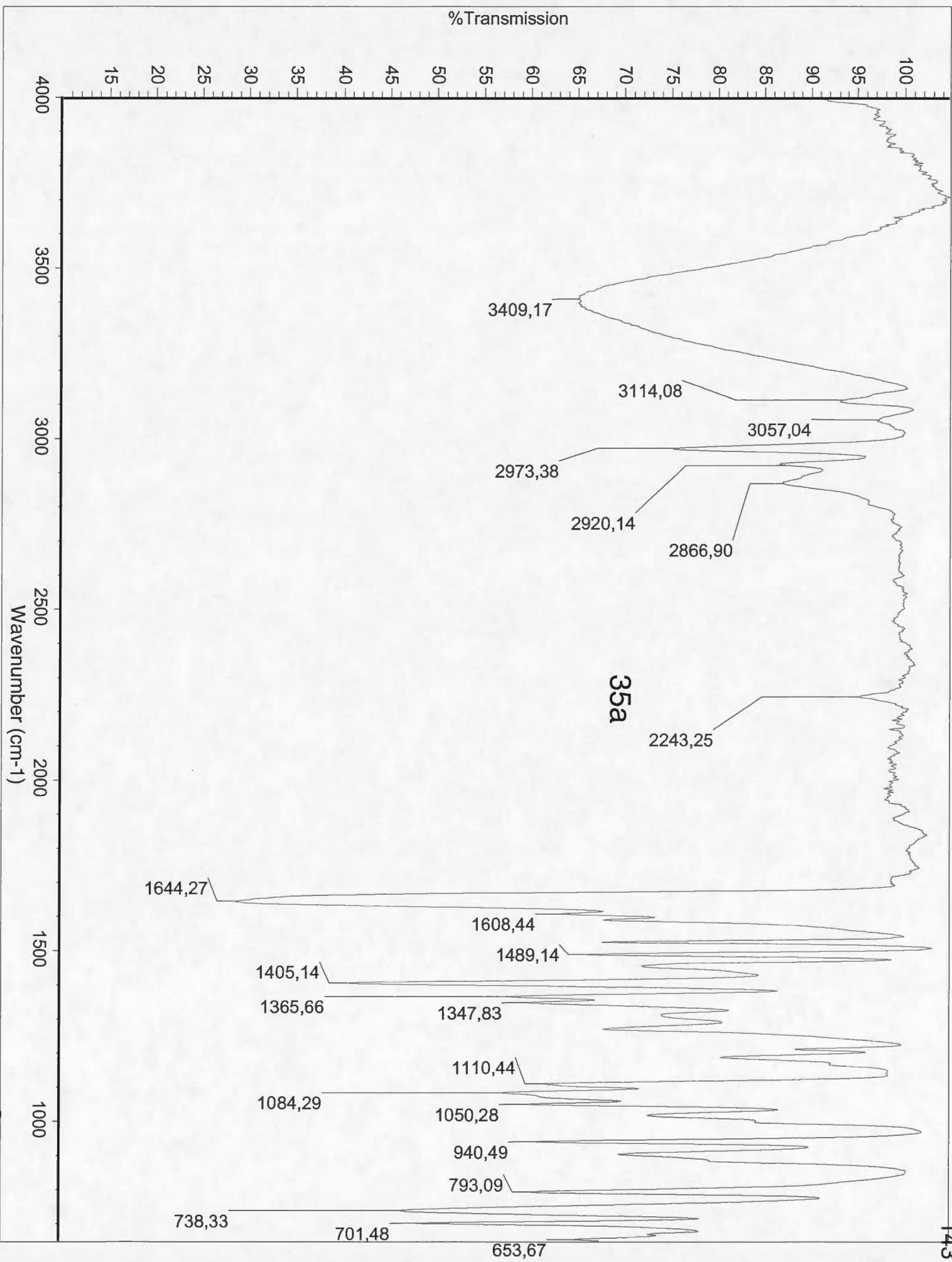




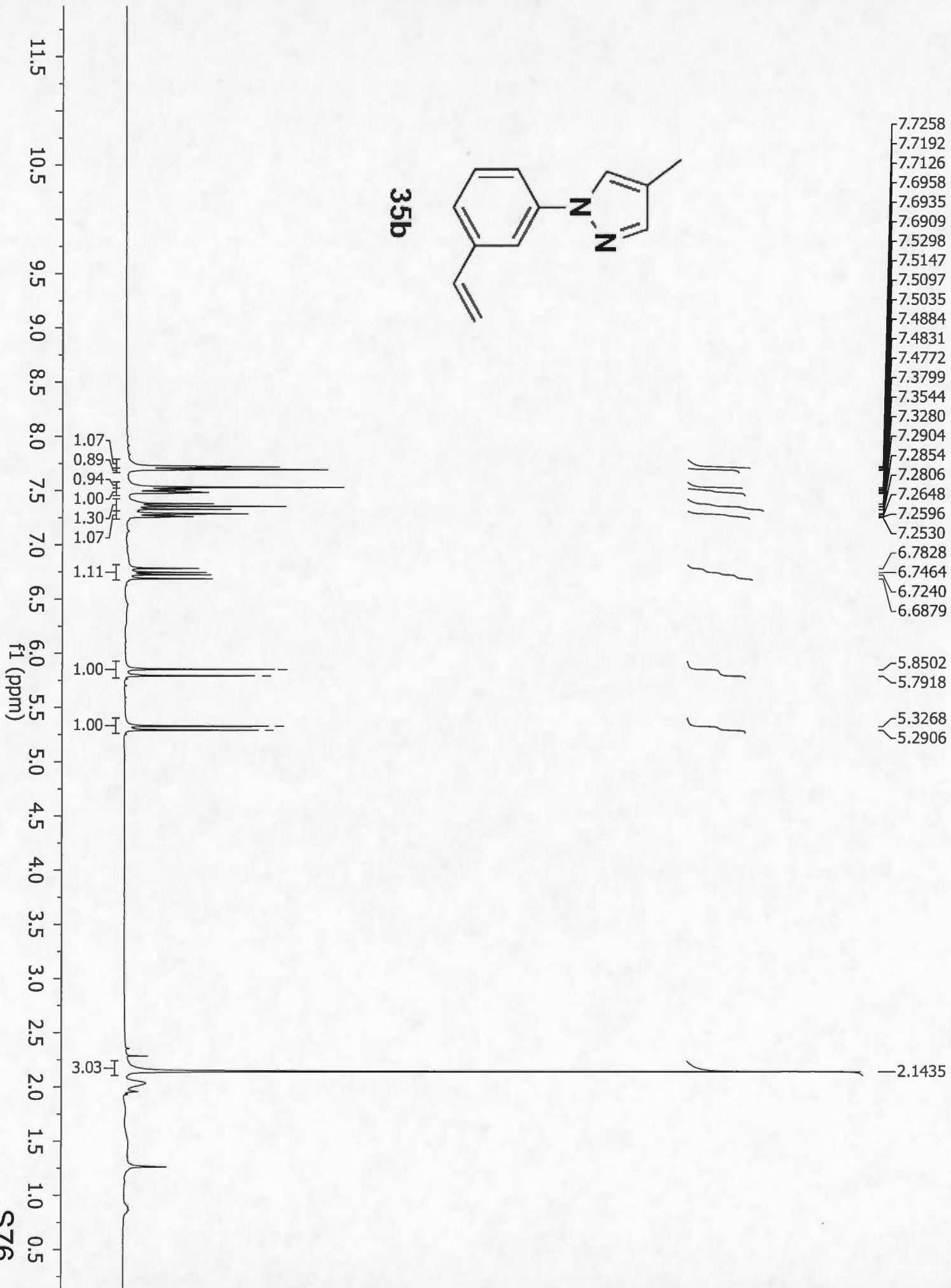
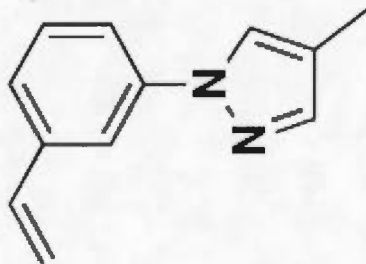




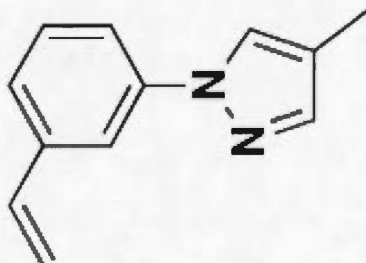




35b



35b

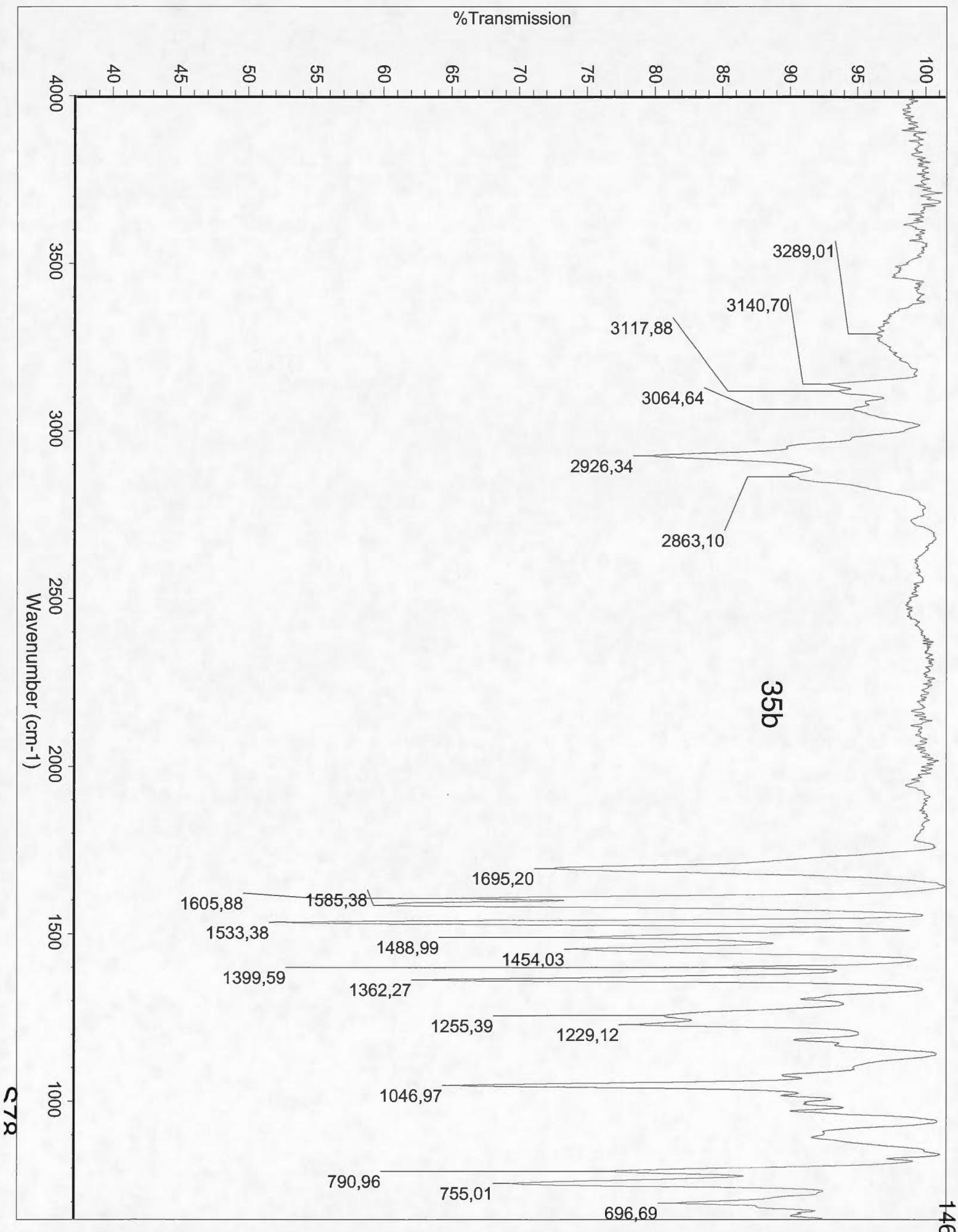


141.9116  
140.6495  
139.0503  
136.2946  
129.5525  
125.4962  
123.9293  
118.3510  
118.0213  
116.6479  
115.1449

—9.0582

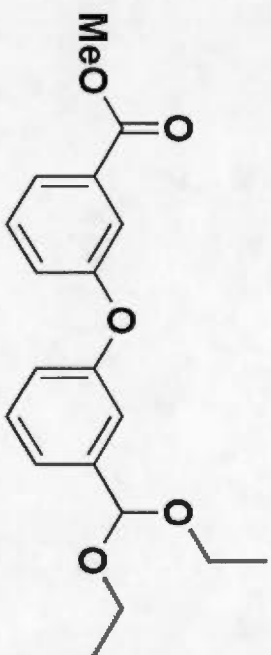
f1 (ppm)

S77

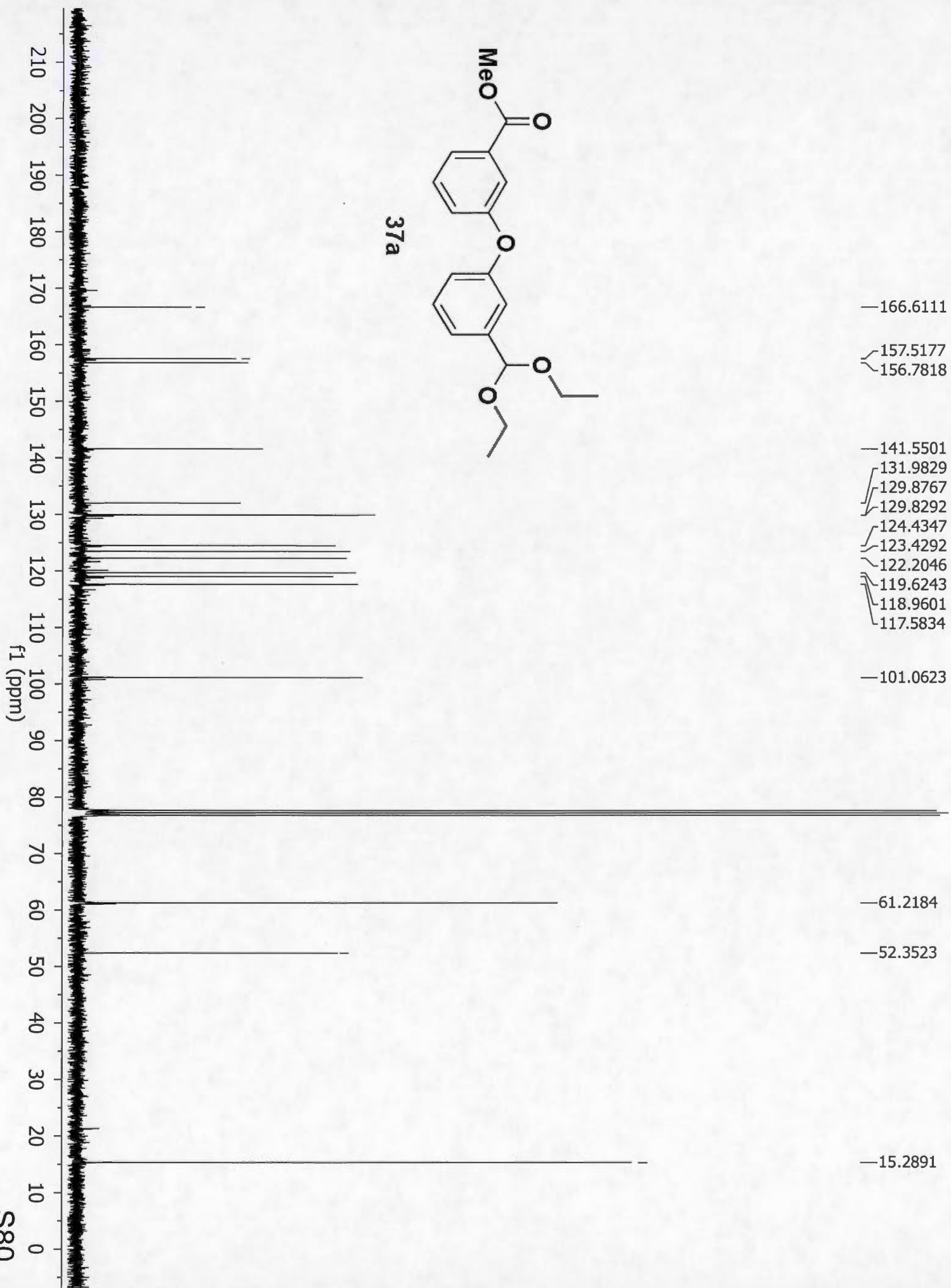


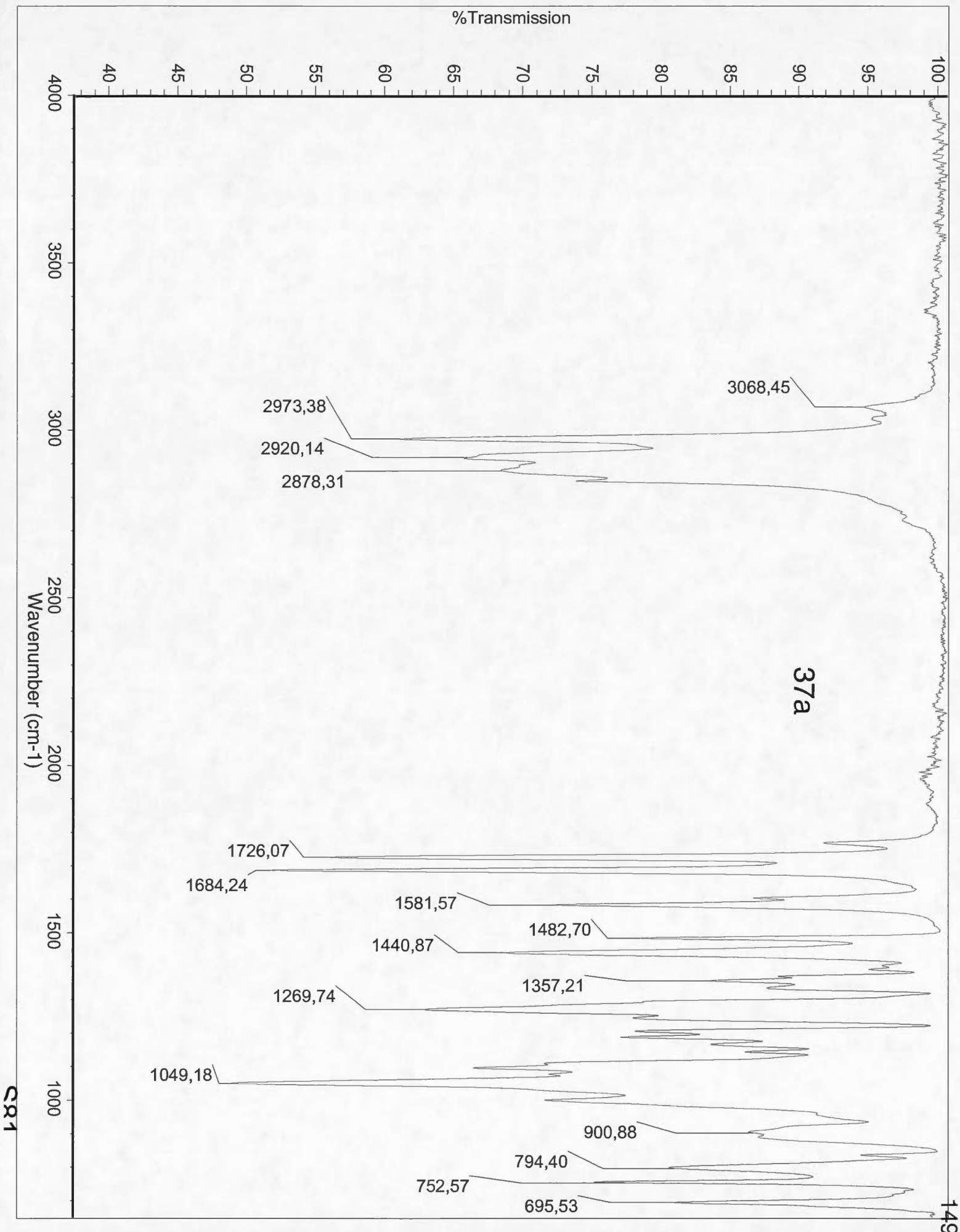


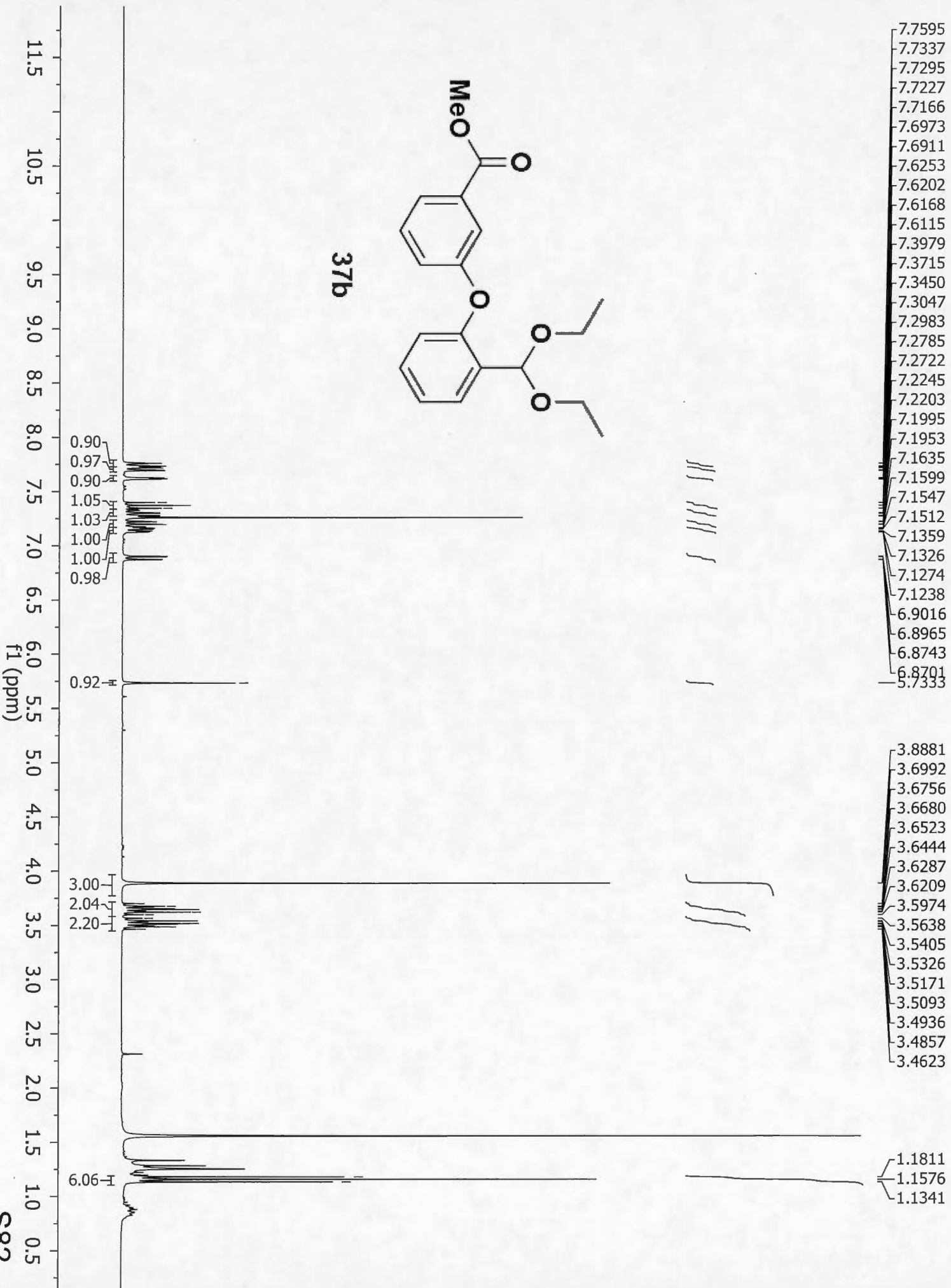




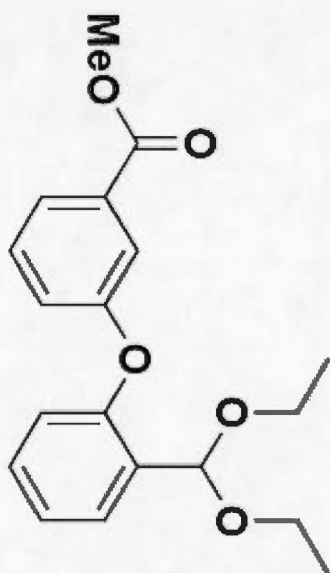
37a



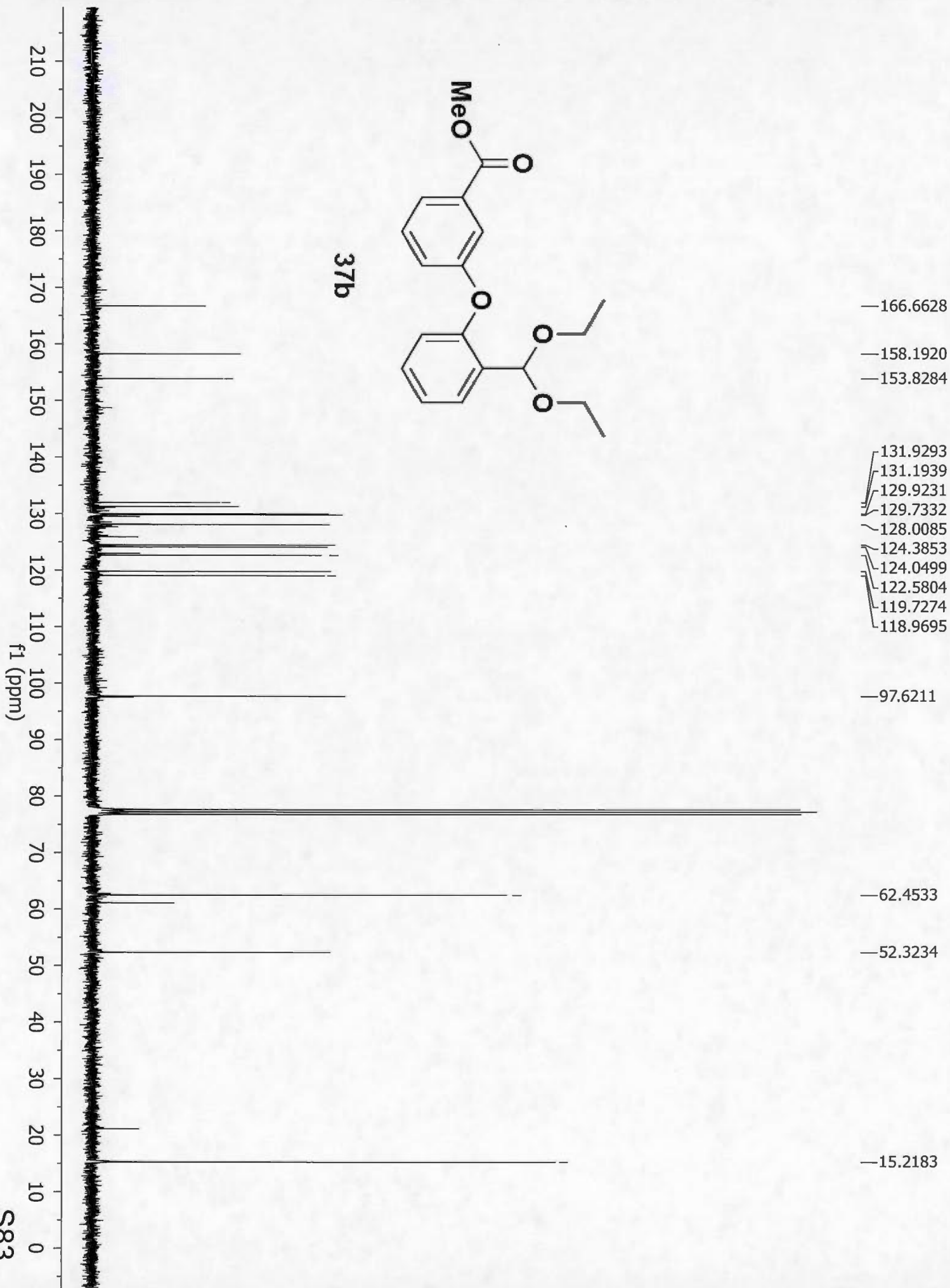


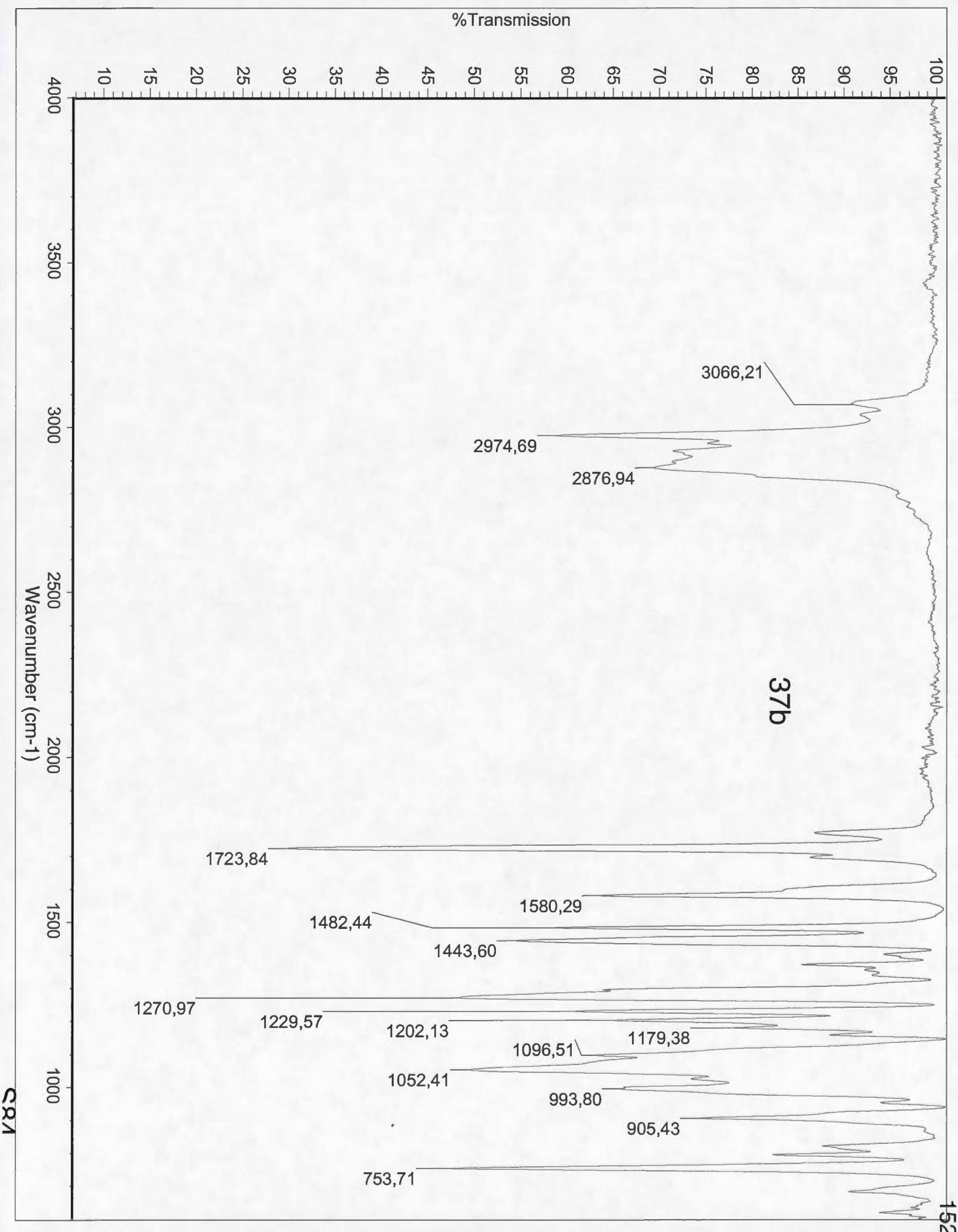


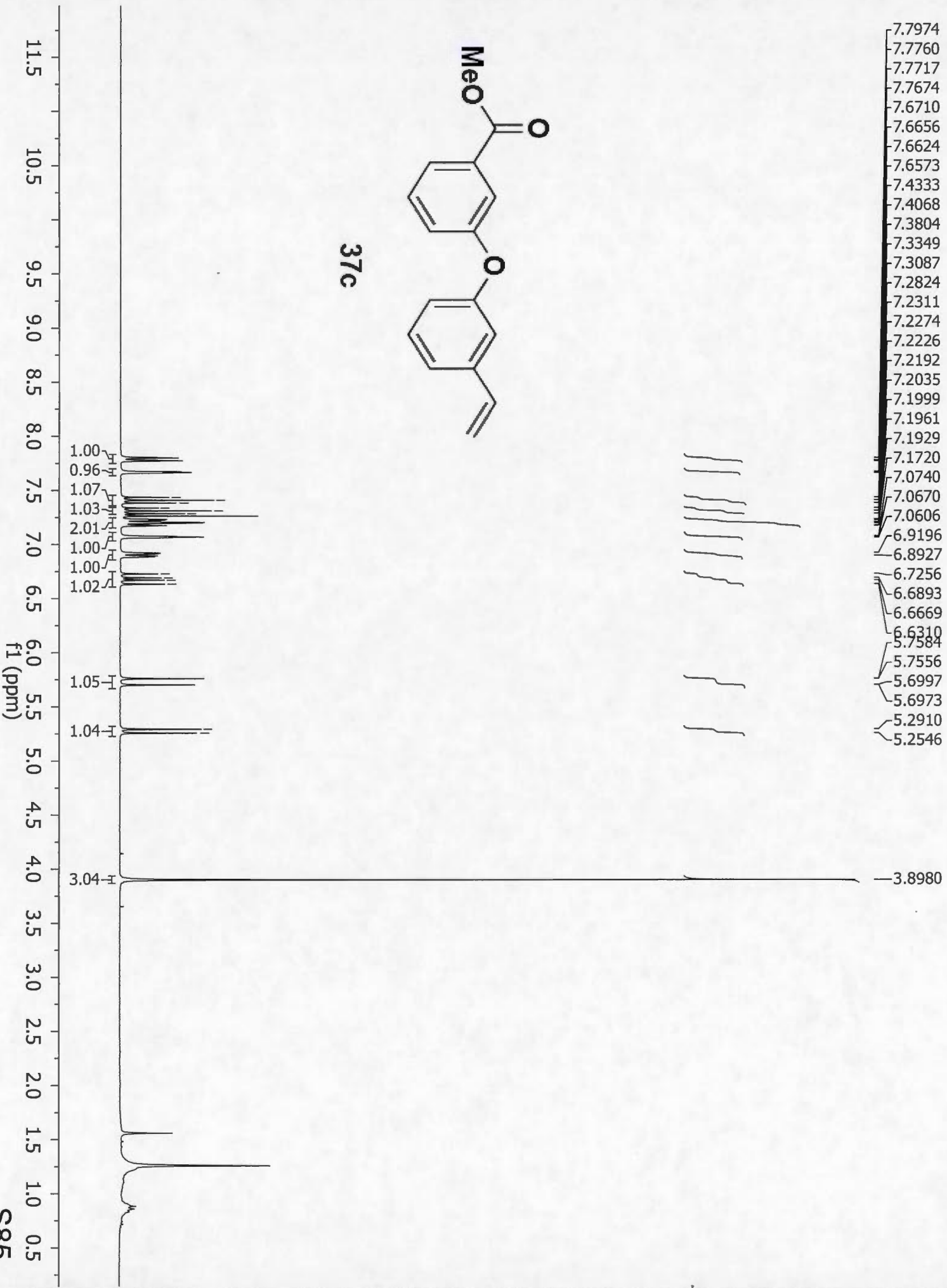


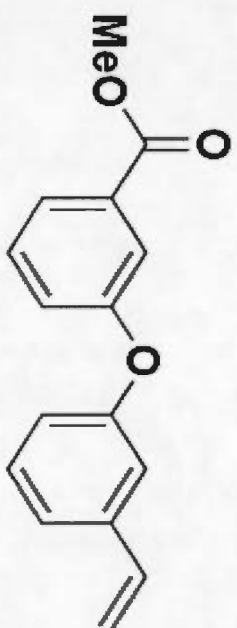


37b

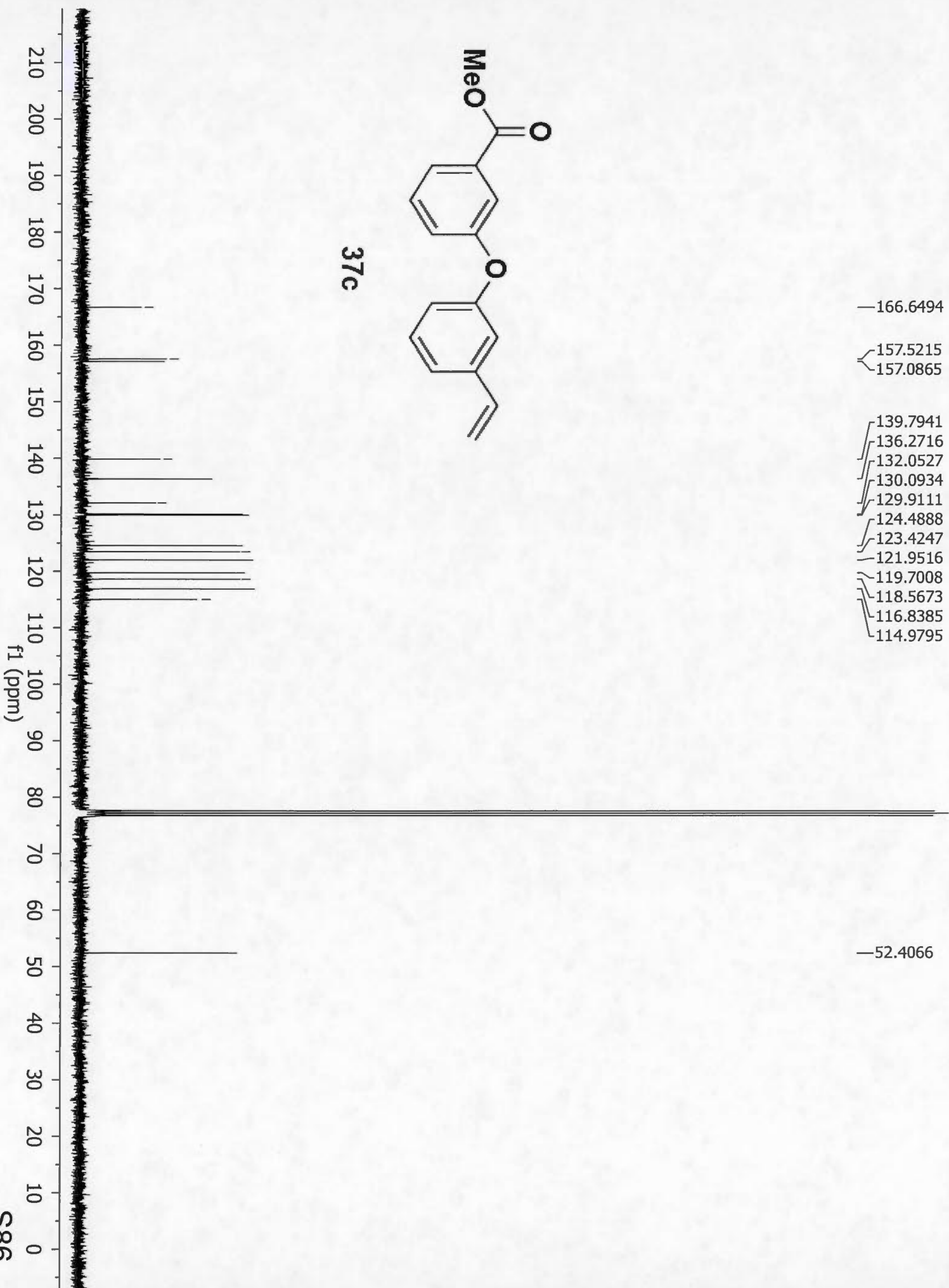




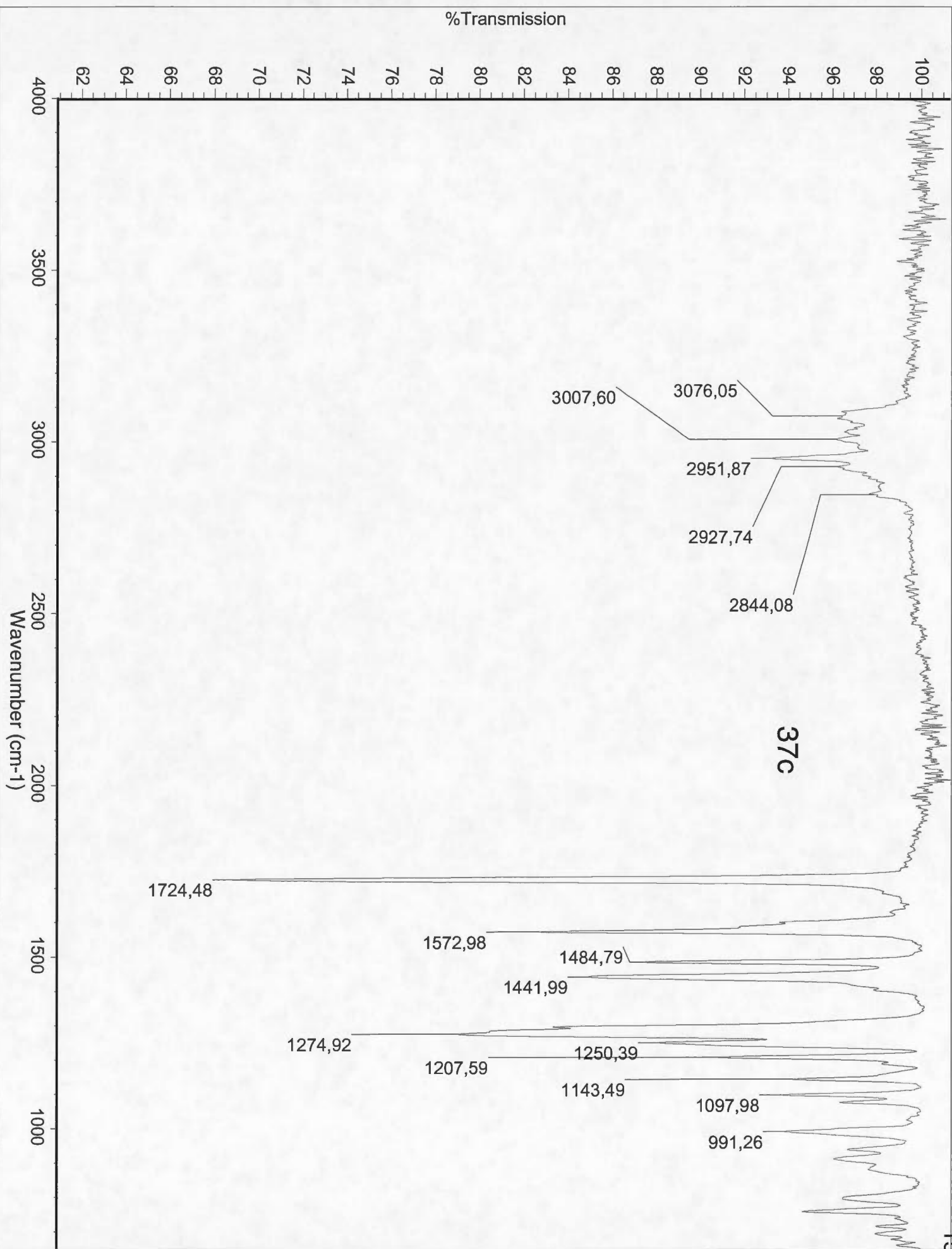


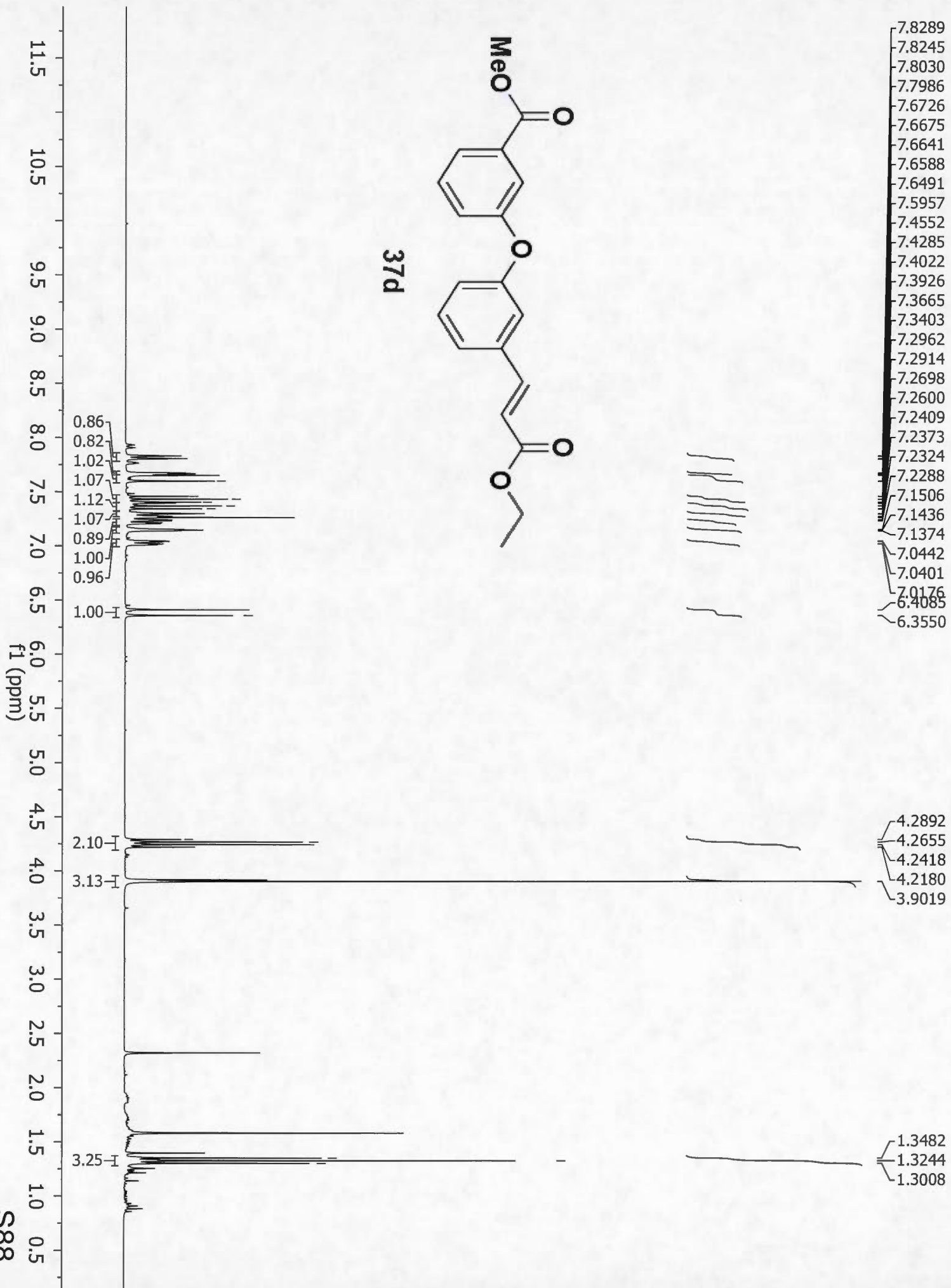


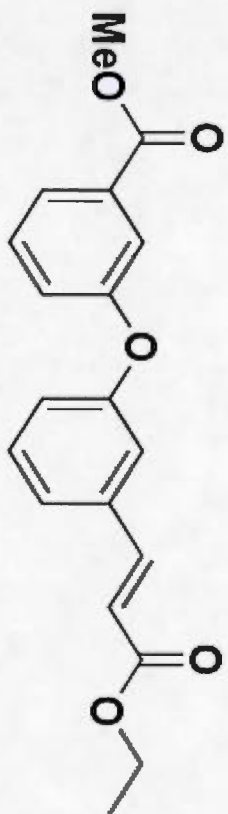
37c











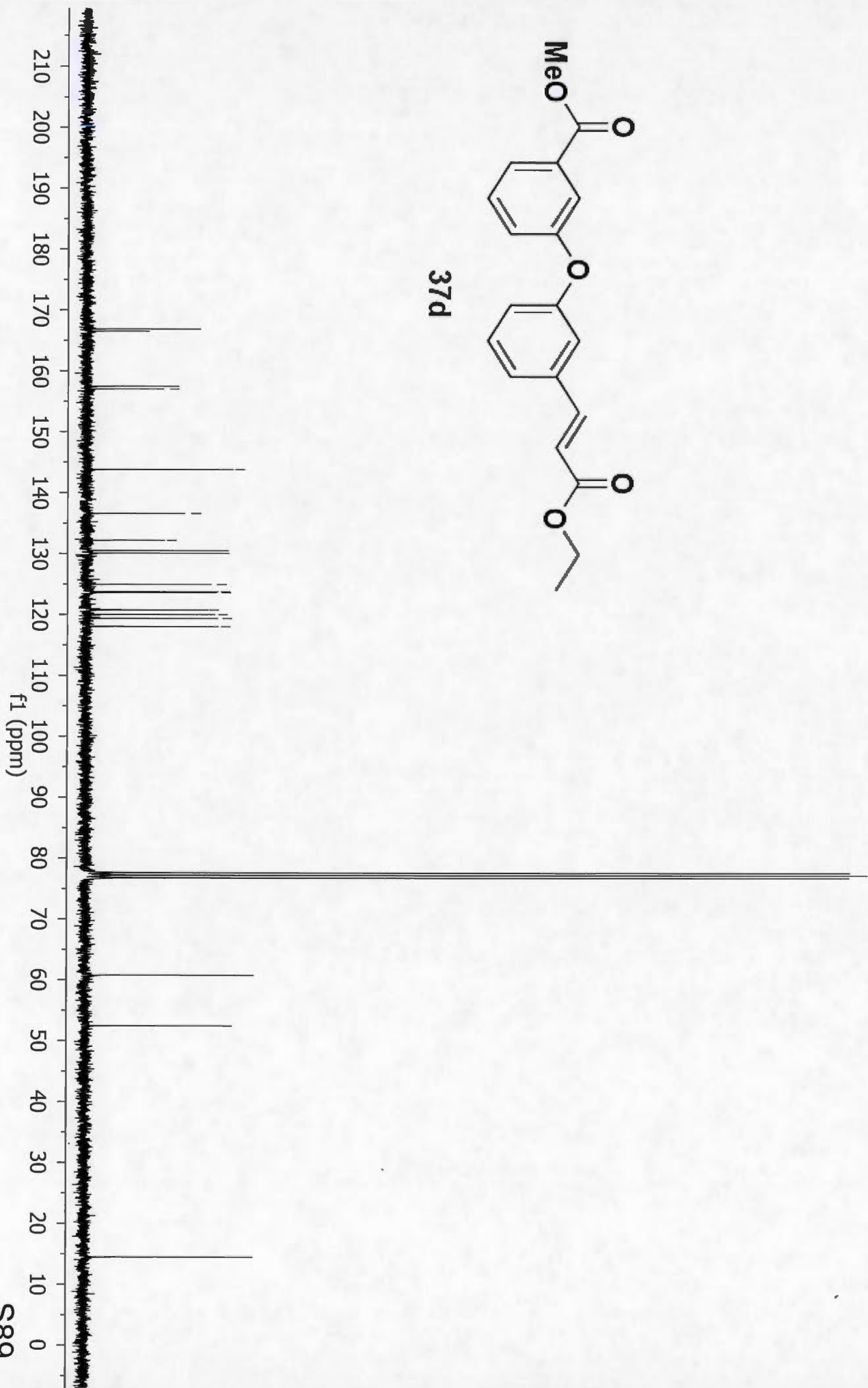
37d

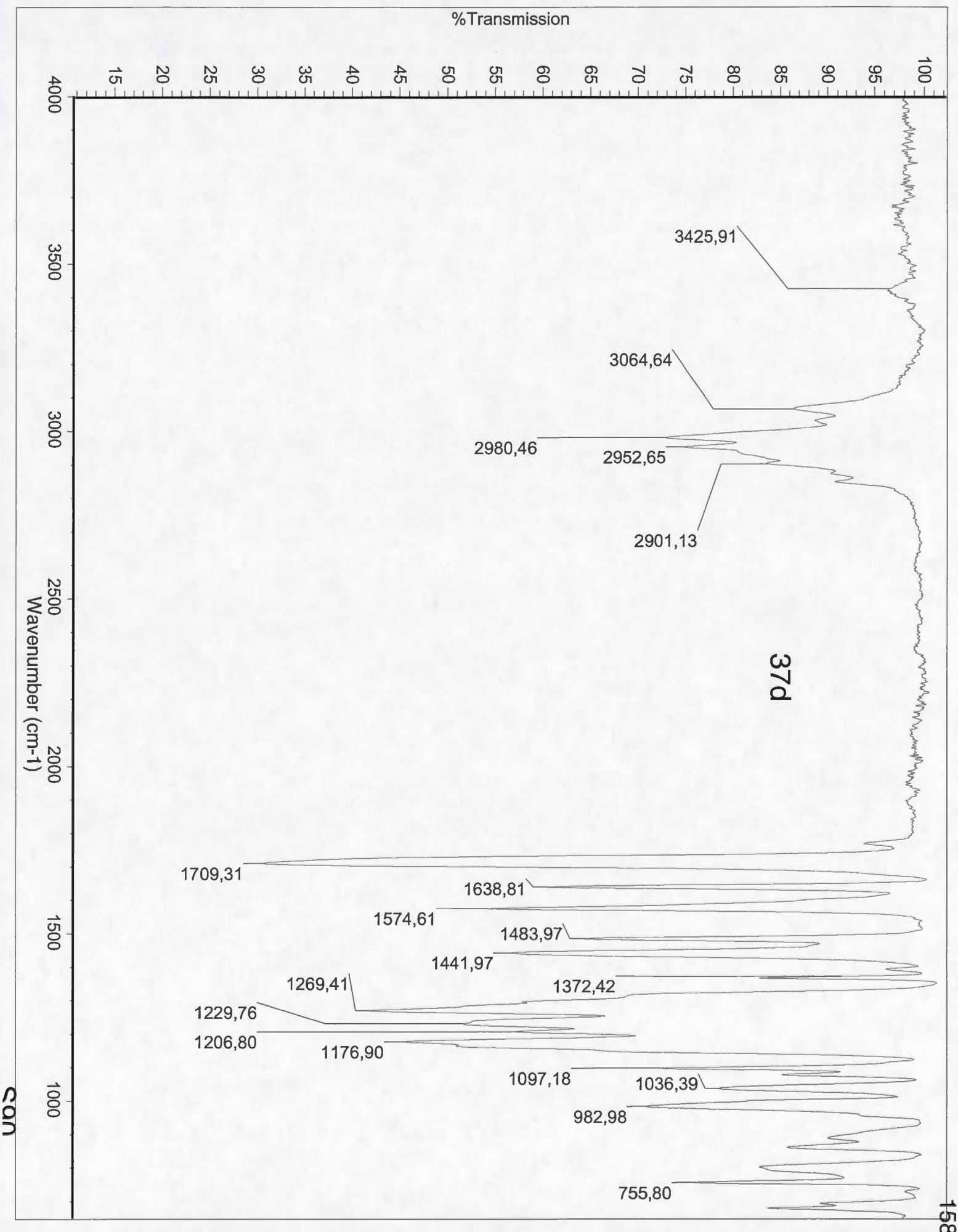
- 166.8552
- 166.5076
- 157.4695
- 157.0022
- 143.8026
- 136.5622
- 132.1849
- 130.4937
- 130.4700
- 130.0595
- 124.9233
- 123.7340
- 123.5839
- 120.7147
- 119.9918
- 119.3255
- 117.9723

60.7379

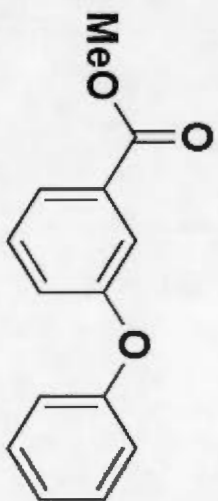
52.4343

14.4299

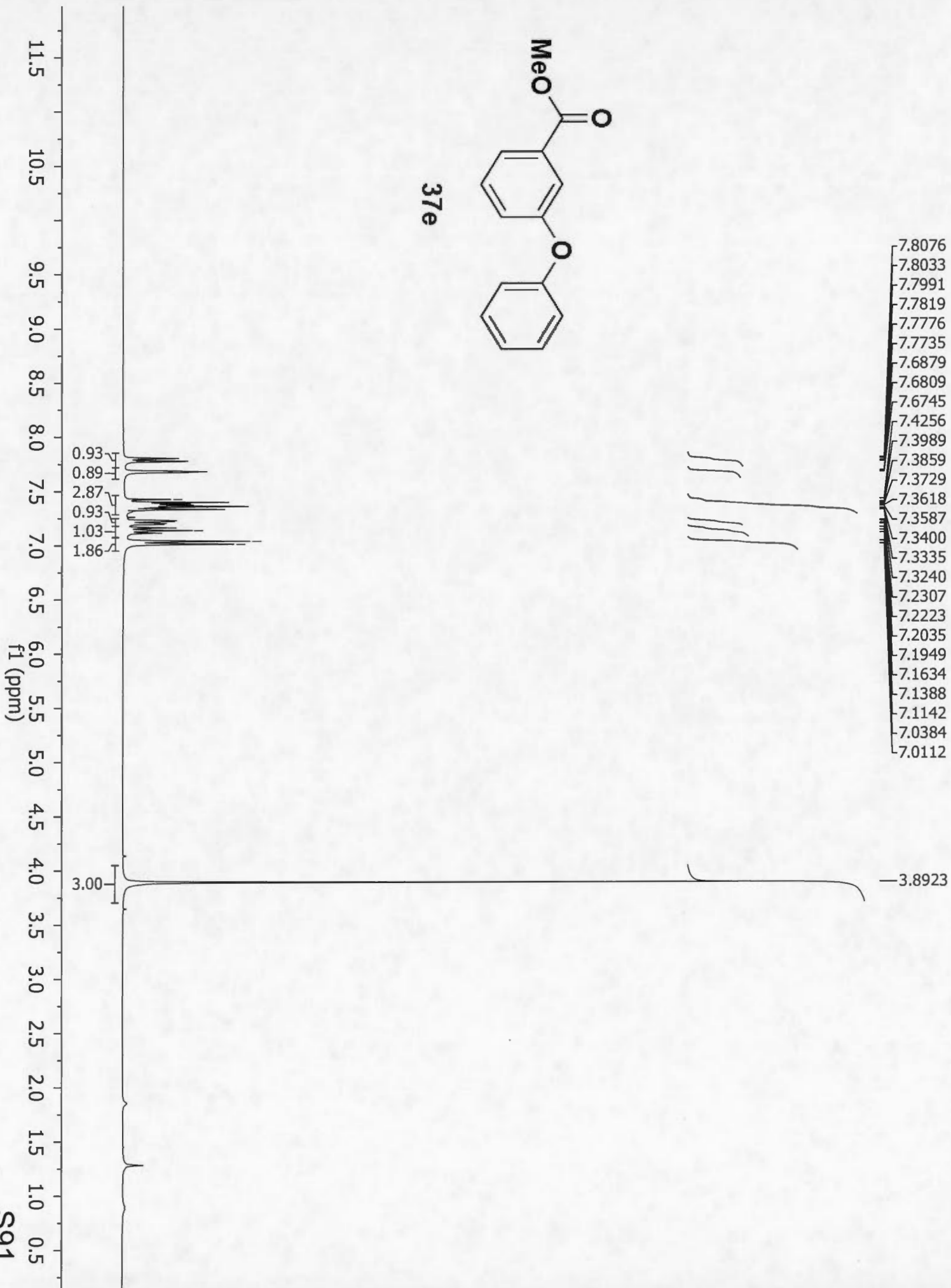


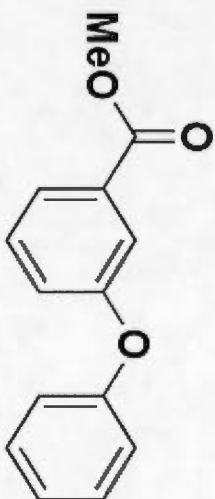




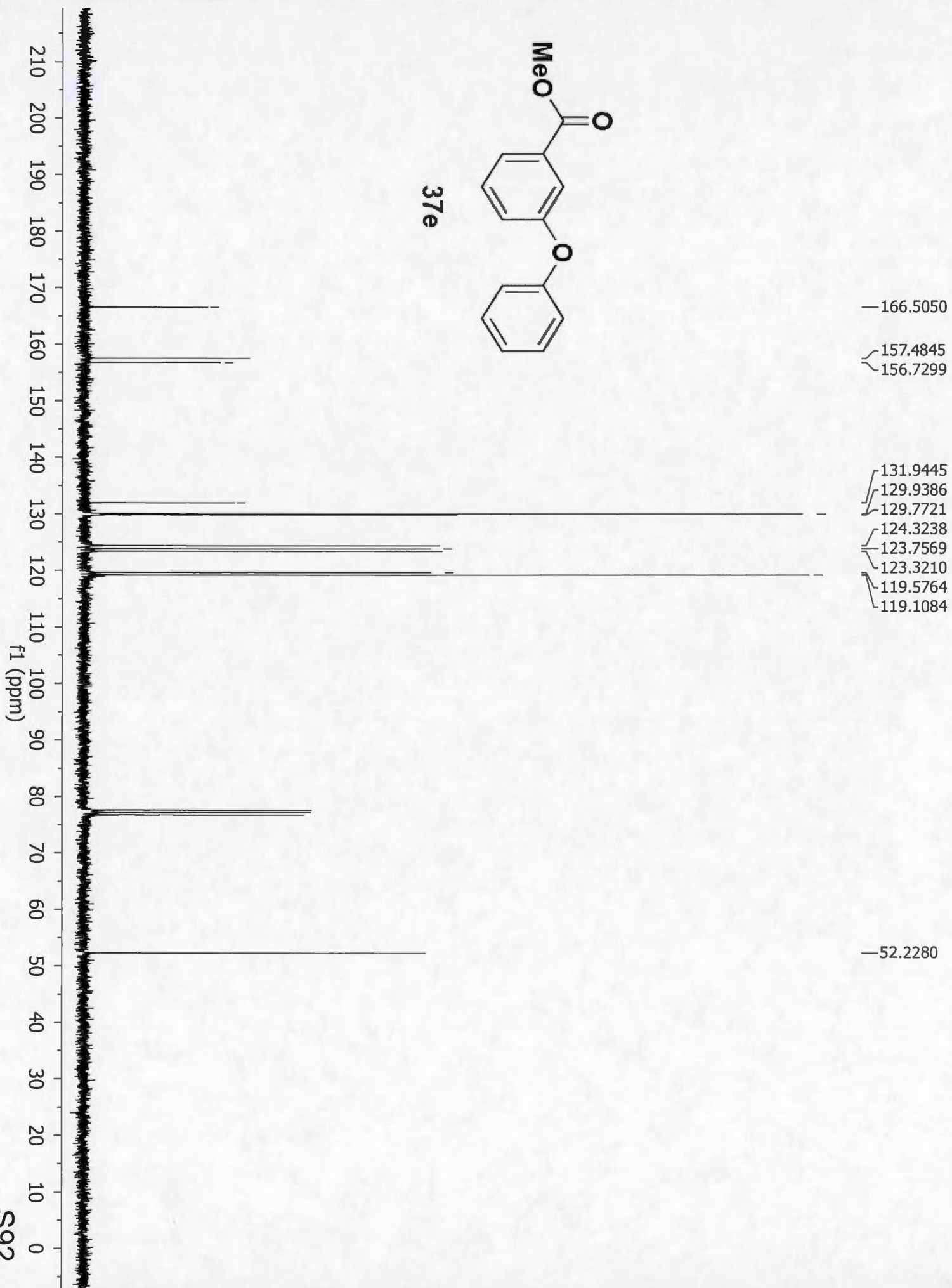


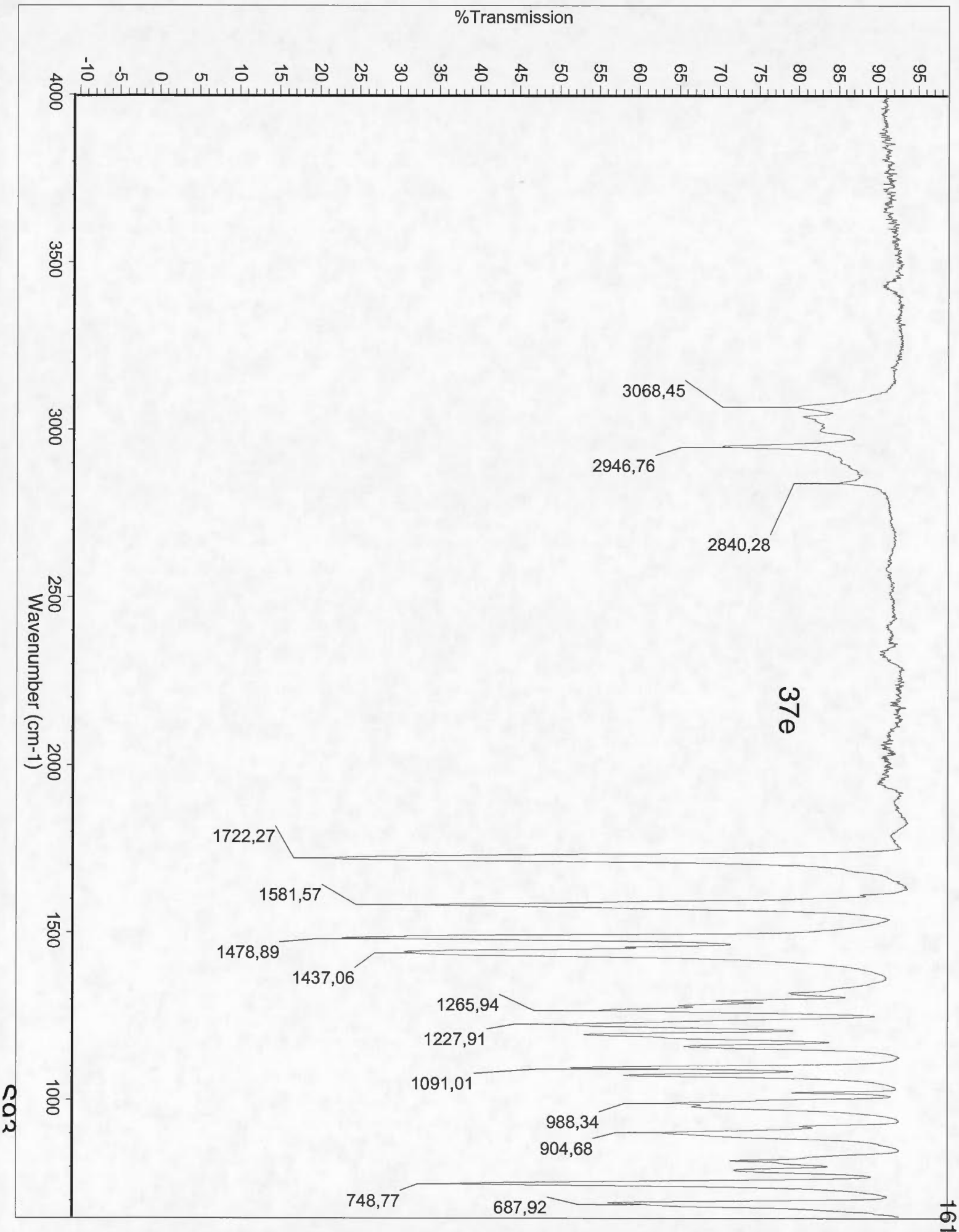
37e

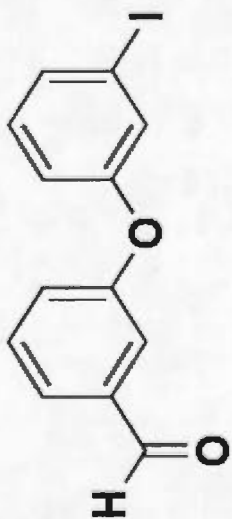




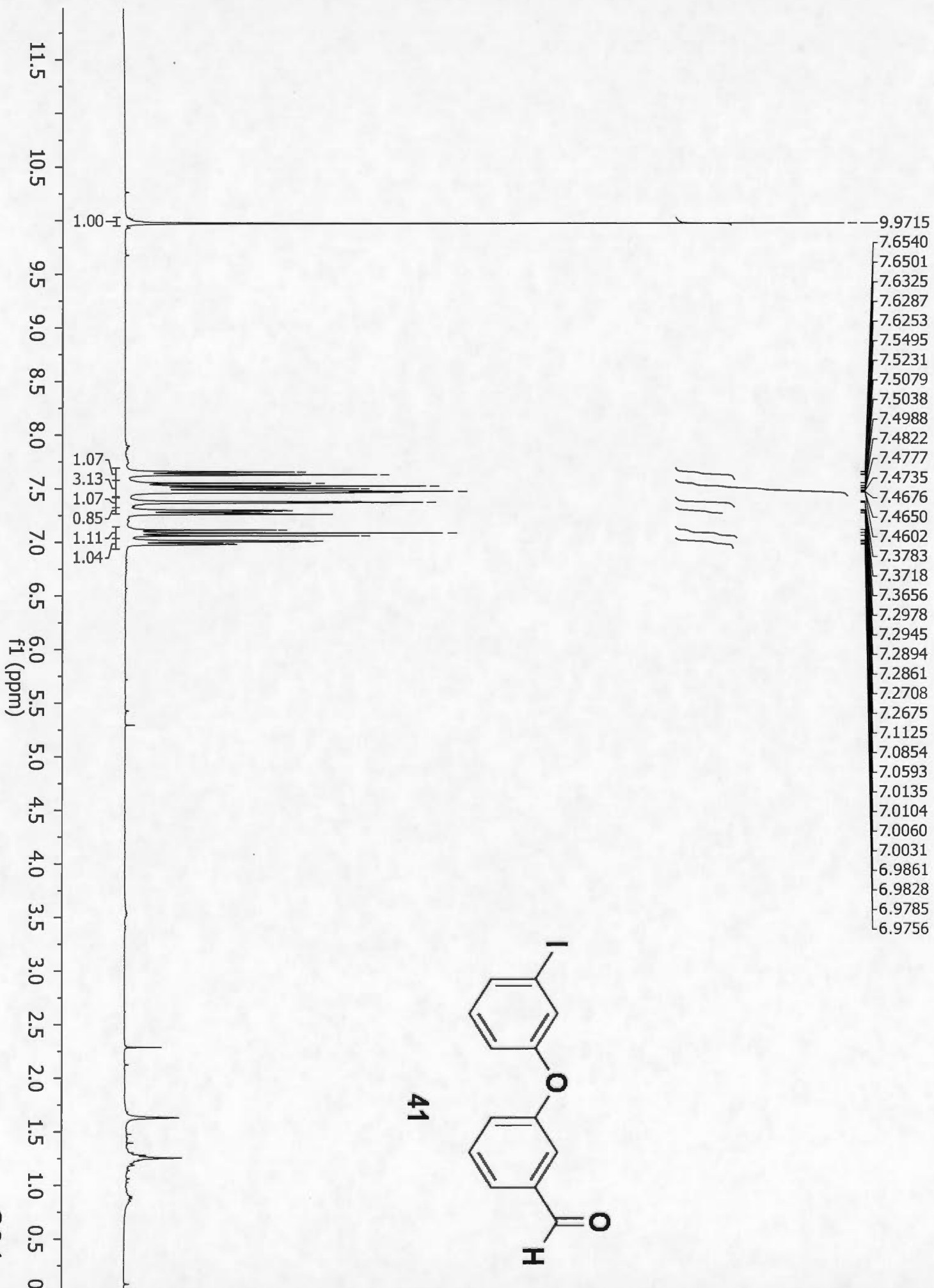
37e



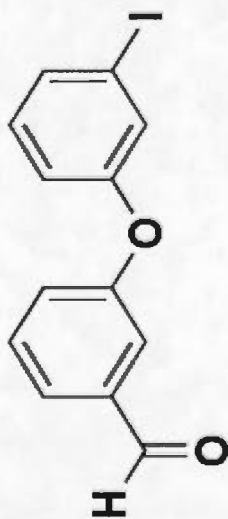




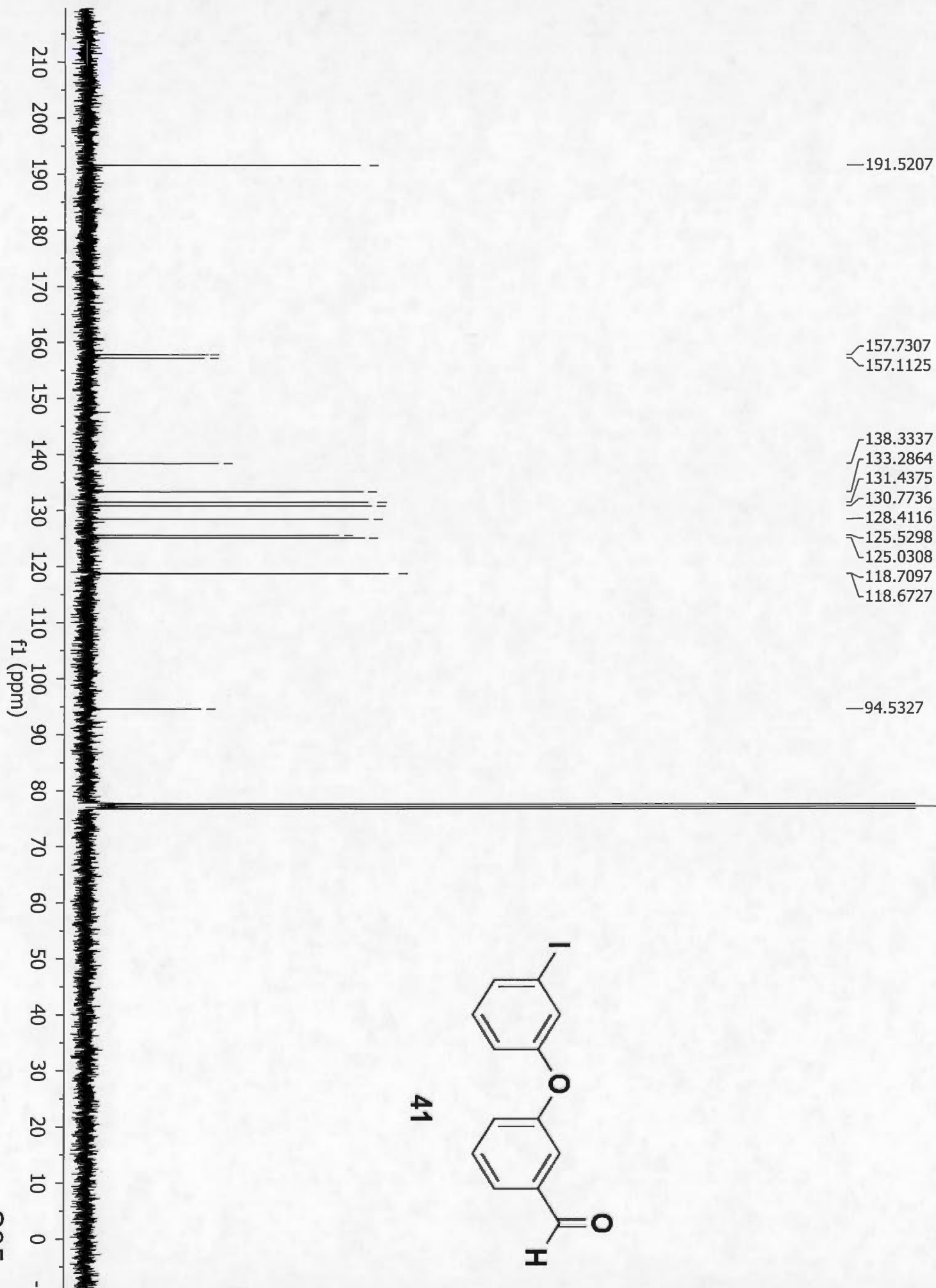
41

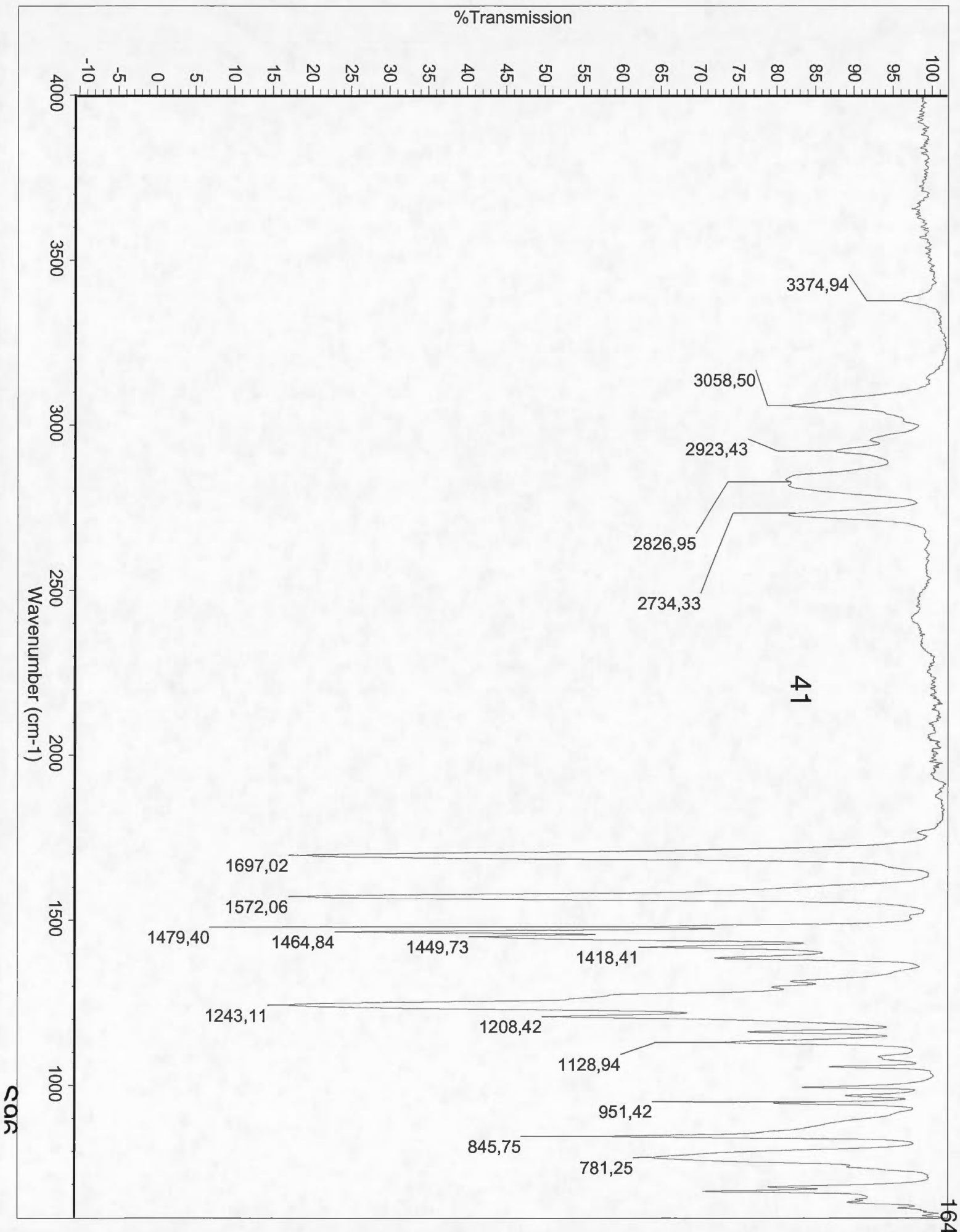


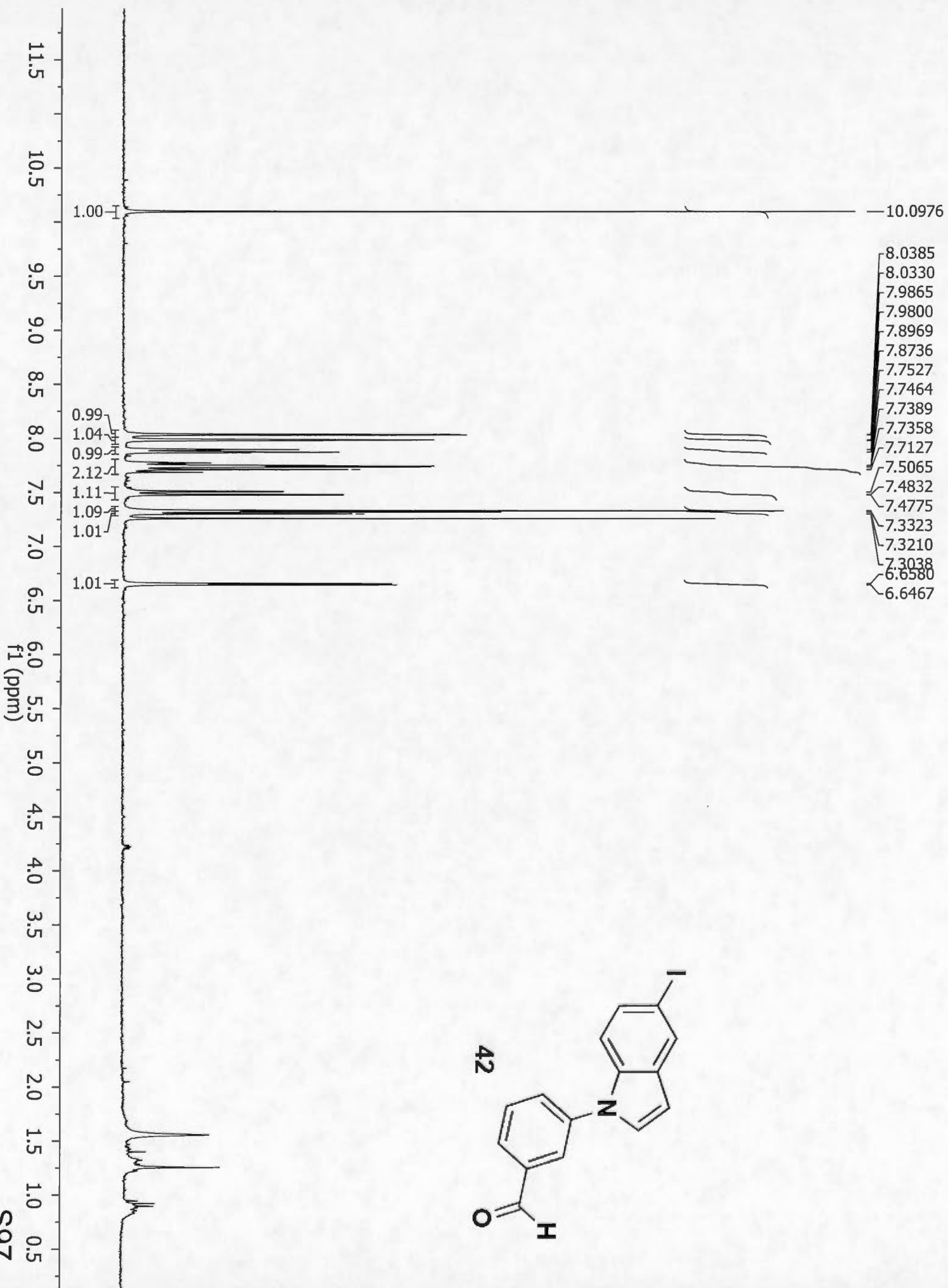
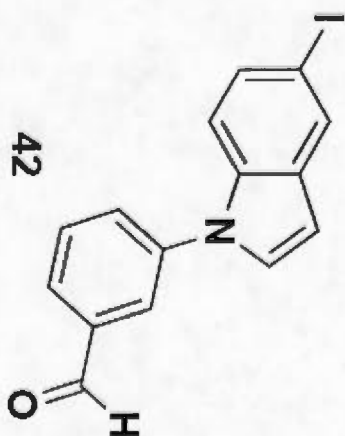


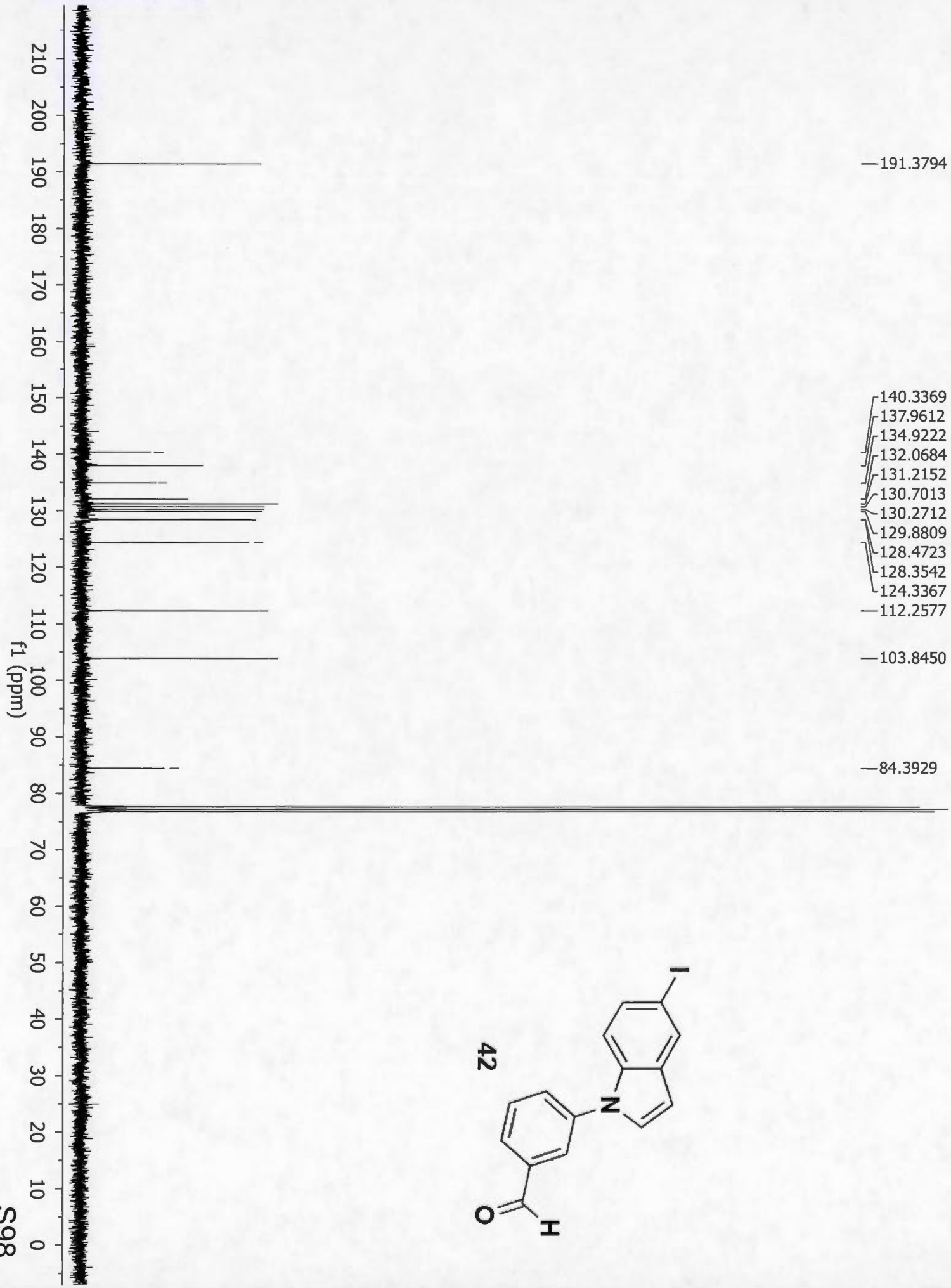
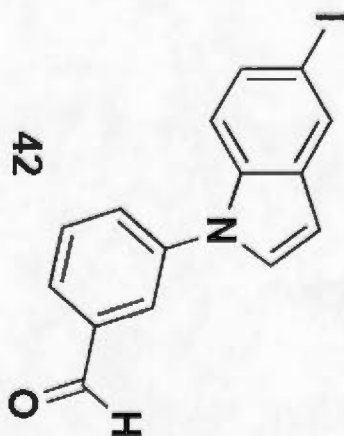


41

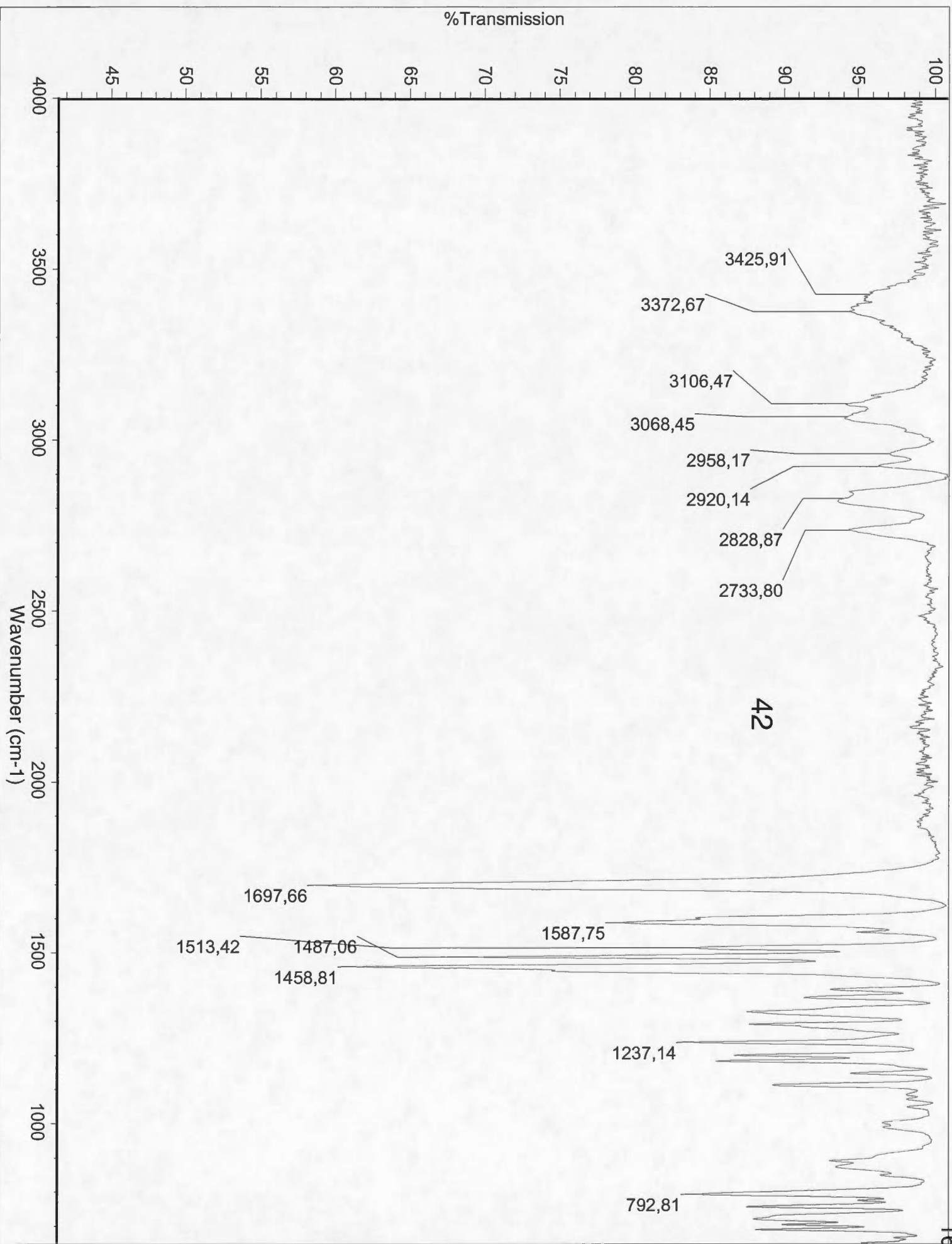


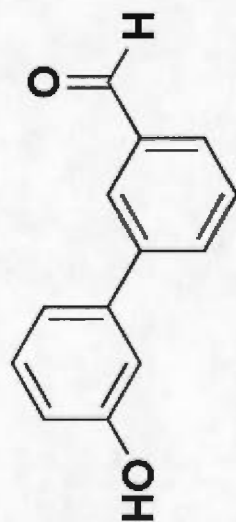




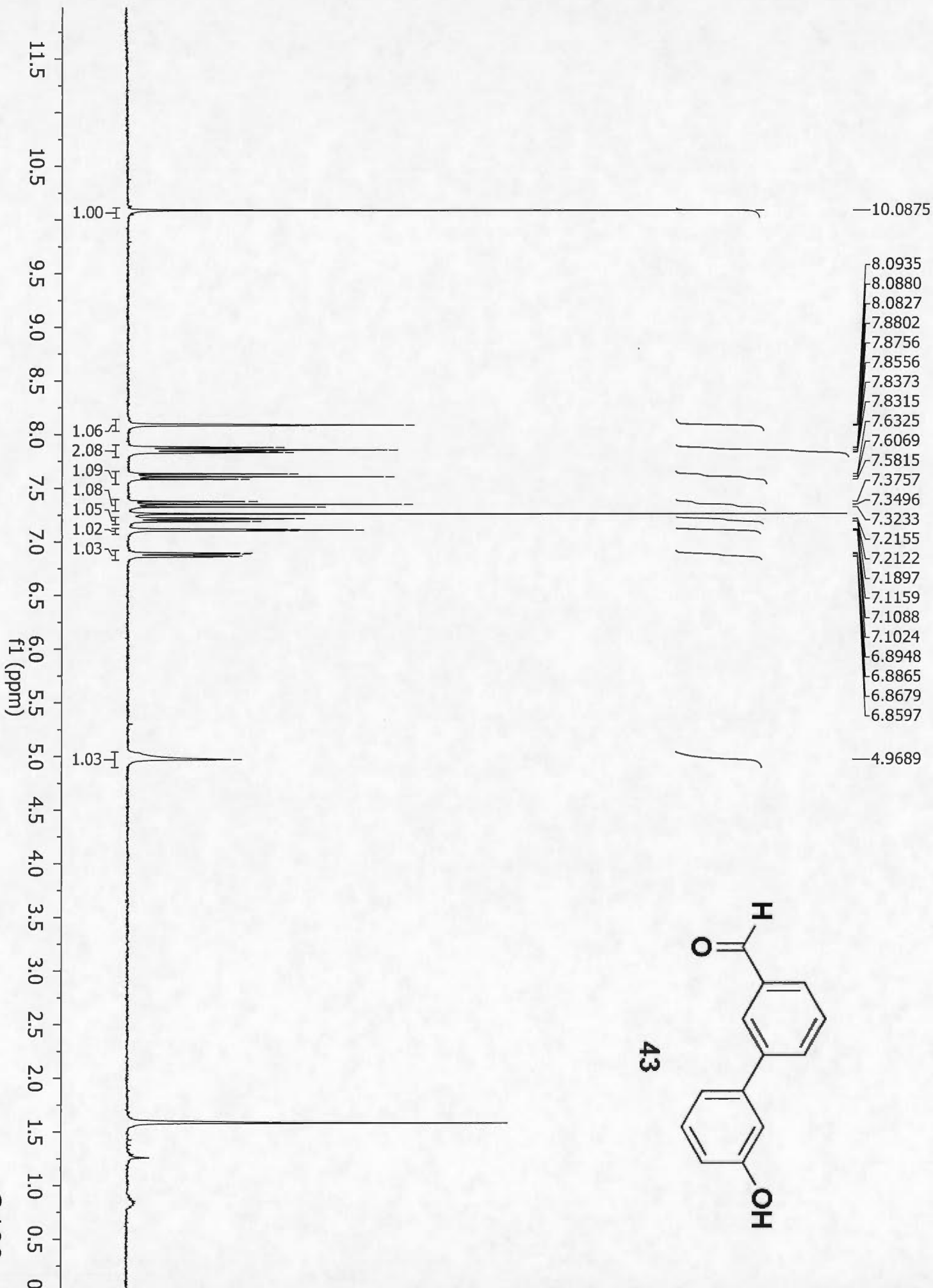


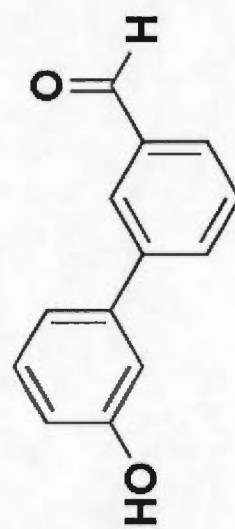




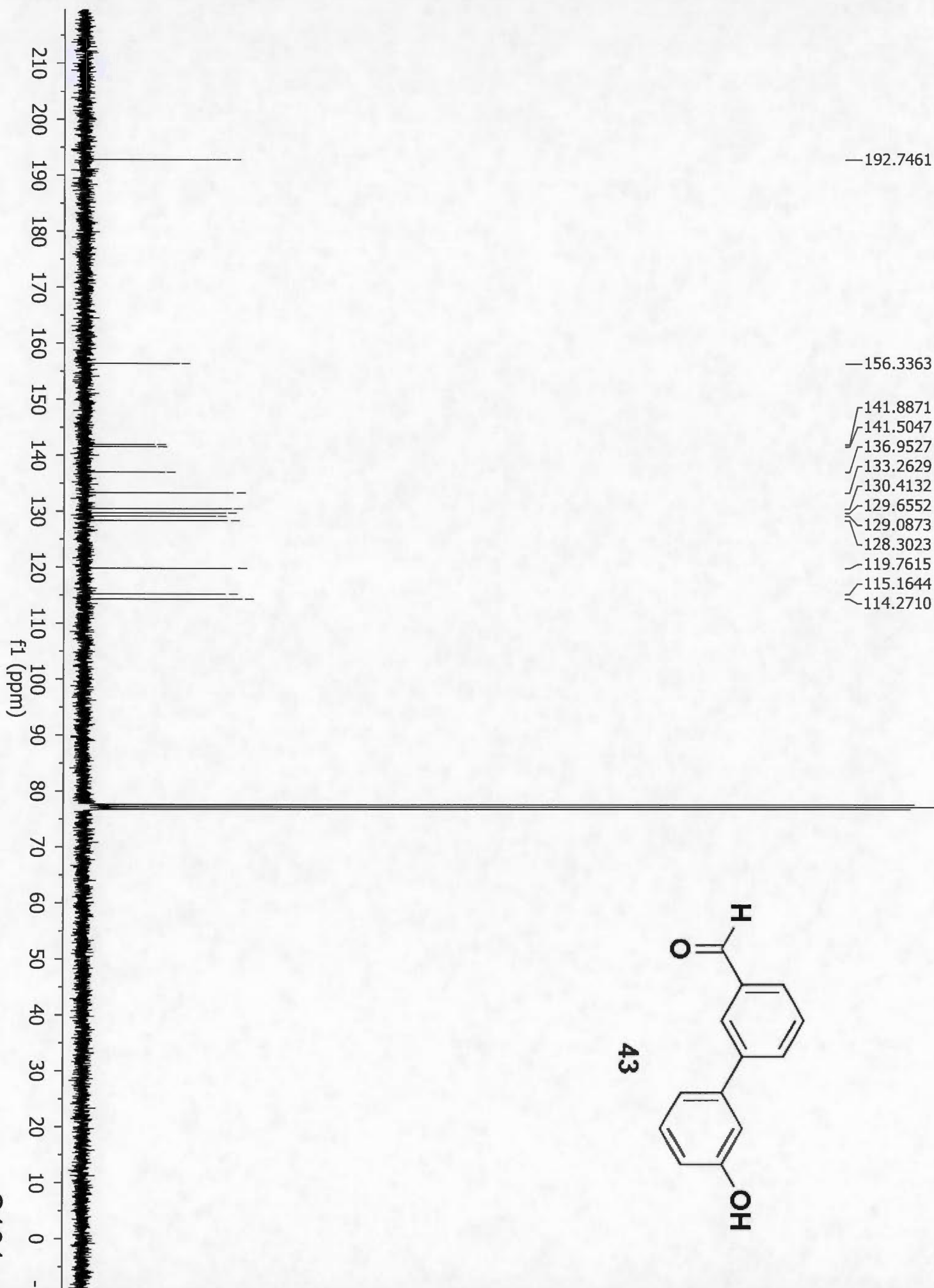


43

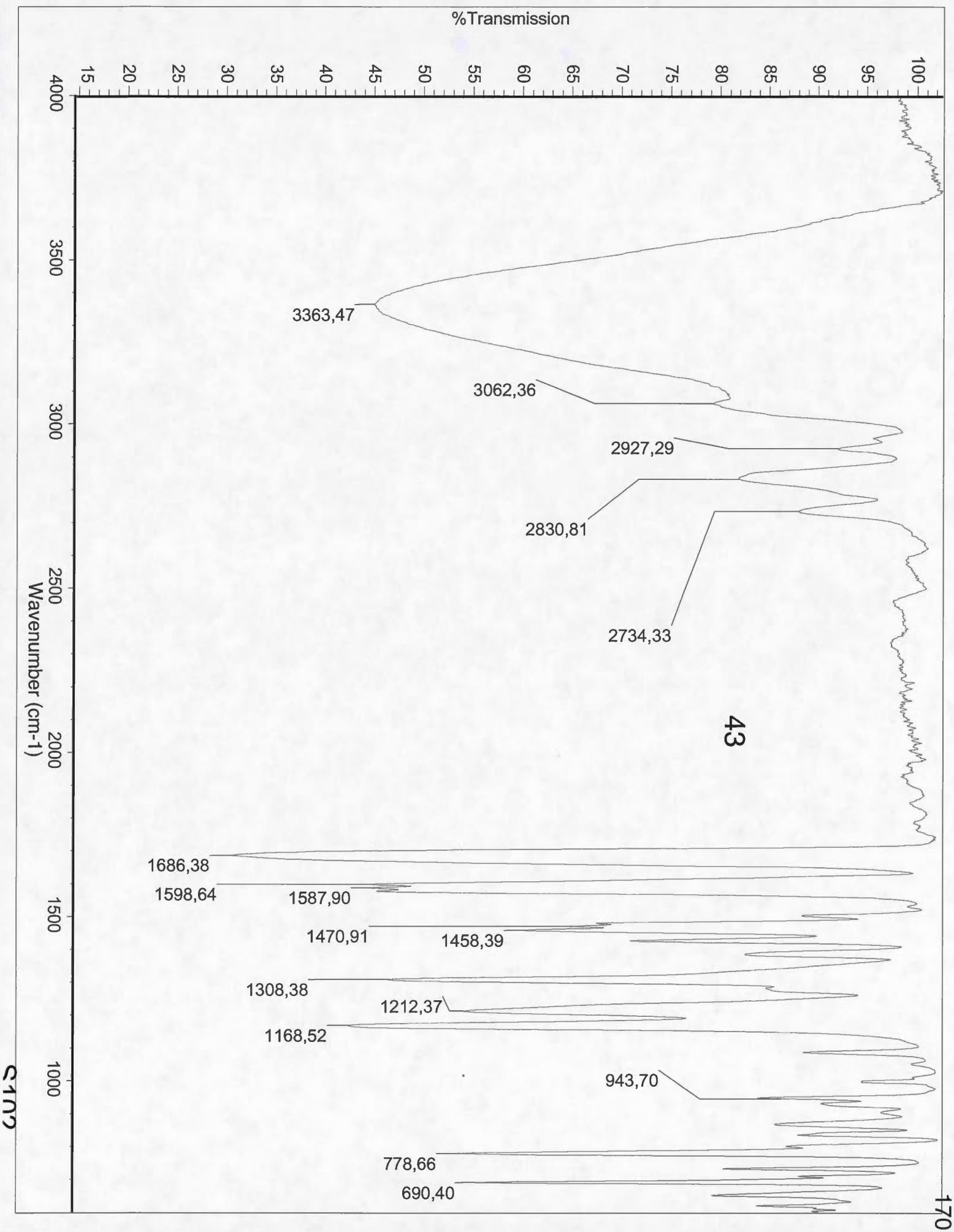




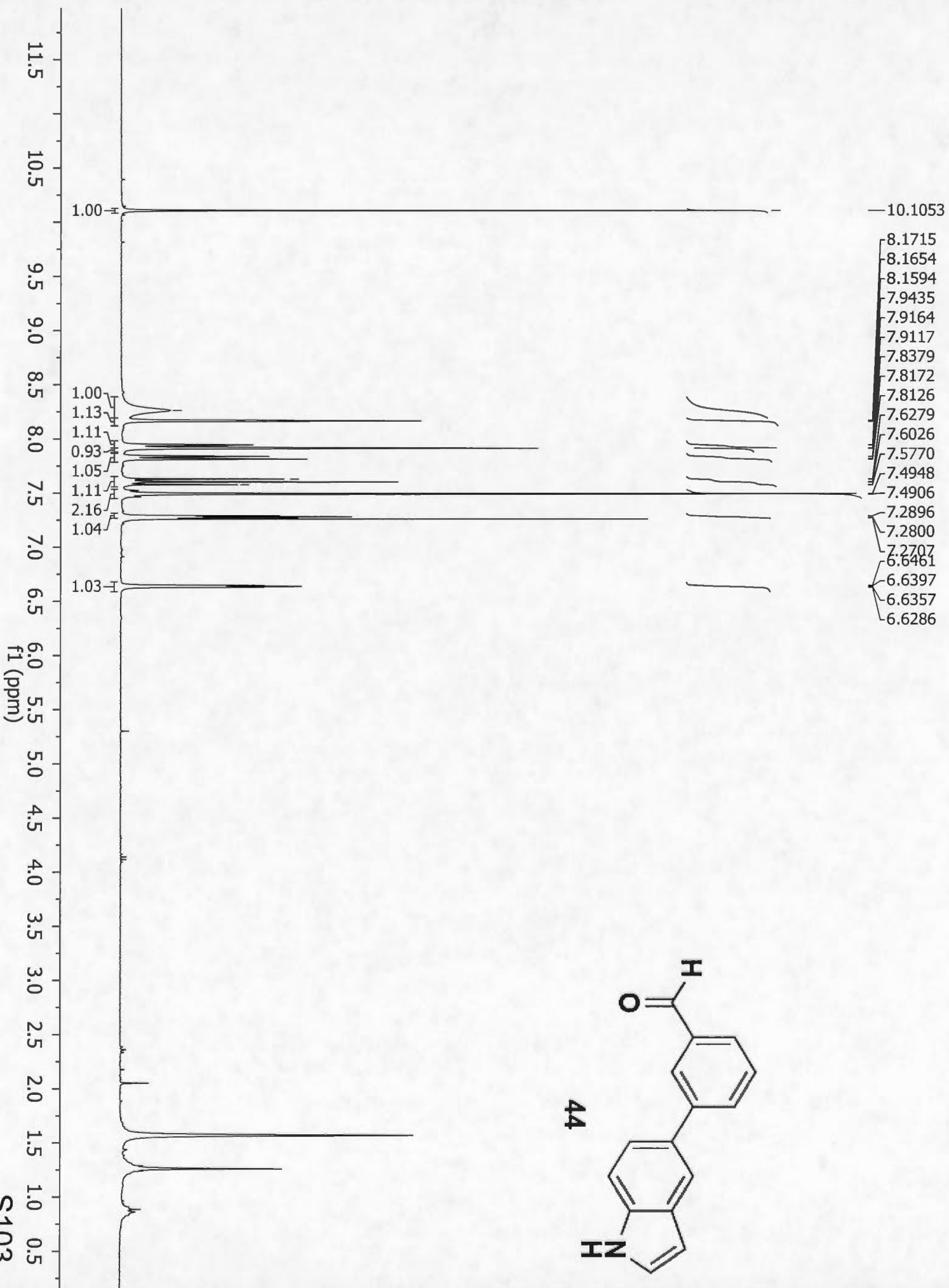
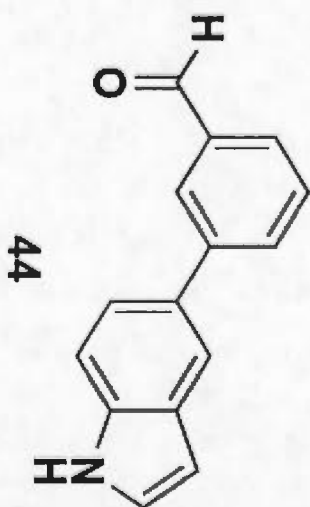
43

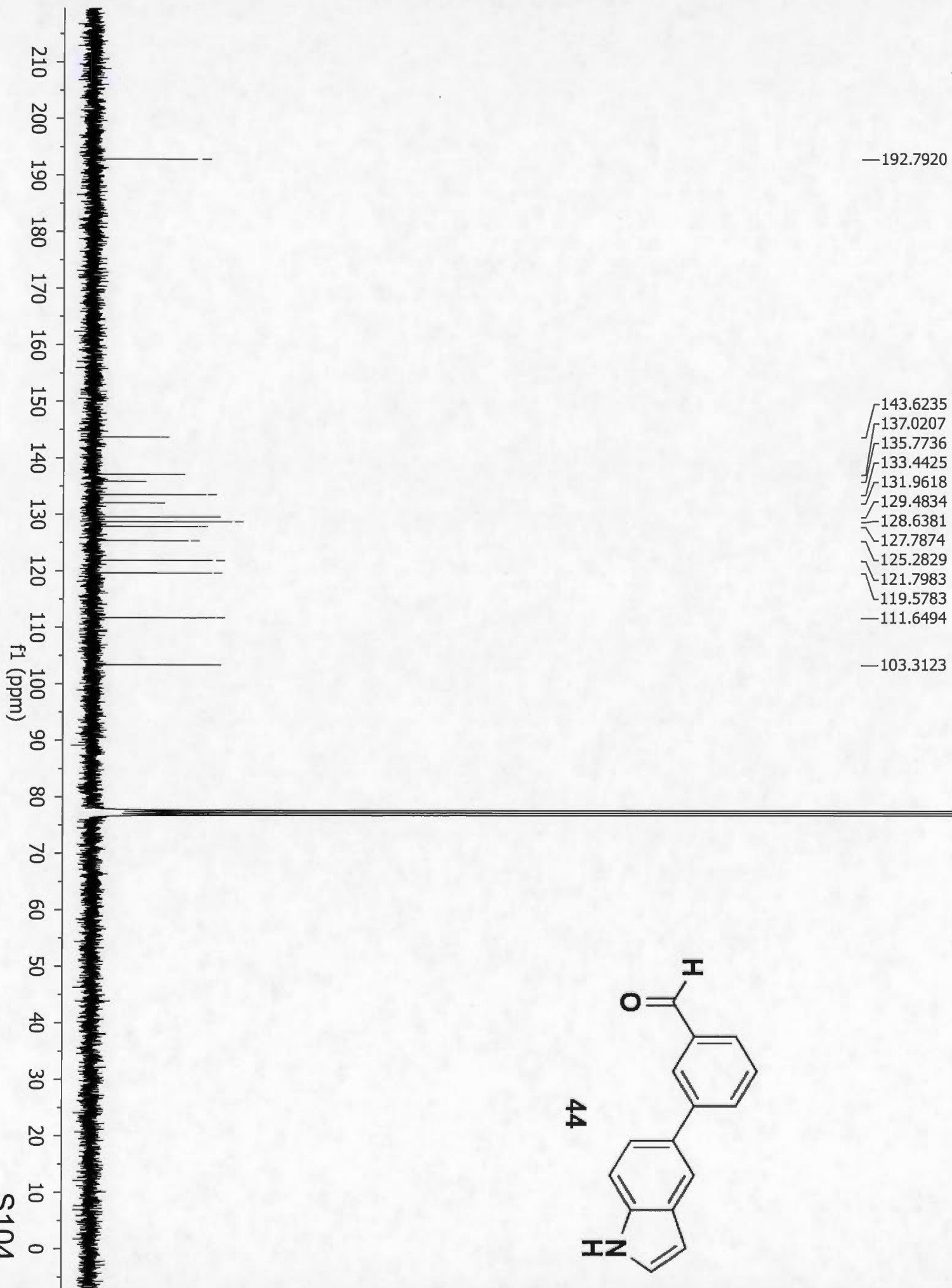
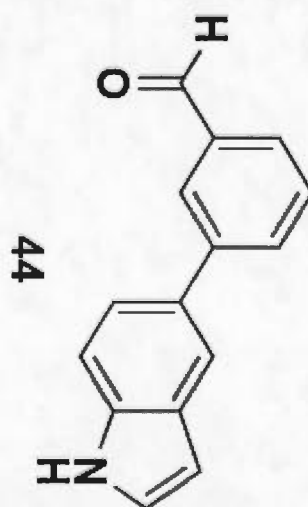


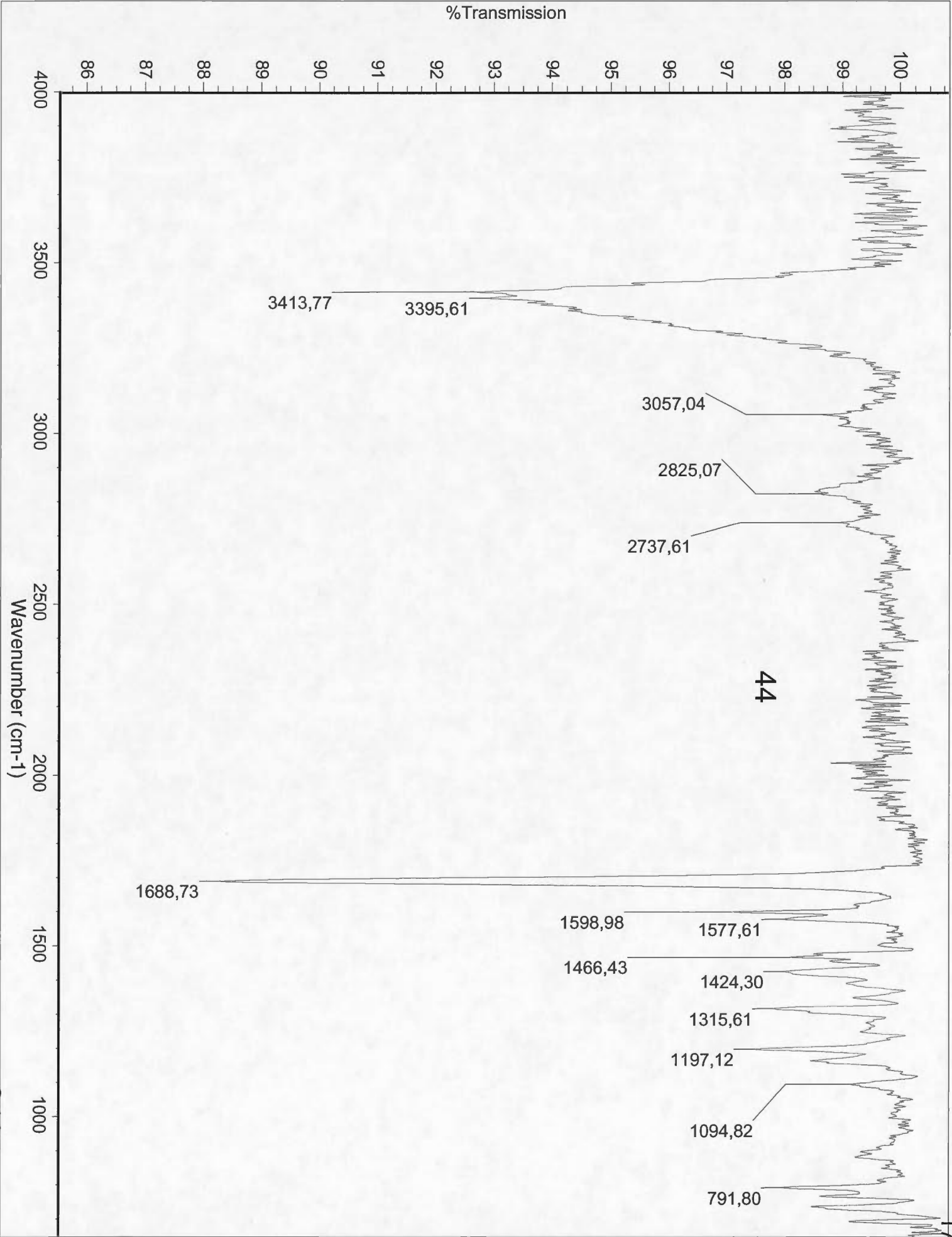
S101







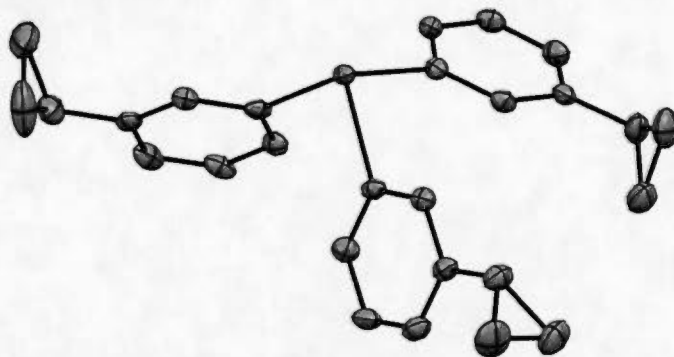




### Crystallographic analysis

Single-crystal X-ray diffraction data were acquired at the Mo  $K\alpha$  or Cu  $K\alpha$  wavelength. The structure for **21b** was reported previously (CCDC code 949485).<sup>i</sup>

### Tris(3-cyclopropylphenyl)bismuthine (1f)



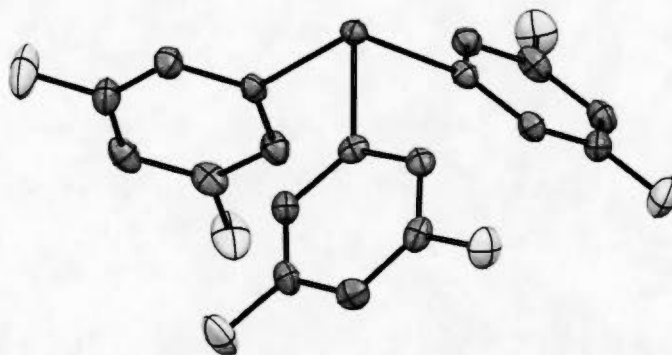
**Figure S1.** ORTEP view at 50% ellipsoid probability for compound **1f**.

A colorless plate-like specimen of  $C_{27}H_{27}Bi$ , approximate dimensions 0.058 mm x 0.146 mm x 0.267 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 1464 frames were collected. The total exposure time was 4.07 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 24743 reflections to a maximum  $\theta$  angle of  $27.65^\circ$  ( $0.77 \text{ \AA}$  resolution), of which 5061 were independent (average redundancy 4.889, completeness = 99.7%,  $R_{int} = 5.73\%$ ,  $R_{sig} = 4.73\%$ ) and 4135 (81.70%) were greater than  $2\sigma(F^2)$ . The final cell constants of  $a = 17.2733(15) \text{ \AA}$ ,  $b = 6.1956(5) \text{ \AA}$ ,  $c = 20.4121(18) \text{ \AA}$ ,  $\beta = 92.6470(10)^\circ$ , volume =  $2182.1(3) \text{ \AA}^3$ , are based upon the refinement of the XYZ-centroids of 7035 reflections above  $20 \sigma(I)$  with  $4.721^\circ < 2\theta < 52.57^\circ$ . Data were corrected for absorption effects using the multi-scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.648. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.4835 and 0.7456. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group  $P 1 21/n 1$ , with  $Z = 4$  for the formula unit,  $C_{27}H_{27}Bi$ . The final anisotropic full-matrix least-squares refinement on  $F^2$  with 253 variables converged at  $R1 = 2.58\%$ , for the observed data and  $wR2 = 5.06\%$  for all data. The goodness-of-fit was 1.016. The largest peak in the final difference electron density synthesis was  $0.659 \text{ e}/\text{\AA}^3$  and the largest hole was  $-0.848 \text{ e}/\text{\AA}^3$  with an RMS deviation of  $0.129 \text{ e}/\text{\AA}^3$ . On the basis of the final model, the calculated density was  $1.706 \text{ g}/\text{cm}^3$  and  $F(000)$ , 1088 e $^-$ .

<sup>i</sup> Petiot, P.; Gagnon, A. *Eur. J. Org. Chem.* **2013**, 5282-5289.



Tris(3,5-difluorophenyl)bismuthine (**1i**)



**Figure S2.** ORTEP view at 50% ellipsoid probability for compound **1i**.

A colorless plate-like specimen of  $C_{18}H_9BiF_6$ , approximate dimensions 0.055 mm x 0.122 mm x 0.189 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 5856 frames were collected. The total exposure time was 8.13 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 22071 reflections to a maximum  $\theta$  angle of  $68.28^\circ$  ( $0.83 \text{ \AA}$  resolution), of which 2867 were independent (average redundancy 7.698, completeness = 98.6%,  $R_{\text{int}} = 4.33\%$ ,  $R_{\text{sig}} = 2.53\%$ ) and 2746 (95.78%) were greater than  $2\sigma(F^2)$ . The final cell constants of  $a = 5.16200(10) \text{ \AA}$ ,  $b = 15.7296(3) \text{ \AA}$ ,  $c = 19.6908(3) \text{ \AA}$ ,  $\beta = 97.0350(10)^\circ$ , volume =  $1586.78(5) \text{ \AA}^3$ , are based upon the refinement of the XYZ-centroids of 9954 reflections above  $20 \sigma(I)$  with  $7.214^\circ < 2\theta < 136.4^\circ$ . Data were corrected for absorption effects using the multi-scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.523. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.3941 and 0.7531. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group  $P 1 21/c 1$ , with  $Z = 4$  for the formula unit,  $C_{18}H_9BiF_6$ . The final anisotropic full-matrix least-squares refinement on  $F^2$  with 226 variables converged at  $R1 = 1.89\%$ , for the observed data and  $wR2 = 4.92\%$  for all data. The goodness-of-fit was 1.088. The largest peak in the final difference electron density synthesis was  $0.564 \text{ e/\AA}^3$  and the largest hole was  $-0.898 \text{ e/\AA}^3$  with an RMS deviation of  $0.111 \text{ e/\AA}^3$ . On the basis of the final model, the calculated density was  $2.295 \text{ g/cm}^3$  and  $F(000)$ , 1016 e $^-$ .

Tris(4-methoxyphenyl)bismuthine (1k)

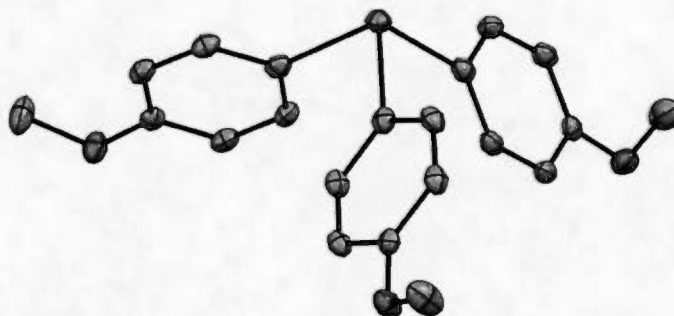


Figure S3. ORTEP view at 50% ellipsoid probability for compound **1k**.

A colorless rhomb-like specimen of  $C_{21}H_{21}BiO_3$ , approximate dimensions 0.204 mm x 0.395 mm x 0.417 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 1464 frames were collected. The total exposure time was 0.41 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a trigonal unit cell yielded a total of 11061 reflections to a maximum  $\theta$  angle of  $27.53^\circ$  ( $0.77 \text{ \AA}$  resolution), of which 1424 were independent (average redundancy 7.768, completeness = 99.9%,  $R_{\text{int}} = 5.08\%$ ,  $R_{\text{sig}} = 2.50\%$ ) and 1356 (95.22%) were greater than  $2\sigma(F^2)$ . The final cell constants of  $a = 13.0227(11) \text{ \AA}$ ,  $b = 13.0227(11) \text{ \AA}$ ,  $c = 18.9137(13) \text{ \AA}$ , volume =  $2777.9(5) \text{ \AA}^3$ , are based upon the refinement of the XYZ-centroids of 5605 reflections above  $20 \sigma(I)$  with  $5.617^\circ < 2\theta < 54.88^\circ$ . Data were corrected for absorption effects using the multi-scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.683. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.5092 and 0.7456. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group  $R\bar{3}$ , with  $Z = 6$  for the formula unit,  $C_{21}H_{21}BiO_3$ . The final anisotropic full-matrix least-squares refinement on  $F^2$  with 77 variables converged at  $R1 = 1.80\%$ , for the observed data and  $wR2 = 3.81\%$  for all data. The goodness-of-fit was 1.103. The largest peak in the final difference electron density synthesis was  $0.575 \text{ e}/\text{\AA}^3$  and the largest hole was  $-0.819 \text{ e}/\text{\AA}^3$  with an RMS deviation of  $0.091 \text{ e}/\text{\AA}^3$ . On the basis of the final model, the calculated density was  $1.902 \text{ g}/\text{cm}^3$  and  $F(000)$ , 1524 e $^-$ .

Tris(2-(diethoxymethyl)phenyl)bismuthine (1n)

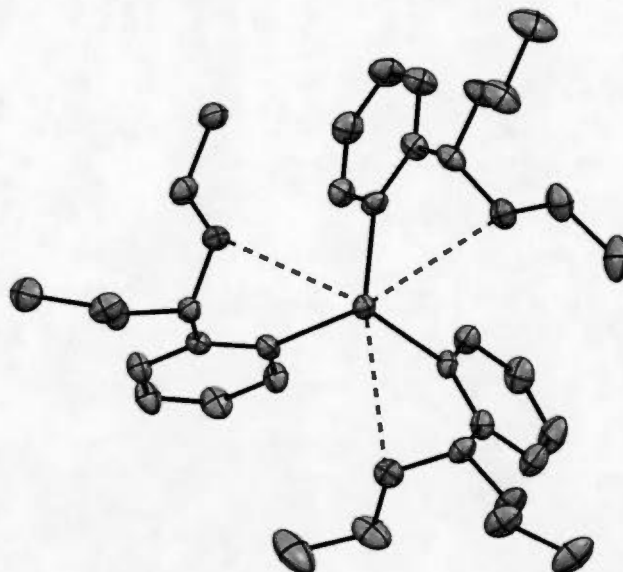
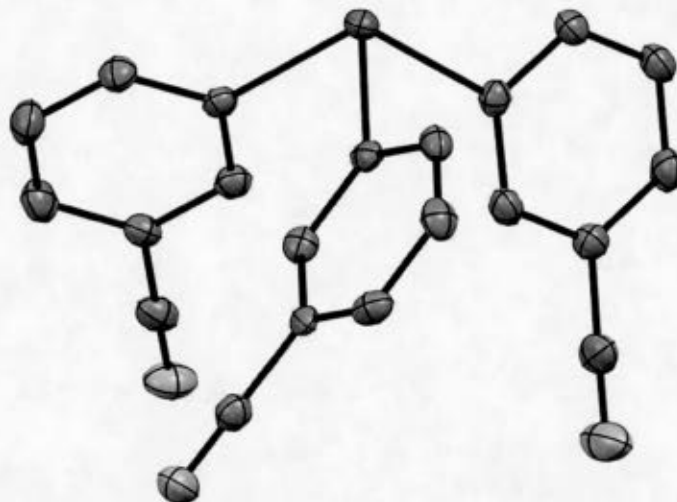


Figure S4. ORTEP view at 50% ellipsoid probability for compound 1n.

A colorless plate-like specimen of  $C_{33}H_{45}BiO_6$ , approximate dimensions 0.039 mm x 0.067 mm x 0.220 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 5856 frames were collected. The total exposure time was 6.30 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 149815 reflections to a maximum  $\theta$  angle of  $68.39^\circ$  ( $0.83 \text{ \AA}$  resolution), of which 18042 were independent (average redundancy 8.304, completeness = 99.5%,  $R_{\text{int}} = 6.47\%$ ,  $R_{\text{sig}} = 3.25\%$ ) and 13630 (75.55%) were greater than  $2\sigma(F^2)$ . The final cell constants of  $a = 12.2493(2) \text{ \AA}$ ,  $b = 40.0209(5) \text{ \AA}$ ,  $c = 23.2425(3) \text{ \AA}$ ,  $\beta = 119.9300(10)^\circ$ , volume =  $9874.6(3) \text{ \AA}^3$ , are based upon the refinement of the XYZ-centroids of 9835 reflections above  $20 \sigma(I)$  with  $8.329^\circ < 2\theta < 136.6^\circ$ . Data were corrected for absorption effects using the numerical method (SADABS). The ratio of minimum to maximum apparent transmission was 0.404. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.2896 and 0.7169. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group  $P 1 21/c 1$ , with  $Z = 12$  for the formula unit,  $C_{33}H_{45}BiO_6$ . The final anisotropic full-matrix least-squares refinement on  $F^2$  with 1099 variables converged at  $R1 = 2.67\%$ , for the observed data and  $wR2 = 6.33\%$  for all data. The goodness-of-fit was 1.004. The largest peak in the final difference electron density synthesis was  $0.550 \text{ e}/\text{\AA}^3$  and the largest hole was  $-1.148 \text{ e}/\text{\AA}^3$  with an RMS deviation of  $0.085 \text{ e}/\text{\AA}^3$ . On the basis of the final model, the calculated density was  $1.507 \text{ g}/\text{cm}^3$  and  $F(000)$ , 4488 e $^-$ .



**3,3',3''-bismuthinetriyltribenzonitrile (1s)**



**Figure S5.** ORTEP view at 50% ellipsoid probability for compound **1s**.

A clear colorless block-like specimen of  $C_{21}H_{12}BiN_3$ , approximate dimensions 0.142 mm x 0.330 mm x 0.532 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 1464 frames were collected. The total exposure time was 0.41 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 20762 reflections to a maximum  $\theta$  angle of  $27.50^\circ$  ( $0.77 \text{ \AA}$  resolution), of which 4081 were independent (average redundancy 5.087, completeness = 99.6%,  $R_{\text{int}} = 5.36\%$ ,  $R_{\text{sig}} = 3.99\%$ ) and 3398 (83.26%) were greater than  $2\sigma(F^2)$ . The final cell constants of  $a = 8.5793(8) \text{ \AA}$ ,  $b = 11.6696(11) \text{ \AA}$ ,  $c = 17.9621(16) \text{ \AA}$ ,  $\beta = 96.1690(10)^\circ$ , volume =  $1787.9(3) \text{ \AA}^3$ , are based upon the refinement of the XYZ-centroids of 6528 reflections above  $20 \sigma(I)$  with  $4.775^\circ < 2\theta < 54.58^\circ$ . Data were corrected for absorption effects using the multi-scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.494. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.3684 and 0.7456. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group  $P 1 21/c 1$ , with  $Z = 4$  for the formula unit,  $C_{21}H_{12}BiN_3$ . The final anisotropic full-matrix least-squares refinement on  $F^2$  with 226 variables converged at  $R1 = 2.25\%$ , for the observed data and  $wR2 = 4.67\%$  for all data. The goodness-of-fit was 1.017. The largest peak in the final difference electron density synthesis was  $0.501 \text{ e}/\text{\AA}^3$  and the largest hole was  $-0.842 \text{ e}/\text{\AA}^3$  with an RMS deviation of  $0.120 \text{ e}/\text{\AA}^3$ . On the basis of the final model, the calculated density was  $1.914 \text{ g}/\text{cm}^3$  and  $F(000)$ , 968 e $^-$ .



Trimethyl 4,4',4''-bismuthinetriyltribenzoate (1t)

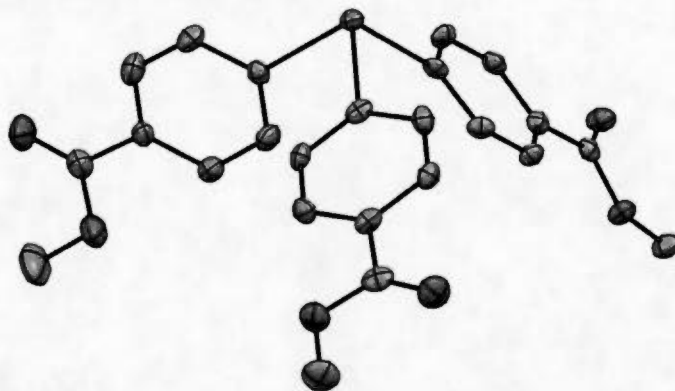


Figure S6. ORTEP view at 50% ellipsoid probability for compound 1t.

A clear colorless plate-like specimen of  $C_{24}H_{21}BiO_6$ , approximate dimensions 0.038 mm x 0.266 mm x 0.307 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 1464 frames were collected. The total exposure time was 4.07 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a triclinic unit cell yielded a total of 12705 reflections to a maximum  $\theta$  angle of  $27.80^\circ$  ( $0.76 \text{ \AA}$  resolution), of which 5155 were independent (average redundancy 2.465, completeness = 97.5%,  $R_{\text{int}} = 5.08\%$ ,  $R_{\text{sig}} = 6.80\%$ ) and 4499 (87.27%) were greater than  $2\sigma(F^2)$ . The final cell constants of  $a = 5.7360(7) \text{ \AA}$ ,  $b = 10.0706(12) \text{ \AA}$ ,  $c = 19.648(2) \text{ \AA}$ ,  $\alpha = 96.3650(10)^\circ$ ,  $\beta = 90.7050(10)^\circ$ ,  $\gamma = 98.8130(10)^\circ$ , volume =  $1114.2(2) \text{ \AA}^3$ , are based upon the refinement of the XYZ-centroids of 5695 reflections above  $20 \sigma(I)$  with  $5.520^\circ < 2\theta < 53.85^\circ$ . Data were corrected for absorption effects using the multi-scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.611. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.4554 and 0.7456. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group  $P -1$ , with  $Z = 2$  for the formula unit,  $C_{24}H_{21}BiO_6$ . The final anisotropic full-matrix least-squares refinement on  $F^2$  with 283 variables converged at  $R1 = 3.64\%$ , for the observed data and  $wR2 = 8.12\%$  for all data. The goodness-of-fit was 1.060. The largest peak in the final difference electron density synthesis was  $1.828 \text{ e}^-/\text{\AA}^3$  and the largest hole was  $-1.530 \text{ e}^-/\text{\AA}^3$  with an RMS deviation of  $0.175 \text{ e}^-/\text{\AA}^3$ . On the basis of the final model, the calculated density was  $1.831 \text{ g/cm}^3$  and  $F(000)$ , 592  $e^-$ .

Tris(2-formylphenyl)bismuthine (21c)

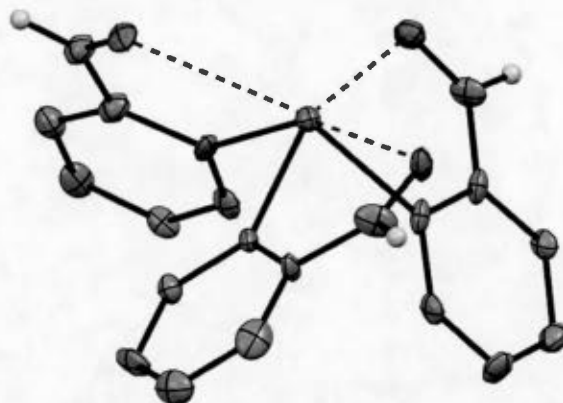
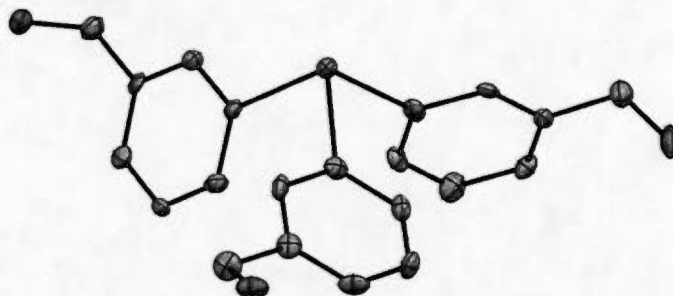


Figure S7. ORTEP view at 50% ellipsoid probability for compound **21c**.

A plate-like specimen of  $C_{21}H_{15}BiO_3$ , approximate dimensions 0.053 mm x 0.104 mm x 0.147 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. The total exposure time was 4.07 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 15775 reflections to a maximum  $\theta$  angle of  $27.51^\circ$  ( $0.77 \text{ \AA}$  resolution), of which 5124 were independent (average redundancy 3.08, completeness = 99.9%,  $R_{\text{int}} = 7.17\%$ ,  $R_{\text{sig}} = 6.93\%$ ) and 3198 (62.41%) were greater than  $2\sigma(F^2)$ . The final cell constants of  $a = 5.3061(7) \text{ \AA}$ ,  $b = 16.256(2) \text{ \AA}$ ,  $c = 10.1005(16) \text{ \AA}$ ,  $\beta = 98.294(2)^\circ$ , volume =  $862.1(2) \text{ \AA}^3$ , are based upon the refinement of the XYZ-centroids of 3696 reflections above  $20 \sigma(I)$  with  $4.783^\circ < 2\theta < 54.03^\circ$ . Data were corrected for absorption effects using TWINABS. The ratio of minimum to maximum apparent transmission was 0.730. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.4831 and 0.7456. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group  $P 1 c 1$ , with  $Z = 2$  for the formula unit,  $C_{21}H_{15}BiO_3$ . The final anisotropic full-matrix least-squares refinement on  $F^2$  with 215 variables converged at  $R1 = 3.61\%$ , for the observed data and  $wR2 = 9.93\%$  for all data. The goodness-of-fit was 1.048. The largest peak in the final difference electron density synthesis was  $1.284 \text{ e}/\text{\AA}^3$  and the largest hole was  $-1.685 \text{ e}/\text{\AA}^3$  with an RMS deviation of  $0.198 \text{ e}/\text{\AA}^3$ . On the basis of the final model, the calculated density was  $2.020 \text{ g}/\text{cm}^3$  and  $F(000)$ , 496 e.

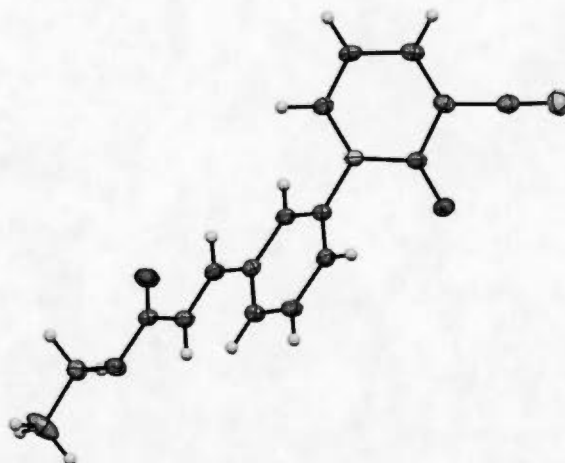
Tris(3-(hydroxymethyl)phenyl)bismuthine (21e)



**Figure S8.** ORTEP view at 50% ellipsoid probability for compound **21e**.

A colorless rhomb-like specimen of  $C_{21}H_{21}BiO_3$ , approximate dimensions 0.045 mm x 0.054 mm x 0.140 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 1464 frames were collected. The total exposure time was 4.07 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 20965 reflections to a maximum  $\theta$  angle of  $27.63^\circ$  ( $0.77 \text{ \AA}$  resolution), of which 4213 were independent (average redundancy 4.976, completeness = 99.0%,  $R_{\text{int}} = 8.70\%$ ,  $R_{\text{sig}} = 6.72\%$ ) and 3219 (76.41%) were greater than  $2\sigma(F^2)$ . The final cell constants of  $a = 25.024(2) \text{ \AA}$ ,  $b = 6.1831(6) \text{ \AA}$ ,  $c = 23.770(2) \text{ \AA}$ ,  $\beta = 94.7380(10)^\circ$ , volume =  $3665.3(6) \text{ \AA}^3$ , are based upon the refinement of the XYZ-centroids of 3217 reflections above  $20 \sigma(I)$  with  $4.935^\circ < 2\theta < 45.59^\circ$ . Data were corrected for absorption effects using the multi-scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.702. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.5231 and 0.7456. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group  $C 1 2/c 1$ , with  $Z = 8$  for the formula unit,  $C_{21}H_{21}BiO_3$ . The final anisotropic full-matrix least-squares refinement on  $F^2$  with 235 variables converged at  $R1 = 3.35\%$ , for the observed data and  $wR2 = 7.22\%$  for all data. The goodness-of-fit was 1.015. The largest peak in the final difference electron density synthesis was  $0.819 \text{ e}/\text{\AA}^3$  and the largest hole was  $-1.464 \text{ e}/\text{\AA}^3$  with an RMS deviation of  $0.191 \text{ e}/\text{\AA}^3$ . On the basis of the final model, the calculated density was  $1.922 \text{ g}/\text{cm}^3$  and  $F(000)$ , 2032 e.



**(*E*)-Ethyl 3-(3-(3-cyano-2-oxypyridin-1(2*H*)-yl)phenyl)acrylate (17a)****Figure S9.** ORTEP view at 50% ellipsoid probability for compound **17a**.

A colorless plate-like specimen of  $C_{17}H_{14}N_2O_3$ , approximate dimensions 0.028 mm x 0.139 mm x 0.546 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 5856 frames were collected. The total exposure time was 10.47 hours. The frames were integrated with the Bruker SAINT software package using a narrow-frame algorithm. The integration of the data using a triclinic unit cell yielded a total of 14337 reflections to a maximum  $\theta$  angle of  $68.31^\circ$  ( $0.83 \text{ \AA}$  resolution), of which 4311 were independent (average redundancy 3.326, completeness = 98.8%,  $R_{\text{int}} = 6.23\%$ ,  $R_{\text{sig}} = 4.18\%$ ) and 3773 (87.52%) were greater than  $2\sigma(F^2)$ . The final cell constants of  $a = 7.0152(3) \text{ \AA}$ ,  $b = 7.0736(4) \text{ \AA}$ ,  $c = 15.1206(8) \text{ \AA}$ ,  $\alpha = 83.644(3)^\circ$ ,  $\beta = 80.150(3)^\circ$ ,  $\gamma = 73.302(3)^\circ$ , volume =  $706.58(6) \text{ \AA}^3$ , are based upon the refinement of the XYZ-centroids of 4294 reflections above  $20 \sigma(I)$  with  $5.942^\circ < 2\theta < 136.3^\circ$ . Data were corrected for absorption effects using TWINABS. The ratio of minimum to maximum apparent transmission was 0.857. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.5767 and 0.7531. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group  $P -1$ , with  $Z = 2$  for the formula unit,  $C_{17}H_{14}N_2O_3$ . The final anisotropic full-matrix least-squares refinement on  $F^2$  with 202 variables converged at  $R1 = 4.72\%$ , for the observed data and  $wR2 = 13.54\%$  for all data. The goodness-of-fit was 1.090. The largest peak in the final difference electron density synthesis was  $0.219 \text{ e}/\text{\AA}^3$  and the largest hole was  $-0.196 \text{ e}/\text{\AA}^3$  with an RMS deviation of  $0.046 \text{ e}/\text{\AA}^3$ . On the basis of the final model, the calculated density was  $1.383 \text{ g}/\text{cm}^3$  and  $F(000)$ , 308 e $^-$ .



**Table S1.** Crystal and solution data for **1f**, **1i**, **1k**, **1n**, **1s**, **1t**, **21c**, **21e** and **17a**.

Name	<b>1f</b>	<b>1i</b>	<b>1k</b>	<b>1n</b>	<b>1s</b>
CCDC deposition number	1470682	1470683	1470684	1470685	1470686
Chemical formula	C <sub>27</sub> H <sub>27</sub> Bi	C <sub>18</sub> H <sub>9</sub> BiF <sub>6</sub>	C <sub>21</sub> H <sub>21</sub> BiO <sub>3</sub>	C <sub>33</sub> H <sub>45</sub> BiO <sub>6</sub>	C <sub>21</sub> H <sub>12</sub> BiN <sub>3</sub>
Formula weight (g/mol)	560.46	548.23	530.36	746.67	515.32
Temperature (K)	150(2)	150(2)	150(2)	150(2)	150(2)
Wavelength (Å)	0.71073	1.54178	0.71073	1.54178	0.71073
Crystal size (mm)	0.058 x 0.146 x 0.267	0.055 x 0.122 x 0.189	0.204 x 0.395 x 0.417	0.039 x 0.067 x 0.220	0.142 x 0.330 x 0.532
Crystal system	monoclinic	monoclinic	trigonal	monoclinic	monoclinic
Space group	P 2 <sub>1</sub> /n	P 2 <sub>1</sub> /c	R -3	P 2 <sub>1</sub> /c	P 2 <sub>1</sub> /c
<i>a</i> (Å)	17.2733(15)	5.16200(10)	13.0227(11)	12.2493(2)	8.5793(8)
<i>b</i> (Å)	6.1956(5)	15.7296(3)	13.0227(11)	40.0209(5)	11.6696(11)
<i>c</i> (Å)	20.4121(18)	19.6908(3)	18.9137(13)	23.2425(3)	17.9621(16)
$\alpha$ (°)	90	90	90	90	90
$\beta$ (°)	92.6470(10)	97.0350(10)	90	119.9300(10)	96.1690(10)
$\gamma$ (°)	90	90	120	90	90
Volume (Å <sup>3</sup> )	2182.1(3)	1586.78(5)	2777.9(5)	9874.6(3)	1787.9(3)
<i>Z</i>	4	4	6	12	4
Density (calculated, g/cm <sup>3</sup> )	1.706	2.295	1.902	1.507	1.914
Absorption coefficient (mm <sup>-1</sup> )	8.089	22.413	9.537	10.830	9.868
<i>F</i> (000)	1088	1016	1524	4488	968
Theta range for data collection (°)	1.51 to 27.65	3.61 to 68.28	2.10 to 27.53	2.21 to 68.39	2.09 to 27.50
Index ranges	-22 < <i>h</i> <= 22	-6 < <i>h</i> <= 6	-16 < <i>h</i> <= 16	-14 < <i>h</i> <= 14	-11 < <i>h</i> <= 11
	-7 < <i>k</i> <= 8	-18 < <i>k</i> <= 18	-16 < <i>k</i> <= 16	-48 < <i>k</i> <= 48	-15 < <i>k</i> <= 15
	-26 < <i>l</i> <= 26	-23 < <i>l</i> <= 23	-24 < <i>l</i> <= 24	-27 < <i>l</i> <= 28	-23 < <i>l</i> <= 23
Reflections collected	24743	22071	11061	149815	20762
Independent reflections ( <i>R</i> <sub>int</sub> )	5061 (0.0573)	2867 (0.0433)	1424 (0.0508)	18042 (0.0647)	4081 (0.0536)
Coverage of independent reflections	99.7%	98.6%	99.9%	99.5%	99.6%
Max. and min. transmission	0.7456 and 0.4835	0.7531 and 0.3941	0.7456 and 0.5092	0.7169 and 0.2896	0.7456 and 0.3684
Data / restraints / parameters	5061 / 0 / 253	2867 / 0 / 226	1424 / 0 / 77	18042 / 0 / 1099	4081 / 0 / 226
Goodness-of-fit on <i>F</i> <sup>2</sup>	1.016	1.088	1.103	1.004	1.017
Final <i>R</i> indices [ <i>I</i> > 2σ( <i>I</i> )]	<i>R</i> <sub>1</sub> = 0.0258	<i>R</i> <sub>1</sub> = 0.0189	<i>R</i> <sub>1</sub> = 0.0180	<i>R</i> <sub>1</sub> = 0.0267	<i>R</i> <sub>1</sub> = 0.0225
	<i>wR</i> <sub>2</sub> = 0.0472	<i>wR</i> <sub>2</sub> = 0.0486	<i>wR</i> <sub>2</sub> = 0.0377	<i>wR</i> <sub>2</sub> = 0.0565	<i>wR</i> <sub>2</sub> = 0.0439
	<i>R</i> <sub>1</sub> = 0.0387	<i>R</i> <sub>1</sub> = 0.0202	<i>R</i> <sub>1</sub> = 0.0198	<i>R</i> <sub>1</sub> = 0.0431	<i>R</i> <sub>1</sub> = 0.0321
	<i>wR</i> <sub>2</sub> = 0.0506	<i>wR</i> <sub>2</sub> = 0.0492	<i>wR</i> <sub>2</sub> = 0.0381	<i>wR</i> <sub>2</sub> = 0.0633	<i>wR</i> <sub>2</sub> = 0.0467
Largest diff. peak and hole (e Å <sup>-3</sup> )	0.659 and -0.848	0.564 and -0.898	0.575 and -0.819	0.550 and -1.148	0.501 and -0.842
Rms deviation from mean (e Å <sup>-3</sup> )	0.129	0.111	0.091	0.085	0.120
Absolute structure parameter					
(Flack)					

Table S1 (cont'd)

Name	1t	21c	21e	17a
CCDC deposition number	1470687	1470689	1470690	1470691
Chemical formula	C <sub>24</sub> H <sub>21</sub> BiO <sub>6</sub>	C <sub>21</sub> H <sub>15</sub> BiO <sub>3</sub>	C <sub>21</sub> H <sub>21</sub> BiO <sub>3</sub>	C <sub>17</sub> H <sub>14</sub> N <sub>2</sub> O <sub>3</sub>
Formula weight (g/mol)	614.39	524.31	530.36	294.30
Temperature (K)	150(2)	150(2)	150(2)	150(2)
Wavelength (Å)	0.71073	0.71073	0.71073	1.54178
Crystal size (mm)	0.038 x 0.266 x 0.307	0.053 x 0.104 x 0.147	0.045 x 0.054 x 0.140	0.028 x 0.139 x 0.546
Crystal system	triclinic	monoclinic	monoclinic	triclinic
Space group	P -1	P c	C 2/c	P -1
a (Å)	5.7360(7)	5.3061(7)	25.024(2)	7.0152(3)
b (Å)	10.0706(12)	16.256(2)	6.1831(6)	7.0736(4)
c (Å)	19.648(2)	10.1005(16)	23.770(2)	15.1206(8)
α (°)	96.3650(10)	90	90	83.644(3)
β (°)	90.7050(10)	98.294(2)	94.7380(10)	80.150(3)
γ (°)	98.8130(10)	90	90	73.302(3)
Volume (Å <sup>3</sup> )	1114.2(2)	862.1(2)	3665.3(6)	706.58(6)
Z	2	2	8	2
Density (calculated, g/cm <sup>3</sup> )	1.831	2.020	1.922	1.383
Absorption coefficient (mm <sup>-1</sup> )	7.949	10.242	9.637	0.791
F(000)	592	496	2032	308
Theta range for data collection (°)	2.06 to 27.80	1.25 to 27.51	1.63 to 27.63	2.97 to 68.31
Index ranges	-7<h<=7 -12<=k<=13 -25<=l<=25	-6<h<=6 -21<=k<=0 -13<=l<=5	-32<=h<=32 -7<=k<=8 -31<=l<=30	-8<h<=8 -8<=k<=8 0<=l<=18
Reflections collected	12705	15775	20965	14337
Independent reflections ( <i>R</i> <sub>int</sub> )	5155 (0.0508)	5124 (0.7117)	4213 (0.0870)	4311 (0.623)
Coverage of independent reflections	97.5%	99.9%	99.0%	98.8%
Max. and min. transmission	0.7456 and 0.4554	0.7456 and 0.4831	0.7456 and 0.5231	0.7531 and 0.5767
Data / restraints / parameters	5155 / 0 / 283	5124 / 2 / 215	4213 / 0 / 235	4311 / 0 / 202
Goodness-of-fit on <i>F</i> <sup>2</sup>	1.060	1.048	1.015	1.090
Final <i>R</i> indices [ <i>I</i> >2σ( <i>I</i> )]	<i>R</i> <sub>1</sub> = 0.0364 <i>wR</i> <sub>2</sub> = 0.0786	<i>R</i> <sub>1</sub> = 0.0361 <i>wR</i> <sub>2</sub> = 0.0881	<i>R</i> <sub>1</sub> = 0.0335 <i>wR</i> <sub>2</sub> = 0.0644	<i>R</i> <sub>1</sub> = 0.0472 <i>wR</i> <sub>2</sub> = 0.1276
Final <i>R</i> indices (all data)	<i>R</i> <sub>1</sub> = 0.0455 <i>wR</i> <sub>2</sub> = 0.0812	<i>R</i> <sub>1</sub> = 0.0684 <i>wR</i> <sub>2</sub> = 0.0993	<i>R</i> <sub>1</sub> = 0.0547 <i>wR</i> <sub>2</sub> = 0.0722	<i>R</i> <sub>1</sub> = 0.0547 <i>wR</i> <sub>2</sub> = 0.1354
Largest diff. peak and hole (e Å <sup>-3</sup> )	1.828 and -1.530	1.284 and -1.685	0.819 and -1.464	0.219 and -0.196
Rms deviation from mean (e Å <sup>-3</sup> )	0.175	0.198	0.191	0.046
Absolute structure parameter (Flack)		0.07(13)		

**Table S2.** Bi–C bond lengths and C–Bi–C angles in **1f**, **1i**, **1k**, **1n**, **1s**, **1t**, **21c**, and **21e**.

Compound	Bi–C (Å)			C–Bi–C (°)		
<b>1f</b>	2.253(3)	2.254(4)	2.256(4)	92.48(13)	94.93(13)	95.72(14)
<b>1i</b>	2.255(3)	2.263(3)	2.264(3)	92.94(12)	93.10(12)	96.19(12)
<b>1k*</b>	2.251(3)			93.80(9)		
<b>1n</b>	2.262(4)	2.272(4)	2.273(4)	93.42(13)	93.60(13)	96.12(14)
<b>1s</b>	2.263(3)	2.269(3)	2.273(3)	91.77(12)	93.07(12)	98.06(12)
<b>1t</b>	2.255(5)	2.260(5)	2.261(5)	92.10(18)	93.32(19)	94.81(17)
<b>21c</b>	2.232(18)	2.315(16)	2.324(16)	91.2(6)	92.2(6)	92.4(6)
<b>21e</b>	2.241(6)	2.254(6)	2.264(6)	91.5(2)	94.6(2)	95.6(2)

\* By symmetry, all Bi–C distances and C–Bi–C angles are equal.

## **ANNEXE C**

**Manuscript 2 : Chemoselective Copper-Catalyzed *O*-Arylation of Small Tyrosine-Containing Peptides using Functionnalized Organobismuth Reagent**

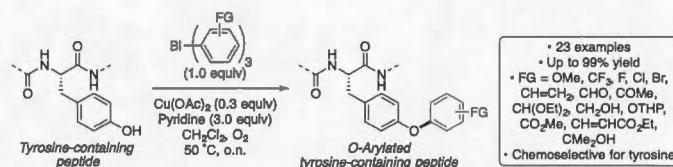


# Chemoselective Copper-Catalyzed *O*-Arylation of Small Tyrosine-Containing Peptides using Highly Functionalized Triarylbismuthines

Martin Hébert, Adrien Le Roch, Marie-Jeanne Archambault, Francis Pinsonneault, Hugo Lachance and Alexandre Gagnon\*

Département de chimie, Université du Québec à Montréal, C.P. 8888, Succ. Centre-Ville, Montréal, Québec, Canada, H3C 3P8

Supporting Information Placeholder



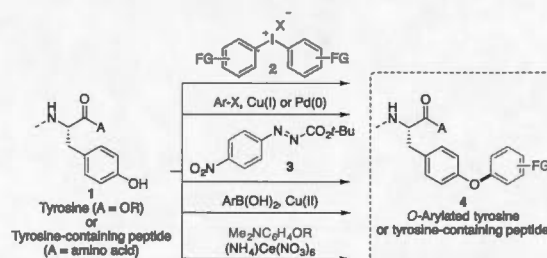
**ABSTRACT:** The modification of peptides is a valuable strategy to study the function of proteins and to modulate their biophysical properties and biological activities. Tyrosine is an attractive residue for protein modification because the phenolic group has a distinct reactivity from other amino acids and because tyrosine is often present at the surface of proteins. We report herein a copper-catalyzed *O*-arylation reaction of small tyrosine-containing peptides involving highly functionalized triarylbismuthines. The reaction allows the transfer of highly functionalized aryl groups and shows excellent chemoselectivity towards tyrosine over other amino acids.

The modification of peptides by covalent bond formation has emerged as a powerful strategy to study the function of proteins.<sup>1</sup> Modification of single amino acids on peptides can lead to changes in conformation, biophysical properties, pharmacodynamic profile and biological activity.<sup>2</sup> Because of their greater accessibility, most of the protein modifications target lysines and cysteines. However, the higher reactivity of these residues can lead to reaction at multiple sites. The modification of tyrosine residues in peptides is particularly attractive since the reactivity of phenols is substantially distinct from that of functional groups contained in other amino acids. Moreover, due to their amphiphilic nature, the phenolic side chains of tyrosine are often present at the surface of proteins, making them accessible for chemical modification.

The *O*-arylation of tyrosine is an interesting strategy for peptide modification. Methods to *O*-arylate tyrosine include reaction with iodonium reagents,<sup>3</sup> copper<sup>4</sup> and palladium-catalyzed<sup>5</sup> coupling with aryl halides and arylation with *p*-nitro phenylazocarboxylates (**Scheme 1**).<sup>6</sup> To our knowledge, the *O*-arylation of tyrosine-containing peptides has only been accomplished using two approaches, namely, the oxidative coupling with *p*-dimethylaminoanisole<sup>7</sup> and the arylation with arylboronic acids (**Scheme 1**).<sup>8</sup> Other transformations that target tyrosine include the *O*-alkylation of the phenol using allylpalladium complexes,<sup>9</sup> reaction at the *ortho* position using Mannich-type reactions<sup>10</sup> and coupling with diazonium reagents.<sup>11</sup> However, there is still a need for *O*-

arylation reaction methods that allow the chemoselective installation of highly functionalized aryl groups on tyrosine and tyrosine-containing peptides.

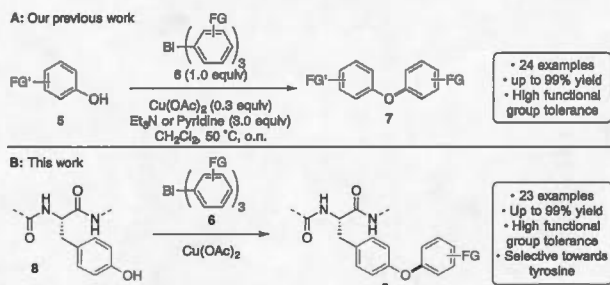
## Scheme 1. Methods for the *O*-Arylation of Tyrosine and Tyrosine-Containing Peptides



Triarylbismuthines are a class of organometallic reagents that can be easily prepared from inexpensive and non-toxic bismuth salts.<sup>12</sup> These reagents are particularly attractive since they are air and moisture-stable, have low toxicity and show remarkable functional group tolerance. In the 1980s, Barton and Finet reported a series of seminal papers on the use of triarylbismuthines as arylating agents.<sup>13</sup> Triarylbismuthines have also found wide applications in the preparation of transition metal complexes,<sup>14</sup> in total synthesis,<sup>15</sup> in palladium-catalyzed cross-coupling reactions<sup>16</sup> and in other reactions.<sup>17</sup> Barton reported the *N*-arylation of unprotected amino acids esters using triphenylbismuth diacetate.<sup>18</sup> However, the arylation of side chains of amino acids using trivalent bismuth reagents has never been reported.

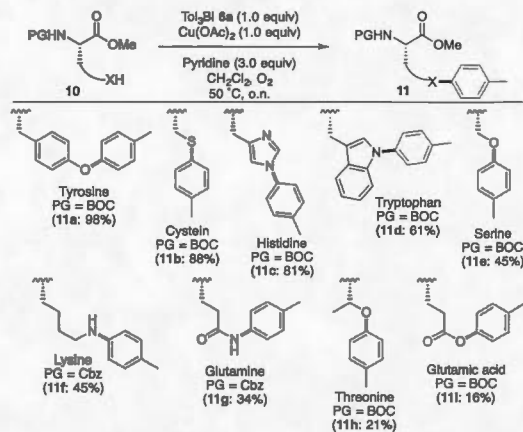
We reported over the past years a portfolio of reactions for the construction C–C,<sup>19</sup> C–N<sup>20</sup> and C–O<sup>21</sup> bonds involving organobismuth reagents.<sup>22</sup> These protocols operate under mild conditions, show high functional groups tolerance and allow the transfer of functionalized aryl and alkyl groups to medicinally relevant scaffolds. We recently demonstrated that highly functionalized diarylethers can be prepared via the copper-catalyzed *O*-arylation of phenols using triarylbi-muthines (Scheme 2A).<sup>21c</sup> We report herein our studies on the copper-catalyzed *O*-arylation of tyrosine-containing peptides using triarylbi-muthines (Scheme 2B).<sup>23</sup>

### Scheme 2. Copper-Catalyzed Arylation of Phenols Using Triarylbi-muthines and Application to the Arylation of Tyrosine-Containing Peptides



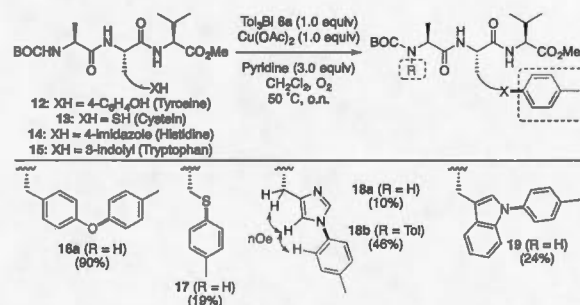
We began by evaluating the reactivity of different amino acids possessing a nucleophilic side chain under the conditions developed for the *O*-arylation of phenols (Scheme 3). Using 1.0 equivalent of tritolybi-muthine, 0.3 equivalents of copper acetate, and 3.0 equivalents of pyridine in dichloromethane under oxygen at 50 °C overnight, we found that tyrosine afforded the highest yield of the corresponding arylated product, followed by cysteine and histidine (products 11a,b,c). We previously demonstrated that indoles can be efficiently arylated using triarylbi-muthines under copper catalysis. Here, tryptophan gave the desired *N*-tolyl derivative 11d in 61% yield which is slightly lower than what we observed for other simpler indoles. Serine, lysine and glutamine could also be arylated using our protocol, affording the desired arylated amino acids 11e–g in yields ranging from 34% to 45%. Previous results from our group on the copper-catalyzed *O*-arylation of 1,2-aminoalcohols using triarylbi-muthines showed that the arylation of secondary alcohols is much more challenging than primary alcohols.<sup>20b</sup> Therefore, as expected, threonine was arylated in a much lower yield than serine (21% for 11h vs 45% for 11e). Finally, glutamic acid provided the desired *O*-tolyl product 11i in only 16% yield while aspartic acid failed to provide the desired arylation product, showing that carboxylic acids are poorly reactive under these reaction conditions. All together, these results show that while this copper-catalyzed protocol can be used to install aryl groups on the side chain of numerous amino acids, tyrosine is the most reactive amino acid.

### Scheme 3. Copper-Catalyzed Arylation of Amino Acids Using Tritolybi-muthine



We then verified the reactivity of the four most reactive amino acids in the context of tripeptides. To achieve this, we prepared tripeptides where tyrosine, cysteine, histidine and tryptophan are flanked by an alanine on the *N*-terminus and a valine on the *C*-terminus and submitted them to our copper-catalyzed arylation conditions (Scheme 4). The results show that tyrosine retains its reactivity when incorporated in a tripeptide (16a). On the contrary, we found that cysteine, histidine and tryptophan lose most of their reactivity when incorporated in a tripeptide (17, 18a, 19). In the case of the histidine-containing peptide, *n*Oe studies demonstrated that the arylation occurred at the distal nitrogen. Interestingly, a product of double arylation where a tolyl group is added at the BOC-amide was also isolated from the reaction of BOC-Ala-His-Val-OMe 18b. These results confirm that tyrosine is the most reactive amino acid under the copper-catalyzed arylation using triarylbi-muthines.

### Scheme 4. Copper-Catalyzed Arylation of Tripeptides Using Tritolybi-muthine

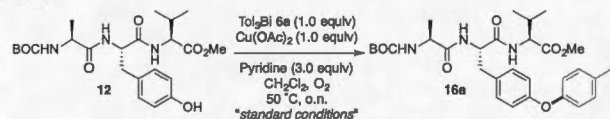


Optimization of the reaction conditions for the tolylation of BOC-Ala-Tyr-Val-OMe 12 showed that other solvents such as toluene, acetonitrile, DMF and THF provide lower yields of the desired arylated peptide 16a (Table 1, Entries 2–5). Use of only 1.0 equivalent of pyridine (Entry 6) or replacement by triethylamine (Entry 7) was found to be well tolerated. Reduction of the catalyst loading to 0.3 equivalents was well tolerated also (Entry 8) but further reduction to 0.1 equivalent was detrimental to the reaction (Entry 9). It is well known that in copper-catalyzed reactions involving triarylbi-muth reagents, only one aryl group is usually transferred. In this case, product 16a was obtained in 75% yield upon running the reaction with 0.7 equivalents of tritolybi-muth (Entry 10). Finally, an erosion in the yield of the reaction was observed upon



conducting the reaction under air (Entry 11) or argon (Entry 12), showing that oxygen is essential to obtain optimal yields. A drastic drop in the yield was also observed when the reaction was performed at room temperature (Entry 13).

**Table 1. Optimization of the Reaction Conditions for the Copper-Catalyzed Arylation of BOC-Ala-Tyr-Val-OMe**



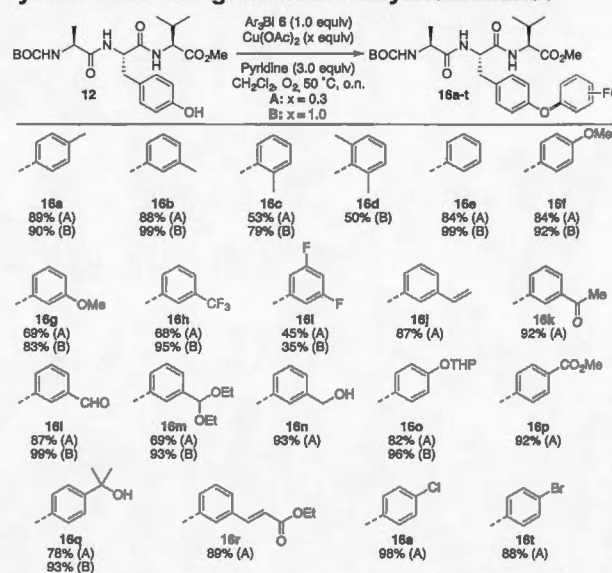
Entry	Change from "standard conditions"	Yield (%) <sup>a</sup>
1	No change	90
2	Toluene instead of DCM	79
3	Acetonitrile instead of DCM	27
4	DMF instead of DCM	39
5	THF instead of DCM	77
6	1.0 Equiv of Pyridine instead of 3.0 equiv	84
7	Et <sub>3</sub> N instead of Pyridine	83
8	0.3 Equiv Cu(OAc) <sub>2</sub> instead of 1.0 equiv	89
9	0.1 Equiv Cu(OAc) <sub>2</sub> instead of 1.0 equiv	56
10	0.7 Equiv Tol <sub>3</sub> Bi instead of 1.0 equiv	75
11	Air instead of O <sub>2</sub>	74
12	Argon instead of O <sub>2</sub>	55
13	r.t. instead of 50 °C	25

<sup>a</sup> Yield are for isolated pure product.

We then performed the arylation reaction on tyrosine-containing tripeptide **12** using diversely substituted triarylbiomethines **6** (Scheme 5). The reaction was conducted using 1.0 equivalent of triarylbiomethine, 3.0 equivalents of pyridine, and 0.3 equivalents of copper acetate in dichloromethane at 50 °C under oxygen (Method A). In cases where lower yields were obtained, the reaction was also performed using 1.0 equivalent of copper acetate (Method B). The results show that *para*-, *meta*-, and unsubstituted aryl groups can be transferred with similar efficiency (**16a,b,e**). Interestingly, only a minor reduction in the yield was observed in the transfer of an *ortho*-tolyl group (**16c**), showing that *ortho*-substitution is well tolerated. The method even allowed the installation of a highly hindered 2,6-dimethylphenyl fragment in 50% yield (**16d**). Triarylbiomethines possessing a methoxy group at the *para* or *meta* position afforded the corresponding compounds **16f** and **16g**, showing that the presence of electron donating and withdrawing groups is well tolerated. The presence of a trifluoromethyl on the aryl group was also tolerated (**16h**). However, the presence of fluorine atoms at the 3- and 5-positions led to a substantial drop in the yield of the reaction (**16i**). The method showed remarkable functional group tolerance as indicated by the efficient coupling of triarylbiomethine reagents possessing a vinyl moiety (**16j**), a methylketone (**16k**), an aldehyde (**16l**), an acetal (**16m**), a primary alcohol (**16n**), a THP-protected phenol (**16o**), a methyl

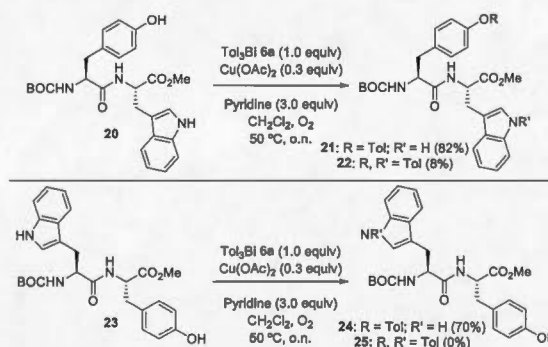
ester (**16p**), a tertiary alcohol (**16q**), an  $\alpha,\beta$ -unsaturated ester (**16r**), a chloride (**16s**) and a bromide (**16t**).

**Scheme 5. Copper-Catalyzed Arylation of BOC-Ala-Tyr-Val-OMe Using Various Triarylbiomethines**



**Method A:** Ar<sub>3</sub>Bi (1.0 equiv), Pyridine (3.0 equiv), Cu(OAc)<sub>2</sub> (0.3 equiv), CH<sub>2</sub>Cl<sub>2</sub>, O<sub>2</sub>, 50 °C, o.n.; **Method B:** Ar<sub>3</sub>Bi (1.0 equiv), Pyridine (3.0 equiv), Cu(OAc)<sub>2</sub> (1.0 equiv), CH<sub>2</sub>Cl<sub>2</sub>, O<sub>2</sub>, 50 °C, o.n.

**Scheme 6. Chemoselectivity Study in the Arylation of BOC-Tyr-Trp-OMe and BOC-Trp-Tyr-OMe**



Next, we evaluated the chemoselectivity of the arylation protocol by performing the reaction on BOC-Tyr-Trp-OMe **20** (Scheme 6). Using 0.3 equivalents of copper acetate and tritolybiomethine, we obtained the corresponding *O*-tolyl peptide **21** in 82% yield along with 8% of the ditolyl product **22**. To test whether the position of the tyrosine had an impact on the outcome of the process, the reaction was also performed on BOC-Trp-Tyr-OMe **23**. In this case, only the *O*-tolyl product **24** was obtained in a slightly reduced yield. Moreover, since none of the ditolyl product **25** was obtained, this suggests that the reactivity of tyrosine is moderately affected by its position in the peptide and/or by their neighbouring amino acids. Surprisingly, we could not isolate any of the product from mono arylation on the indole. The results from arylation on peptide **20** and **23** demonstrate that tyrosine is more reactive than tryptophan in the copper-catalyzed arylation reaction using triarylbiomethines.

In summary, we developed a chemoselective copper-catalyzed O-arylation reaction of small tyrosine-containing peptides. The reaction operates under mild conditions, allows the introduction of *ortho*-, *meta*-, and *para*-substituted aryl groups and shows exceptional functional group tolerance. We demonstrated that the reaction is selective for tyrosine and that the outcome of the process is moderately affected by the position of the tyrosine in the peptide. The application of this method to the arylation of more elaborated peptides is under investigation in our laboratories and results will be reported in due course.

## ASSOCIATED CONTENT

### Supporting Information

Experimental procedures, characterization data, and copies of NMR and IR spectra. This material is available free of charge via the Internet at <http://pubs.acs.org>.

## AUTHOR INFORMATION

### Corresponding Author

\*E-mail: [gagnon.alexandre@uqam.ca](mailto:gagnon.alexandre@uqam.ca)

### Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENT

This work was supported by the Natural Sciences and Engineering Research Council of Canada (NSERC), the Université du Québec à Montréal (UQAM) and the Centre in Green Chemistry and Catalysis (CGCC). M.H. thanks NSERC and FRQNT for postgraduate scholarships.

## REFERENCES

- (1) Qi, D.; Tann, C.-M.; Haring, D.; Distefano, M. D. *Chem. Rev.* **2001**, *101*, 3081-3111.
- (2) Davis, B. G. *Curr. Opin. Biotechnol.* **2003**, *14*, 379-386.
- (3) (a) Jalalian, N.; Petersen, T. B.; Olofsson, B. *Chem. Eur. J.* **2012**, *18*, 14140-14149; (b) Jalalian, N.; Ishikawa, E. E.; Silva, L. F.; Olofsson, B. *Org. Lett.* **2011**, *13*, 1552-1555.
- (4) (a) Cai, Q.; Zou, B.; Ma, D. *Angew. Chem. Int. Ed.* **2006**, *45*, 1276-1279; (b) Fan, M.; Zhou, W.; Jiang, Y.; Ma, D. *Angew. Chem. Int. Ed.* **2016**, *55*, 6211-6215; (c) Cai, Q.; He, G.; Ma, D. *J. Org. Chem.* **2006**, *71*, 5268-5273.
- (5) Chapman, C. J.; Matsuno, A.; Frost, C. G.; Willis, M. C. *Chem. Commun.* **2007**, 3904-3905.
- (6) (a) Jasch, H.; Höfling, S. B.; Heinrich, M. R. *J. Org. Chem.* **2012**, *77*, 1520-1532; (b) Höfling, S. B.; Barthuschat, A. L.; Heinrich, M. R. *Angew. Chem. Int. Ed.* **2010**, *49*, 9769-9772.
- (7) Seim, K. L.; Obermeyer, A. C.; Francis, M. B. *J. Am. Chem. Soc.* **2011**, *133*, 16970-16976.
- (8) Kilitoglu, B.; Arndt, H.-D. *Synlett* **2009**, 720-723.
- (9) Tilley, S. D.; Francis, M. B. *J. Am. Chem. Soc.* **2006**, *128*, 1080-1081.
- (10) Joshi, N. S.; Whitaker, L. R.; Francis, M. B. *J. Am. Chem. Soc.* **2004**, *126*, 15942-15943.
- (11) Hooker, J. M.; Kovacs, E. W.; Francis, M. B. *J. Am. Chem. Soc.* **2004**, *126*, 3718-3719.
- (12) For reviews on organobismuth reagents, see: (a) Condon, S.; Pichon, C.; Davi, M. *Org. Prep. Proc. Int.* **2014**, *46*, 89-131; (b) Elliott, G. I.; Konopelski, J. P. *Tetrahedron* **2001**, *57*, 5683-5705; (c) Suzuki, H.; Ikegami, T.; Matano, Y. *Synthesis* **1997**, 249-267; (d) Finet, J.-P. *Chem. Rev.* **1989**, *89*, 1487-1501.
- (13) Abramovitch, R. A.; Barton, D. H. R.; Finet, J.-P. *Tetrahedron* **1988**, *44*, 3039-3071 and references cited therein.
- (14) (a) Benjamin, S. L.; Reid, G. *Coord. Chem. Rev.* **2015**, *297-298*, 168-180; (b) Braunschweig, H.; Cogswell, P.; Schwab, K. *Coord. Chem. Rev.* **2011**, *255*, 101-117.
- (15) (a) Nicolaou, K. C.; Sarlah, D.; Wu, T. R.; Zhan, W. *Angew. Chem. Int. Ed.* **2009**, *48*, 6870-6874; (b) Krawczuk, P. J.; Schöne, N.; Baran, P. S. *Org. Lett.* **2009**, *11*, 4774-4776.
- (16) (a) Rao, M. L. N.; Dhanorkar, R. J. *RSC Adv.* **2016**, *6*, 1012-1017; (b) Rao, M. L. N.; Dhanorkar, R. J. *Eur. J. Org. Chem.* **2014**, 5214-5228; (c) Rao, M. L. N.; Dhanorkar, R. J. *RSC Adv.* **2014**, *4*, 13134-13144; (d) Rao, M. L. N.; Jadhav, D. N.; Dasgupta, P. *Eur. J. Org. Chem.* **2013**, 781-788; (e) Rao, M. L. N.; Banerjee, D.; Giri, S. J. *Organomet. Chem.* **2010**, *695*, 1518-1525; (f) Rao, M. L. N.; Jadhav, D. N.; Dasgupta, P. *Org. Lett.* **2010**, *12*, 2048-2051.
- (17) (a) Kobiki, Y.; Kawaguchi, S.-i.; Ogawa, A. *Org. Lett.* **2015**, *17*, 3490-3493; (b) Wang, T.; Sang, S.; Liu, L.; Qiao, H.; Gao, Y.; Zhao, Y. *J. Org. Chem.* **2014**, *79*, 608-617; (c) Sato, I.; Toyota, Y.; Asakura, N. *Eur. J. Org. Chem.* **2007**, 2608-2610; (d) Cho, C. S.; Yoshimori, Y.; Uemura, S. *Bull. Chem. Soc. Jpn.* **1995**, *68*, 950-957.
- (18) Barton, D. H. R.; Finet, J. -P.; Khamsi, J. *Tetrahedron Lett.* **1989**, *30*, 937-940.
- (19) (a) Petiot, P.; Gagnon, A. *Heterocycles* **2014**, *88*, 1615-1624; (b) Petiot, P.; Gagnon, A. *Eur. J. Org. Chem.* **2013**, 5282-5289; (c) Gagnon, A.; Albert, V.; Duplessis, M. *Synlett* **2010**, 2936-2940; (d) Gagnon, A.; Duplessis, M.; Fader, L. *Org. Prep. Proc. Int.* **2010**, *42*, 1-69; (e) Gagnon, A.; Duplessis, M.; Alsabeh, P.; Barabé, F. *J. Org. Chem.* **2008**, *73*, 3604-3607.
- (20) (a) Petiot, P.; Dansereau, J.; Gagnon, A. *RSC Adv.* **2014**, *4*, 22255-22259; (b) Gagnon, A.; St-Onge, M.; Little, K.; Duplessis, M.; Barabé, F. *J. Am. Chem. Soc.* **2007**, *129*, 44-45.
- (21) (a) Ahmad, T.; Dansereau, J.; Hébert, M.; Grand-Maitre, C.; Larivée, A.; Siddiqui, A.; Gagnon, A. *Tetrahedron Lett.* **2016**, doi.org/10.1016/j.tetlet.2016.08.021; (b) Petiot, P.; Dansereau, J.; Hébert, M.; Khene, I.; Ahmad, T.; Samaali, S.; Leroy, M.; Pinsonneault, F.; Legault, C. Y.; Gagnon, A. *Org. Biomol. Chem.* **2015**, *13*, 1322-1327; (c) Crifar, C.; Petiot, P.; Ahmad, T.; Gagnon, A. *Chem. Eur. J.* **2014**, *20*, 2755-2760.
- (22) Hébert, M.; Petiot, P.; Benoit, E.; Dansereau, J.; Ahmad, T.; Le Roch, A.; Ottenwaelder, X.; Gagnon, A. *J. Org. Chem.* **2016**, *81*, 5401-5416.
- (23) For a review on modification of peptides by transition metal catalysis, see: Antos, J. M.; Francis, M. B. *Curr. Opin. Chem. Biol.* **2006**, *10*, 253-262.



## Supporting Information

### Chemoselective Copper-Catalyzed O-Arylation of Small Tyrosine-Containing Peptides Using Fonctionnalized Organobismuth Reagents

Martin Hébert,<sup>a</sup> Adrien Le Roch,<sup>a</sup> Marie-Jeanne Archambault,<sup>a</sup> Francis Pinsonneault,<sup>a</sup> Hugo Lachance,<sup>a</sup>  
and Alexandre Gagnon<sup>a,b\*</sup>

<sup>a</sup> Université du Québec à Montréal, Département de chimie, C.P. 8888, Succursale Centre-Ville,  
Montréal, Québec, Canada, H3C 3P8. E-mail: [gagnon.alexandre@uqam.ca](mailto:gagnon.alexandre@uqam.ca)

<sup>b</sup> Centre in Green Chemistry and Catalysis

### Table of Contents

#### EXPERIMENTAL SECTION

General Information:.....S1

General procedure for the synthesis of triarylbismuthines:.....S2

General procedure for the arylation of amino acids and peptides:.....S2

EXPERIMENTAL PROCEDURES AND CHARACTERIZATION.....S2

#### Spectral data:

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **6s**.....S32

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **6t**.....S34

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **11a**.....S36

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **11b**.....S38

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **11c**.....S40

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **11d**.....S42

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **11e**.....S44

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **11f**.....S46

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **11g**.....S48

<sup>1</sup>H, <sup>13</sup>C NMR spectra of **11h**.....S50

$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>11i</b> .....	S52
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>12i</b> .....	S54
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>12</b> .....	S56
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>13i</b> .....	S58
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>13ii</b> .....	S60
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>13</b> .....	S62
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>14i</b> .....	S64
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>14</b> .....	S66
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>15i</b> .....	S68
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>15</b> .....	S70
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16a</b> .....	S72
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16b</b> .....	S74
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16c</b> .....	S76
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16d</b> .....	S78
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16e</b> .....	S80
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16f</b> .....	S82
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16g</b> .....	S84
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16h</b> .....	S86
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16i</b> .....	S88
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16j</b> .....	S90
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16k</b> .....	S92
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16l</b> .....	S94
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16m</b> .....	S96
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16n</b> .....	S98
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16o</b> .....	S100
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16p</b> .....	S102
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16q</b> .....	S104
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16r</b> .....	S106
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16s</b> .....	S108

$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>16t</b> .....	S110
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>17</b> .....	S112
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>18a</b> .....	S114
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>18b</b> .....	S116
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>19</b> .....	S118
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>20</b> .....	S120
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>21</b> .....	S122
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>22</b> .....	S124
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>23</b> .....	S126
$^1\text{H}$ , $^{13}\text{C}$ NMR spectra of <b>24</b> .....	S128

## EXPERIMENTAL SECTION

**General Information:** Unless otherwise indicated, all reactions were run under argon in non-flame dried glassware. For reactions performed under oxygen, 99.6% extra dry oxygen was used. Unless otherwise stated, commercial reagents were used without further purification. Grignard reagents were prepared by conventional methods using metallic magnesium or via Knochel's procedure.<sup>1</sup> Anhydrous bismuth chloride >98% was purchased from Aldrich. Triphenylbismuth diacetate was purchased from Aldrich. Triarylbiomuthines **6a**, **6c**, **6g**, **6o** and **6p** were prepared according to a procedure that we previously reported.<sup>2</sup> Triarylbiomuthines **6b**, **6l** and **6m** were prepared according to methods previously reported by us.<sup>3</sup> Triarylbiomuthines **6d**, **6f**, **6h**, **6i**, **6q** and **6r** were prepared according to a procedure that we previously reported.<sup>4</sup> Organobiomuthines **6e**, **6j**, **6k** and **6n** were prepared according to methods we previously reported.<sup>5</sup> Anhydrous solvents were obtained using a MBRAUN (model MB-SPS 800) encapsulated solvent purification system and were further dried over 4 Å molecular sieves. The progress of reactions was monitored by analytical thin-layer chromatography using silica gel 60 F254 precoated plates. Flash chromatography was performed employing 230-400 mesh silica (Silicycle) using the indicated solvent system according to standard techniques. Melting points were taken on an Electrothermal Mel-TEMP and are uncorrected. Nuclear magnetic resonance spectra (<sup>1</sup>H, <sup>13</sup>C) were recorded on a Bruker Avance-III 300 MHz NanoBay or a Bruker Avance-III HD 600 MHz spectrometer. Chemical shifts for <sup>1</sup>H-NMR spectra are recorded in parts per million from tetramethylsilane with the solvent resonance as the internal standard (chloroform, δ 7.26 ppm; methanol, δ 3.31 ppm). Data is reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, qt = quintuplet, dd = doublet of doublet, dt = doublet of triplet, m = multiplet), coupling constant J in Hz and integration. Chemical shifts for <sup>13</sup>C spectra are recorded in parts per million from tetramethylsilane using the central peak of deuteriochloroform (δ 77.16 ppm) or the central peak of tetradeuteromethanol (δ 49.00 ppm) as the internal standard. IR spectra were recorded on a Thermo Scientific Nicolet 6700 PT-IR from thin films and are reported in reciprocal centimeters (cm<sup>-1</sup>). HRMS were performed at Université du Québec à Montréal (nanoQAM) on Agilent Technologies, LC 1200 Series / 6210 TOF LCMS analyzer using the electrospray (ESI) mode.

<sup>1</sup> Dohle, W.; Lindsay, D. M.; Knochel, P. *Org. Lett.* **2001**, *18*, 2871-2873.

<sup>2</sup> Petiot, P.; Gagnon, A. *Eur. J. Org. Chem.* **2013**, *2013*, 5282-5289.

<sup>3</sup> Crifar, C.; Petiot, P.; Ahmad, T.; Gagnon, A. *Chem. Eur. J.* **2014**, *20*, 2755-2760.

<sup>4</sup> Petiot, P.; Dansereau, J.; Gagnon, A. *RSC Adv.* **2014**, *4*, 22255-22259.

<sup>5</sup> Hébert, M.; Petiot, P.; Benoit, E.; Dansereau, J.; Ahmad, T.; Le Roch, A.; Ottenwaelder, X.; Gagnon, A. *J. Org. Chem.* **2016**, *81*, 5401-5416.

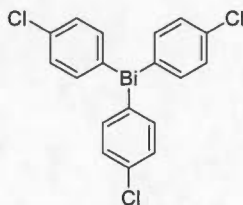


**General procedure for the synthesis of triarylbiomuthines:** Organobismuthines (**6a-i**, **6m**, **6o** and **6s**) were prepared according to the following procedure: In a flask equipped with a magnetic stir bar and a condenser, bismuth chloride (500 mg, 1.6 mmol) was dissolved in anhydrous THF (23 mL) under argon and was cooled to  $-10\text{ }^{\circ}\text{C}$  (ice/acetone bath). The organomagnesium reagent (5.23 mmol) was slowly added dropwise under argon. The reaction mixture was stirred at room temperature for 1 h and heated at  $65\text{ }^{\circ}\text{C}$  for 30 minutes. After cooling to room temperature, the solution was diluted with sat. aq.  $\text{NaHCO}_3$  (100 mL) and extracted with EtOAc (2 X 100 mL). The combined organic phases were washed with sat. aq.  $\text{NaHCO}_3$  (2 X 100 mL), sat. aq.  $\text{NaCl}$  (2 X 100 mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered and concentrated under reduced pressure. The crude product was purified by column chromatography using the indicated solvent system to afford the desired triarylbiomuthine.

**General procedure for the arylation of amino acids and peptides:** Compounds (**11a-i**, **16a-t**, **17**, **18a-b**, **19**, **21**, **22** and **24**) were prepared according to the following procedures: **Method A:** In a sealed tube, the amino acid or peptide (0.1 mmol) was dissolved in non-anhydrous solvent grade dichloromethane (2 mL). The organobismuthine (0.1 mmol) was added followed by copper (II) acetate (0.03 mmol) and pyridine (0.3 mmol). The tube was sealed and heated at  $50\text{ }^{\circ}\text{C}$  under  $\text{O}_2$  for 16 h. The reaction mixture was cooled to room temperature and silica gel was added. The mixture was concentrated under reduced pressure and the crude product was purified by flash column chromatography using the indicated solvent system to afford the corresponding product. **Method B:** Same as method **A** except that 0.1 mmol of  $\text{Cu}(\text{OAc})_2$  was used.

## EXPERIMENTAL PROCEDURES AND CHARACTERIZATION

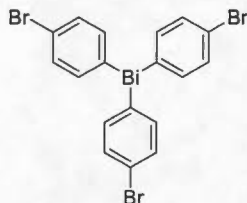
### Tris(4-chlorophenyl)bismuthine (**6s**)



The general procedure was followed on a 3.68 mmol scale starting from bismuth chloride and 4-chlorophenylmagnesium bromide. The crude product was purified on silica gel (2% EtOAc/hexanes) to afford tris(4-chlorophenyl)bismuthine **6s** as a white solid (1.6824 g, 84%); mp  $106\text{--}107\text{ }^{\circ}\text{C}$ ;  $R_f$  0.24 (2% EtOAc/hexanes).  $^1\text{H-NMR}$  (300 MHz)  $\delta$  7.64–7.60 (m, 6H), 7.39–7.35 (m, 6H);  $^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )  $\delta$

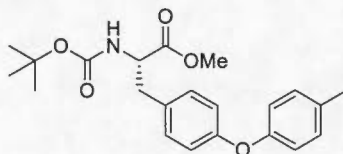
152.9, 138.9, 134.6, 131.1; IR (neat) 3032, 1900, 1556, 1467, 1373, 1176, 1082, 1042, 1001, 805, 711; HRMS (ESI) calcd for  $[C_{18}H_{12}Cl_3Bi + HCOO]^-$ : 586.9790, found 586.9741.

**Tris(4-bromophenyl)bismuthine (6t)**



1,4-Dibromobenzene (3.0974g, 13.128 mmol) was added to a flame dried round bottom flask and a magnetic stir bar. 1,4-Dibromobenzene was dissolved in anhydrous THF (13 mL) and the flask was chased with argon and cooled to  $-78^{\circ}C$  in an acetone/dry ice bath. *n*-BuLi (2.5 M in hexanes) (11.816 mmol) was added dropwise and the reaction mixture was stirred at  $-78^{\circ}C$  for 1 h. In another flame dried round bottom flask,  $BiCl_3$  was added, the flask was chased with argon, the  $BiCl_3$  was dissolved in anhydrous THF (33 mL) and the solution was cooled to  $-78^{\circ}C$  in an acetone/dry ice bath. The lithium-halogen exchange solution was transferred to the  $BiCl_3$  solution using a double tipped needle and argon pressure. The reaction mixture was stirred at  $-78^{\circ}C$  for 3 h and it was then warmed to room temperature and stirred for 16 h. EtOAc (50 mL) was added to the reaction mixture, it was then diluted with sat. aq.  $NaHCO_3$  (100 mL) and extracted with EtOAc (2 X 50 mL). The combined organic phases were washed with sat. aq.  $NaHCO_3$  (2 X 100 mL), sat. aq.  $NaCl$  (2 X 100 mL), dried over  $Na_2SO_4$  and concentrated over reduced pressure. The crude product was purified on silica gel (100% hexanes) to afford tris(4-bromophenyl)bismuthine **6t** as a white solid (1.4211 g, 71%): mp  $132-133^{\circ}C$ ;  $R_f$  0.19 (100% hexanes).  $^1H$ -NMR (300 MHz)  $\delta$  7.56-7.53 (m, 6H), 7.52-7.49 (m, 6H);  $^{13}C$ -NMR (150 MHz,  $CDCl_3$ )  $\delta$  153.5, 139.2, 134.0, 123.1; IR (neat) 3056, 1548, 1467, 1368, 1070, 1037, 993, 792, 686; HRMS (ESI) calcd for  $[C_{18}H_{12}Br_3Bi + HCOO]^-$ : 718.8275, found 718.8284.

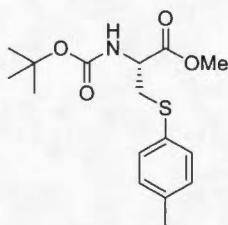
**(S)-Methyl 2-((*tert*-butoxycarbonyl)amino)-3-(4-(*p*-tolylloxy)phenyl)propanoate (11a)**



Method B was followed on a 0.169 mmol scale starting from **10a** and organobismuthine **6a**. The crude product was purified on silica gel (15% EtOAc/hexanes) to afford **11a** as a colorless oil (64.0 mg, 98%):  $R_f$  0.34 (20% EtOAc/hexanes);  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.15-7.12 (m, 2H), 7.08-7.05 (m, 2H), 6.92-6.88 (m, 4H), 5.03-4.75 (m, 1H), 4.60-4.38 (m, 1H), 3.72 (s, 3H), 3.09 (dd,  $J = 13.8, 5.7$  Hz, 1H), 3.00 (dd,  $J = 14.0,$

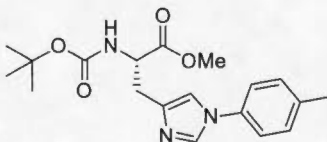
6.1 Hz, 1H), 2.33 (s, 3H), 1.42 (s, 9H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.4, 157.0, 155.2, 154.7, 133.1, 130.6, 130.4, 130.3, 119.3, 118.4, 80.1, 54.6, 52.3, 37.8, 28.4, 20.8; IR (neat) 3371, 3027, 2976, 1744, 1713, 1604, 1498, 1436, 1365, 1235, 1162, 1056, 1015, 873, 814; HRMS (ESI) calcd for  $[\text{C}_{22}\text{H}_{27}\text{NO}_5 + \text{Na}]^+$ : 408.1781, found 408.1795.

**(R)-Methyl 2-((*tert*-butoxycarbonyl)amino)-3-(*p*-tolylthio)propanoate (11b)**



Method B was followed on a 0.215 mmol scale starting from **10b** and organobismuthine **6a**. The crude product was purified on silica gel (0.5% MeOH/ $\text{CH}_2\text{Cl}_2$ ) to afford **11b** as a pale yellow oil (61.6 mg, 88%):  $R_f$  0.23 (0.5% MeOH/ $\text{CH}_2\text{Cl}_2$ );  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.31 (d,  $J$  = 8.1 Hz, 2H), 7.09 (d,  $J$  = 7.9 Hz, 2H), 5.50–4.97 (m, 1H), 4.65–4.20 (m, 1H), 3.53 (s, 3H), 3.31 (d,  $J$  = 4.9 Hz, 2H), 2.30 (s, 3H), 1.41 (s, 9H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  171.2, 155.0, 137.4, 131.9, 131.0, 129.9, 80.1, 53.4, 52.4, 37.9, 28.4, 21.1; IR (neat) 3367, 3068, 2974, 2925, 1744, 1712, 1491, 1364, 1160, 1050, 1013, 915, 854, 805, 727; HRMS (ESI) calcd for  $[\text{C}_{16}\text{H}_{23}\text{NO}_4\text{S} + \text{Na}]^+$ : 348.1240, found 348.1226.

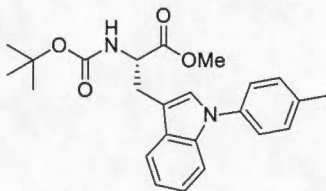
**(S)-Methyl 2-((*tert*-butoxycarbonyl)amino)-3-(1-(*p*-tolyl)-1*H*-imidazol-4-yl)propanoate (11c)**



Method B was followed on a 0.186 mmol scale starting from **10c** and organobismuthine **6a**. The crude product was purified on silica gel (20% EtOAc/hexanes) to afford **11c** as a colorless amorphous paste (54.4 mg, 81%):  $R_f$  0.64 (40% EtOAc/hexanes);  $^1\text{H}$ -NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.72 (s, 1H), 7.23 (d,  $J$  = 8.4 Hz, 2H), 7.20 (d,  $J$  = 8.4 Hz, 2H), 7.01 (s, 1H), 5.91–5.42 (m, 1H), 4.60–4.42 (m, 1H), 3.70 (s, 3H), 3.14 (dd,  $J$  = 14.8, 5.6 Hz, 1H), 3.07 (dd,  $J$  = 14.8, 4.7 Hz, 1H), 2.37 (s, 3H), 1.42 (s, 9H);  $^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  172.6, 155.7, 138.4, 137.5, 135.3, 134.8, 130.4, 121.2, 116.0, 79.7, 53.6, 52.3, 30.3, 28.4, 21.0; IR (neat) 3350, 3129, 2973, 2928, 1741, 1703, 1521, 1483, 1365, 1251, 1159, 1051, 1019, 813; HRMS (ESI) calcd for  $[\text{C}_{19}\text{H}_{25}\text{N}_3\text{O}_4 + \text{H}]^+$ : 360.1918, found 360.1922.

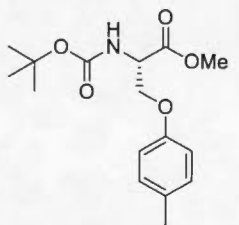


**(S)-Methyl 2-((*tert*-butoxycarbonyl)amino)-3-(1-(*p*-tolyl)-1*H*-indol-3-yl)propanoate (11d)**



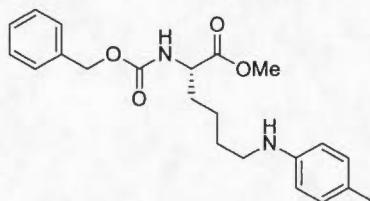
Method B was followed on a 0.157 mmol scale starting from **10d** and organobismuthine **6a**. The crude product was purified on silica gel (20% EtOAc/hexanes) to afford **11d** as a pale yellow solid (38.2 mg, 60%): mp 87-88 °C;  $R_f$  0.74 (40% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.62 (d,  $J$  = 7.2 Hz, 1H), 7.51 (d,  $J$  = 7.6 Hz, 1H), 7.38-7.30 (m, 4H), 7.25-7.17 (m, 2H), 7.16-7.11 (m, 1H), 5.17-4.89 (m, 1H), 4.75-4.54 (m, 1H), 3.72 (s, 3H), 3.37-3.34 (m, 2H), 2.44 (s, 3H), 1.45 (s, 9H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 155.3, 137.1, 136.4, 136.2, 130.2, 126.7, 124.3, 122.6, 120.2, 119.2, 111.1, 110.7, 79.9, 54.3, 52.4, 28.4, 28.0, 21.1; IR (neat) 3334, 3038, 2920, 2848, 1734, 1684, 1513, 1452, 1291, 1246, 1213, 1159, 1015, 821, 726; HRMS (ESI) calcd for  $[\text{C}_{24}\text{H}_{28}\text{N}_2\text{O}_4 + \text{Na}]^+$ : 431.1941, found 431.1945.

**(S)-Methyl 2-((*tert*-butoxycarbonyl)amino)-3-(*p*-tolylloxy)propanoate (11e)**

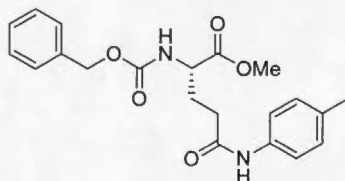


Method B was followed on a 0.228 mmol scale starting from **10e** and organobismuthine **6a**. The crude product was purified on silica gel (20% EtOAc/hexanes) to afford **11e** as a yellow oil (31.1 mg, 45%):  $R_f$  0.41 (20% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.08-7.06 (m, 2H), 6.78-6.76 (m, 2H), 5.51 (d,  $J$  = 8.8 Hz, 1H), 4.64 (td,  $J$  = 8.7, 3.1 Hz, 1H), 4.36 (dd,  $J$  = 9.3, 2.9 Hz, 1H), 4.16 (dd,  $J$  = 9.2, 3.2 Hz, 1H), 3.76 (s, 3H), 2.28 (s, 3H), 1.46 (s, 9H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  170.7, 156.3, 155.5, 130.9, 130.0, 114.7, 80.3, 68.6, 53.8, 52.7, 28.4, 20.6; IR (neat) 3379, 2974, 2925, 1748, 1708, 1503, 1360, 1230, 1156, 1058, 817; HRMS (ESI) calcd for  $[\text{C}_{16}\text{H}_{23}\text{NO}_5 + \text{Na}]^+$ : 332.1468, found 332.1483.

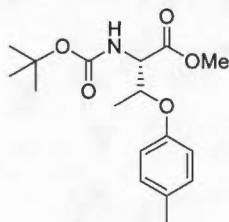


**(S)-Methyl 2-(((benzyloxy)carbonyl)amino)-6-(*p*-tolylamino)hexanoate (11f)**

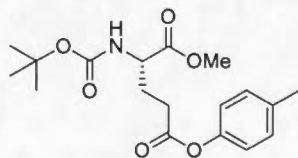
Method B was followed on a 0.306 mmol scale starting from **10f** and organobismuthine **6a**. The crude product was purified on silica gel (30% EtOAc/hexanes) to afford **11f** as an orange oil (40.2 mg, 35%):  $R_f$  0.48 (40% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.38-7.34 (m, 5H), 6.99 (d,  $J = 8.1$  Hz, 2H), 6.52 (d,  $J = 8.1$  Hz, 2H), 5.34 (d,  $J = 8.4$  Hz, 1H), 5.13 (s, 2H), 4.41 (q,  $J = 7.7$  Hz, 1H), 3.74 (s, 3H), 3.08 (t,  $J = 6.8$  Hz, 2H), 2.24 (s, 3H), 1.94-1.85 (m, 1H), 1.73-1.68 (m, 1H), 1.66-1.58 (m, 2H), 1.48-1.40 (m, 2H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  173.0, 156.0, 146.1, 136.3, 129.8, 128.6, 128.3, 128.2, 126.6, 113.0, 67.2, 53.8, 52.5, 44.1, 32.7, 29.2, 22.9, 20.5; IR (neat) 3369, 3030, 2947, 2863, 1711, 1616, 1517, 1448, 1254, 1209, 1042, 908, 806, 749, 692; HRMS (ESI) calcd for  $[\text{C}_{22}\text{H}_{28}\text{N}_2\text{O}_4 + \text{H}]^+$ : 385.2122, found 385.2129.

**(S)-Methyl 2-(((benzyloxy)carbonyl)amino)-5-oxo-5-(*p*-tolylamino)pentanoate (11g)**

Method B was followed on a 0.169 mmol scale starting from **10g** and organobismuthine **6a**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **11g** as a white solid (22.4 mg, 34%): mp 131-132 °C;  $R_f$  0.28 (40% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.14 (s(br), 1H), 7.41 (d,  $J = 8.1$  Hz, 2H), 7.36-7.33 (m, 5H), 7.08 (d,  $J = 8.2$  Hz, 2H), 5.82 (d,  $J = 8.0$  Hz, 1H), 5.08 (s, 2H), 4.40 (dt,  $J = 9.2, 3.9$  Hz, 1H), 3.70 (s, 3H), 2.42-2.40 (m, 2H), 2.30 (s, 3H), 2.28-2.18 (m, 1H), 2.04-1.97 (m, 1H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.4, 170.2, 156.8, 136.1, 135.6, 134.0, 129.6, 128.7, 128.4, 128.3, 120.0, 67.4, 53.5, 52.8, 33.8, 29.5, 21.0; IR (neat) 3334, 3293, 3125, 3034, 2950, 2920, 2848, 1726, 1684, 1658, 1597, 1528, 1433, 1270, 1220, 1159, 1019, 969, 817, 688; HRMS (ESI) calcd for  $[\text{C}_{21}\text{H}_{24}\text{N}_2\text{O}_5 + \text{H}]^+$ : 385.1758, found 385.1776.

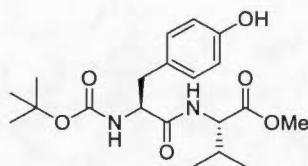
**(2S,3S)-Methyl 2-((*tert*-butoxycarbonyl)amino)-3-(*p*-tolylloxy)butanoate (11h)**

Method B was followed on a 0.214 mmol scale starting from **10h** and organobismuthine **6a**. The crude product was purified on silica gel (20% EtOAc/hexanes) to afford **11h** as a colorless oil (14.4 mg, 21%):  $R_f$  0.32 (20% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.07-7.04 (m, 2H), 6.79-6.76 (m, 2H), 5.40 (d,  $J$  = 9.7 Hz, 1H), 4.89 (dq,  $J$  = 12.5, 2.31 Hz, 1H), 4.47 (dd,  $J$  = 9.7, 2.4 Hz, 1H), 3.67 (s, 3H), 2.27 (s, 3H), 1.49 (s, 9H), 1.32 (d,  $J$  = 6.3 Hz, 3H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  171.2, 156.3, 155.1, 131.2, 130.1, 116.6, 80.2, 74.8, 58.1, 52.6, 28.5, 20.6, 16.5; IR (neat) 3441, 3030, 2977, 2867, 1753, 1715, 1502, 1220, 1159, 1068, 905, 806; HRMS (ESI) calcd for  $[\text{C}_{17}\text{H}_{25}\text{NO}_5 + \text{Na}]^+$ : 346.1625, found 346.1635.

**(S)-1-Methyl 5-*p*-tolyl 2-((*tert*-butoxycarbonyl)amino)pentanedioate (11i)**

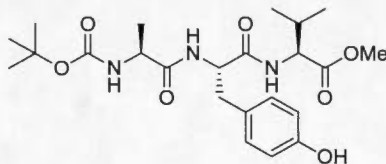
Method B was followed on a 0.191 mmol scale starting from **10i** and organobismuthine **6a**. The crude product was purified on silica gel (20% EtOAc/hexanes) to afford **11i** as a yellow oil (10.7 mg, 16%):  $R_f$  0.26 (20% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.16 (d,  $J$  = 8.2 Hz, 2H), 6.96 (d,  $J$  = 8.4 Hz, 2H), 5.15 (d,  $J$  = 7.9 Hz, 1H), 4.42 (q,  $J$  = 5.3 Hz, 1H), 3.76 (s, 3H), 2.74-2.56 (m, 2H), 2.34 (s, 3H), 2.32-2.26 (m, 1H), 2.13-2.00 (m, 1H), 1.45 (s, 9H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.7, 171.6, 155.5, 148.5, 135.6, 130.0, 121.3, 80.3, 53.0, 52.6, 30.6, 28.4, 27.9, 21.0; IR (neat) 3375, 3032, 2974, 2921, 2848, 1748, 1711.7, 1503, 1446, 1360, 1189, 1164, 1140, 1050, 1013, 805; HRMS (ESI) calcd for  $[\text{C}_{18}\text{H}_{25}\text{NO}_6 + \text{Na}]^+$ : 374.1574, found 374.1579.

**(S)-Methyl 2-((S)-2-((tert-butoxycarbonyl)amino)-3-(4-hydroxyphenyl)propanamido)-3-methylbutanoate (12i)**



Boc-Tyr-OH (3.564 g, 12.67 mmol), H-Val-OMe•HCl (2.124 g, 12.67 mmol) and HCTU (5.767 g, 13.94 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (127 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to 0°C in an ice bath and DiPEA (7.72 mL, 44.34 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was diluted with sat. aq. NaHCO<sub>3</sub> (100 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 X 100 mL). The combined organic phases were washed with sat. aq. NaHCO<sub>3</sub> (2 X 100 mL), sat. aq. NaCl (2 X 100 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was purified on silica gel (50% EtOAc/hexanes) to afford **12i** as a white solid (4.07 g, 81%): mp 72-73 °C; R<sub>f</sub> 0.59 (60% EtOAc/hexanes); <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.01 (d, *J* = 7.9 Hz, 2H), 6.94-6.78 (m, 1H), 6.72 (d, *J* = 8.0 Hz, 2H), 6.47 (s(br), 1H), 5.29-4.85 (m, 1H), 4.59-4.37 (m, 1H), 4.37-4.08 (m, 1H), 3.68 (s, 3H), 2.96 (d, *J* = 6.9 Hz, 2H), 2.12-2.05 (m, 1H), 1.42 (s, 9H), 0.87 (d, *J* = 7.4 Hz, 3H), 0.84 (d, *J* = 7.5 Hz, 3H); <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 171.9, 171.7, 155.7, 155.4, 130.5, 127.9, 115.7, 80.7, 57.5, 56.2, 52.3, 37.4, 31.4, 28.4, 18.9, 17.9; IR (neat) 3306, 3065, 2968, 1734, 1655, 1615, 1515, 1438, 1366, 1224, 1162, 1047, 1018, 828; HRMS (ESI) calcd for [C<sub>20</sub>H<sub>30</sub>N<sub>2</sub>O<sub>6</sub> + H]<sup>+</sup>: 395.2177, found 395.2166.

**(6S,9S,12S)-Methyl 9-(4-hydroxybenzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (12)**



**12i** (2.462 g, 6.24 mmol) was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (37.20 mL) in a round bottom flask with a stir bar. The flask was chased with argon and cooled to 0 °C in an ice bath. TFA (24.80 mL, 323.87 mmol) was added dropwise, the reaction mixture was warmed to room temperature and stirred for 2 h. The reaction mixture was concentrated under reduced pressure, dissolved in CH<sub>2</sub>Cl<sub>2</sub> (100 mL), diluted with sat. aq. NaHCO<sub>3</sub> (100 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 X 100 mL). The combined organic phases were washed with sat. aq.

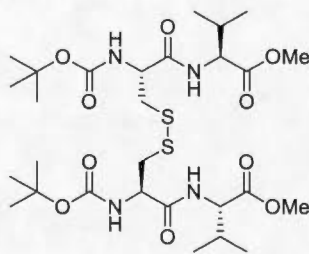


NaHCO<sub>3</sub> (2 X 100 mL), sat. aq. NaCl (2 X 100 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The *N*-deprotected product (1.582 g, 5.37 mmol), Boc-Ala-OH (1.016 g, 5.37 mmol) and HCTU (2.443 g, 5.91 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (54 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to 0°C in an ice bath and DiPEA (3.27 mL, 18.79 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was diluted with sat. aq. NaHCO<sub>3</sub> (100 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 X 100 mL). The combined organic phases were washed with sat. aq. NaHCO<sub>3</sub> (2 X 100 mL), sat. aq. NaCl (2 X 100 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was purified on silica gel (60% EtOAc/hexanes) to afford **12** as a white solid (1.908 g, 76% over 2 steps): mp 95-96 °C; R<sub>f</sub> 0.32 (4% MeOH/CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.02 (d, *J* = 8.4 Hz, 2H), 6.78 (d, *J* = 7.8 Hz, 1H), 6.72 (d, *J* = 8.4 Hz, 2H), 6.48 (d, *J* = 7.5 Hz, 1H), 6.29 (s(br), 1H), 5.03 (d, *J* = 7.1 Hz, 1H), 4.62 (q, *J* = 6.5 Hz, 1H), 4.42 (dd, *J* = 8.6, 5.3 Hz, 1H), 4.14 (quint, *J* = 6.3 Hz, 1H), 3.70 (s, 3H), 2.99-2.97 (m, 2H), 2.14-2.03 (m, 1H), 1.42 (s, 9H), 1.29 (d, *J* = 7.1 Hz, 3H), 0.86 (d, *J* = 7.0 Hz, 3H), 0.83 (d, *J* = 7.0 Hz, 3H); <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 173.2, 171.9, 171.3, 155.7, 155.6, 130.5, 127.3, 115.7, 80.4, 57.6, 54.7, 52.3, 50.3, 37.4, 31.3, 28.4, 18.9, 18.5, 18.0; IR (neat) 3289, 3080, 2971, 1646, 1615, 1515, 1447, 1366, 1226, 1162, 1021, 852, 754; HRMS (ESI) calcd for [C<sub>23</sub>H<sub>35</sub>N<sub>3</sub>O<sub>7</sub> + H]<sup>+</sup>: 466.2548, found 466.2563.

**(2S,2'S)-Dimethyl**

**2,2'-(((2R,2'R)-3,3'-disulfanediylbis(2-((*tert*-**

**butoxycarbonyl)amino)propanoyl))bis(azanediyl))bis(3-methylbutanoate) (13i)**

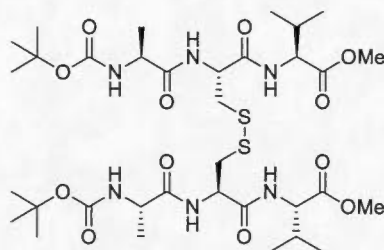


(Boc-Cys-OH)<sub>2</sub> (3.282 g, 7.45 mmol), H-Val-OMe•HCl (2.500 g, 14.90 mmol) and HCTU (6.780 g, 16.39 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (75 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to 0°C in an ice bath and DiPEA (9.08 mL, 52.15 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was diluted with sat. aq. NaHCO<sub>3</sub> (100 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 X 100 mL). The combined organic phases were washed with sat. aq. NaHCO<sub>3</sub> (2 X 100 mL), sat. aq. NaCl (2 X 100 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **13i** as a white solid (4.304 g, 86%): mp 136-137 °C; R<sub>f</sub> 0.36 (40% EtOAc/hexanes);



$^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.37 (d,  $J$  = 8.8 Hz, 2H), 5.56 (d,  $J$  = 8.9 Hz, 2H), 4.70-4.67 (m, 2H), 4.48 (dd,  $J$  = 9.0, 6.3 Hz, 2H), 3.71 (s, 6H), 3.12-2.97 (m, 4H), 2.26-2.12 (m, 2H), 1.45 (s, 18H), 0.95 (d,  $J$  = 7.1 Hz, 6H), 0.94 (d,  $J$  = 6.8 Hz, 6H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.1, 170.5, 155.7, 80.4, 57.9, 54.1, 52.2, 44.9, 31.0, 28.4, 19.3, 18.5; IR (neat) 3322, 3072, 2966, 1740, 1667, 1516, 1315, 1274, 1246, 1209, 1164, 1017, 862, 760, 645; HRMS (ESI) calcd for  $[\text{C}_{28}\text{H}_{50}\text{N}_4\text{O}_{10}\text{S}_2 + \text{H}]^+$ : 667.3041, found 667.3072.

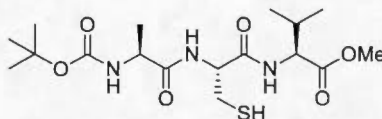
**(6S,6'S,9R,9'R,12S,12'S)-Dimethyl 9,9'-(disulfanediylbis(methylene))bis(12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate) (13ii)**



**13i** (4.275 g, 6.37 mmol) was dissolved in  $\text{CH}_2\text{Cl}_2$  (38.40 mL) in a round bottom flask with a stir bar. The flask was chased with argon and cooled to 0 °C in an ice bath. TFA (25.6 mL, 334.31 mmol) was added dropwise, the reaction mixture was warmed to room temperature and stirred for 2 h. The reaction mixture was concentrated under reduced pressure, dissolved in  $\text{CH}_2\text{Cl}_2$  (100 mL), diluted with sat. aq.  $\text{NaHCO}_3$  (100 mL) and extracted with  $\text{CH}_2\text{Cl}_2$  (2 X 100 mL). The combined organic phases were washed with sat. aq.  $\text{NaHCO}_3$  (2 X 100 mL), sat. aq.  $\text{NaCl}$  (2 X 100 mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered and concentrated under reduced pressure. The *N*-deprotected product (2.735 g, 5.86 mmol), Boc-Ala-OH (1.109 g, 5.86 mmol) and HCTU (2.667 g, 6.45 mmol) were dissolved in  $\text{CH}_2\text{Cl}_2$  (59 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to 0 °C in an ice bath and DiPEA (3.57 mL, 20.51 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was diluted with sat. aq.  $\text{NaHCO}_3$  (100 mL) and extracted with  $\text{CH}_2\text{Cl}_2$  (2 X 100 mL). The combined organic phases were washed with sat. aq.  $\text{NaHCO}_3$  (2 X 100 mL), sat. aq.  $\text{NaCl}$  (2 X 100 mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered and concentrated under reduced pressure. The crude product was purified on silica gel (60% EtOAc/hexanes) to afford **13ii** as a white solid (4.009 g, 78% over 2 steps): mp 107-108 °C;  $R_f$  0.61 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.36 (d,  $J$  = 7.1 Hz, 2H), 7.15 (d,  $J$  = 8.4 Hz, 2H), 5.54-5.48 (m, 2H), 5.38 (d,  $J$  = 6.2 Hz, 2H), 4.47 (dd,  $J$  = 7.7, 5.2 Hz, 2H), 4.35-4.18 (m, 2H), 3.73 (s, 6H), 3.06 (dd,  $J$  = 14.8, 3.0 Hz, 2H), 2.84 (dd,  $J$  = 14.5, 11.0 Hz, 2H), 2.26-2.15 (m, 2H), 1.43 (s, 18H), 1.36 (d,  $J$  = 7.0 Hz, 6H), 0.98 (d,  $J$  = 6.8 Hz, 12H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  173.6, 172.3, 170.3, 155.5, 80.2, 57.8, 52.8, 52.2, 50.7, 46.3, 31.1, 28.5, 19.2, 18.8, 18.1; IR (neat) 3289, 3077, 2970, 2933, 1740, 1712, 1646, 1507,

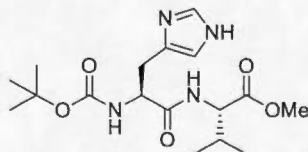
1360, 1246, 1156, 1054, 1025, 858, 731; HRMS (ESI) calcd for  $[C_{34}H_{60}N_6O_{12}S_2 + H]^+$ : 809.3783, found 809.3750.

**(6S,9R,12S)-Methyl 12-isopropyl-9-(mercaptomethyl)-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (13)**



**13ii** (3.712 g, 4.59 mmol) was dissolved in n-propanol (72.80 mL) and water (36.40 mL) in a round bottom flask with a stir bar. The pH was adjusted to 8 using  $NH_4OH$  (30 M) and the flask was chased with argon. Tri-n-butylphosphine (1.74 mL, 6.88 mmol) was added dropwise and the reaction mixture was stirred for 1 h at room temperature. The pH was adjusted to 6 using HCl (12 M), the reaction mixture was diluted with sat. aq. NaCl (100 mL) and extracted with  $CHCl_3$  (3 X 100 mL). The combined organic phases were dried over  $Na_2SO_4$ , filtered and concentrated under reduced pressure. The crude product was purified on silica gel (50% EtOAc/hexanes) to afford **13** as a white solid (2.82 g, 76%): mp 90-91 °C;  $R_f$  0.39 (60% EtOAc/hexanes);  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.18 (d,  $J$  = 7.1 Hz, 1H), 7.01 (d,  $J$  = 8.3 Hz, 1H), 5.10 (d,  $J$  = 6.3 Hz, 1H), 4.68 (q,  $J$  = 6.3 Hz, 1H), 4.48 (dd,  $J$  = 8.4, 5.1 Hz, 1H), 4.20 (quint,  $J$  = 6.5 Hz, 1H), 3.73 (s, 3H), 3.11-3.02 (m, 1H), 2.79-2.69 (m, 1H), 2.24-2.13 (m, 1H), 1.76 (t,  $J$  = 8.6 Hz, 1H), 1.43 (s, 9H), 1.37 (d,  $J$  = 7.1 Hz, 3H), 0.93 (d,  $J$  = 6.6 Hz, 3H), 0.91 (d,  $J$  = 6.7 Hz, 3H);  $^{13}C$ -NMR (75 MHz,  $CDCl_3$ )  $\delta$  173.0, 172.0, 169.7, 155.7, 80.6, 57.7, 54.4, 52.3, 50.5, 31.0, 28.4, 26.6, 19.2, 18.2, 17.9; IR (neat) 3297, 3072, 2966, 2929, 2876, 2553, 1744, 1642, 1511, 1454, 1360, 1454, 1360, 1246, 1205, 1160, 1017, 854, 751; HRMS (ESI) calcd for  $[C_{17}H_{31}N_3O_6S + H]^+$ : 406.2006, found 406.2020.

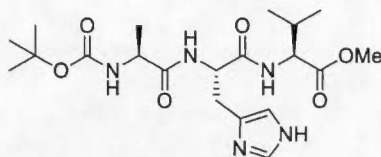
**(S)-Methyl 2-((S)-2-((tert-butoxycarbonyl)amino)-3-(1H-imidazol-4-yl)propanamido)-3-methylbutanoate (14i)**



Boc-His-OH (2.905 g, 11.38 mmol), H-Val-OMe•HCl (1.908 g, 11.38 mmol) and HCTU (5.179 g, 12.52 mmol) were dissolved in  $CH_2Cl_2$  (110 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to 0°C in an ice bath and DiPEA (6.94 mL, 39.83 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was

diluted with sat. aq. NaHCO<sub>3</sub> (100 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 X 100 mL). The combined organic phases were washed with sat. aq. NaHCO<sub>3</sub> (2 X 100 mL), sat. aq. NaCl (2 X 100 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was purified on silica gel (2.5% MeOH/CH<sub>2</sub>Cl<sub>2</sub>) to afford **14i** as a colorless oil (2.3663 g, 56%): *R*<sub>f</sub> 0.44 (10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.60-7.30 (m, 1H), 7.52 (s, 1H), 6.80 (s, 1H), 6.25-5.80 (m, 1H), 4.42-4.38 (m, 2H), 3.69 (s, 3H), 3.11 (dd, *J* = 14.9, 4.2 Hz, 1H), 2.98 (dd, *J* = 14.8, 6.2 Hz, 1H), 2.14-2.04 (m, 1H), 1.42 (s, 9H), 0.81 (d, *J* = 6.9 Hz, 3H), 0.80 (d, *J* = 6.7 Hz, 3H); <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 172.3, 172.2, 156.0, 135.1, 132.2, 119.1, 80.4, 57.7, 54.2, 52.3, 31.0, 29.0, 28.4, 19.0, 17.8; IR (neat) 3292, 3085, 2962, 2920, 2848, 1741, 1661, 1517, 1361, 1251, 1163, 1019, 931, 855, 821, 753, 658; HRMS (ESI) calcd for [C<sub>17</sub>H<sub>28</sub>N<sub>4</sub>O<sub>5</sub> + H]<sup>+</sup>: 369.2132, found 369.2130.

**(6*S*,9*S*,12*S*)-Methyl 9-((1*H*-imidazol-4-yl)methyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (**14**)**

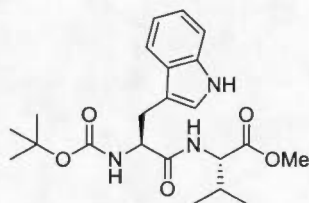


**14i** (252 mg, 0.68 mmol) was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (4.00 mL) in a round bottom flask with a stir bar. The flask was chased with argon and cooled to 0 °C in an ice bath. TFA (2.72 mL, 32.91 mmol) was added dropwise, the reaction mixture was warmed to room temperature and stirred for 2 h. The reaction mixture was concentrated under reduced pressure, dissolved in CH<sub>2</sub>Cl<sub>2</sub> (50 mL), diluted with sat. aq. NaHCO<sub>3</sub> (50 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 X 50 mL). The combined organic phases were washed with sat. aq. NaHCO<sub>3</sub> (2 X 50 mL), sat. aq. NaCl (2 X 50 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The *N*-deprotected product (164 mg, 0.61 mmol), Boc-Ala-OH (115 mg, 0.61 mmol) and HCTU (278 mg, 0.67 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (6.1 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to 0 °C in an ice bath and DiPEA (796 μL, 2.14 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was diluted with sat. aq. NaHCO<sub>3</sub> (50 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 X 50 mL). The combined organic phases were washed with sat. aq. NaHCO<sub>3</sub> (2 X 50 mL), sat. aq. NaCl (2 X 50 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was purified on silica gel (7.5% MeOH/CH<sub>2</sub>Cl<sub>2</sub>) to afford **14** as a white solid (174 mg, 58% over 2 steps): mp 81-82 °C; *R*<sub>f</sub> 0.32 (10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 7.88 (s(br), 1H), 7.78-7.58 (m, 1H), 7.51 (s, 1H), 7.15-6.05



(s(br), 1H), 6.78 (s, 1H), 5.60-5.25 (m, 1H), 4.73 (q,  $J = 5.7$  Hz, 1H), 4.38 (dd,  $J = 8.2, 5.2$  Hz, 1H), 4.18 (quint,  $J = 6.8$  Hz, 1H) 3.70 (s, 3H), 3.11 (dd,  $J = 15.1, 5.1$  Hz, 1H), 3.02 (dd,  $J = 14.9, 5.9$  Hz, 1H), 2.16-2.05 (m, 1H), 1.41 (s, 9H), 1.36 (d,  $J = 7.0$  Hz, 3H), 0.83 (d,  $J = 6.8$  Hz, 3H), 0.81 (d,  $J = 7.0$  Hz, 3H);  $^{13}\text{C}$ -NMR (150 MHz,  $\text{CDCl}_3$ )  $\delta$  173.5, 172.3, 171.3, 155.8, 134.9, 131.9, 119.0, 80.1, 57.9, 53.2, 52.2, 50.6, 30.8, 28.8, 28.4, 19.0, 18.5, 17.9; IR (neat) 3285, 3077, 2973, 2931, 1737, 1650, 1517, 1441, 1365, 1251, 1159, 1015, 840, 753; HRMS (ESI) calcd for  $[\text{C}_{20}\text{H}_{33}\text{N}_5\text{O}_6 + \text{H}]^+$ : 440.2504, found 440.2494.

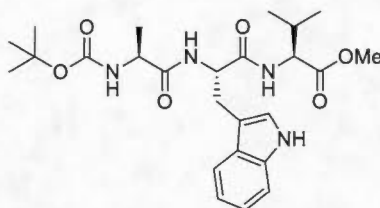
**(S)-Methyl 2-((S)-2-((tert-butoxycarbonyl)amino)-3-(1H-indol-3-yl)propanamido)-3-methylbutanoate (15i)**



Boc-Trp-OH (750 mg, 2.46 mmol), H-Val-OMe•HCl (413 mg, 2.46 mmol) and HCTU (1.121 g, 2.71 mmol) were dissolved in  $\text{CH}_2\text{Cl}_2$  (25 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to  $0^\circ\text{C}$  in an ice bath and DiPEA (1.50 mL, 8.62 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was diluted with sat. aq.  $\text{NaHCO}_3$  (50 mL) and extracted with  $\text{CH}_2\text{Cl}_2$  (2 X 50 mL). The combined organic phases were washed with sat. aq.  $\text{NaHCO}_3$  (2 X 50 mL), sat. aq.  $\text{NaCl}$  (2 X 50 mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered and concentrated under reduced pressure. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **15i** as a white solid (983.7 mg, 96%): mp  $68-69^\circ\text{C}$ ;  $R_f$  0.24 (40% EtOAc/hexanes);  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.20 (s(br), 1H), 7.64 (d,  $J = 7.9$  Hz, 1H), 7.35 (d,  $J = 8.0$  Hz, 1H), 7.19 (dt,  $J = 7.1, 1.0$  Hz, 1H), 7.11 (dt,  $J = 7.1, 0.9$  Hz, 1H), 7.06 (d,  $J = 1.6$  Hz, 1H), 6.35 (d,  $J = 8.3$  Hz, 1H), 5.09 (s(br), 1H), 4.62-4.47 (m, 1H), 4.44 (dd,  $J = 8.6, 4.8$  Hz, 1H), 3.64 (s, 3H), 3.32 (dd,  $J = 14.7, 5.9$  Hz, 1H), 3.20 (dd,  $J = 14.6, 7.0$  Hz, 1H), 2.04-1.93 (m, 1H), 1.41 (s, 9H), 0.76 (d,  $J = 6.8$  Hz, 3H), 0.69 (d,  $J = 6.9$  Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.1, 172.0, 155.6, 136.4, 127.5, 123.3, 122.2, 119.7, 118.8, 111.4, 110.2, 80.3, 57.2, 55.3, 52.1, 31.1, 28.3, 28.2, 18.7, 17.6; IR (neat) 3326, 3056, 2966, 2938, 1659, 1507, 1364, 1246, 1152, 1005, 854, 739, 539; HRMS (ESI) calcd for  $[\text{C}_{22}\text{H}_{31}\text{N}_3\text{O}_5 + \text{Na}]^+$ : 440.2156, found 440.2175.

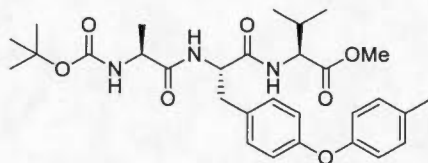


**(6S,9S,12S)-Methyl 9-((1*H*-indol-3-yl)methyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (15)**



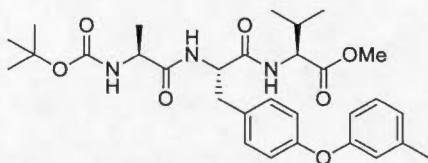
**15i** (765 mg, 1.84 mmol) was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (10.80 mL) in a round bottom flask with a stir bar. The flask was chased with argon and cooled to 0 °C in an ice bath. TFA (7.20 mL, 94.03 mmol) was added dropwise, the reaction mixture was warmed to room temperature and stirred for 2 h. The reaction mixture was concentrated under reduced pressure, dissolved in CH<sub>2</sub>Cl<sub>2</sub> (100 mL), diluted with sat. aq. NaHCO<sub>3</sub> (100 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 X 100 mL). The combined organic phases were washed with sat. aq. NaHCO<sub>3</sub> (2 X 100 mL), sat. aq. NaCl (2 X 100 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The *N*-deprotected product (528 mg, 1.67 mmol), Boc-Ala-OH (316 mg, 1.67 mmol) and HCTU (759 mg, 1.83 mmol) were dissolved in CH<sub>2</sub>Cl<sub>2</sub> (17.00 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to 0°C in an ice bath and DiPEA (1.02 mL, 5.83 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was diluted with sat. aq. NaHCO<sub>3</sub> (50 mL) and extracted with CH<sub>2</sub>Cl<sub>2</sub> (2 X 50 mL). The combined organic phases were washed with sat. aq. NaHCO<sub>3</sub> (2 X 50 mL), sat. aq. NaCl (2 X 50 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered and concentrated under reduced pressure. The crude product was purified on silica gel (60% EtOAc/hexanes) to afford **15** as a white solid (550.4 mg, 61% over 2 steps): mp 175-176 °C; *R<sub>f</sub>* 0.24 (60% EtOAc/hexanes); <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 8.22 (s(br), 1H), 7.69 (d, *J* = 7.7 Hz, 1H), 7.35 (d, *J* = 7.8 Hz, 1H), 7.19 (dt, *J* = 7.1, 1.2 Hz, 1H) 7.13 (dd, *J* = 7.7, 1.2 Hz, 1H), 7.11-7.10 (m, 1H), 6.84 (d, *J* = 7.6 Hz, 1H), 6.32 (d, *J* = 8.3 Hz, 1H), 4.92 (d, *J* = 7.0 Hz, 1H), 4.77-4.70 (m, 1H), 4.36 (dd, *J* = 8.4, 5.1 Hz, 1H), 4.12 (quint, *J* = 7.1 Hz, 1H), 3.64 (s, 3H), 3.34 (dd, *J* = 14.6, 5.3 Hz, 1H), 3.16 (dd, *J* = 14.5, 7.8 Hz, 1H), 2.07-1.96 (m, 1H), 1.38 (s, 9H), 1.31 (d, *J* = 7.1 Hz, 3H), 0.78 (d, *J* = 6.9 Hz, 3H), 0.75 (d, *J* = 6.9 Hz, 3H); <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 172.9, 171.9, 171.3, 155.5, 136.3, 127.6, 123.7, 122.2, 119.7, 118.8, 111.4, 110.3, 80.3, 57.7, 54.1, 52.2, 50.6, 31.1, 28.3, 28.0, 18.9, 18.5, 17.9; IR (neat) 3391, 3346, 3258, 3064, 2969, 2943, 1745, 1696, 1669, 1639, 1521, 1486, 1361, 1251, 1159, 1091, 1019, 848, 741, 673; HRMS (ESI) calcd for [C<sub>25</sub>H<sub>36</sub>N<sub>4</sub>O<sub>6</sub> + H]<sup>+</sup>: 489.2708, found 489.2727.

**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-(4-(*p*-tolylloxy)benzyl)-3-oxa-5,8,11-triazatridecan-13-oate (16a)**



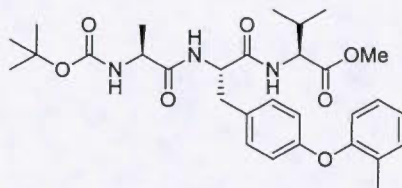
Method B was followed on a 0.107 mmol scale starting from **12** and organobismuthine **6a**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16a** as a white solid (53.3 mg, 90%): mp 77-78 °C;  $R_f$  0.51 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.14-7.09 (m, 4H), 6.89-6.85 (m, 5H), 6.59 (d,  $J$  = 8.4 Hz, 1H), 5.10 (d,  $J$  = 7.2 Hz, 1H), 4.69 (q,  $J$  = 7.1 Hz, 1H), 4.42 (dd,  $J$  = 8.6, 5.2 Hz, 1H), 4.17 (quint,  $J$  = 7.4 Hz, 1H), 3.70 (s, 3H) 3.11-2.91 (m, 2H), 2.31 (s, 3H), 2.14-2.03 (m, 1H), 1.41 (s, 9H), 1.30 (d,  $J$  = 7.1 Hz, 3H), 0.85 (d,  $J$  = 6.8 Hz, 3H), 0.83 (d,  $J$  = 6.8 Hz, 3H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 171.8, 170.7, 156.9, 155.5, 154.7, 133.0, 130.7, 130.6, 130.3, 119.1, 118.4, 80.3, 57.4, 54.6, 52.2, 50.4, 37.5, 31.3, 28.4, 20.8, 19.0, 18.5, 17.9; IR (neat) 3282, 3068, 2968, 2929, 1742, 1644, 1499, 1445, 1391, 1366, 1162, 1011, 870, 813, 753; HRMS (ESI) calcd for  $[\text{C}_{30}\text{H}_{41}\text{N}_3\text{O}_7 + \text{H}]^+$ : 556.3017, found 556.3013.

**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-(4-(*m*-tolylloxy)benzyl)-3-oxa-5,8,11-triazatridecan-13-oate (16b)**



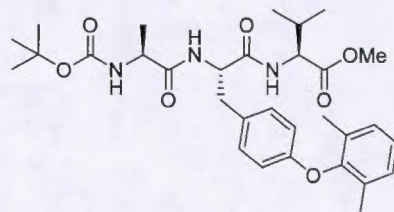
Method B was followed on a 0.0644 mmol scale starting from **12** and organobismuthine **6b**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16b** as a white solid (35.8 mg, 99%): mp 72-73 °C;  $R_f$  0.51 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.21 (d,  $J$  = 7.7 Hz, 1H), 7.17-7.15 (d,  $J$  = 8.0 Hz, 2H), 6.90 (d,  $J$  = 7.8 Hz, 3H), 6.81-6.75 (m, 3H), 6.40 (d,  $J$  = 8.5 Hz, 1H), 4.95 (d,  $J$  = 7.1 Hz, 1H), 4.65 (q,  $J$  = 7.2 Hz, 1H), 4.43 (dd,  $J$  = 8.6, 5.2 Hz, 1H), 4.15 (quint,  $J$  = 7.1 Hz, 1H), 3.70 (s, 3H), 3.07-3.03 (m, 2H), 2.32 (s, 3H), 2.13-2.04 (m, 1H), 1.42 (s, 9H), 1.32 (d,  $J$  = 7.1 Hz, 3H), 0.86 (d,  $J$  = 7.5 Hz, 3H), 0.83 (d,  $J$  = 6.4 Hz, 3H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.7, 171.8, 170.6, 157.2, 156.5, 155.5, 140.0, 131.0, 130.7, 129.6, 124.2, 119.7, 119.0, 116.0, 80.5, 57.5, 54.6, 52.3, 50.5, 37.5, 31.3, 28.4, 21.5, 19.0, 18.4, 17.9; IR (neat) 3281, 3072, 2965, 2924, 2852, 1741, 1642, 1505, 1365, 1258, 1216, 1163, 1015, 931, 855, 779, 684; HRMS (ESI) calcd for  $[\text{C}_{30}\text{H}_{41}\text{N}_3\text{O}_7 + \text{H}]^+$ : 556.3017, found 556.3012.

**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-(4-(*o*-tolylloxy)benzyl)-3-oxa-5,8,11-triazatridecan-13-oate (16c)**



Method A was followed on a 0.107 mmol scale starting from **12** and organobismuthine **6c**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16c** as a brown-yellow solid (31.7 mg, 53%): mp 81-82 °C;  $R_f$  0.43 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.22 (d,  $J = 7.3$  Hz, 1H), 7.16-7.10 (m, 3H), 7.07-7.01 (m, 1H), 6.88-6.79 (m, 4H), 6.55-6.51 (m, 1H), 5.04 (d,  $J = 7.1$  Hz, 1H), 4.67 (q,  $J = 7.1$  Hz, 1H), 4.43 (dd,  $J = 8.6, 5.1$  Hz, 1H), 4.16 (quint,  $J = 7.4$  Hz, 1H), 3.69 (s, 3H), 3.04-3.01 (m, 2H), 2.21 (s, 3H), 2.14-2.03 (m, 1H), 1.41 (s, 9H), 1.30 (d,  $J = 7.0$  Hz, 3H), 0.85 (d,  $J = 6.8$  Hz, 3H), 0.83 (d,  $J = 6.9$  Hz, 3H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 171.8, 170.7, 157.0, 155.5, 154.6, 131.5, 130.7, 130.3, 129.9, 127.2, 124.0, 119.6, 117.6, 80.3, 57.5, 54.6, 53.5, 52.2, 50.4, 37.4, 31.2, 28.4, 19.0, 18.5, 17.9, 16.2; IR (neat) 3280, 3072, 2966, 2933, 1642, 1511, 1356, 1238, 1160, 1021, 878, 747; HRMS (ESI) calcd for  $[\text{C}_{30}\text{H}_{41}\text{N}_3\text{O}_7 + \text{H}]^+$ : 556.3017, found 556.3014.

**(6S,9S,12S)-Methyl 9-(4-(2,6-dimethylphenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16d)**

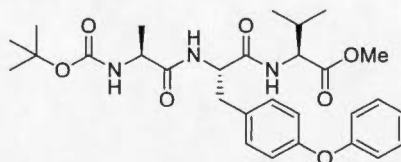


Method B was followed on a 0.0642 mmol scale starting from **12** and organobismuthine **6d**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16d** as a yellow oil (18.2 mg, 50%):  $R_f$  0.49 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.11-7.08 (m, 2H), 7.07-7.04 (m, 2H), 6.95-6.93 (m, 1H), 6.72 (d,  $J = 7.6$  Hz, 1H), 6.69-6.65 (m, 2H), 6.37 (d,  $J = 6.2$  Hz, 1H), 4.89 (d,  $J = 5.1$  Hz, 1H), 4.61 (q,  $J = 7.0$  Hz, 1H), 4.41 (dd,  $J = 8.5, 5.0$  Hz, 1H), 4.13 (quint,  $J = 6.1$  Hz, 1H), 3.70 (s, 3H), 3.01 (d,  $J = 6.8$  Hz, 2H), 2.10 (s, 6H), 2.04-1.98 (m, 1H), 1.41 (s, 9H), 1.29 (d,  $J = 7.1$  Hz, 3H), 0.85 (d,  $J = 6.8$  Hz, 3H), 0.81 (d,  $J = 6.9$  Hz, 3H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.7, 171.8, 170.7, 157.1, 155.6, 151.2, 131.5, 130.7, 129.2, 129.1, 125.2, 114.9, 80.5, 57.5, 54.6, 52.2, 50.5, 37.2, 31.2, 28.4, 19.0, 18.4, 17.9, 16.5; IR (neat) 3283,



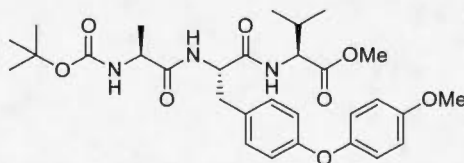
3074, 2969, 2930, 2876, 2855, 1747, 1699, 1650, 1547, 1506, 1472, 1393, 1368, 1223, 1186, 1170, 1026, 872, 771; HRMS (ESI) calcd for  $[C_{31}H_{43}N_3O_7 + H]^+$ : 570.3174, found 570.3153.

**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-(4-phenoxybenzyl)-3-oxa-5,8,11-triazatridecan-13-oate (16e)**



Method B was followed on a 0.107 mmol scale starting from **12** and organobismuthine **6e**. The crude product was purified on silica gel (50% EtOAc/hexanes) to afford **16e** as a light brown solid (48.6 mg, 83%): mp 88-89 °C;  $R_f$  0.46 (60% EtOAc/hexanes);  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.32-7.27 (m, 2H), 7.16-7.13 (m, 2H), 7.10-7.04 (m, 1H), 6.99-6.94 (m, 3H), 6.92-6.87 (m, 2H), 6.68 (d,  $J$  = 5.5 Hz, 1H), 5.17 (d,  $J$  = 7.3 Hz, 1H), 4.74 (q,  $J$  = 7.2 Hz, 1H), 4.44 (dd,  $J$  = 8.4, 5.2 Hz, 1H), 4.19 (quint,  $J$  = 7.2 Hz, 1H), 3.69 (s, 3H), 3.02 (d,  $J$  = 6.9 Hz, 2H), 2.12-2.03 (m, 1H), 1.40 (s, 9H), 1.30 (d,  $J$  = 7.1 Hz, 3H), 0.86 (d,  $J$  = 6.8 Hz, 3H), 0.83 (d,  $J$  = 6.8 Hz, 3H);  $^{13}C$ -NMR (75 MHz,  $CDCl_3$ )  $\delta$  172.9, 171.8, 170.8, 157.2, 156.3, 155.5, 131.2, 130.8, 129.8, 123.3, 119.0, 118.9, 80.2, 57.4, 54.5, 52.2, 50.3, 37.6, 31.3, 28.4, 19.0, 18.6, 17.9; IR (neat) 3281, 3072, 2966, 2925, 1638, 1486, 1230, 1160, 1017, 866, 751; HRMS (ESI) calcd for  $[C_{29}H_{39}N_3O_7 + H]^+$ : 542.2861, found 542.2866.

**(6S,9S,12S)-Methyl 12-isopropyl-9-(4-(4-methoxyphenoxy)benzyl)-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16f)**

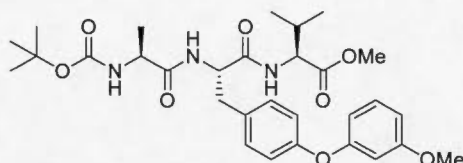


Method B was followed on a 0.0651 mmol scale starting from **12** and organobismuthine **6f**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16f** as a white solid (34.5 mg, 92%): mp 63-64 °C;  $R_f$  0.51 (60% EtOAc/hexanes);  $^1H$ -NMR (600 MHz,  $CDCl_3$ )  $\delta$  7.11 (d,  $J$  = 8.4 Hz, 2H), 6.95-6.94 (m, 2H), 6.87-6.84 (m, 2H), 6.84-6.83 (m, 2H), 6.82-6.80 (m, 1H), 6.53-6.38 (m, 1H), 5.05-4.90 (m, 1H), 4.65 (q,  $J$  = 7.4 Hz, 1H), 4.42 (dd,  $J$  = 8.6, 5.2 Hz, 1H), 4.20-4.10 (m, 1H), 3.79 (s, 3H), 3.69 (s, 3H), 3.04 (dd,  $J$  = 13.9, 6.1 Hz, 1H), 3.00 (dd,  $J$  = 13.8, 7.3 Hz, 1H), 2.11-2.06 (m, 1H), 1.41 (s, 9H), 1.31 (d,  $J$  = 7.0 Hz, 3H), 0.85 (d,  $J$  = 6.8 Hz, 3H), 0.83 (d,  $J$  = 6.9 Hz, 3H);  $^{13}C$ -NMR (75 MHz,  $CDCl_3$ )  $\delta$  172.8, 171.8, 170.7, 157.7, 156.0, 155.5,



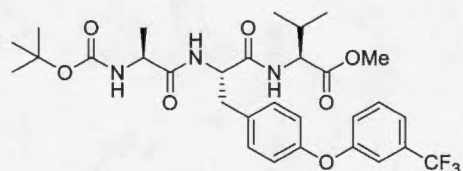
150.2, 130.6, 130.4, 120.9, 117.8, 115.0, 80.4, 57.5, 55.8, 54.7, 52.3, 50.4, 37.3, 31.3, 28.4, 19.0, 18.4, 17.9; IR (neat) 3277, 3068, 2962, 2925, 2872, 2852, 1740, 1691, 1646, 1495, 1368, 1225, 1164, 1029, 825; HRMS (ESI) calcd for  $[C_{30}H_{41}N_3O_8 + H]^+$ : 572.2966, found 572.2998.

**(6S,9S,12S)-Methyl 12-isopropyl-9-(4-(3-methoxyphenoxy)benzyl)-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16g)**



Method B was followed on a 0.108 mmol scale starting from **12** and organobismuthine **6g**. The crude product was purified on silica gel (3% MeOH/ $CH_2Cl_2$ ) to afford **16g** as an orange-brown solid (51.4 mg, 83%): mp 69-70 °C;  $R_f$  0.32 (5% MeOH/ $CH_2Cl_2$ );  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.22-7.14 (m, 3H), 6.93-6.90 (m, 2H), 6.85 (d,  $J$  = 7.9 Hz, 1H), 6.63 (ddd,  $J$  = 8.3, 2.3, 1.0 Hz, 1H), 6.56-6.53 (m, 2H), 6.53-6.47 (m, 1H), 5.05 (d,  $J$  = 7.1 Hz, 1H), 4.68 (q,  $J$  = 7.2 Hz, 1H), 4.43 (dd,  $J$  = 8.6, 5.2 Hz, 1H), 4.16 (quint,  $J$  = 7.6 Hz, 1H), 3.76 (s, 3H), 3.69 (s, 3H), 3.04 (d,  $J$  = 6.9 Hz, 2H), 2.14-2.03 (m, 1H), 1.41 (s, 9H), 1.31 (d,  $J$  = 7.1 Hz, 3H), 0.86 (d,  $J$  = 6.9 Hz, 3H), 0.83 (d,  $J$  = 6.8 Hz, 3H);  $^{13}C$ -NMR (75 MHz,  $CDCl_3$ )  $\delta$  172.8, 171.8, 170.6, 161.1, 158.5, 156.1, 155.5, 131.4, 130.8, 130.2, 119.2, 111.0, 108.9, 105.0, 80.4, 57.5, 55.5, 54.6, 52.3, 50.4, 37.5, 31.3, 28.4, 19.0, 18.5, 17.9; IR (neat) 3285, 3072, 2966, 2921, 2848, 1642, 1483, 1213, 1164, 1131, 1042, 952, 854, 768, 682; HRMS (ESI) calcd for  $[C_{30}H_{41}N_3O_8 + H]^+$ : 572.2966, found 572.2973.

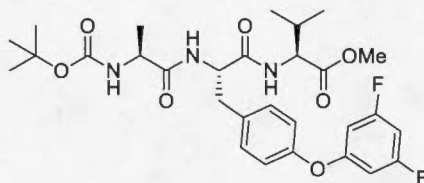
**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-(4-(3-(trifluoromethyl)phenoxy)benzyl)-3-oxa-5,8,11-triazatridecan-13-oate (16h)**



Method B was followed on a 0.107 mmol scale starting from **12** and organobismuthine **6h**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16h** as a pale yellow solid (44.4 mg, 68%): mp 78-79 °C;  $R_f$  0.44 (60% EtOAc/hexanes);  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.40 (t,  $J$  = 7.9 Hz, 1H), 7.31 (d,  $J$  = 7.8 Hz, 1H), 7.20-7.18 (m, 3H), 7.13-7.10 (m, 1H), 6.98 (d,  $J$  = 8.1 Hz, 1H), 6.94-6.89 (m, 2H), 6.76-6.68 (m, 1H), 5.16 (d,  $J$  = 7.4 Hz, 1H), 4.75 (q,  $J$  = 7.1 Hz, 1H), 4.45 (dd,  $J$  = 8.7, 5.2 Hz, 1H), 4.20 (quint,  $J$  = 6.3 Hz, 1H), 3.68 (s, 3H), 3.04 (d,  $J$  = 6.9 Hz, 2H), 2.14-2.01 (m, 1H), 1.40 (s, 9H), 1.30 (d,  $J$  = 7.0 Hz, 3H), 0.85 (d,  $J$

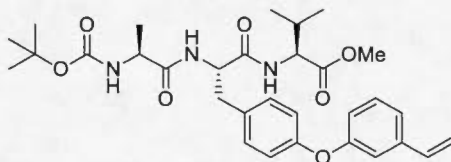
= 6.9 Hz, 3H), 0.83 (d,  $J$  = 6.9 Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  173.0, 171.9, 170.8, 157.9, 155.5, 155.2, 132.5, 132.4, 132.1, 131.1, 130.4, 125.6, 122.0, 121.6, 119.8, 119.73, 119.67, 119.6, 119.5, 115.44, 115.40, 115.34, 115.30, 113.4, 80.2, 57.4, 54.5, 52.2, 50.4, 37.7, 31.3, 28.4, 19.0, 18.5, 17.9; IR (neat) 3285, 3072, 2970, 2933, 1646, 1503, 1442, 1324, 1230, 1156, 1123, 1062, 915, 694; HRMS (ESI) calcd for  $[\text{C}_{30}\text{H}_{38}\text{F}_3\text{N}_3\text{O}_7 + \text{H}]^+$ : 610.2735, found 610.2732.

**(6S,9S,12S)-Methyl 9-(4-(3,5-difluorophenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16i)**



Method A was followed on a 0.085 mmol scale starting from **12** and organobismuthine **6i**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16i** as a white solid (22.2 mg, 45%): mp 141-142 °C;  $R_f$  0.34 (40% EtOAc/hexanes);  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.22 (d,  $J$  = 8.5 Hz, 2H), 6.97-6.94 (m, 2H), 6.86 (d,  $J$  = 7.9 Hz, 1H), 6.53-6.52 (m, 1H), 6.51-6.44 (m, 3H), 5.03 (d,  $J$  = 7.2 Hz, 1H), 4.70 (q,  $J$  = 7.2 Hz, 1H), 4.44 (dd,  $J$  = 8.6, 5.0 Hz, 1H), 4.16 (quint,  $J$  = 6.9 Hz, 1H), 3.69 (s, 3H), 3.06 (d,  $J$  = 6.9 Hz, 2H), 2.15-2.04 (m, 1H), 1.41 (s, 9H), 1.31 (d,  $J$  = 7.0 Hz, 3H), 0.86 (d,  $J$  = 6.8 Hz, 3H), 0.83 (d,  $J$  = 6.9 Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 171.8, 170.5, 165.5, 165.3, 162.2, 162.0, 160.1, 159.8, 159.8, 155.5, 154.5, 133.0, 131.1, 120.3, 101.7, 101.3, 98.7, 98.4, 98.1, 80.4, 57.4, 54.6, 52.3, 50.5, 37.6, 31.3, 28.4, 19.0, 18.4, 17.9; IR (neat) 3323, 3283, 3091, 2966, 2931, 2971, 1741, 1692, 1642, 1601, 1517, 1460, 1388, 1216, 1167, 1114, 988, 847; 707, 665; HRMS (ESI) calcd for  $[\text{C}_{29}\text{H}_{37}\text{F}_2\text{N}_3\text{O}_7 + \text{H}]^+$ : 578.2672, found 578.2726.

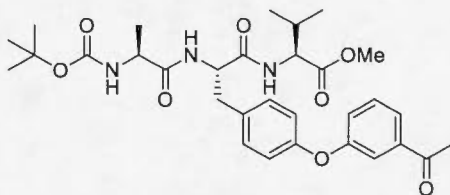
**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-(4-(3-vinylphenoxy)benzyl)-3-oxa-5,8,11-triazatridecan-13-oate (16j)**



Method A was followed on a 0.0775 mmol scale starting from **12** and organobismuthine **6j**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16j** as a white solid (38.3 mg, 87%): mp 80-81 °C;  $R_f$  0.53 (60% EtOAc/hexanes);  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.25 (t,  $J$  = 7.8 Hz, 1H), 7.17-7.11 (m,

3H), 7.04 (t,  $J = 2.0$  Hz, 1H), 6.93-6.84 (m, 4H), 6.65 (dd,  $J = 17.6, 10.9$  Hz, 1H), 6.64-6.54 (m, 1H), 5.70 (d,  $J = 17.5$  Hz, 1H), 5.23 (d,  $J = 10.9$  Hz, 1H), 5.11 (d,  $J = 7.2$  Hz, 1H), 4.71 (q,  $J = 7.2$  Hz, 1H), 4.44 (dd,  $J = 8.6, 5.2$  Hz, 1H), 4.18 (quint,  $J = 7.2$  Hz, 1H), 3.69 (s, 3H), 3.03 (d,  $J = 6.9$  Hz, 2H), 2.14-2.03 (m, 1H), 1.41 (s, 9H), 1.31 (d,  $J = 7.1$  Hz, 3H), 0.86 (d,  $J = 6.8$  Hz, 3H), 0.83 (d,  $J = 6.9$  Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 171.8, 170.7, 157.5, 156.3, 155.5, 139.6, 136.4, 131.3, 130.8, 129.9, 121.4, 119.0, 118.3, 116.6, 114.8, 80.3, 57.4, 54.6, 52.2, 50.4, 37.6, 31.3, 28.5, 19.0, 18.5, 17.9; IR (neat) 3282, 3072, 2969, 2931, 1741, 1688, 1646, 1505, 1437, 1361, 1243, 1163, 1015, 931, 855, 786.8; HRMS (ESI) calcd for  $[\text{C}_{31}\text{H}_{41}\text{N}_3\text{O}_7 + \text{H}]^+$ : 568.3017, found 568.3030.

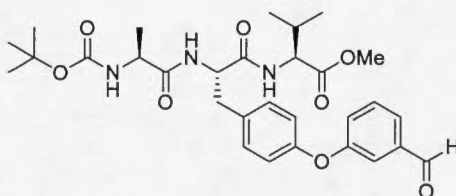
**(6S,9S,12S)-Methyl 9-(4-(3-acetylphenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16k)**



Method A was followed on a 0.0704 mmol scale starting from **12** and organobismuthine **6k**. The crude product was purified on silica gel (50% EtOAc/hexanes) to afford **16k** as an off-white solid (37.9 mg, 92%): mp 60-61 °C;  $R_f$  0.33 (60% EtOAc/hexanes);  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.65 (d,  $J = 7.7$  Hz, 1H), 7.53 (t,  $J = 2.0$  Hz, 1H), 7.39 (t,  $J = 8.0$  Hz, 1H), 7.19-7.16 (m, 2H), 6.92-6.89 (m, 2H), 6.87 (d,  $J = 7.6$  Hz, 1H), 6.58 (d,  $J = 8.6$  Hz, 1H), 5.16 (d,  $J = 7.1$  Hz, 1H), 4.70 (q,  $J = 7.2$  Hz, 1H), 4.44 (dd,  $J = 8.6, 5.2$  Hz, 1H), 4.16 (quint,  $J = 6.9$  Hz, 1H), 3.69 (s, 3H), 3.05 (d,  $J = 6.8$  Hz, 2H), 2.56 (s, 3H), 2.15-2.04 (m, 1H), 1.39 (s, 9H), 1.31 (d,  $J = 7.1$  Hz, 3H), 0.86 (d,  $J = 6.8$  Hz, 3H), 0.83 (d,  $J = 6.8$  Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  197.6, 172.9, 171.9, 170.7, 157.9, 155.6, 138.9, 132.0, 131.0, 130.1, 123.3, 123.2, 119.4, 118.0, 80.3, 57.5, 54.5, 52.2, 50.4, 37.4, 31.3, 28.4, 26.9, 24.8, 19.0, 18.3, 17.9; IR (neat) 3293, 3068, 2969, 2923, 871, 2848, 1741, 1684, 1642, 1505, 1433, 1365, 1224, 1159, 1019, 897, 859, 783; HRMS (ESI) calcd for  $[\text{C}_{31}\text{H}_{41}\text{N}_3\text{O}_8 + \text{H}]^+$ : 584.2966, found 584.3011.

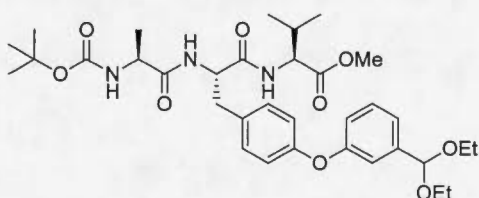


**(6S,9S,12S)-Methyl 9-(4-(3-formylphenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16l)**



Method B was followed on a 0.0752 mmol scale starting from **12** and organobismuthine **6l**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16l** as a white solid (42.7 mg, 99%): mp 65-66 °C;  $R_f$  0.47 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  9.94 (s, 1H), 7.58 (td,  $J$  = 7.6, 1.3 Hz, 1H), 7.48 (t,  $J$  = 7.9 Hz, 1H), 7.41 (dd,  $J$  = 2.5, 1.4 Hz, 1H), 7.26 (ddd,  $J$  = 8.1, 2.7, 1.2 Hz, 1H), 7.23-7.18 (m, 2H), 6.96-6.91 (m, 2H), 6.86 (d,  $J$  = 7.9 Hz, 1H), 6.54 (d,  $J$  = 8.6 Hz, 1H), 5.11 (d,  $J$  = 7.1 Hz, 1H), 4.70 (q,  $J$  = 7.1 Hz, 1H), 4.44 (dd,  $J$  = 8.6, 5.1 Hz, 1H), 4.16 (quint,  $J$  = 6.9 Hz, 1H), 3.69 (s, 3H), 3.09-3.06 (m, 2H), 2.16-2.05 (m, 1H), 1.40 (s, 9H), 1.31 (d,  $J$  = 7.1 Hz, 3H), 0.87 (d,  $J$  = 6.8 Hz, 3H), 0.84 (d,  $J$  = 6.9 Hz, 3H);  $^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )  $\delta$  191.7, 172.9, 171.9, 170.6, 158.6, 155.6, 155.3, 138.2, 132.5, 131.1, 130.6, 124.9, 124.6, 119.8, 118.0, 80.4, 57.5, 54.6, 52.2, 50.4, 37.5, 31.3, 28.4, 19.0, 18.3, 17.9; IR (neat) 3285, 3081, 2966, 2925, 2852, 2729, 1740, 1695, 1650, 1503, 1450, 1360, 1250, 1164, 1013, 751; HRMS (ESI) calcd for  $[\text{C}_{30}\text{H}_{39}\text{N}_3\text{O}_8 + \text{H}]^+$ : 570.2810, found 570.2797.

**(6S,9S,12S)-Methyl 9-(4-(3-(diethoxymethyl)phenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16m)**

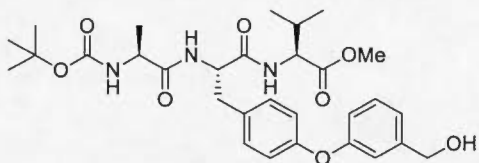


Method B was followed on a 0.0649 mmol scale starting from **12** and organobismuthine **6m**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16m** as a white solid (39.0 mg, 93%): mp 53-54 °C;  $R_f$  0.49 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.31-7.26 (m, 1H), 7.20 (d,  $J$  = 7.7 Hz, 1H), 7.17-7.13 (m, 3H), 6.90 (d,  $J$  = 8.0 Hz, 3H), 6.83 (d,  $J$  = 7.8 Hz, 1H), 6.50 (d,  $J$  = 8.6 Hz, 1H), 5.45 (s, 1H), 5.04 (d,  $J$  = 7.1 Hz, 1H), 4.67 (q,  $J$  = 7.2 Hz, 1H), 4.42 (dd,  $J$  = 8.7, 5.3 Hz, 1H), 4.15 (quint,  $J$  = 5.9 Hz, 1H), 3.69 (s, 3H), 3.65-3.46 (m, 4H), 3.06-3.03 (m, 2H), 2.14-2.03 (m, 1H), 1.41 (s, 9H), 1.31 (d,  $J$  = 7.1 Hz, 3H), 1.21 (t,  $J$  = 7.1 Hz, 6H), 0.86 (d,  $J$  = 7.2 Hz, 3H), 0.83 (d,  $J$  = 6.6 Hz, 3H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8,



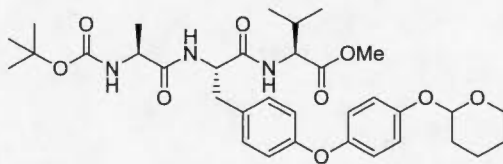
171.8, 170.6, 157.2, 156.4, 155.5, 141.4, 131.2, 130.8, 129.6, 121.7, 119.0, 118.6, 117.5, 101.1, 80.4, 61.2, 57.5, 54.6, 52.2, 50.4, 37.4, 31.3, 28.4, 19.0, 18.4, 17.9, 15.3; IR (neat) 3281, 3072, 2970, 2925, 2876, 1740, 1642, 1503, 1438, 1364, 1246, 1164, 1050, 903, 854, 780; HRMS (ESI) calcd for  $[C_{34}H_{49}N_4O_4 + NH_4]^+$ : 675.3838, found 675.3817.

**(6S,9S,12S)-Methyl 9-(4-(3-(hydroxymethyl)phenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16n)**



Method A was followed on a 0.107 mmol scale starting from **12** and organobismuthine **6n**. The crude product was purified on silica gel (70% EtOAc/hexanes) to afford **16n** as a white solid (56.5 mg, 93%): mp 73-74 °C;  $R_f$  0.58 (100% EtOAc);  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.32-7.20 (m, 1H), 7.12 (d,  $J$  = 8.5 Hz, 2H), 7.03 (d,  $J$  = 7.6 Hz, 1H), 6.98-6.94 (m, 2H), 6.92-6.85 (m, 3H), 6.73 (d,  $J$  = 6.9 Hz, 1H), 5.28-5.17 (m, 1H), 4.70 (q,  $J$  = 7.2 Hz, 1H), 4.63-4.62 (m, 2H), 4.43 (dd,  $J$  = 8.7, 5.3 Hz, 1H), 4.16-4.12 (m, 1H), 3.68 (s, 3H), 3.07-2.94 (m, 2H), 2.72 (s(br), 1H), 2.14-2.05 (m, 1H), 1.39 (s, 9H), 1.28 (d,  $J$  = 7.1 Hz, 3H), 0.86 (d,  $J$  = 6.8 Hz, 3H), 0.83 (d,  $J$  = 6.9 Hz, 3H);  $^{13}C$ -NMR (75 MHz,  $CDCl_3$ )  $\delta$  172.9, 172.0, 170.7, 157.9, 156.0, 155.5, 143.3, 131.5, 130.8, 129.9, 121.4, 119.5, 117.8, 116.7, 80.3, 64.8, 57.5, 54.6, 52.3, 50.3, 37.5, 31.3, 28.4, 19.0, 18.5, 18.0; IR (neat) 3281, 3072, 2962, 2920, 2852, 1737, 1642, 1502, 1441, 1365, 1247, 1163, 1019, 927, 855, 775, 692; HRMS (ESI) calcd for  $[C_{30}H_{41}N_3O_8 + H]^+$ : 572.2966, found 572.2981.

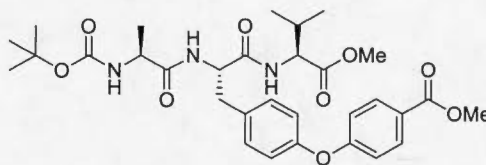
**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-(4-(4-((tetrahydro-2H-pyran-2-yl)oxy)phenoxy)benzyl)-3-oxa-5,8,11-triazatridecan-13-oate (16o)**



Method A was followed on a 0.107 mmol scale starting from **12** and organobismuthine **6o**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16o** as a light brown solid (57.5 mg, 82%): mp 79-80 °C;  $R_f$  0.41 (60% EtOAc/hexanes);  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.11 (d,  $J$  = 8.5 Hz, 2H), 7.02-6.98 (m, 2H), 6.94-6.90 (m, 2H), 6.85-6.83 (m, 3H), 6.54-6.51 (m, 1H), 5.33 (t,  $J$  = 3.4 Hz, 1H), 5.07 (d,  $J$  = 7.2 Hz, 1H), 4.66 (q,  $J$  = 7.1 Hz, 1H), 4.42 (dd,  $J$  = 8.5, 5.1 Hz, 1H), 4.16 (quint,  $J$  = 7.5 Hz, 1H), 3.96-3.88 (m, 1H),

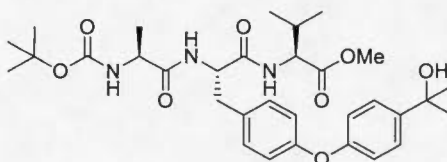
3.68 (s, 3H), 3.62-3.56 (m, 1H), 3.08-2.95 (m, 2H), 2.11-2.03 (m, 1H), 2.01-1.96 (m, 1H), 1.87-1.82 (m, 2H), 1.70-1.57 (m, 4H), 1.40 (s, 9H), 1.30 (d,  $J = 7.1$  Hz, 3H), 0.85 (d,  $J = 6.7$  Hz, 3H), 0.82 (d,  $J = 6.9$  Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 171.8, 170.7, 157.4, 155.4, 153.4, 151.0, 130.6, 130.4, 120.6, 118.0, 117.8, 97.1, 80.3, 62.2, 57.5, 54.6, 52.2, 50.4, 37.4, 31.3, 30.5, 28.4, 25.3, 19.0, 18.5, 17.9; IR (neat) 3285, 3068, 2935, 2874, 1741, 1642, 1494, 1365, 1216, 1163, 1110, 1019, 962, 916, 874, 829; HRMS (ESI) calcd for  $[\text{C}_{34}\text{H}_{47}\text{N}_3\text{O}_9 + \text{H}]^+$ : 642.3385, found 642.3387.

**(6S,9S,12S)-Methyl 12-isopropyl-9-(4-(4-(methoxycarbonyl)phenoxy)benzyl)-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16p)**



Method A was followed on a 0.0604 mmol scale starting from **12** and organobismuthine **6p**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16p** as a white solid (33.4 mg, 92%): mp 79-80 °C;  $R_f$  0.36 (60% EtOAc/hexanes);  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.98-7.94 (m, 2H), 7.20 (d,  $J = 8.4$  Hz, 2H), 6.95 (d,  $J = 7.7$  Hz, 4H), 6.93-6.88 (m, 1H), 6.70-6.52 (m, 1H), 5.12-5.10 (m, 1H), 4.72 (q,  $J = 6.2$  Hz, 1H), 4.44 (dd,  $J = 8.4, 5.1$  Hz, 1H), 4.18 (quint,  $J = 7.6$  Hz, 1H), 3.88 (s, 3H), 3.68 (s, 3H), 3.05 (d,  $J = 7.0$  Hz, 2H), 2.15-2.04 (m, 1H), 1.40 (s, 9H), 1.31 (d,  $J = 7.0$  Hz, 3H), 0.86 (d,  $J = 6.8$  Hz, 3H), 0.83 (d,  $J = 6.9$  Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.9, 171.9, 170.6, 166.7, 161.8, 155.5, 154.7, 132.7, 131.8, 131.1, 124.6, 120.3, 117.3, 80.3, 57.4, 54.6, 52.3, 52.1, 50.3, 37.6, 31.3, 28.4, 19.0, 18.5, 17.9; IR (neat) 3289, 3076, 2969, 1718, 1646, 1498, 1433, 1239, 1159, 1099, 1015, 851, 764; HRMS (ESI) calcd for  $[\text{C}_{31}\text{H}_{41}\text{N}_3\text{O}_9 + \text{H}]^+$ : 600.2916, found 600.2914.

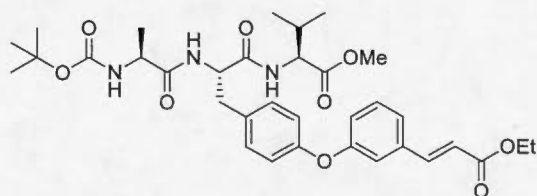
**(6S,9S,12S)-Methyl 9-(4-(4-(2-hydroxypropan-2-yl)phenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16q)**



Method B was followed on a 0.1065 mmol scale starting from **12** and organobismuthine **6q**. The crude product was purified on silica gel (50% EtOAc/hexanes) to afford **16q** as an off-white solid (56.6 mg, 93%): mp 78-79 °C;  $R_f$  0.52 (60% EtOAc/hexanes);  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.44-7.41 (m, 2H), 7.16-7.13 (m,

2H), 6.95-6.92 (m, 2H), 6.91-6.88 (m, 2H), 6.84 (d,  $J = 7.9$  Hz, 1H), 6.52 (d,  $J = 8.4$  Hz, 1H), 5.04 (d,  $J = 7.2$  Hz, 1H), 4.67 (q,  $J = 7.1$  Hz, 1H), 4.42 (dd,  $J = 8.6, 5.1$  Hz, 1H), 4.15 (quint,  $J = 7.4$  Hz, 1H), 3.69 (s, 3H), 3.03 (d,  $J = 7.1$  Hz, 2H), 2.14-2.03 (m, 1H), 2.00 (s(br), 1H), 1.57 (s, 6H), 1.41 (s, 9H), 1.30 (d,  $J = 7.1$  Hz, 3H), 0.86 (d,  $J = 6.8$  Hz, 3H), 0.83 (d,  $J = 6.9$  Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 171.8, 170.6, 156.5, 155.9, 155.5, 144.2, 131.2, 130.7, 126.0, 119.0, 118.5, 80.4, 72.4, 57.5, 54.6, 52.3, 50.4, 37.4, 32.0, 31.3, 28.4, 19.0, 18.5, 17.9; IR (neat) 3289, 3068, 2969, 2931, 2871, 1741, 1692, 1646, 1498, 1361, 1235, 1163, 1015, 950, 855, 753; HRMS (ESI) calcd for  $[\text{C}_{32}\text{H}_{45}\text{N}_3\text{O}_8 - \text{H}]^-$ : 598.3136, found 598.3120.

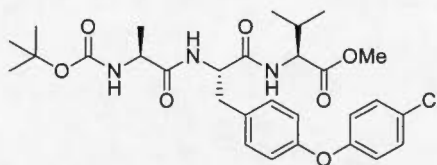
**(6S,9S,12S)-Methyl 9-(4-(3-((*E*)-3-ethoxy-3-oxoprop-1-en-1-yl)phenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16r)**



Method A was followed on a 0.0408 mmol scale starting from **12** and organobismuthine **6r**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **16r** as a white solid (23.6 mg, 89%): mp 68-69 °C;  $R_f$  0.46 (60% EtOAc/hexanes);  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.61 (d,  $J = 16.0$  Hz, 1H), 7.32 (t,  $J = 7.8$  Hz, 1H), 7.24-7.22 (m, 1H), 7.19 (d,  $J = 8.5$  Hz, 2H), 7.15-7.13 (m, 1H), 7.02-6.98 (m, 1H), 6.93-6.90 (m, 2H), 6.84 (d,  $J = 7.9$  Hz, 1H), 6.54-6.42 (m, 1H), 6.37 (d,  $J = 16.0$  Hz, 1H), 5.04 (d,  $J = 7.2$  Hz, 1H), 4.68 (q,  $J = 7.1$  Hz, 1H), 4.44 (dd,  $J = 8.6, 5.1$  Hz, 1H), 4.24 (q,  $J = 7.1$  Hz, 2H), 4.16 (quint,  $J = 6.8$  Hz, 1H), 3.69 (s, 3H), 3.05 (d,  $J = 6.9$  Hz, 2H), 2.15-2.06 (m, 1H), 1.41 (s, 9H), 1.33-1.28 (m, 6H), 0.86 (d,  $J = 6.7$  Hz, 3H), 0.84 (d,  $J = 6.9$  Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 171.8, 170.6, 166.9, 158.0, 155.8, 155.5, 144.0, 136.4, 131.8, 130.9, 130.3, 123.2, 120.5, 119.4, 119.2, 117.6, 80.4, 60.7, 57.4, 54.7, 52.3, 50.5, 37.5, 31.3, 28.4, 19.0, 18.4, 17.9, 14.4; IR (neat) 3289, 3068, 2969, 2931, 1707, 1642, 1498, 1441, 1365, 1232, 1163, 1023, 977, 855, 783; HRMS (ESI) calcd for  $[\text{C}_{34}\text{H}_{45}\text{N}_3\text{O}_9 + \text{H}]^+$ : 640.3229, found 640.3238.

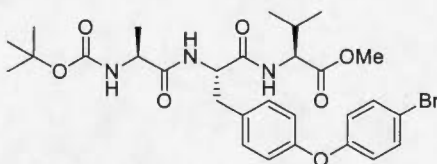


**(6S,9S,12S)-Methyl 9-(4-(4-chlorophenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16s)**



Method A was followed on a 0.0749 mmol scale starting from **12** and organobismuthine **6s**. The crude product was purified on silica gel (50% EtOAc/hexanes) to afford **16s** as a white solid (42.3 mg, 98%): mp 151-152 °C;  $R_f$  0.53 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.29-7.24 (m, 2H), 7.19-7.16 (m, 2H), 6.94-6.87 (m, 4H), 6.78 (d,  $J$  = 7.8 Hz, 1H), 6.42 (d,  $J$  = 8.6 Hz, 1H), 4.96 (d,  $J$  = 7.2 Hz, 1H), 4.66 (q,  $J$  = 7.1 Hz, 1H), 4.43 (dd,  $J$  = 8.6, 5.1 Hz, 1H), 4.15 (quint,  $J$  = 6.9 Hz, 1H), 3.70 (s, 3H), 3.05 (d,  $J$  = 6.9 Hz, 2H), 2.17-2.04 (m, 1H), 1.42 (s, 9H), 1.31 (d,  $J$  = 7.1 Hz, 3H), 0.86 (d,  $J$  = 6.9 Hz, 3H), 0.83 (d,  $J$  = 6.9 Hz, 3H);  $^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 171.9, 170.5, 156.05, 156.03, 155.6, 131.7, 130.9, 129.8, 128.4, 120.1, 119.2, 80.5, 57.5, 54.7, 52.3, 50.4, 37.5, 31.3, 28.4, 19.0, 18.4, 17.9; IR (neat) 3277, 3072, 2958, 2921, 2852, 1740, 1687, 1642, 1499, 1479, 1360, 1234, 1164, 1086, 1001, 825, 756; HRMS (ESI) calcd for  $[\text{C}_{29}\text{H}_{38}\text{ClN}_3\text{O}_7 + \text{H}]^+$ : 576.2471, found 576.2486.

**(6S,9S,12S)-Methyl 9-(4-(4-bromophenoxy)benzyl)-12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-3-oxa-5,8,11-triazatridecan-13-oate (16t)**

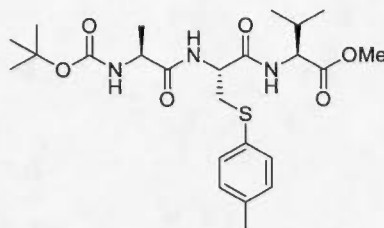


Method A was followed on a 0.0749 mmol scale starting from **12** and organobismuthine **6t**. The crude product was purified on silica gel (50% EtOAc/hexanes) to afford **16t** as a white solid (40.9 mg, 88%): mp 158-159 °C;  $R_f$  0.41 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.43-7.38 (m, 2H), 7.20-7.15 (m, 2H), 6.91-6.85 (m, 4H), 6.81 (d,  $J$  = 7.9 Hz, 1H), 6.45 (d,  $J$  = 8.6 Hz, 1H), 4.98 (d,  $J$  = 7.1 Hz, 1H), 4.67 (q,  $J$  = 7.1 Hz, 1H), 4.43 (dd,  $J$  = 8.6, 5.1 Hz, 1H), 4.15 (quint,  $J$  = 7.1 Hz, 1H), 3.69 (s, 3H), 3.04 (d,  $J$  = 6.9 Hz, 2H), 2.15-2.04 (m, 1H), 1.41 (s, 9H), 1.31 (d,  $J$  = 7.1 Hz, 3H), 0.86 (d,  $J$  = 6.8 Hz, 3H), 0.83 (d,  $J$  = 6.9 Hz, 3H);  $^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )  $\delta$  172.8, 171.9, 170.6, 156.6, 155.9, 155.5, 132.8, 131.8, 130.9, 120.5, 119.3, 115.7, 80.4, 57.5, 54.6, 52.3, 50.4, 37.5, 31.3, 28.4, 19.0, 18.4, 17.9; IR (neat) 3281, 3077, 2962, 2917, 2852, 1740,



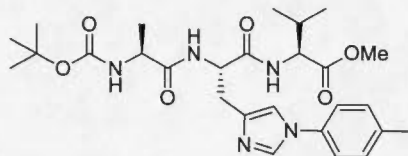
1687, 1642, 1499, 1479, 1364, 1234, 1164, 1070, 1005, 821, 747; HRMS (ESI) calcd for  $[C_{29}H_{38}BrN_3O_7 + H]^+$ : 620.1954, found 620.1966.

**(6S,9R,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-((*p*-tolylthio)methyl)-3-oxa-5,8,11-triazatridecan-13-oate (17)**



Method B was followed on a 0.122 mmol scale starting from **13** and organobismuthine **6a**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **17** as an off-white solid (11.7 mg, 19%): mp 112–113 °C;  $R_f$  0.56 (60% EtOAc/hexanes);  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.35 (d,  $J$  = 7.7 Hz, 2H), 7.13 (d,  $J$  = 7.6 Hz, 2H), 6.95–6.75 (m, 2H), 4.76 (d,  $J$  = 4.4 Hz, 1H), 4.53–4.42 (m, 2H), 4.13 (quint,  $J$  = 7.3 Hz, 1H), 3.73 (s, 3H), 3.38 (dd,  $J$  = 14.0, 5.9 Hz, 1H), 3.18 (dd,  $J$  = 14.0, 6.7 Hz, 1H), 2.33 (s, 3H), 2.20–2.09 (m, 1H), 1.45 (s, 9H), 1.33 (d,  $J$  = 7.1 Hz, 3H), 0.90 (d,  $J$  = 6.5 Hz, 3H), 0.89 (d,  $J$  = 6.6 Hz, 3H);  $^{13}C$ -NMR (150 MHz,  $CDCl_3$ )  $\delta$  172.8, 171.9, 169.8, 155.5, 137.4, 131.0, 130.9, 130.2, 80.5, 57.7, 52.9, 52.3, 50.5, 36.1, 31.3, 28.5, 21.2, 19.0, 18.3, 18.0; IR (neat) 3289, 3072, 2966, 2921, 2872, 1744, 1646, 1511, 1454, 1360, 1246, 1205, 1164, 1021, 858, 805; HRMS (ESI) calcd for  $[C_{24}H_{37}N_3O_6S - H]^-$ : 494.2330, found 494.2323.

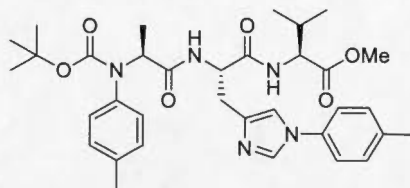
**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-((1-(*p*-tolyl)-1*H*-imidazol-4-yl)methyl)-3-oxa-5,8,11-triazatridecan-13-oate (18a)**



Method B was followed on a 0.0974 mmol scale starting from **14** and organobismuthine **6a**. The crude product was purified on silica gel (5% MeOH/ $CH_2Cl_2$ ) to afford **18a** as an off-white solid (5.5 mg, 11%): mp 54–55 °C;  $R_f$  0.40 (100% EtOAc);  $^1H$ -NMR (600 MHz,  $CDCl_3$ )  $\delta$  8.14 (s(br), 1H), 7.93–7.89 (m, 1H), 7.55–7.45 (m, 1H), 7.31–7.27 (m, 2H), 7.25–7.19 (m, 2H), 7.13 (d,  $J$  = 4.0 Hz, 1H), 5.25 (s(br), 1H), 4.87–4.75 (m, 1H), 4.40 (dd,  $J$  = 8.4, 5.2 Hz, 1H), 4.30–4.20 (m, 1H), 3.68 (s, 3H), 3.30–3.17 (m, 1H), 3.17–3.06 (m, 1H), 2.40 (s, 3H), 2.11–2.08 (m, 1H), 1.44–1.42 (m, 3H), 1.41 (s, 9H), 0.85–0.74 (m, 6H);  $^{13}C$ -NMR (150 MHz,  $CDCl_3$ )  $\delta$  173.2, 172.0, 170.9, 155.6, 138.4, 137.9, 134.33, 134.26, 130.7, 121.6, 117.1, 79.9, 57.7, 53.5, 52.2, 50.7,

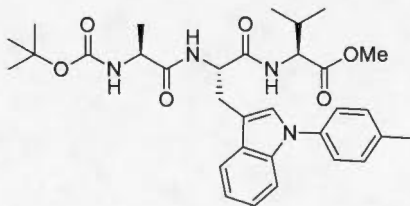
30.9, 29.3, 28.5, 21.1, 19.04, 19.01, 17.8; IR (neat) 3315, 3032, 2965, 2928, 2874, 2855, 1741, 1686, 1512, 1458, 1368, 1320, 1255, 1206, 1166, 1073, 1026, 909, 858, 817, 757, 734, 524; HRMS (ESI) calcd for  $[C_{27}H_{39}N_5O_6 + H]^+$ : 530.2973, found 530.2989.

**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-5-(*p*-tolyl)-9-((1-(*p*-tolyl)-1*H*-imidazol-4-yl)methyl)-3-oxa-5,8,11-triazatridecan-13-oate (18b)**



Method B was followed on a 0.0974 mmol scale starting from **14** and organobismuthine **6a**. The crude product was purified on silica gel (5% MeOH/ $CH_2Cl_2$ ) to afford **18b** as a white solid (27.9 mg, 46%): mp 65–66 °C;  $R_f$  0.64 (100% EtOAc);  $^1H$ -NMR (600 MHz,  $CDCl_3$ )  $\delta$  8.17–8.09 (m, 1H), 8.09 (d,  $J$  = 4.5 Hz, 1H), 7.92 (d,  $J$  = 6.2 Hz, 1H), 7.31–7.26 (m, 4H), 7.22 (s, 1H), 7.13–7.10 (m, 2H), 7.08–7.07 (m, 2H), 5.02 (m, 1H), 4.54 (q,  $J$  = 7.1 Hz, 1H), 4.40 (dd,  $J$  = 8.0, 5.5 Hz, 1H), 3.68 (s, 3H), 3.40–3.30 (m, 1H), 3.16 (dd,  $J$  = 14.6, 7.1 Hz, 1H), 2.40 (s, 3H), 2.31 (s, 3H), 2.22–2.17 (m, 1H), 1.33 (s, 9H), 1.31–1.30 (m, 3H), 0.91 (d,  $J$  = 6.8 Hz, 3H), 0.89 (d,  $J$  = 6.9 Hz, 3H);  $^{13}C$ -NMR (150 MHz,  $CDCl_3$ )  $\delta$  172.4, 172.1, 171.0, 155.0, 138.2, 136.7, 133.9, 130.7, 130.2, 129.5, 129.4, 128.3, 125.7, 121.8, 117.7, 80.9, 58.2, 58.1, 53.2, 52.1, 30.7, 29.9, 28.4, 21.17, 21.16, 19.2, 18.1, 15.6; IR (neat) 3315, 3032, 2965, 2928, 2874, 2855, 1741, 1686, 1512, 1458, 1368, 1320, 1255, 1206, 1166, 1073, 1026, 909, 858, 817, 757, 734, 524; HRMS (ESI) calcd for  $[C_{34}H_{45}N_5O_6 + H]^+$ : 620.3443, found 620.3397.

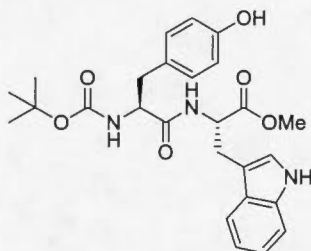
**(6S,9S,12S)-Methyl 12-isopropyl-2,2,6-trimethyl-4,7,10-trioxo-9-((1-(*p*-tolyl)-1*H*-indol-3-yl)methyl)-3-oxa-5,8,11-triazatridecan-13-oate (19)**



Method B was followed on a 0.102 mmol scale starting from **15** and organobismuthine **6a**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **19** as a yellow oil (14.0 mg, 24%):  $R_f$  0.46 (60% EtOAc/hexanes);  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  7.73 (d,  $J$  = 6.9 Hz, 1H), 7.50 (dd,  $J$  = 6.6, 1.5 Hz, 1H),

7.36-7.33 (m, 2H), 7.29-7.26 (m, 2H), 7.23-7.14 (m, 3H), 6.83 (d,  $J = 7.7$  Hz, 1H), 6.49 (d,  $J = 6.9$  Hz, 1H), 5.04 (d,  $J = 7.1$  Hz, 1H), 4.89 (q,  $J = 7.8$  Hz, 1H), 4.37 (dd,  $J = 8.4, 5.2$  Hz, 1H), 4.10 (quint,  $J = 6.8$  Hz, 1H), 3.62 (s, 3H), 3.37 (dd,  $J = 14.5, 5.7$  Hz, 1H), 3.23 (dd,  $J = 14.6, 7.9$  Hz, 1H), 2.41 (s, 3H), 1.97-1.91 (m, 1H), 1.38 (s, 9H), 1.30 (d,  $J = 7.1$  Hz, 3H), 0.68 (d,  $J = 7.7$  Hz, 3H), 0.66 (d,  $J = 8.5$  Hz, 3H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  173.0, 172.0, 171.2, 155.5, 137.1, 136.3, 136.2, 130.2, 128.7, 127.0, 124.1, 122.7, 120.4, 119.2, 111.3, 110.8, 80.2, 57.6, 53.7, 52.1, 50.5, 30.8, 28.3, 27.8, 21.1, 18.7, 18.2, 17.8; IR (neat) 3358, 3048, 2970, 2925, 2872, 1740, 1695, 1650, 1516, 1458, 1360, 1246, 1164, 1017, 825, 731; HRMS (ESI) calcd for  $[\text{C}_{32}\text{H}_{42}\text{N}_4\text{O}_6 + \text{H}]^+$ : 579.3177, found 579.3155.

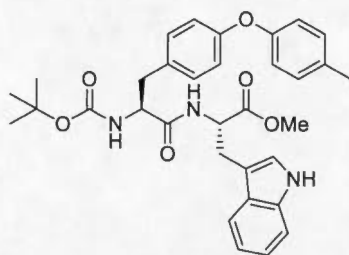
**(S)-Methyl 2-((S)-2-((tert-butoxycarbonyl)amino)-3-(4-hydroxyphenyl)propanamido)-3-(1H-indol-3-yl)propanoate (20)**



Boc-Tyr-OH (608 mg, 2.16 mmol), H-Trp-OMe•HCl (471 mg, 2.16 mmol) and HCTU (981 mg, 2.37 mmol) were dissolved in  $\text{CH}_2\text{Cl}_2$  (22 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to  $0^\circ\text{C}$  in an ice bath and DiPEA (1.32 mL, 7.55 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was diluted with sat. aq.  $\text{NaHCO}_3$  (50 mL) and extracted with  $\text{CH}_2\text{Cl}_2$  (2 X 50 mL). The combined organic phases were washed with sat. aq.  $\text{NaHCO}_3$  (2 X 50 mL), sat. aq.  $\text{NaCl}$  (2 X 50 mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered and concentrated under reduced pressure. The crude product was purified on silica gel (50% EtOAc/hexanes) to afford **20** as a white solid (739.1 mg, 71%): mp  $88-89^\circ\text{C}$ ;  $R_f$  0.39 (60% EtOAc/hexanes);  $^1\text{H}$ -NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.45 (s(br), 1H), 7.39 (d,  $J = 7.7$  Hz, 1H), 7.31 (d,  $J = 8.1$  Hz, 1H), 7.15 (t,  $J = 7.1$  Hz, 1H), 7.06 (t,  $J = 7.4$  Hz, 1H), 6.99-6.82 (m, 1H), 6.79 (s, 1H), 6.61 (d,  $J = 8.1$  Hz, 2H), 6.42-6.15 (m, 1H), 5.30-4.92 (m, 1H), 4.92-4.66 (m, 1H), 4.64-3.82 (m, 1H), 3.52 (s, 3H), 3.21 (d,  $J = 4.6$  Hz, 2H), 3.00-2.63 (m, 2H), 1.40 (s, 9H);  $^{13}\text{C}$ -NMR (75 MHz,  $\text{CDCl}_3$ )  $\delta$  172.0, 171.5, 155.7, 155.4, 136.2, 130.5, 127.8, 127.5, 123.3, 122.3, 119.7, 118.4, 115.7, 111.6, 109.3, 80.6, 56.1, 53.2, 52.5, 37.9, 28.3, 27.6; IR (neat) 3323, 3057, 2977, 2928, 1658, 1513, 1437, 1365, 1228, 1159, 1011, 840, 737; HRMS (ESI) calcd for  $[\text{C}_{26}\text{H}_{31}\text{N}_3\text{O}_6 + \text{H}]^+$ : 482.2286, found 482.2295.

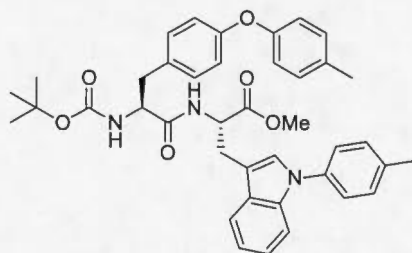


**(S)-Methyl 2-((S)-2-((*tert*-butoxycarbonyl)amino)-3-(4-(*p*-tolylloxy)phenyl)propanamido)-3-(1*H*-indol-3-yl)propanoate (21)**



Method B was followed on a 0.106 mmol scale starting from **20** and organobismuthine **6a**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **21** as an off-white solid (49.0 mg, 82%): mp 58-59 °C;  $R_f$  0.48 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  8.22 (s(br), 1H), 7.40 (d,  $J$  = 7.8 Hz, 1H), 7.33 (d,  $J$  = 8.0 Hz, 1H), 7.17 (t,  $J$  = 7.1 Hz, 1H), 7.13-7.05 (m, 5H), 6.89-6.84 (m, 5H), 6.39 (d,  $J$  = 7.6 Hz, 1H), 5.00-4.98 (m, 1H), 4.87 (q,  $J$  = 5.4 Hz, 1H), 4.32-4.31 (m, 1H), 3.64 (s, 3H), 3.27-3.25 (m, 2H), 2.98-2.96 (m, 2H), 2.32 (s, 3H), 1.38 (s, 9H);  $^{13}\text{C-NMR}$  (75 MHz,  $\text{CDCl}_3$ )  $\delta$  171.9, 171.0, 156.9, 155.4, 154.8, 136.2, 133.0, 131.0, 130.7, 130.3, 127.6, 123.1, 122.3, 119.8, 119.1, 118.5, 111.5, 109.8, 80.2, 55.9, 53.1, 52.5, 37.8, 28.3, 27.8, 20.8; IR (neat) 3330, 3052, 2921, 2856, 1736, 1659, 1503, 1454, 1364, 1234, 1160, 1107, 1017, 813, 743; HRMS (ESI) calcd for  $[\text{C}_{33}\text{H}_{37}\text{N}_3\text{O}_6 + \text{H}]^+$ : 572.2755, found 572.2803.

**(S)-Methyl 2-((S)-2-((*tert*-butoxycarbonyl)amino)-3-(4-(*p*-tolylloxy)phenyl)propanamido)-3-(1-(*p*-tolyl)-1*H*-indol-3-yl)propanoate (22)**



Method B was followed on a 0.105 mmol scale starting from **20** and organobismuthine **6a**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **22** as an orange-pink solid (5.5 mg, 8%): mp 134-135 °C;  $R_f$  0.69 (60% EtOAc/hexanes);  $^1\text{H-NMR}$  (300 MHz,  $\text{CDCl}_3$ )  $\delta$  7.49 (d,  $J$  = 8.1 Hz, 1H), 7.46 (d,  $J$  = 7.9 Hz, 1H), 7.36-7.28 (m, 4H), 7.20 (dt,  $J$  = 8.3, 0.8 Hz, 1H), 7.15-7.06 (m, 6H), 6.85 (dd,  $J$  = 8.4, 1.4 Hz, 4H), 6.43 (d,  $J$  = 7.7 Hz, 1H), 4.94-4.88 (m, 2H), 4.31-4.29 (m, 1H), 3.67 (s, 3H), 3.31 (d,  $J$  = 5.4 Hz, 2H), 2.98 (d,  $J$  = 6.3 Hz, 2H), 2.43 (s, 3H), 2.32 (s, 3H), 1.36 (s, 9H);  $^{13}\text{C-NMR}$  (150 MHz,  $\text{CDCl}_3$ )  $\delta$  171.8, 170.9, 156.9, 155.3, 154.7, 137.1, 136.4, 136.2, 133.0, 130.9, 130.7, 130.33, 130.26, 128.9, 126.9, 124.3, 122.7, 120.3,



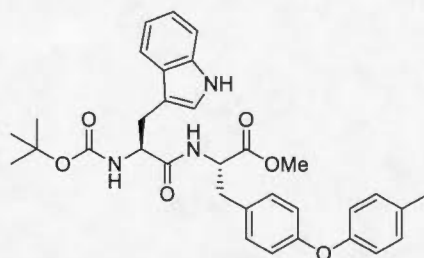
119.2, 118.9, 118.5, 110.9, 110.6, 80.2, 55.9, 53.0, 52.5, 37.7, 28.3, 27.8, 21.2, 20.8; IR (neat) 3326, 3036, 2921, 2852, 1740, 1691, 1654, 1516, 1458, 1385, 1238, 1168, 1013, 821, 735, 629; HRMS (ESI) calcd for  $[C_{40}H_{43}N_3O_6 + H]^+$ : 662.3225, found 662.3228.

**(S)-Methyl 2-((S)-2-((tert-butoxycarbonyl)amino)-3-(1H-indol-3-yl)propanamido)-3-(4-hydroxyphenyl)propanoate (23)**

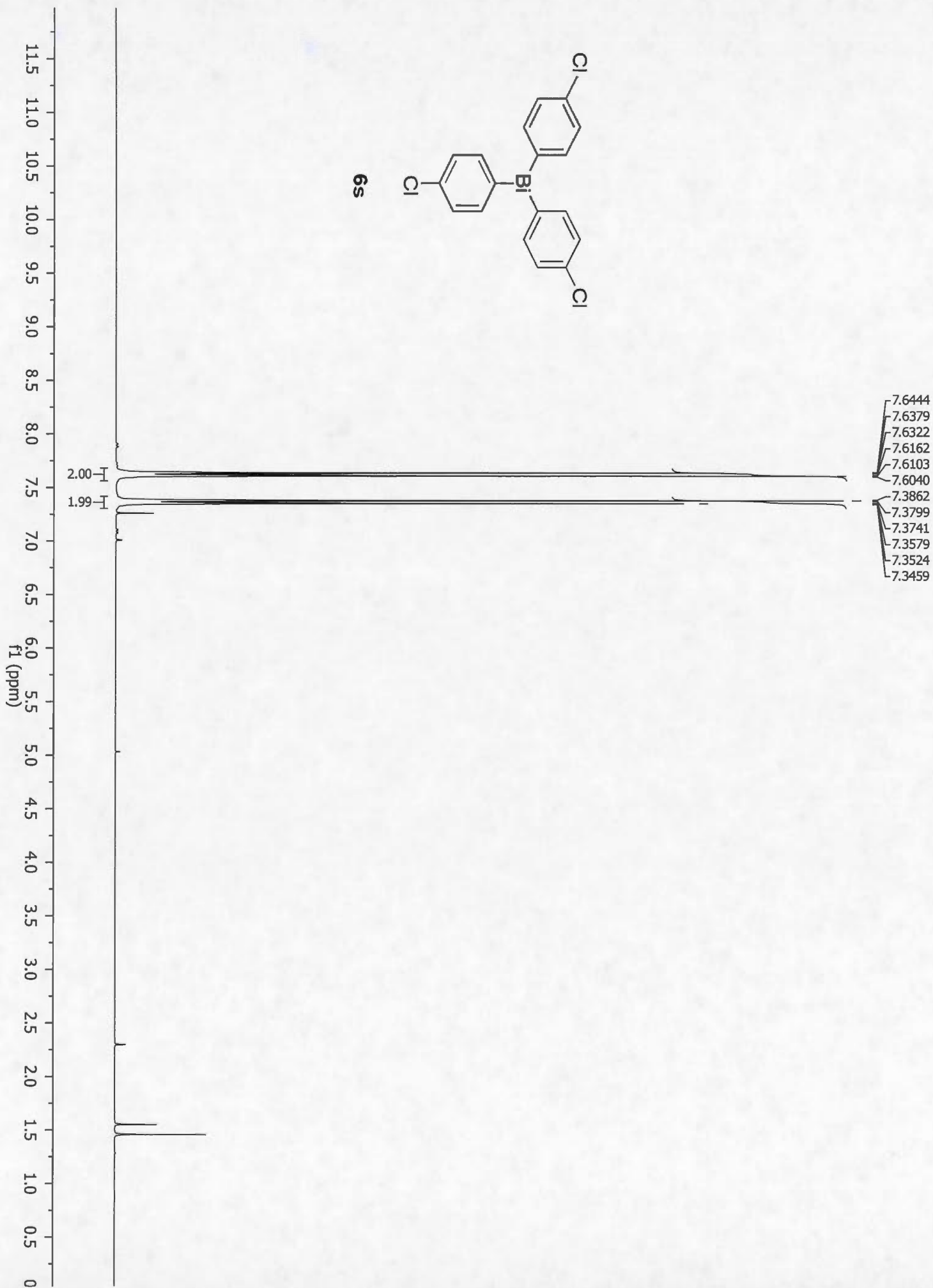


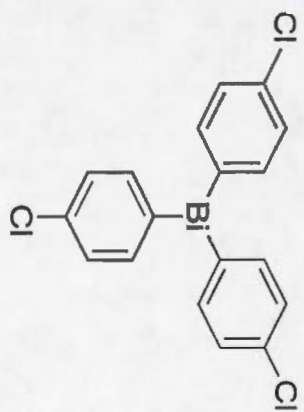
Boc-Trp-OH (927 mg, 3.05 mmol), H-Tyr-OMe•HCl (705 mg, 3.05 mmol) and HCTU (1.385 g, 3.35 mmol) were dissolved in  $CH_2Cl_2$  (30 mL) in a round bottom flask with a stir bar. The flask was chased with argon, the mixture was cooled to 0°C in an ice bath and DIPEA (1.86 mL, 10.66 mmol) was added dropwise. The reaction mixture was warmed up to room temperature and was stirred for 16 h. The reaction mixture was diluted with sat. aq.  $NaHCO_3$  (100 mL) and extracted with  $CH_2Cl_2$  (2 X 100 mL). The combined organic phases were washed with sat. aq.  $NaHCO_3$  (2 X 100 mL), sat. aq.  $NaCl$  (2 X 100 mL), dried over  $Na_2SO_4$ , filtered and concentrated under reduced pressure. The crude product was purified on silica gel (60% EtOAc/hexanes) to afford **23** as a white solid (1.105 g, 74%): mp 105-106°C °C;  $R_f$  0.49 (60% EtOAc/hexanes);  $^1H$ -NMR (300 MHz,  $CDCl_3$ )  $\delta$  8.51 (s(br), 1H), 7.59 (d,  $J$  = 7.8 Hz, 1H), 7.36 (s, 1H), 7.30 (d,  $J$  = 8.0 Hz, 1H), 7.16 (t,  $J$  = 7.2 Hz, 1H), 7.07 (t,  $J$  = 7.7 Hz, 1H), 6.82 (s, 1H), 6.68-6.55 (m, 4H), 6.44 (d,  $J$  = 7.7 Hz, 1H), 5.26 (s(br), 1H), 4.71 (q,  $J$  = 6.5 Hz, 1H), 4.58-4.27 (m, 1H), 3.59 (s, 3H), 3.38-3.18 (m, 1H), 3.11 (dd,  $J$  = 14.5, 6.8 Hz, 1H), 2.91-2.78 (m, 2H), 1.41 (s, 9H);  $^{13}C$ -NMR (75 MHz,  $CDCl_3$ )  $\delta$  171.7, 171.6, 155.8, 155.5, 136.3, 130.4, 127.6, 127.0, 123.5, 122.3, 119.8, 118.8, 115.6, 111.4, 110.1, 80.6, 55.5, 53.5, 52.4, 37.2, 28.4, 28.2; IR (neat) 3327, 3053, 2977, 2931, 1658, 1509, 1433, 1365, 1224, 1159, 1007, 844, 737; HRMS (ESI) calcd for  $[C_{26}H_{31}N_3O_6 + H]^+$ : 482.2286, found 482.2285.

(S)-Methyl 2-((S)-2-((*tert*-butoxycarbonyl)amino)-3-(1*H*-indol-3-yl)propanamido)-3-(4-(*p*-tolxyloxy)phenyl)propanoate (**24**)

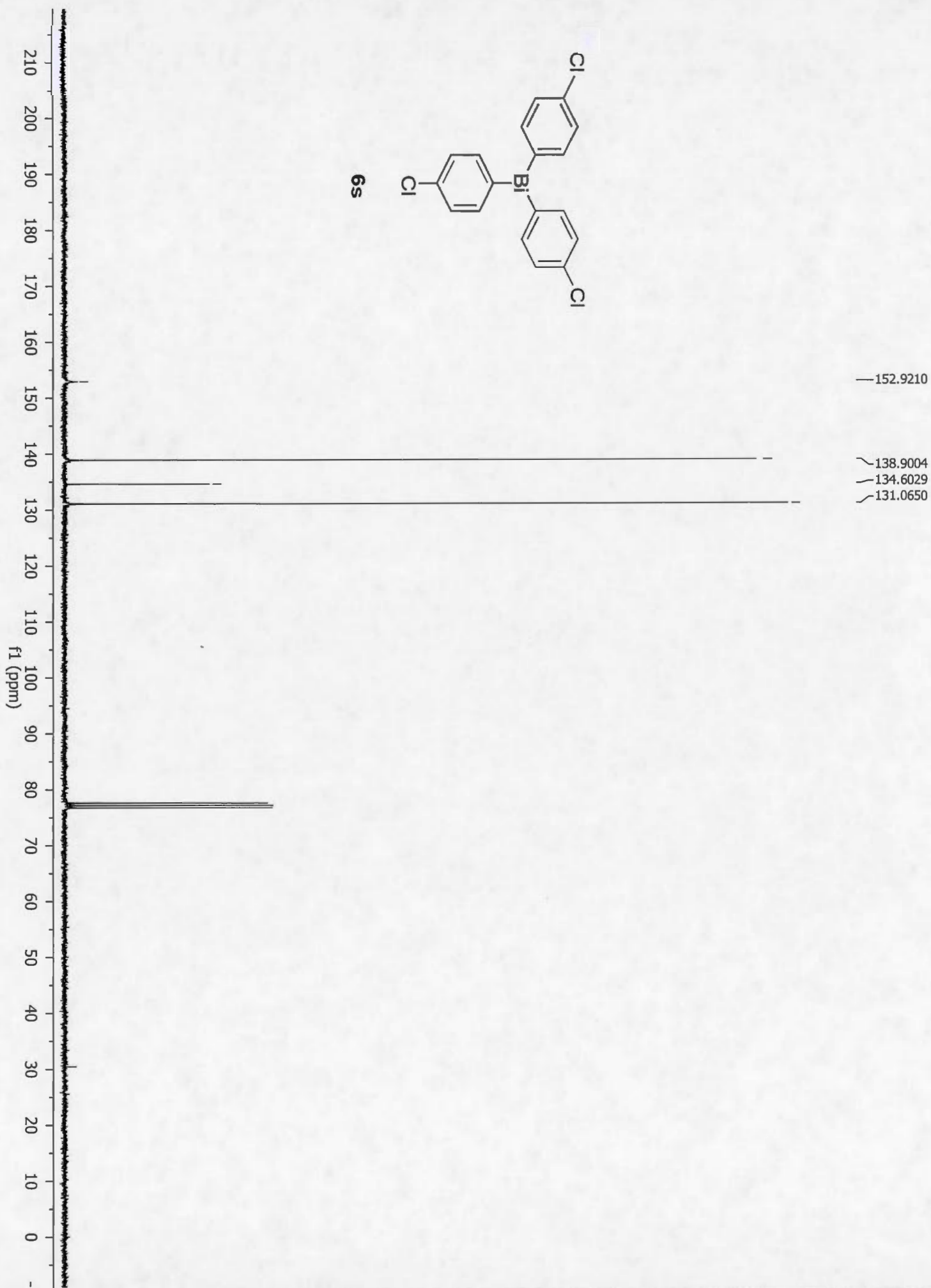


Method A was followed on a 0.104 mmol scale starting from **23** and organobismuthine **6a**. The crude product was purified on silica gel (40% EtOAc/hexanes) to afford **24** as an off-white solid (41.4 mg, 70%): mp 51-52 °C;  $R_f$  0.37 (4% MeOH/CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H-NMR (300 MHz, CDCl<sub>3</sub>) δ 8.31 (s(br), 1H), 7.65 (d,  $J$  = 7.6 Hz, 1H), 7.32 (d,  $J$  = 7.5 Hz, 1H), 7.16 (dt,  $J$  = 7.1, 1.2 Hz, 1H), 7.14-7.05 (m, 3H), 7.00 (s, 1H), 6.90-6.83 (m, 2H), 6.73 (s, 4H), 6.31 (d,  $J$  = 7.4 Hz, 1H), 5.32-5.00 (m, 1H), 4.71 (q,  $J$  = 5.5 Hz, 1H), 4.54-4.31 (m, 1H), 3.61 (s, 3H), 3.39-3.21 (m, 1H), 3.14 (dd,  $J$  = 14.5, 7.0 Hz, 1H) 2.90 (dd,  $J$  = 5.6, 2.2 Hz, 2H), 2.34 (s, 3H), 1.42 (s, 9H); <sup>13</sup>C-NMR (75 MHz, CDCl<sub>3</sub>) δ 171.4, 157.0, 155.5, 154.6, 136.4, 133.1, 130.47, 130.36, 130.0, 127.6, 123.5, 122.4, 119.9, 119.3, 119.0, 118.3, 111.3, 110.5, 80.3, 55.3, 53.4, 52.3, 37.2, 28.4, 20.8; IR (neat) 3370, 3064, 2961, 2924, 2857, 1743, 1662, 1504, 1458, 1441, 1368, 1239, 1172, 1101, 1014, 874, 874, 740; HRMS (ESI) calcd for [C<sub>33</sub>H<sub>37</sub>N<sub>3</sub>O<sub>6</sub> + H]<sup>+</sup>: 572.2755, found 572.2783.

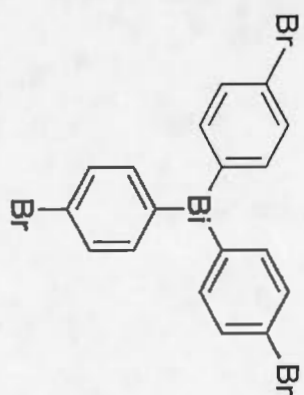




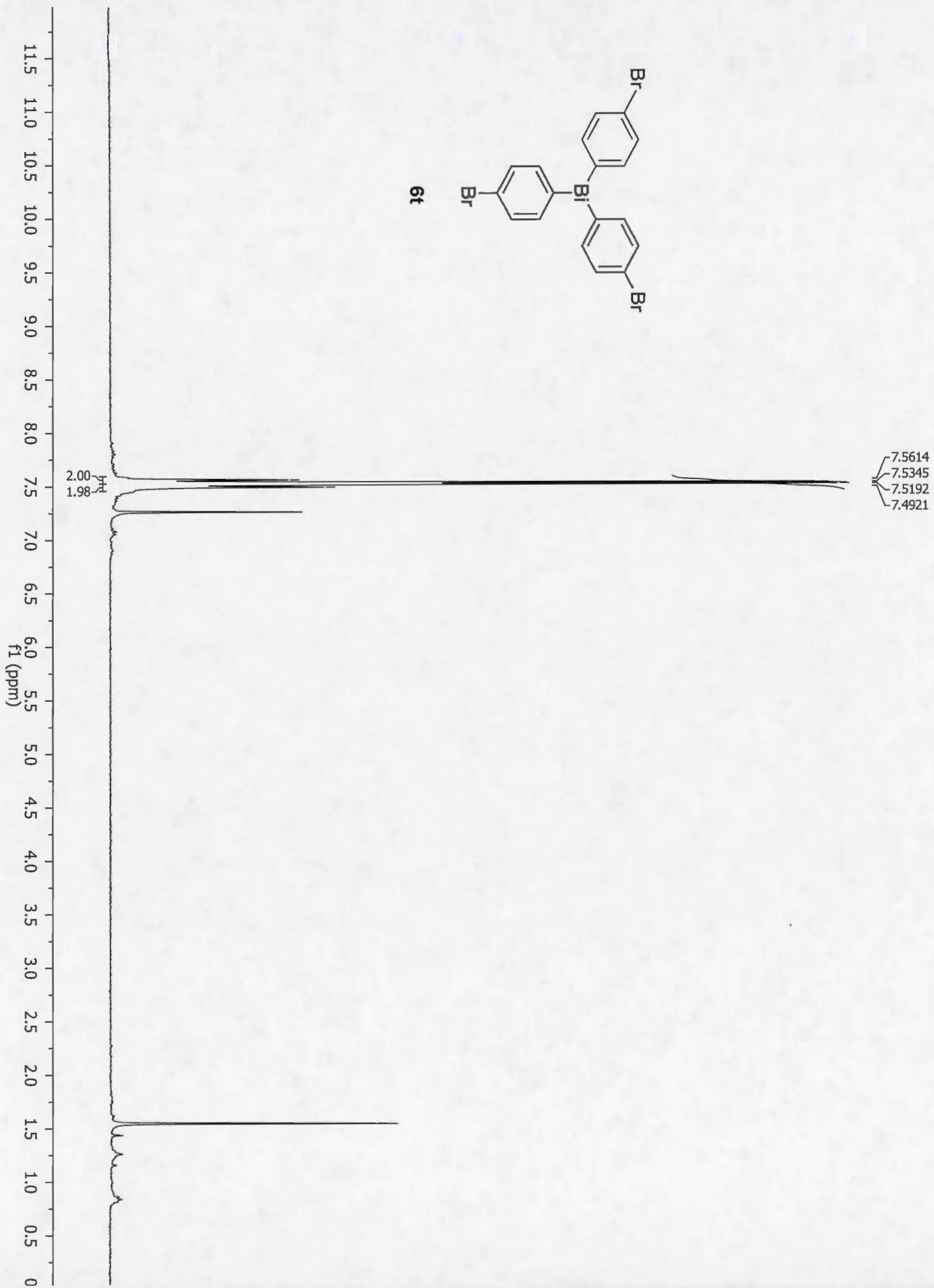
6s

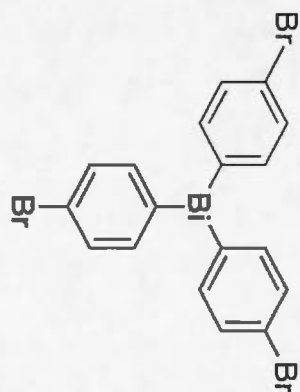




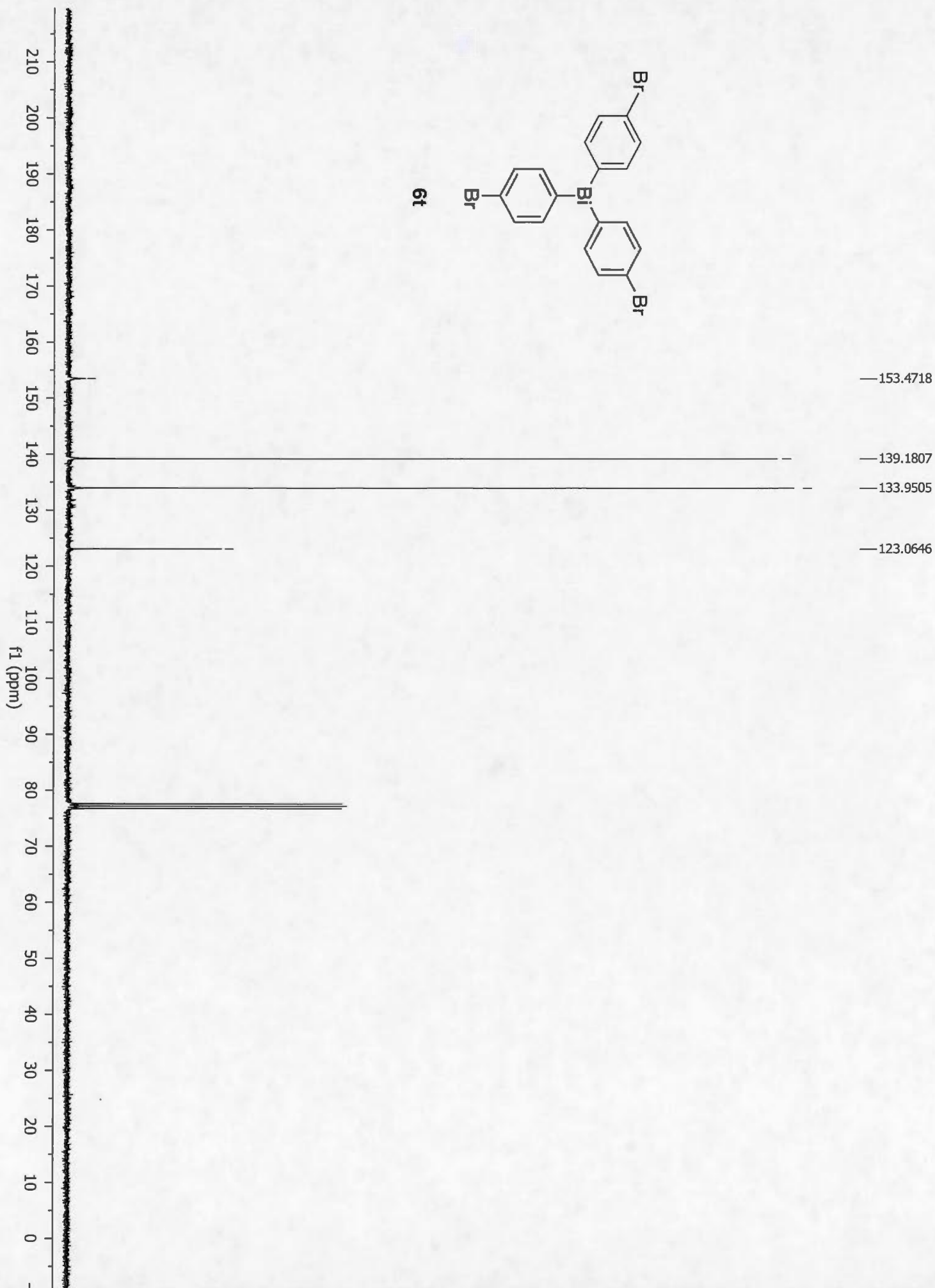


**6t**

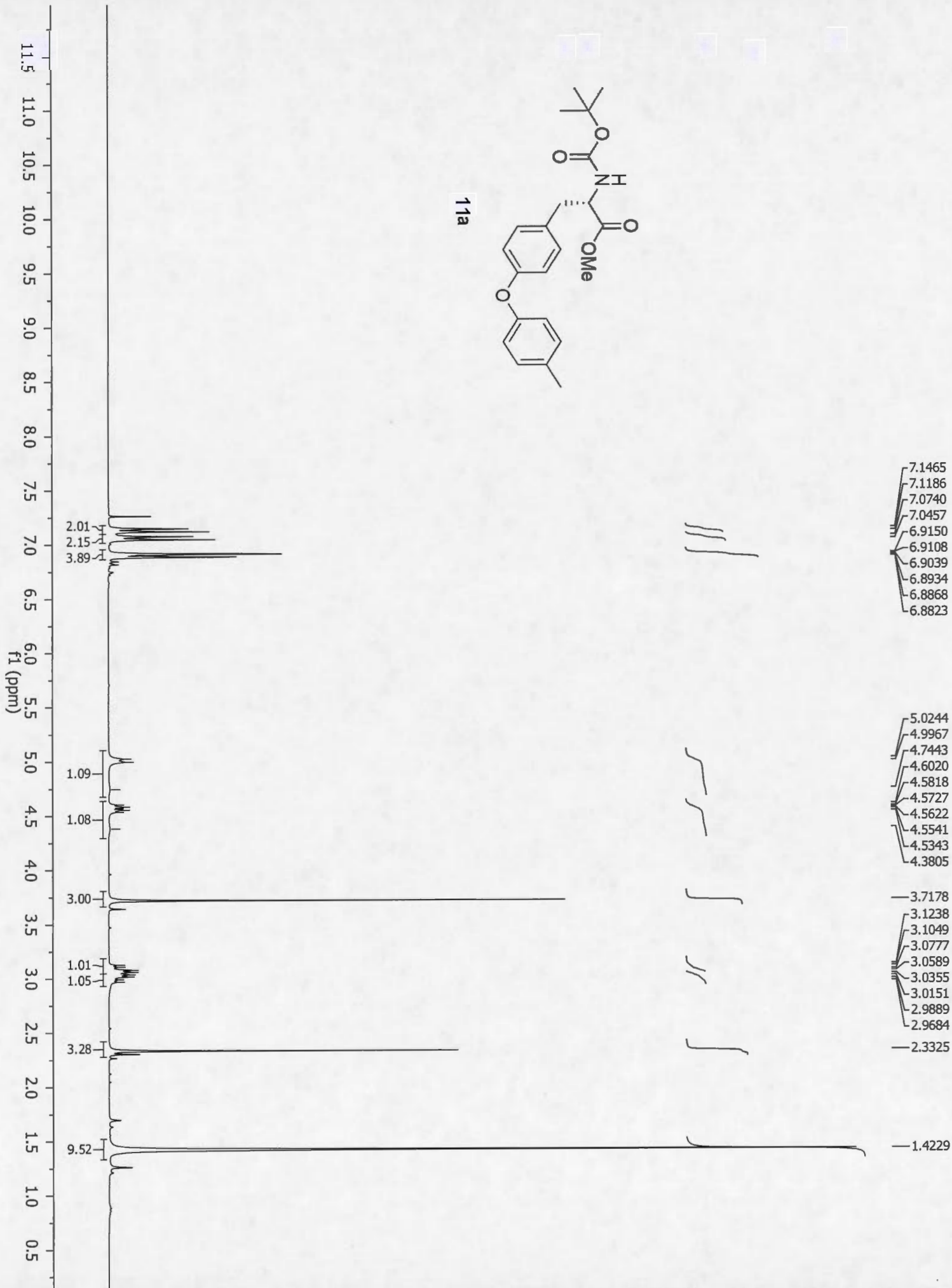
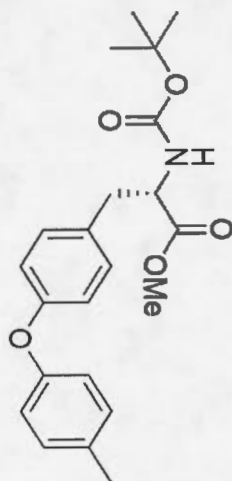


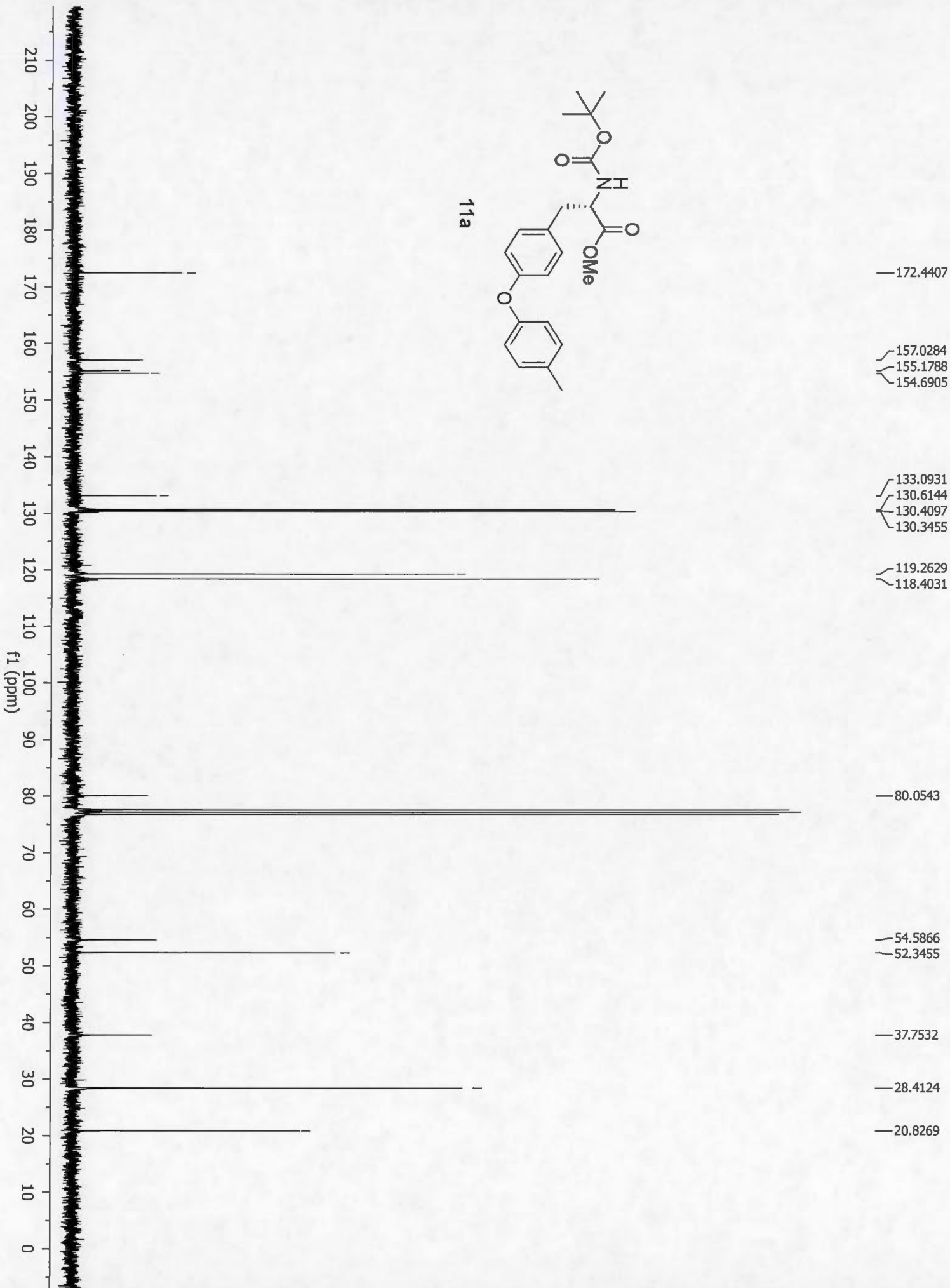


6t



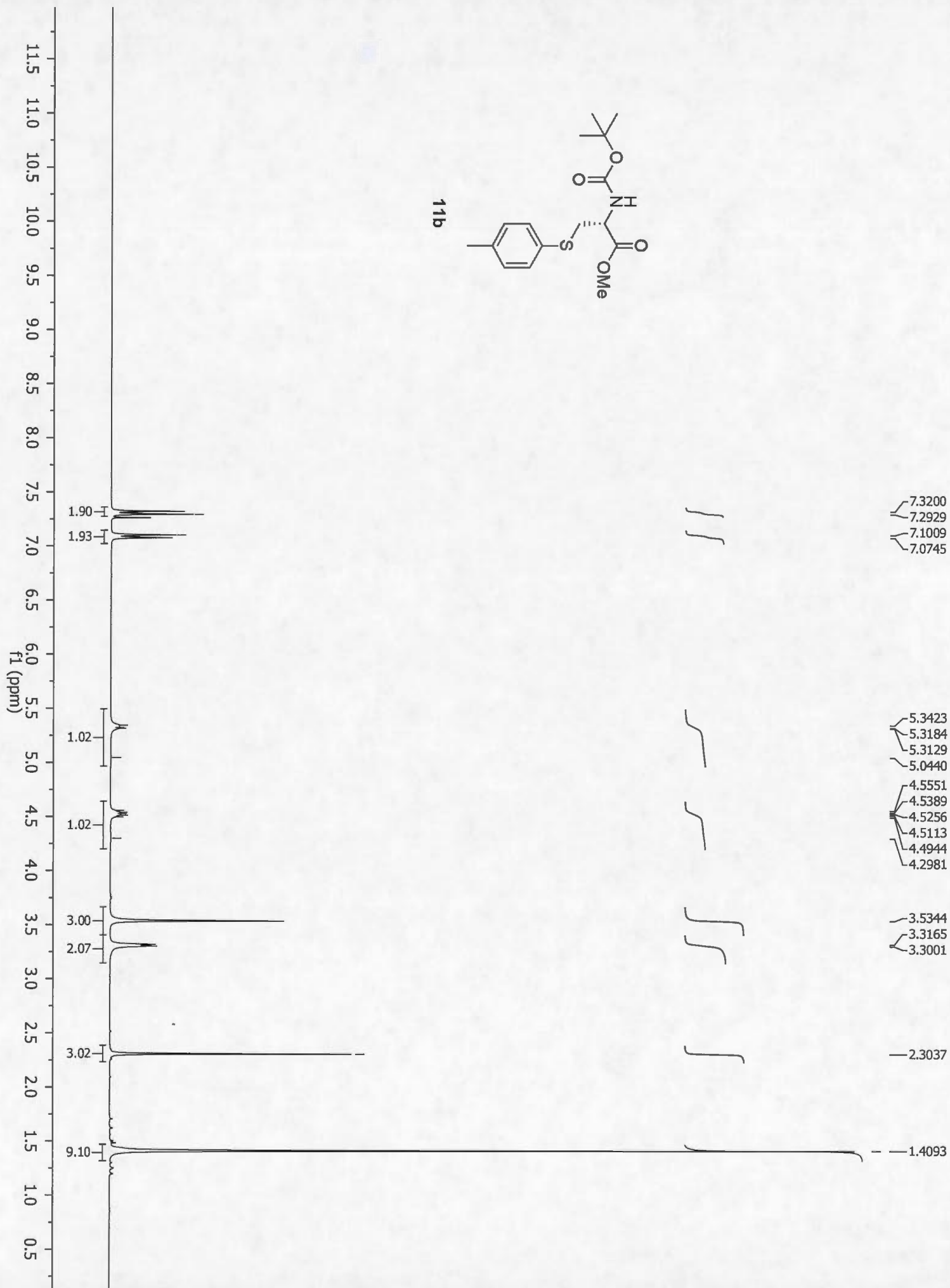
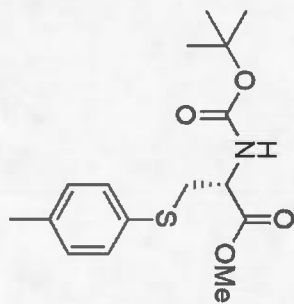
11a



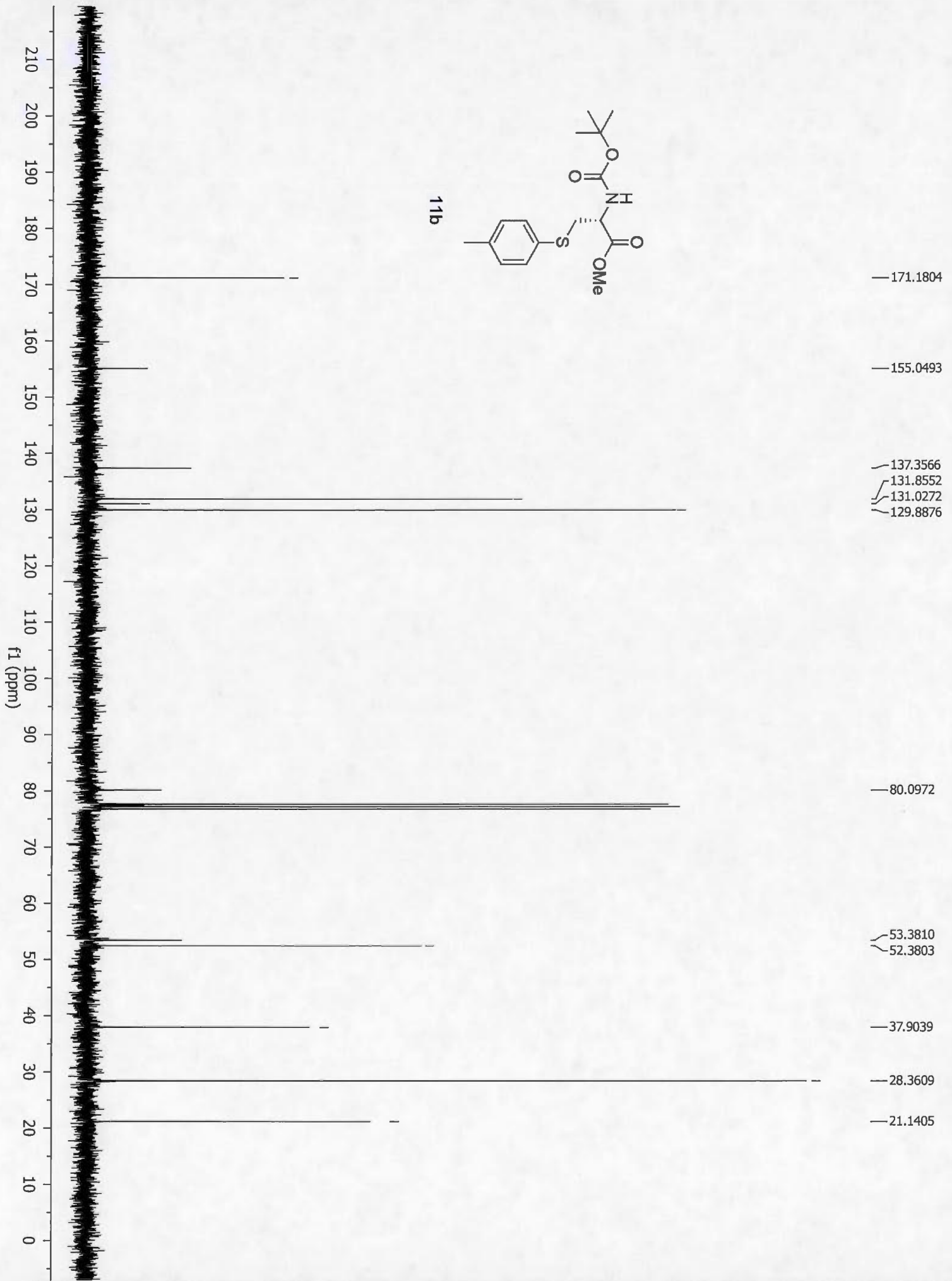
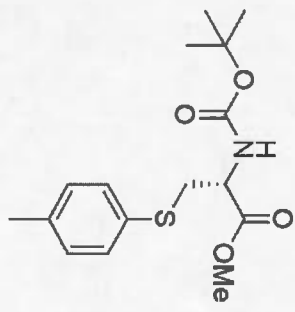




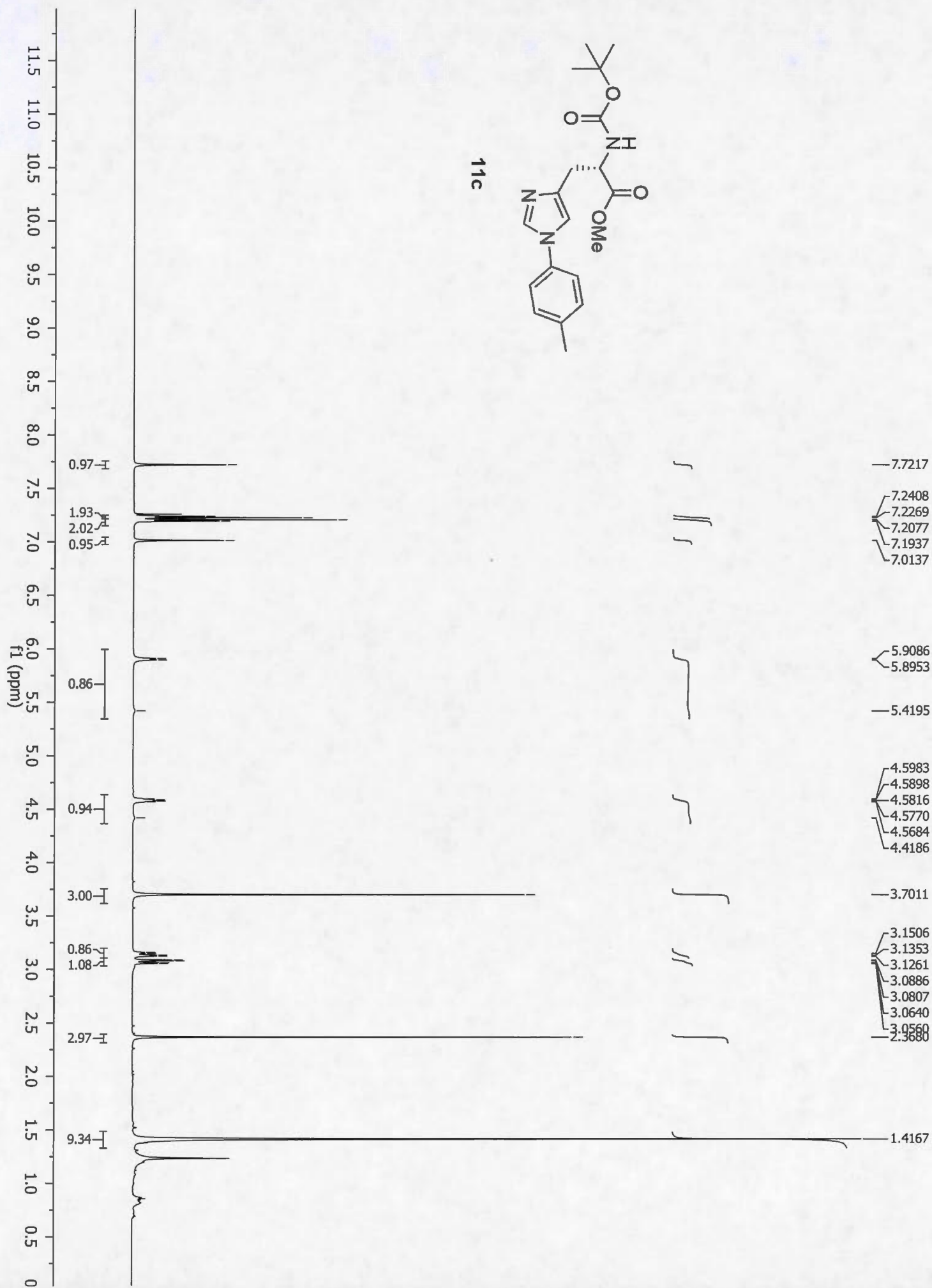
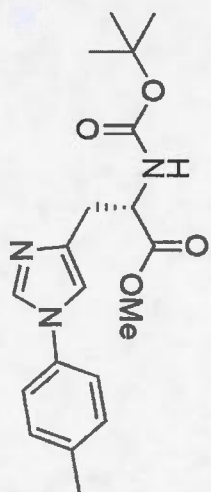
11b

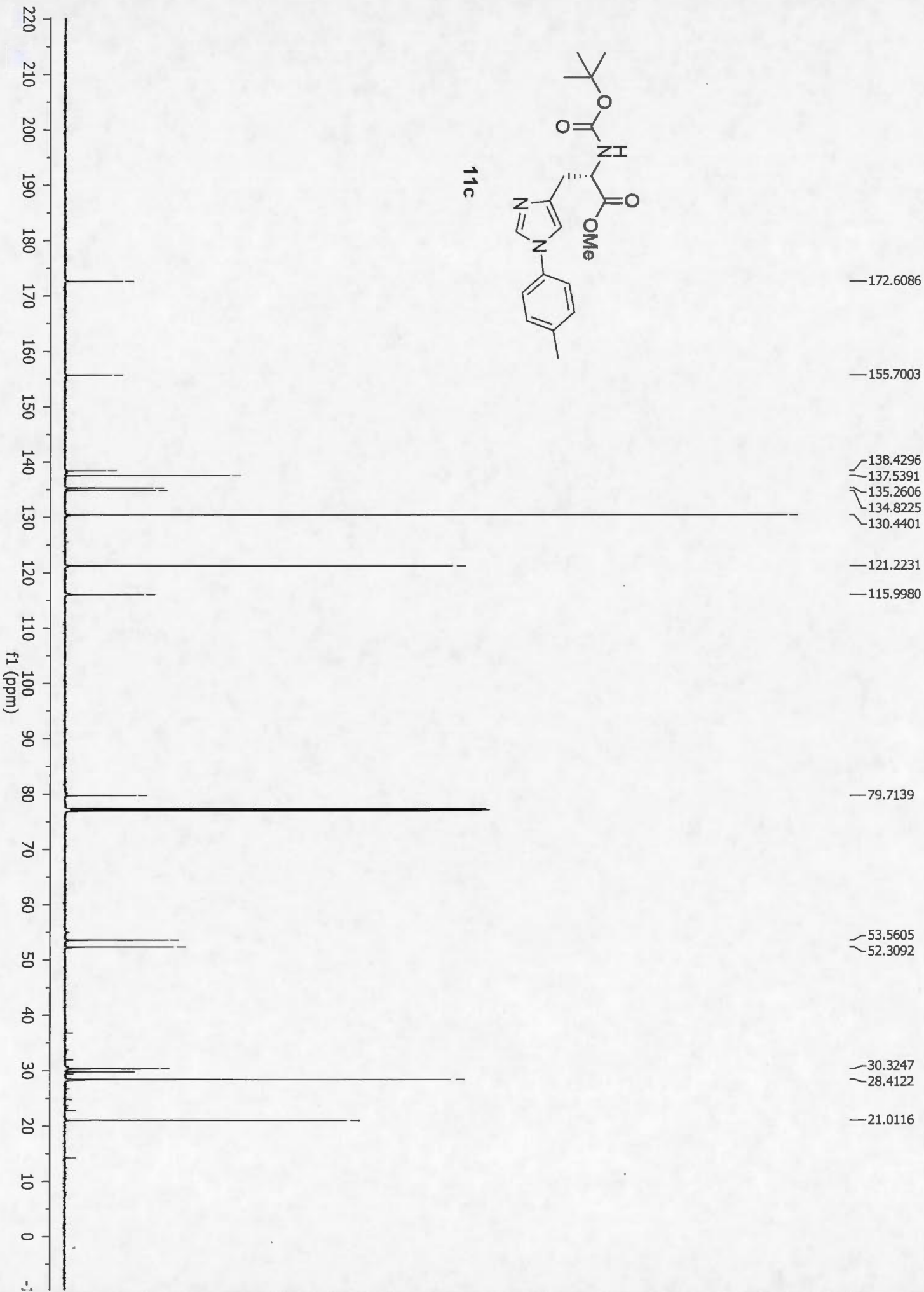


11b

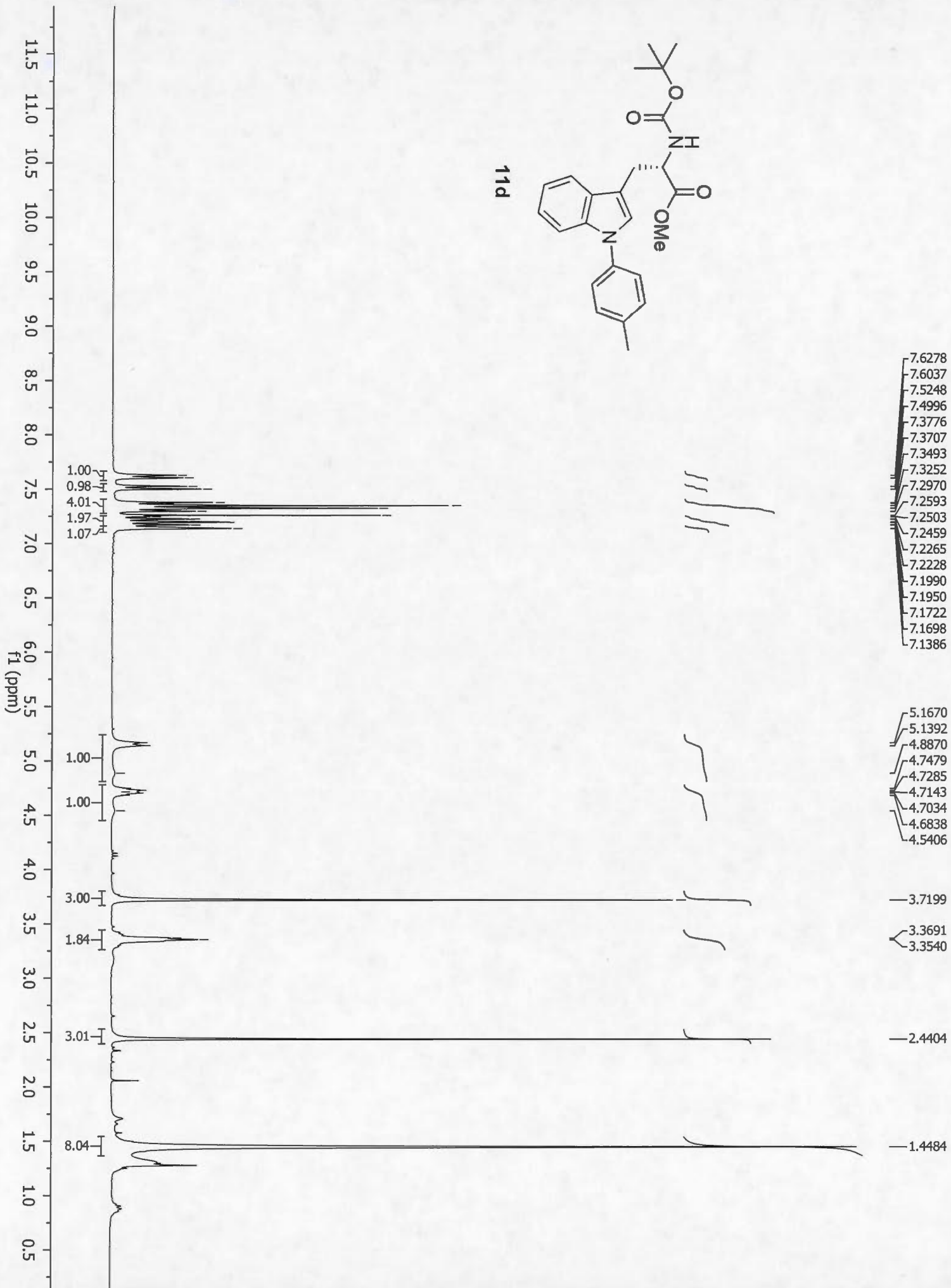


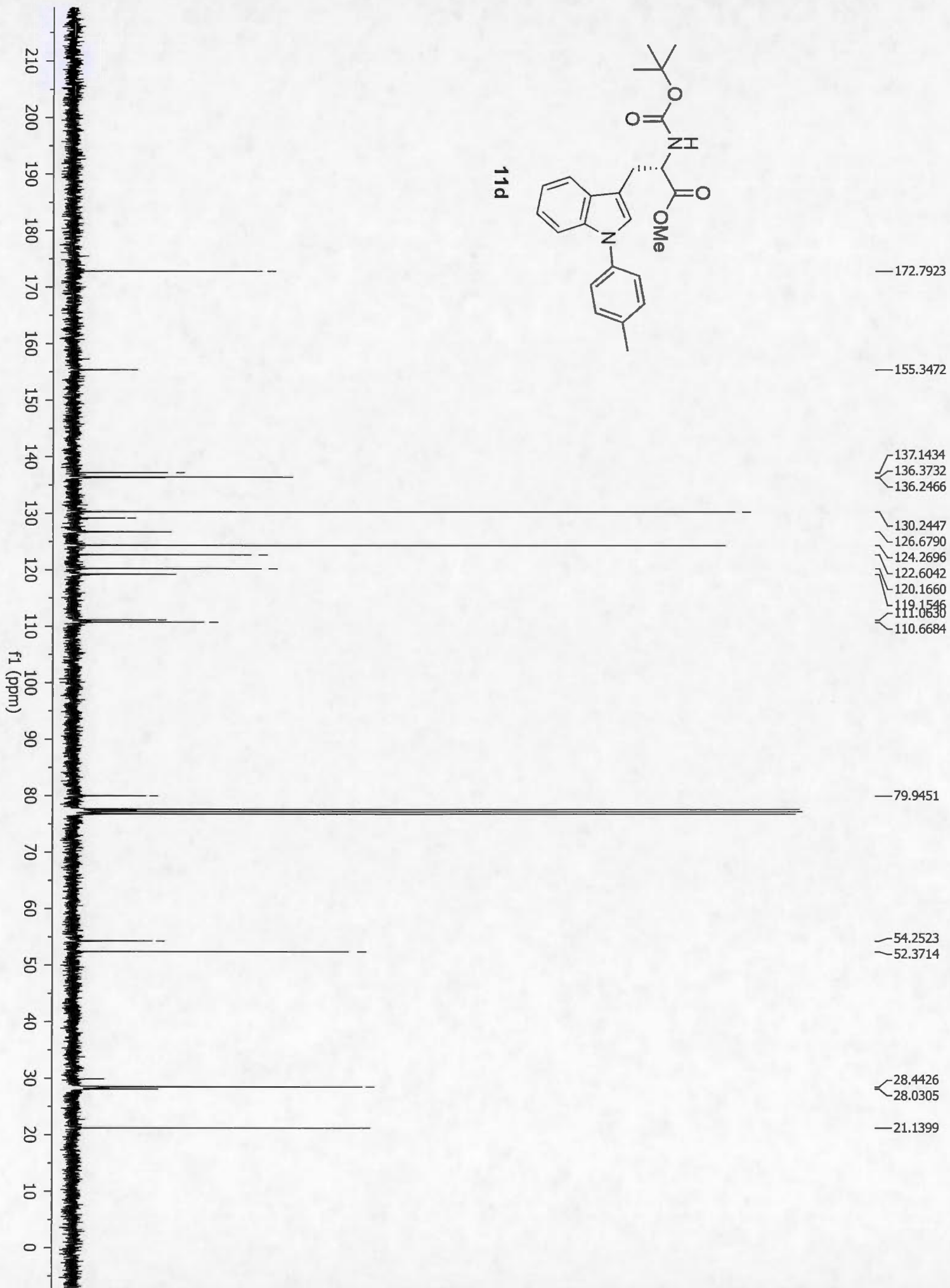
11c



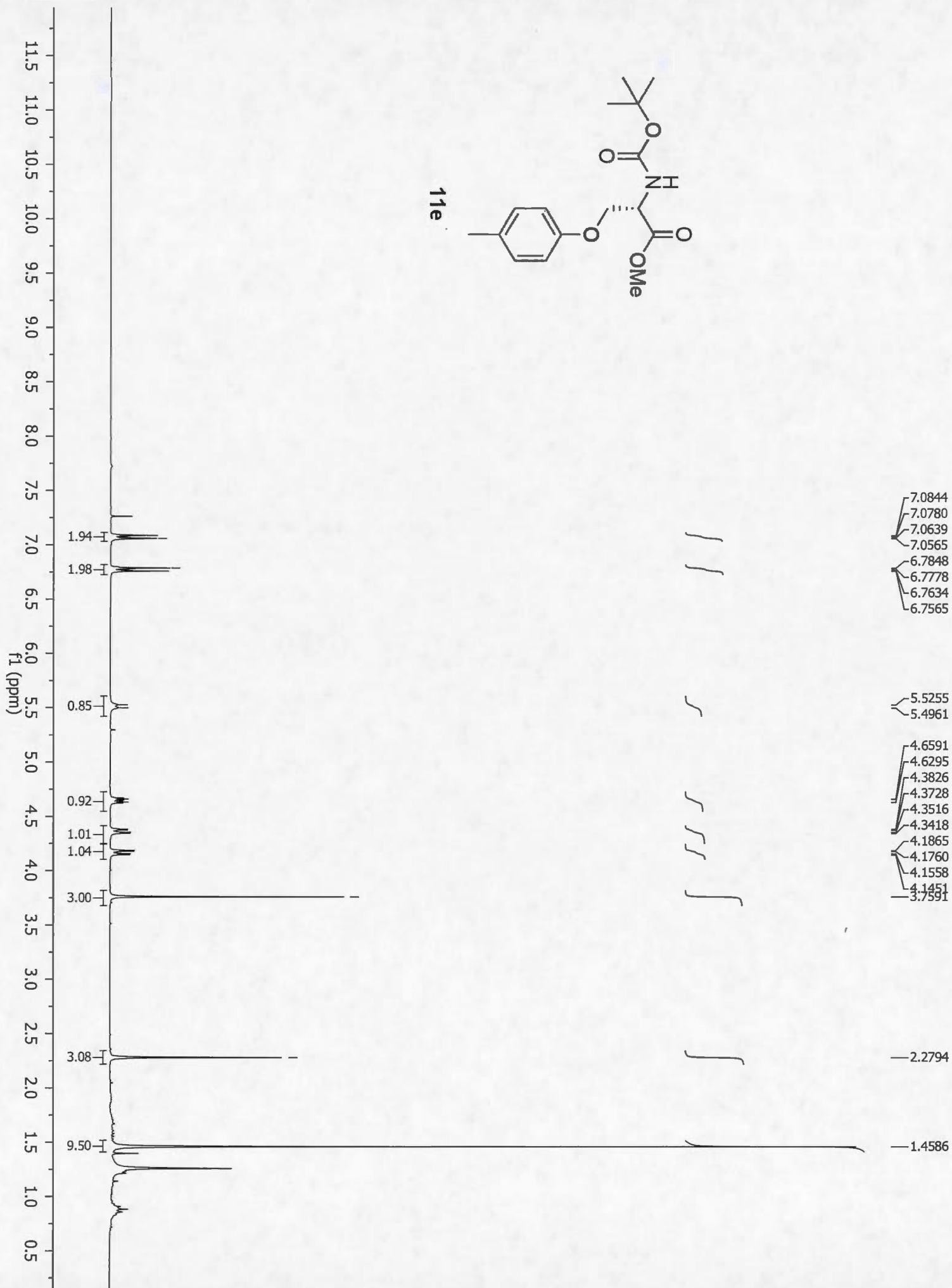
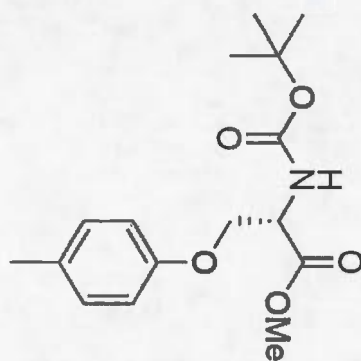




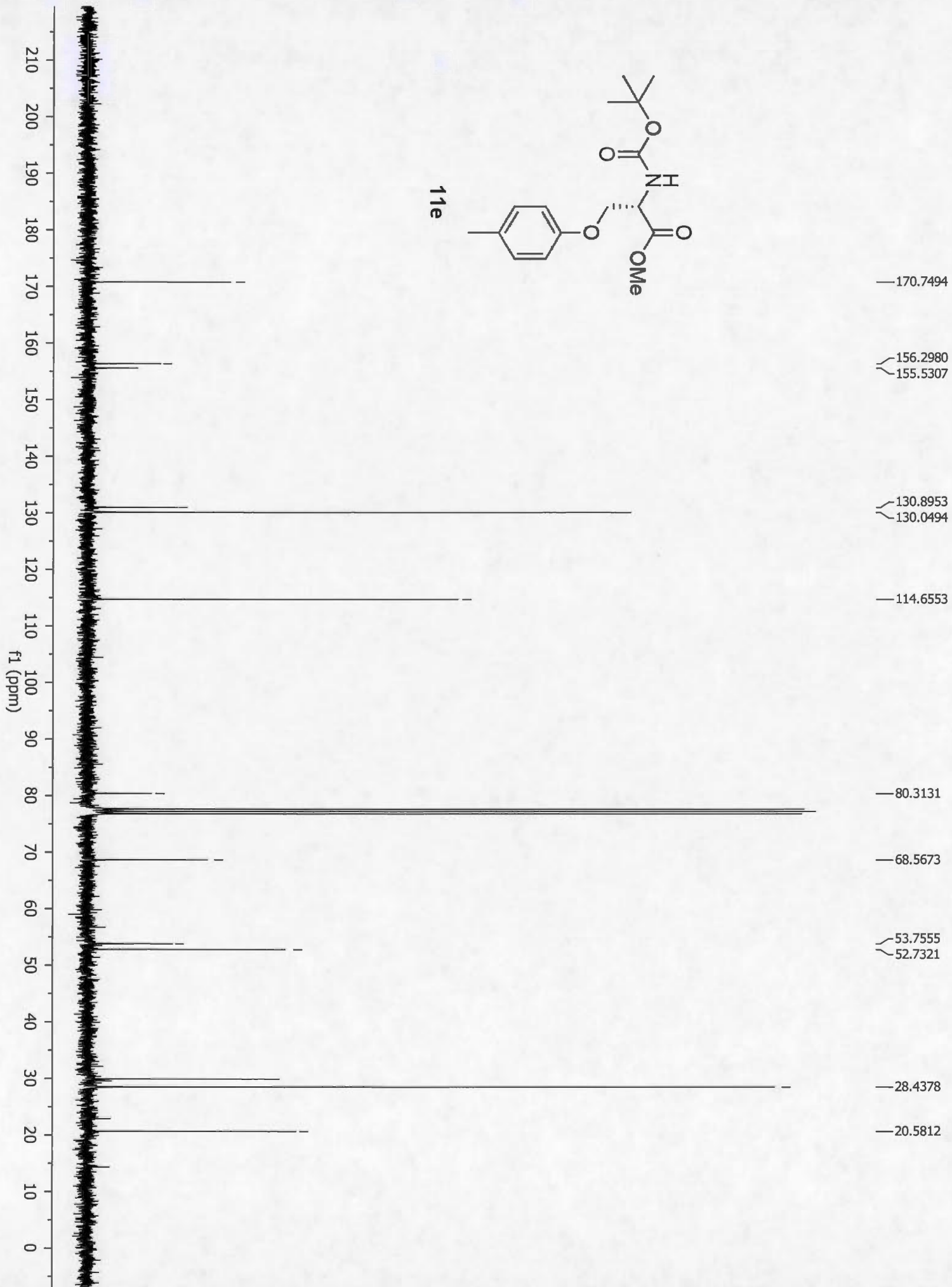
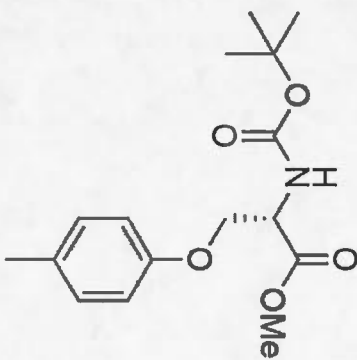




11e

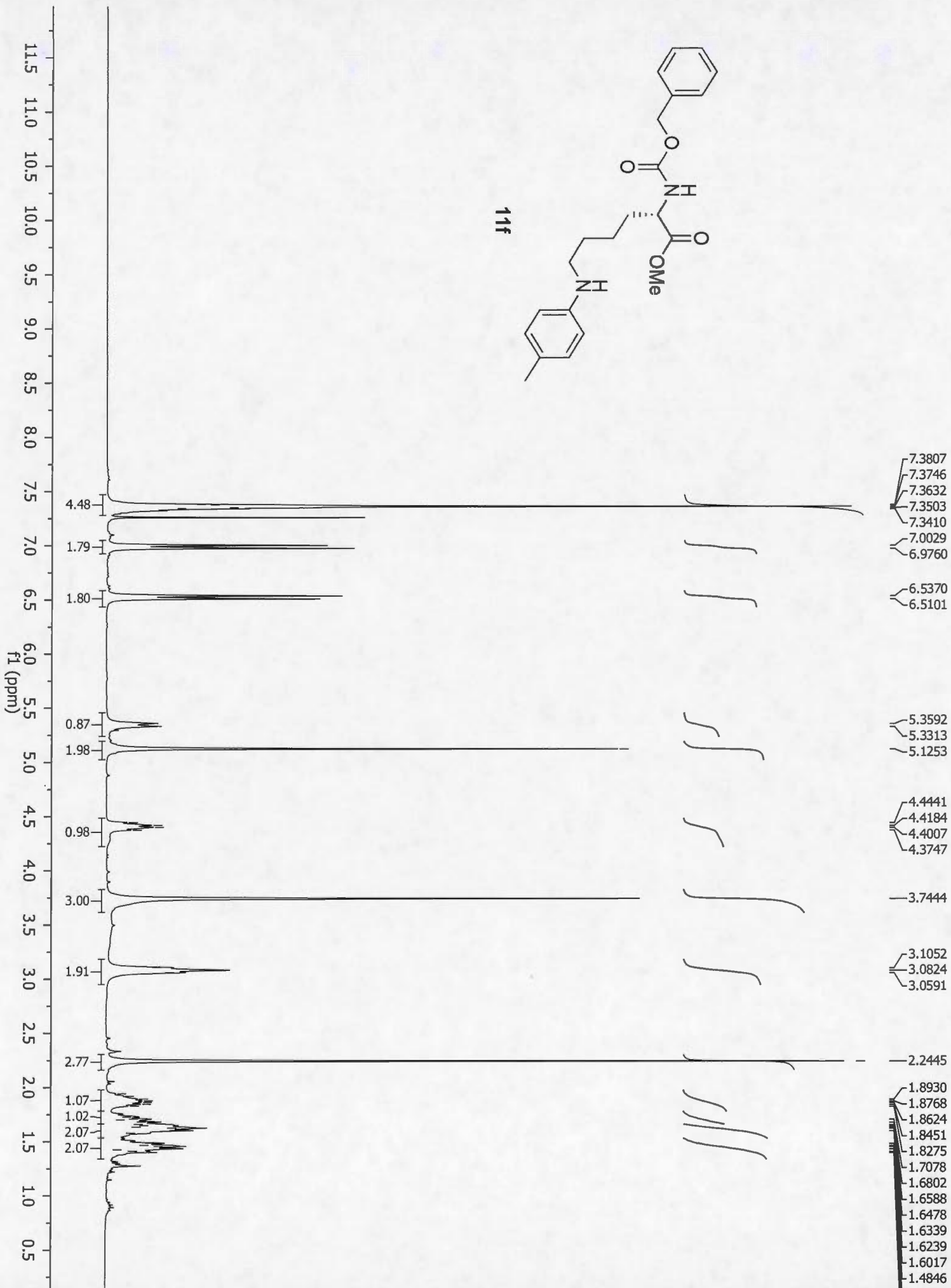
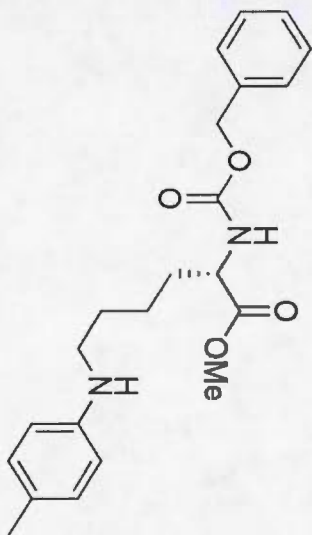


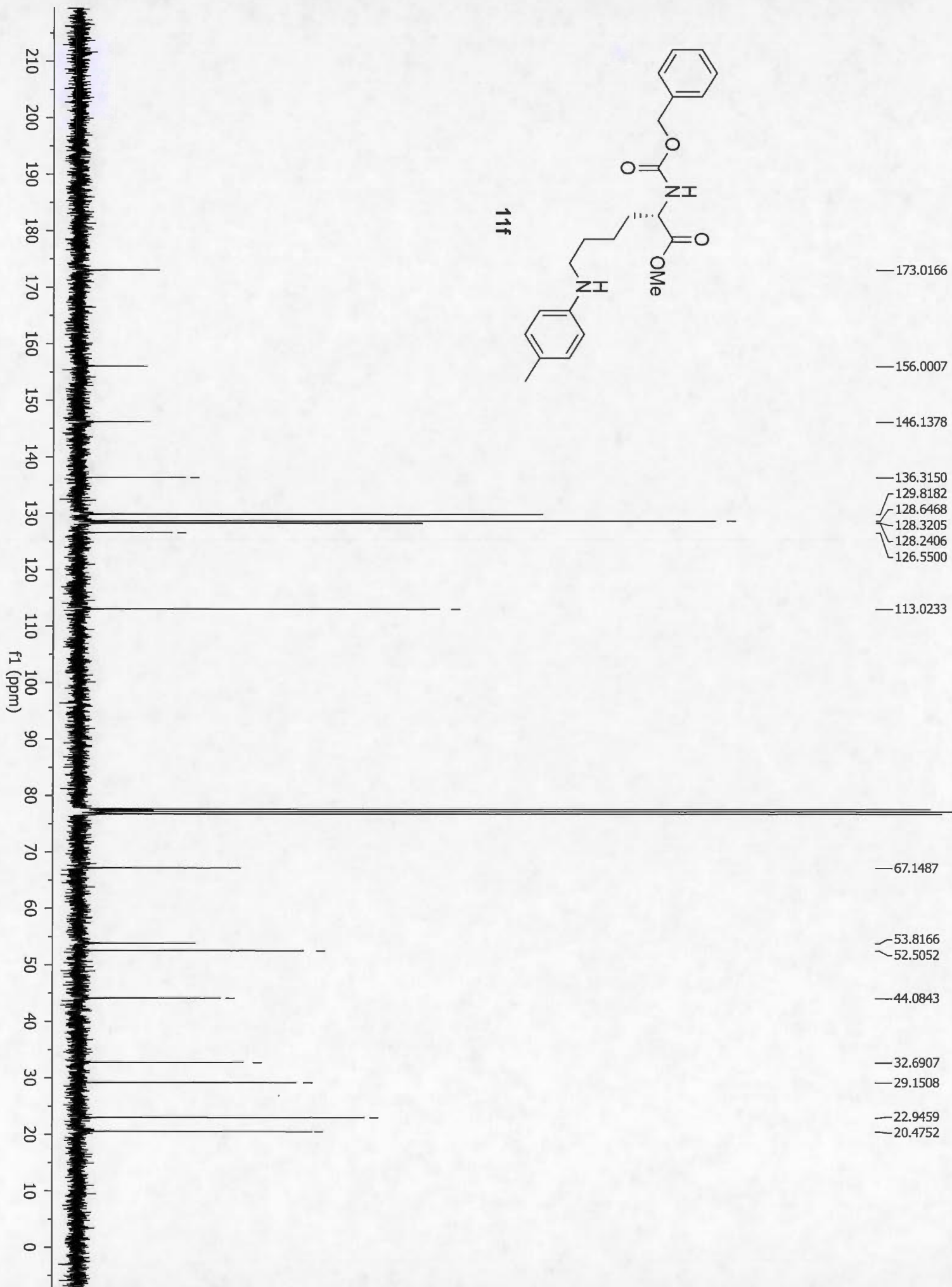
11e

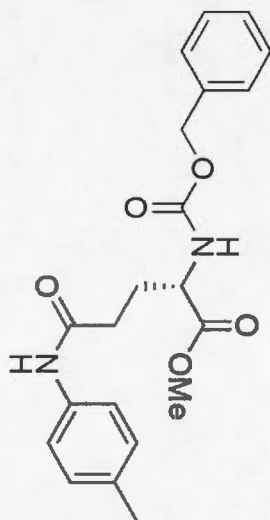




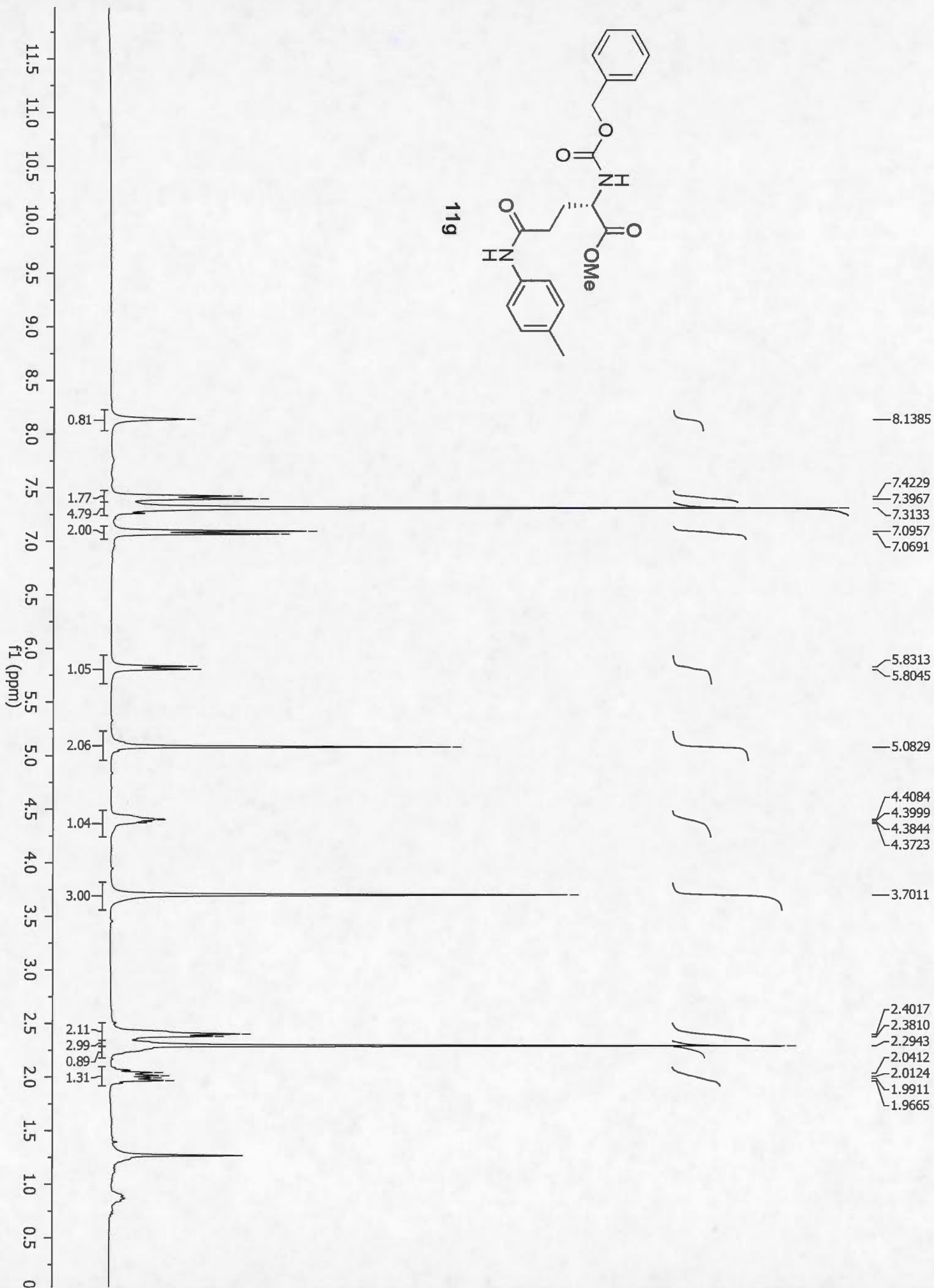
11f

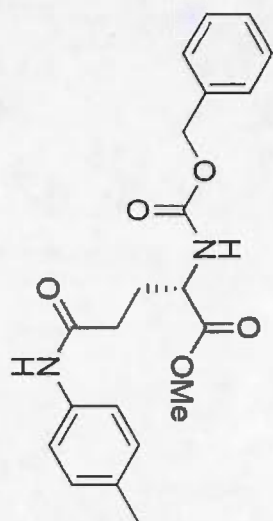




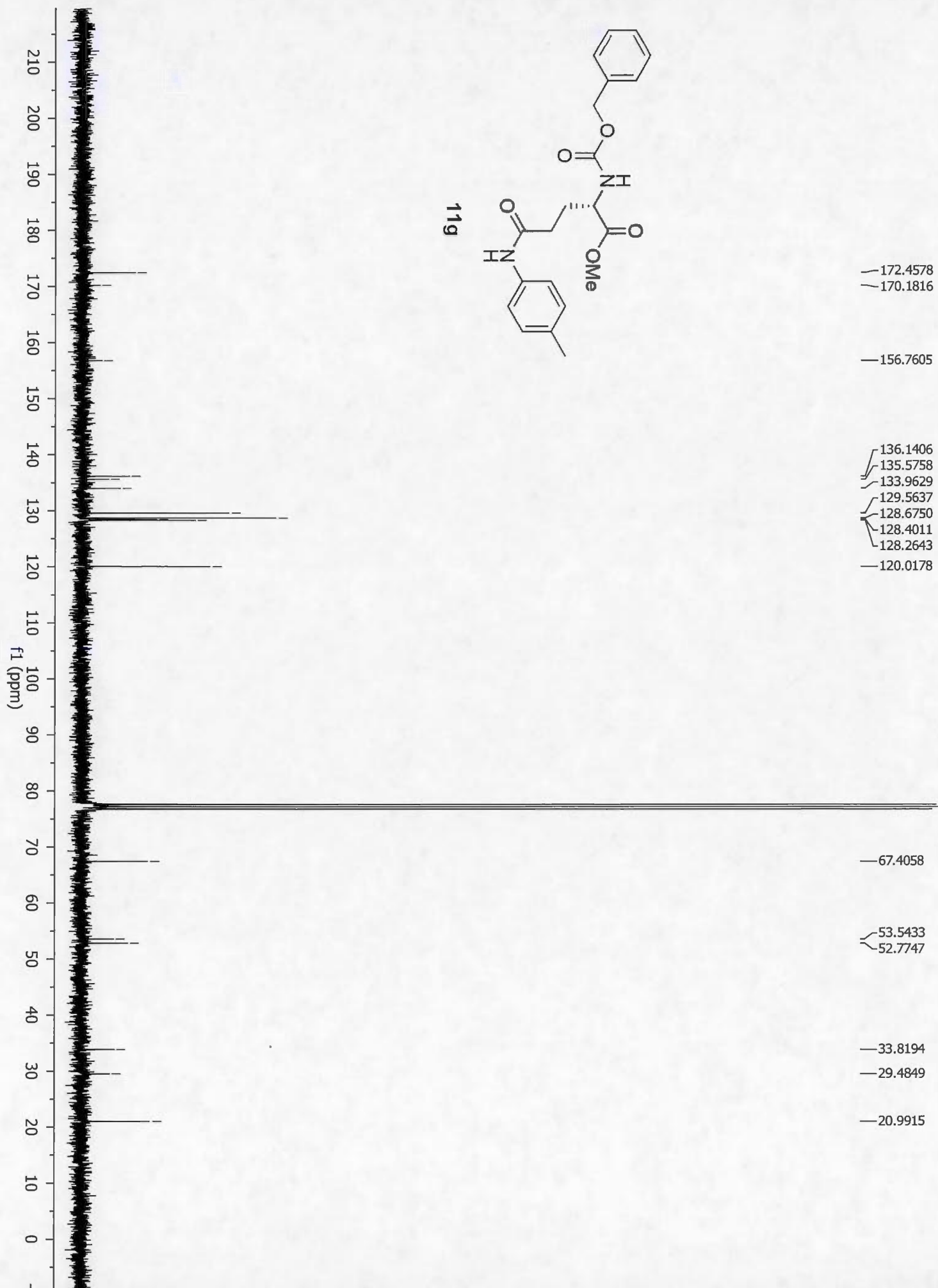


11g

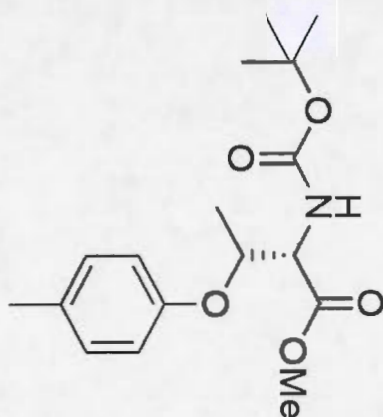




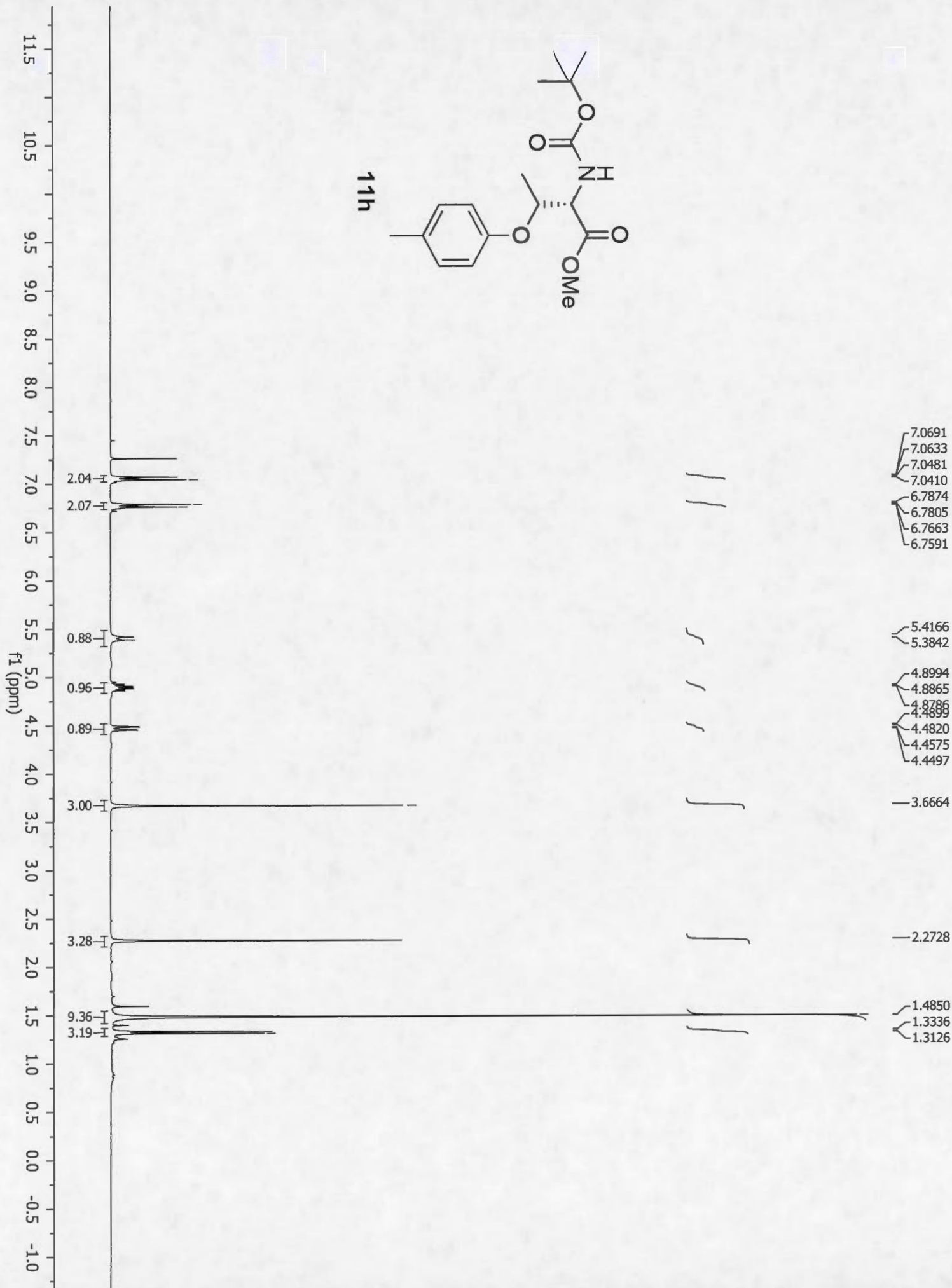
11g

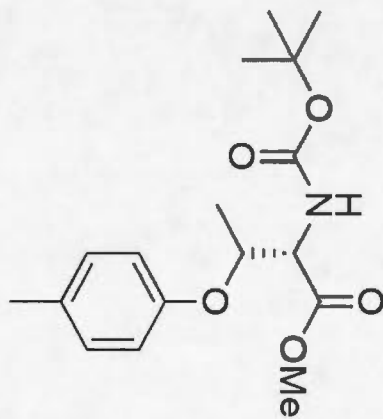




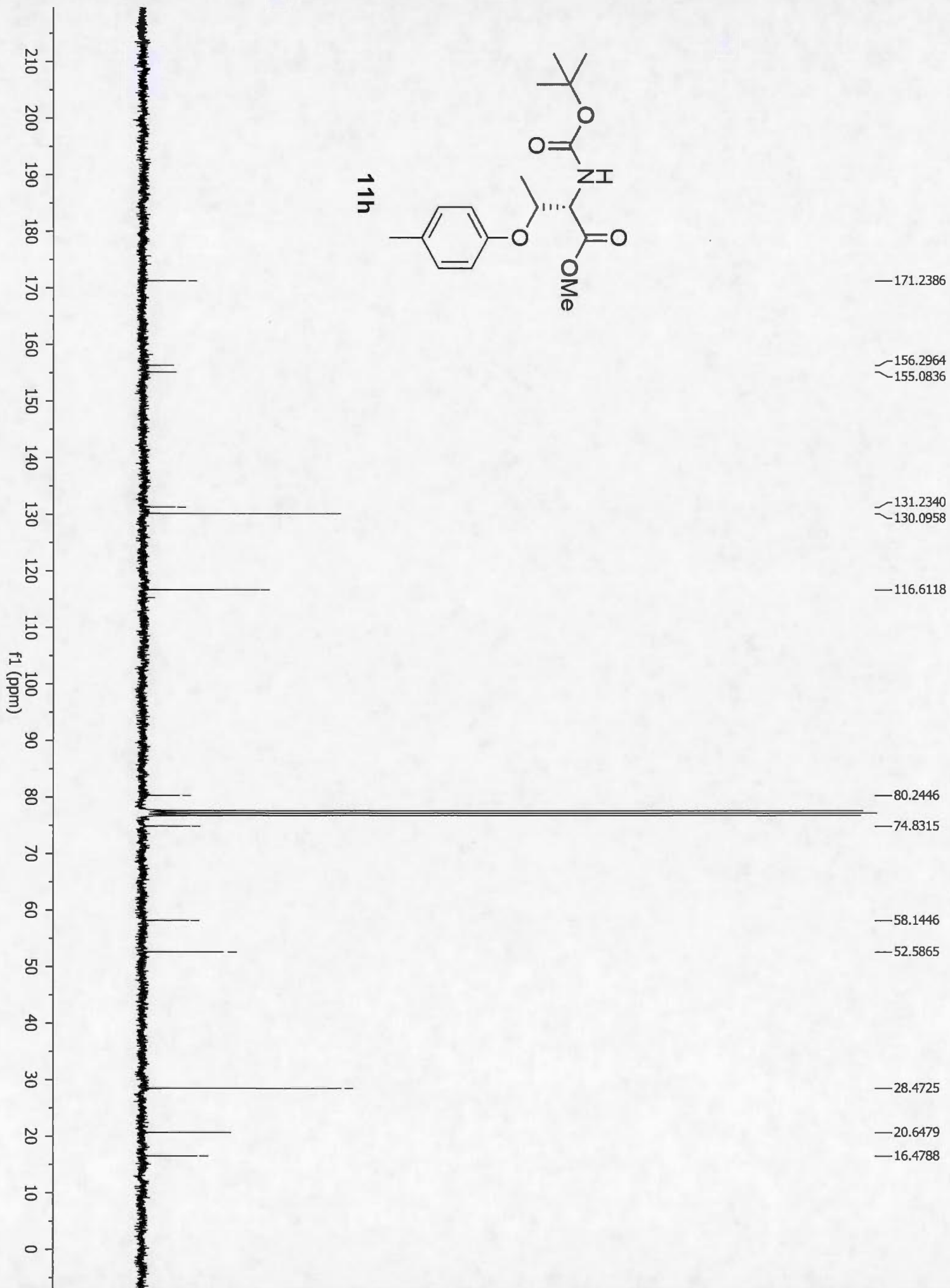


11h

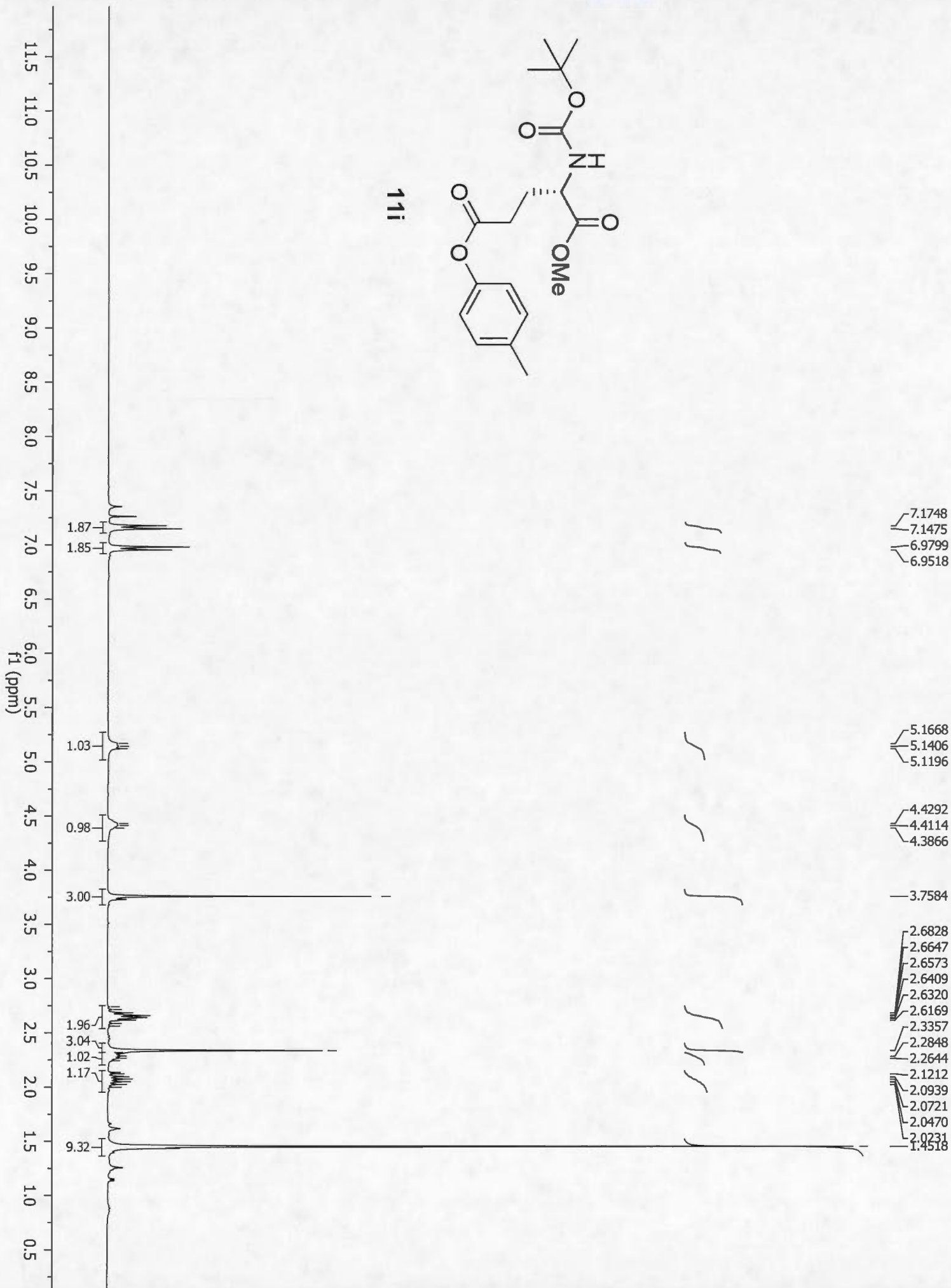
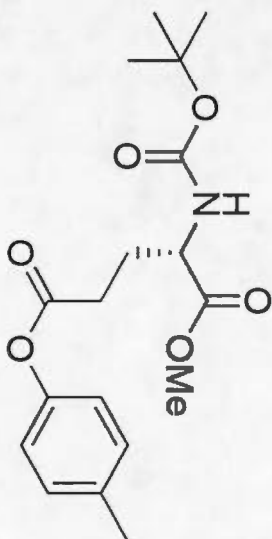


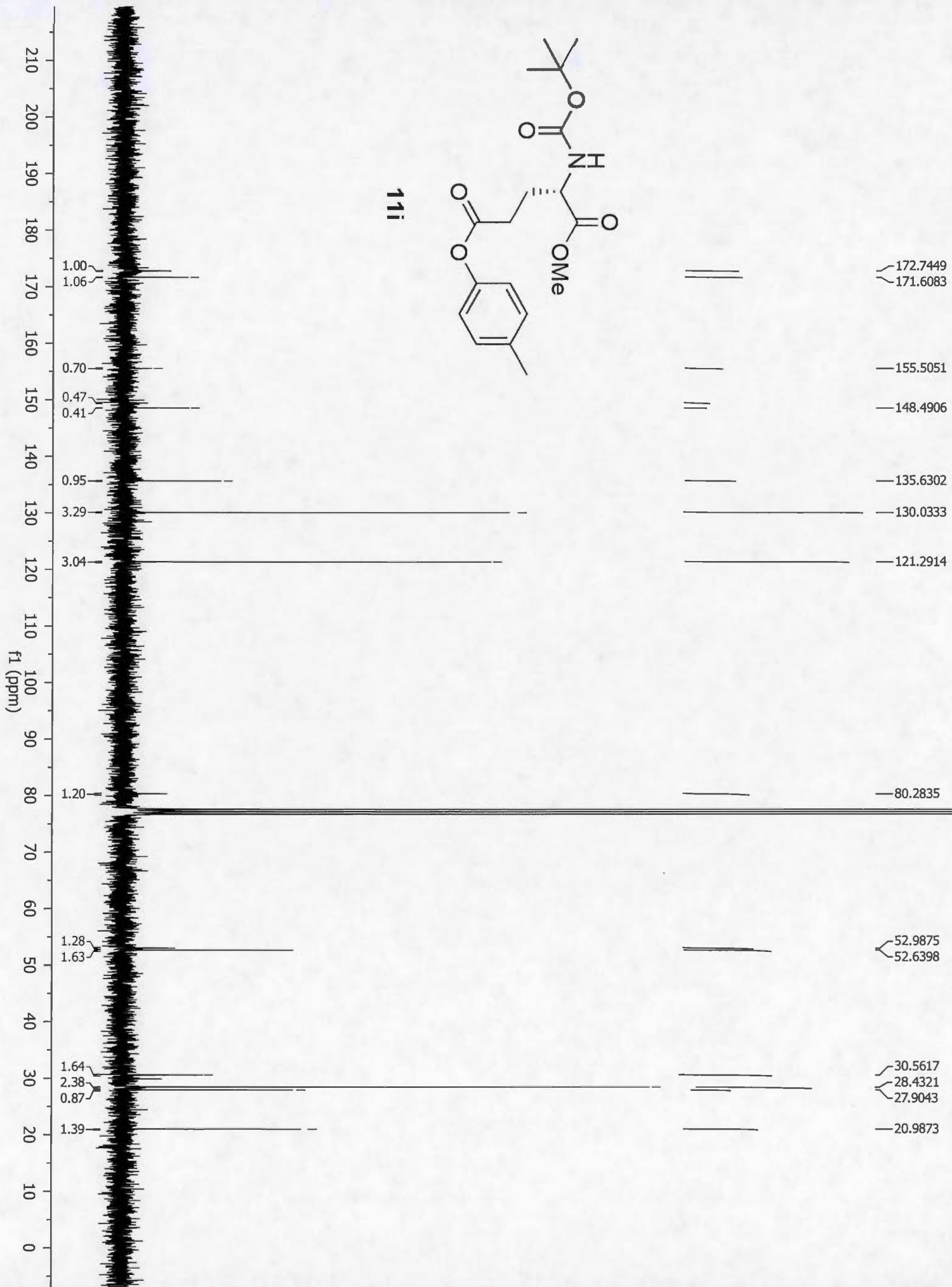


11h

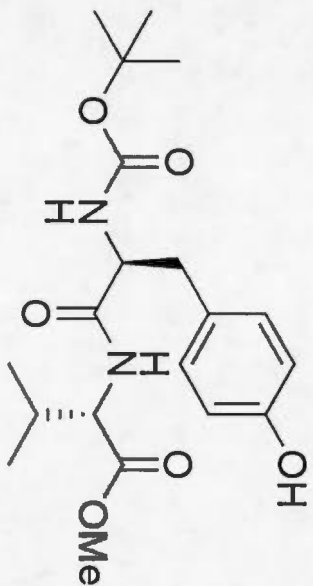


11i

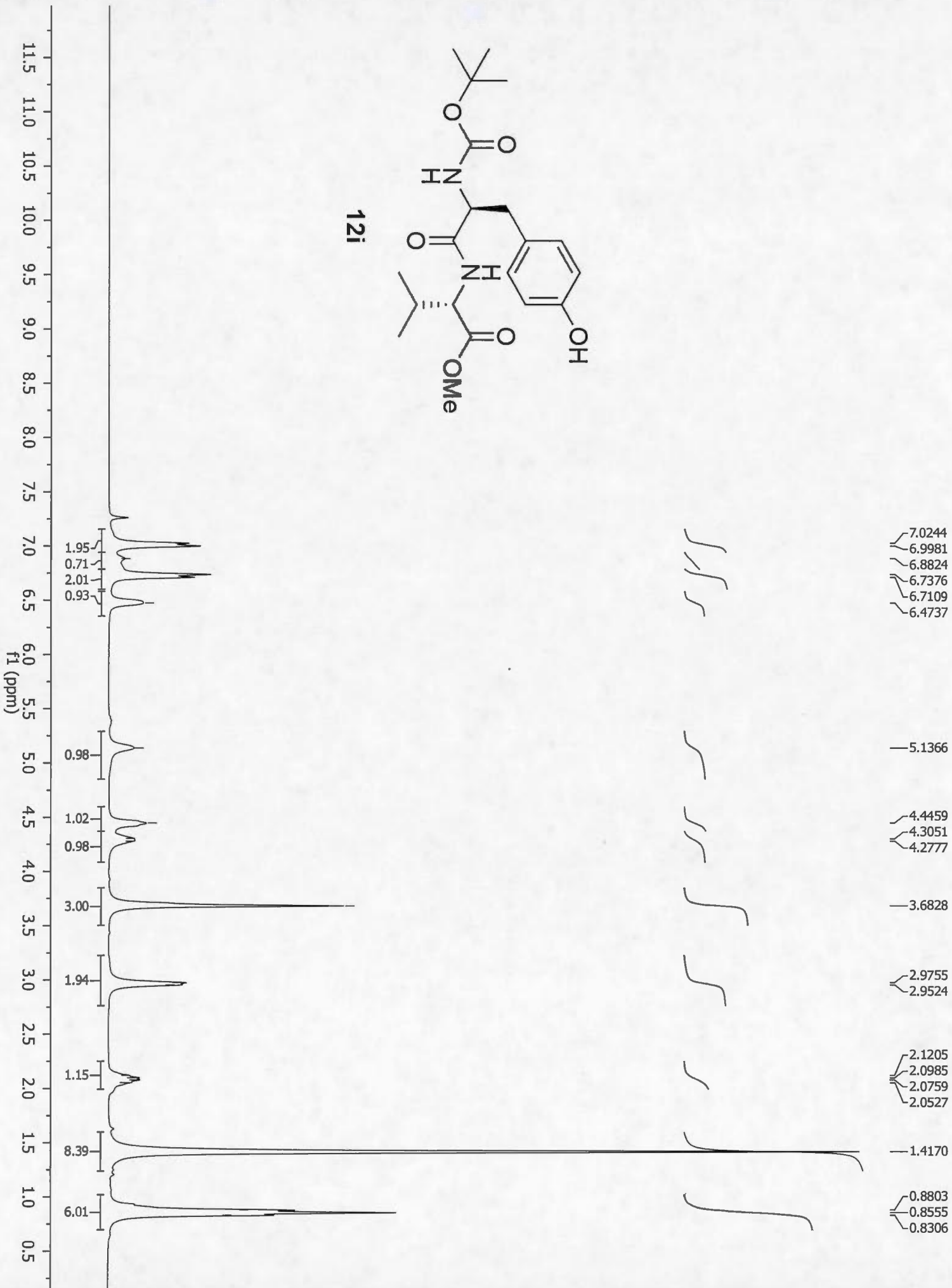


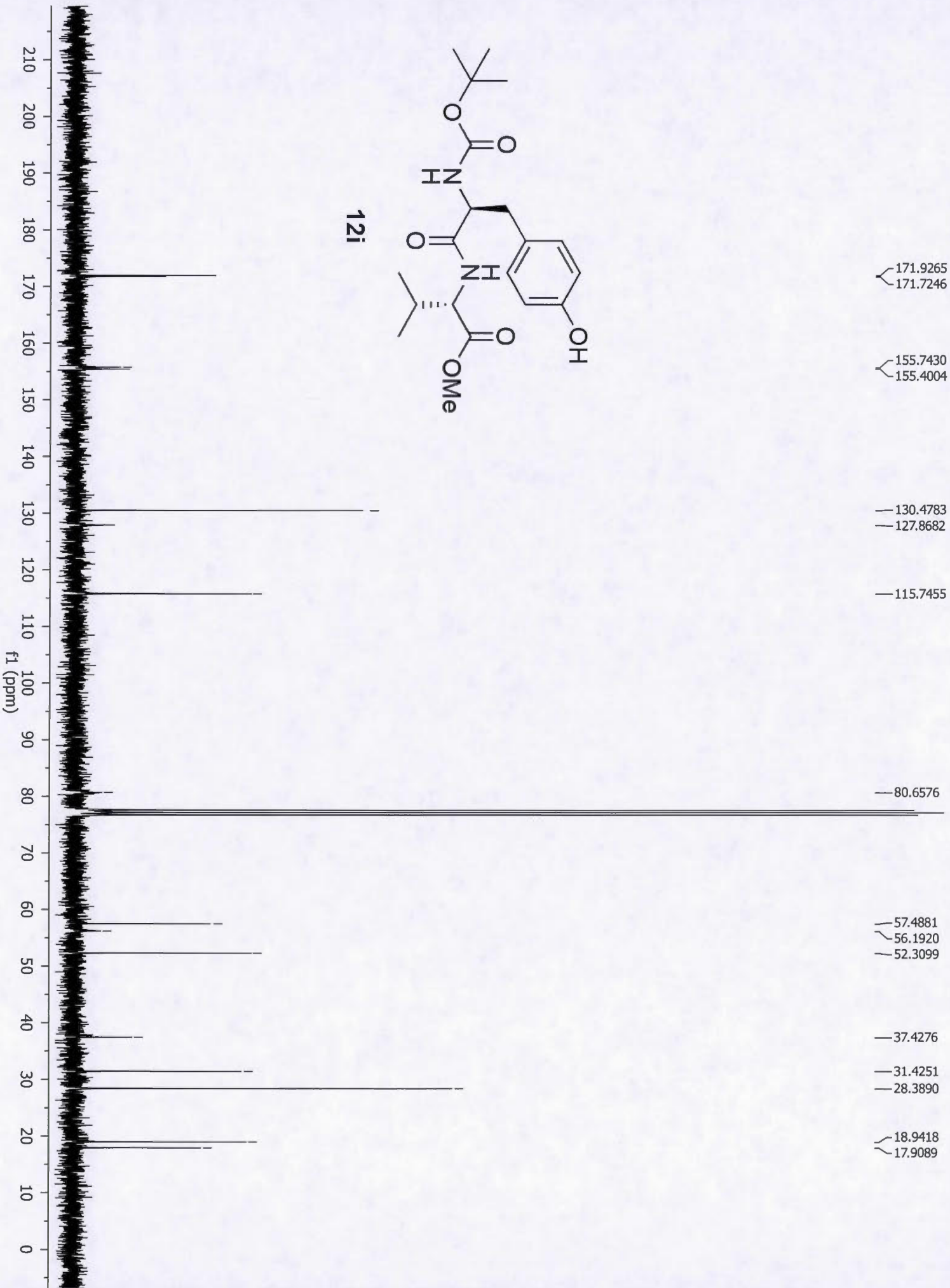


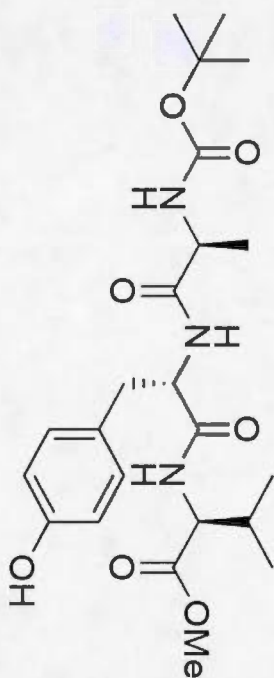




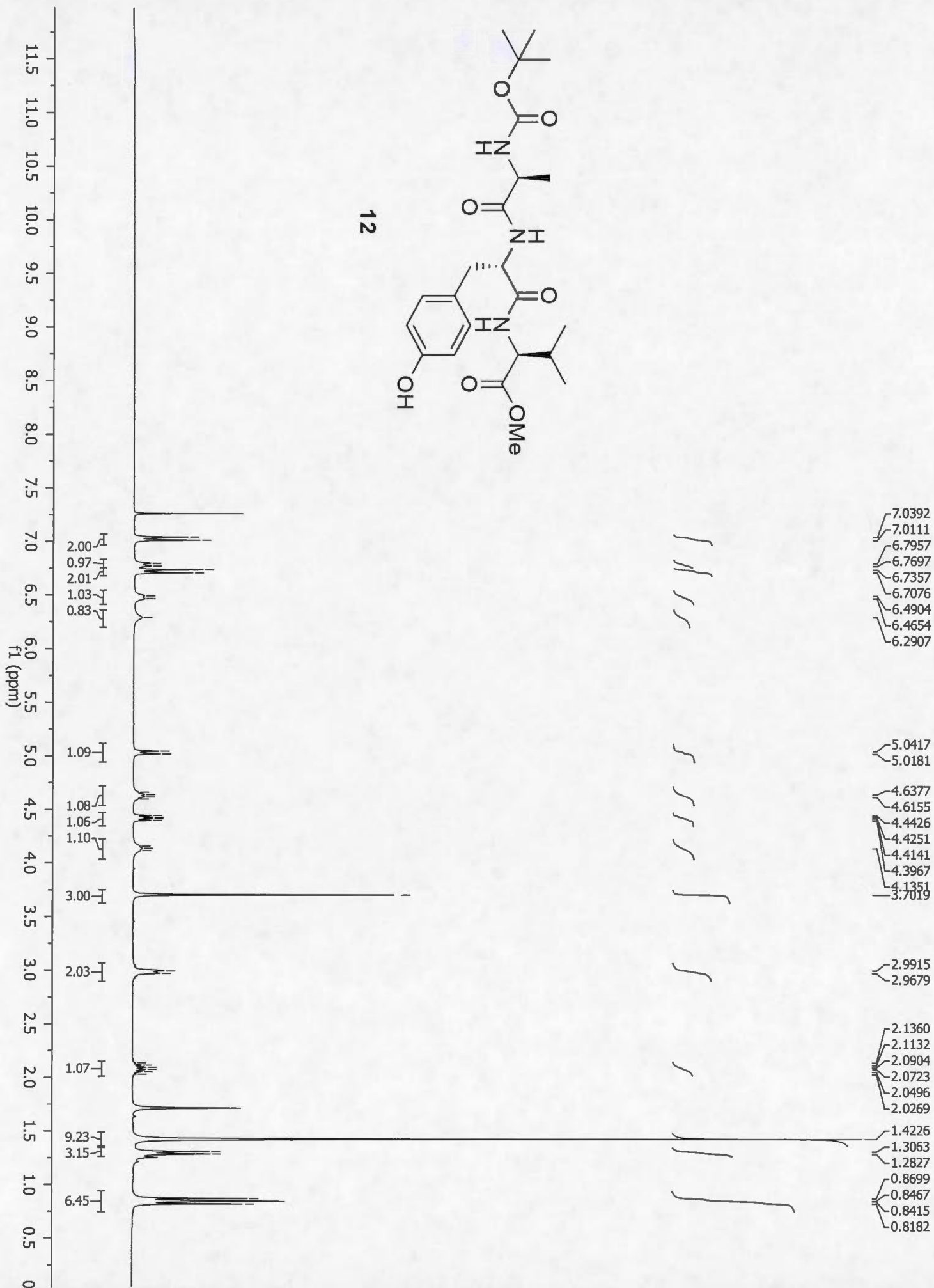
12i

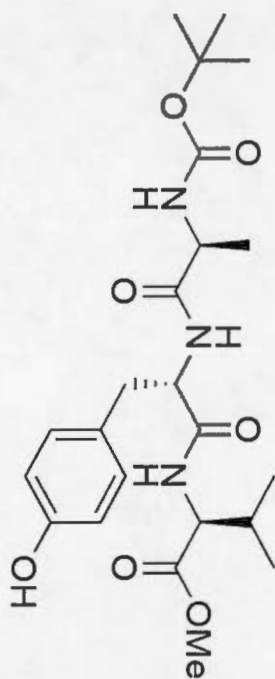




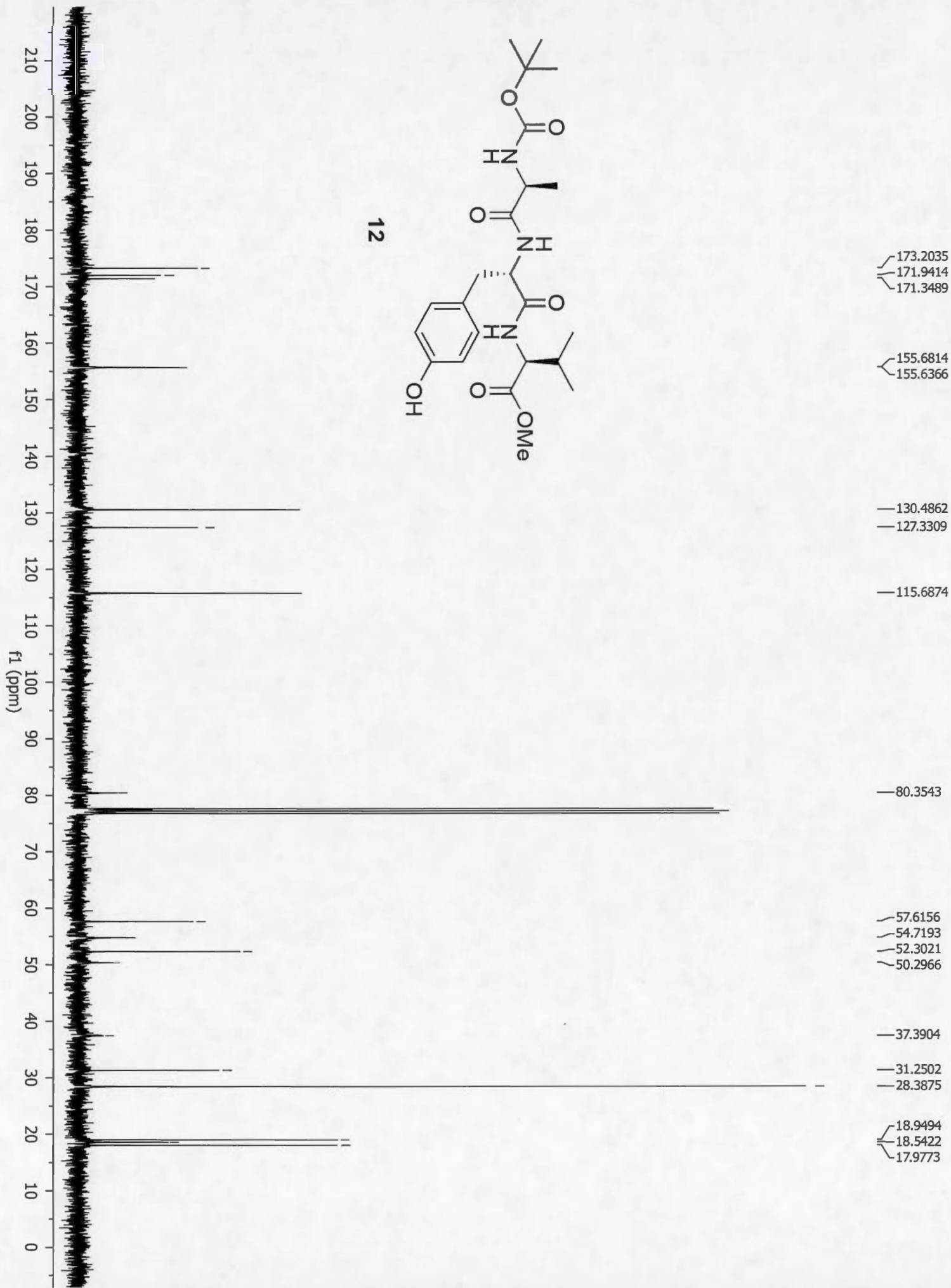


12

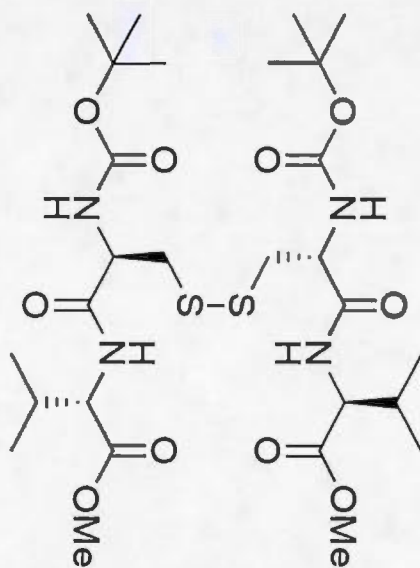




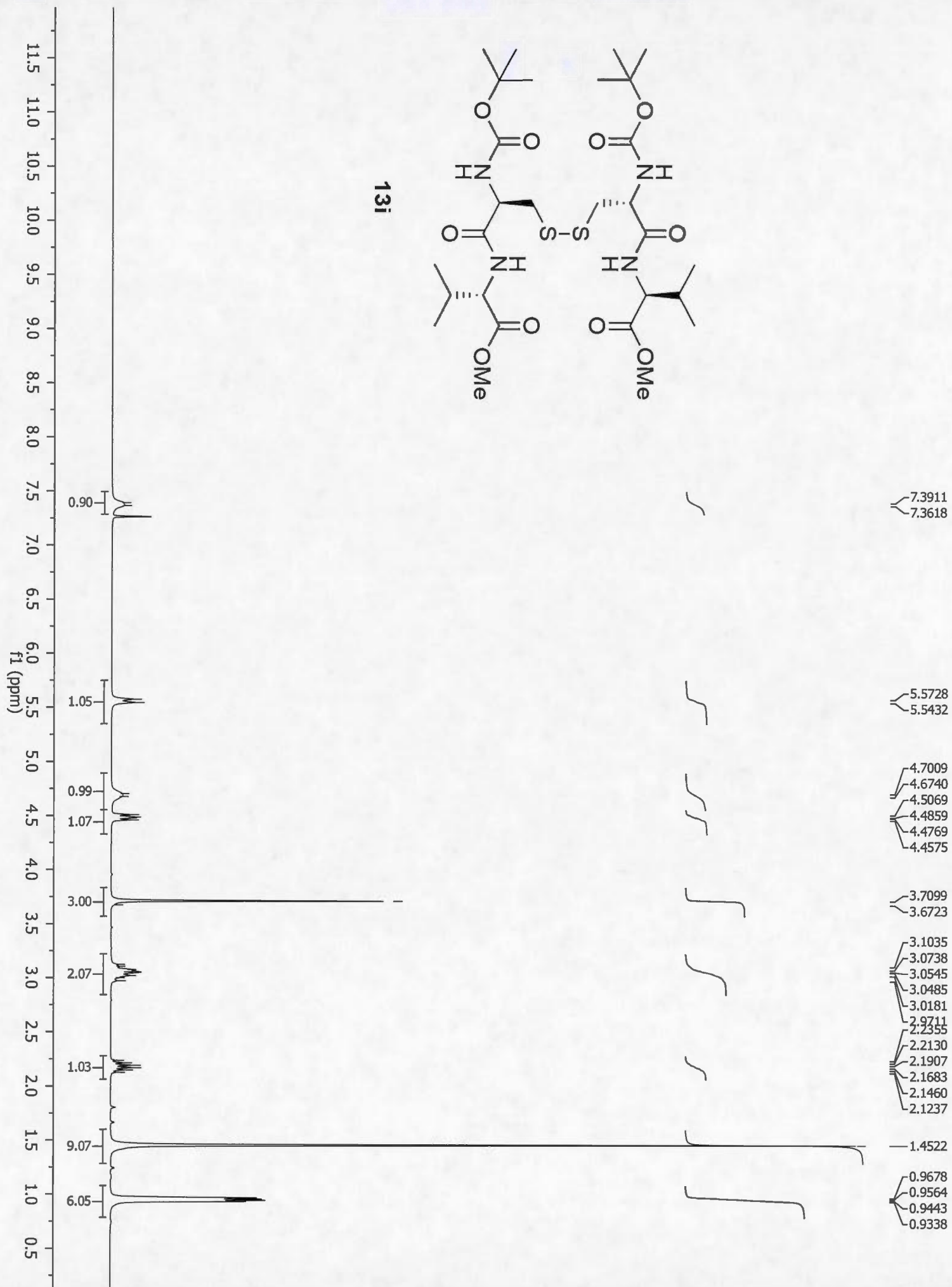
12

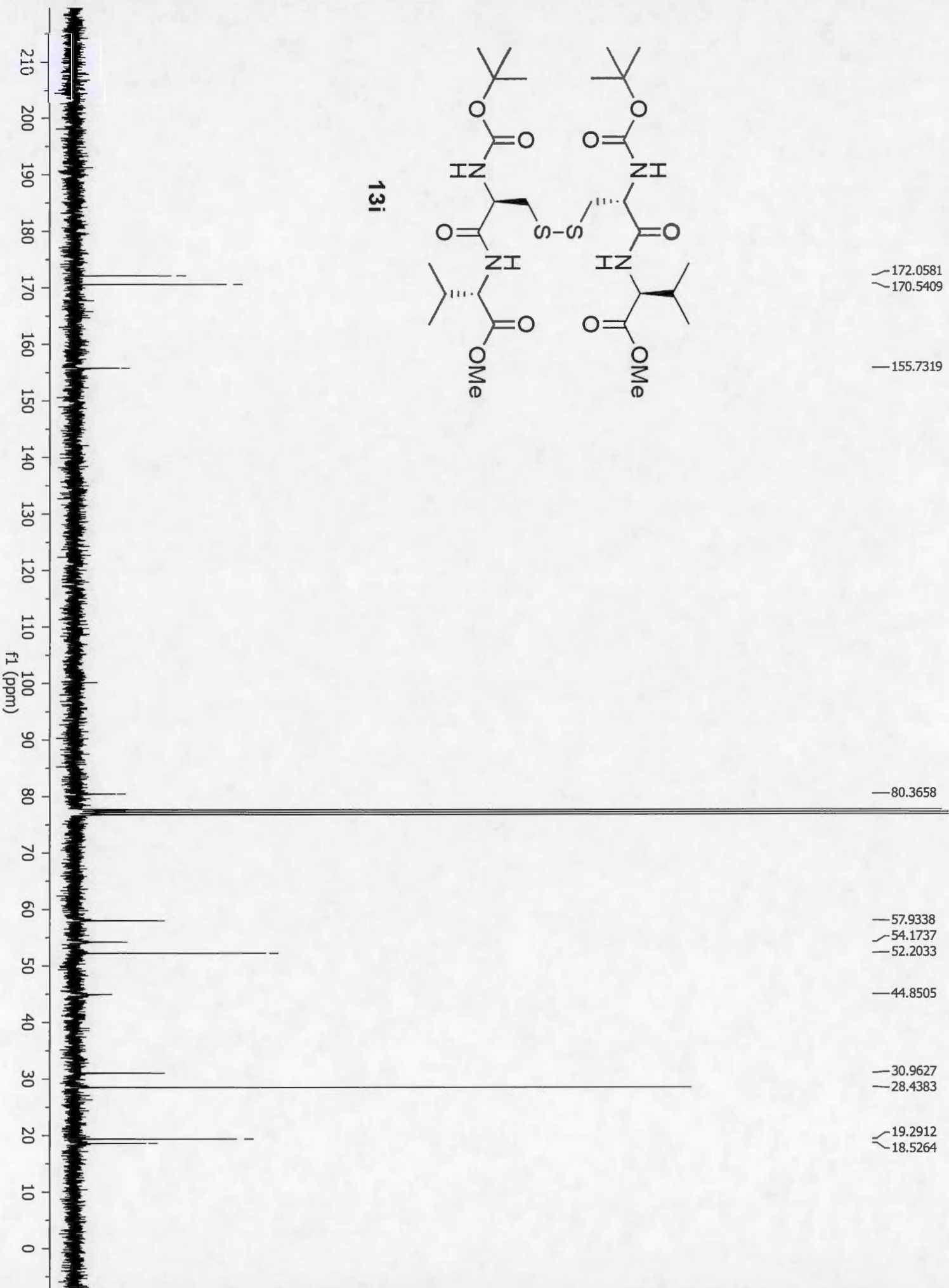


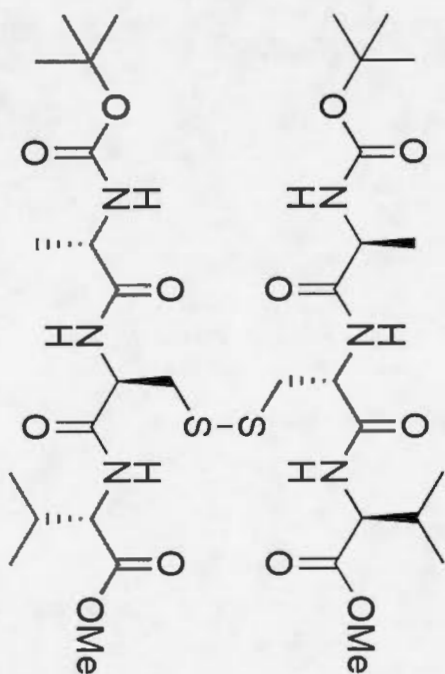




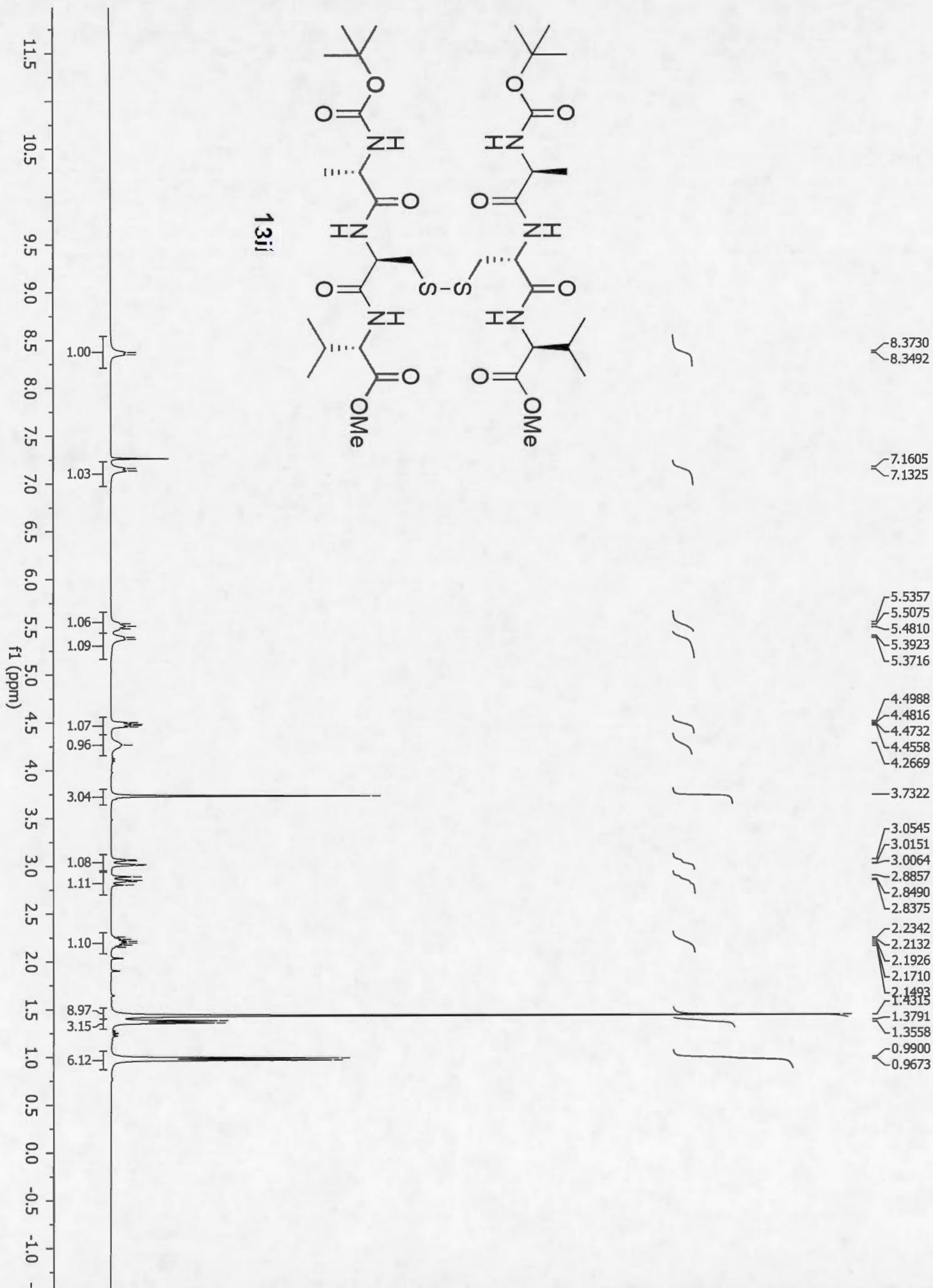
131

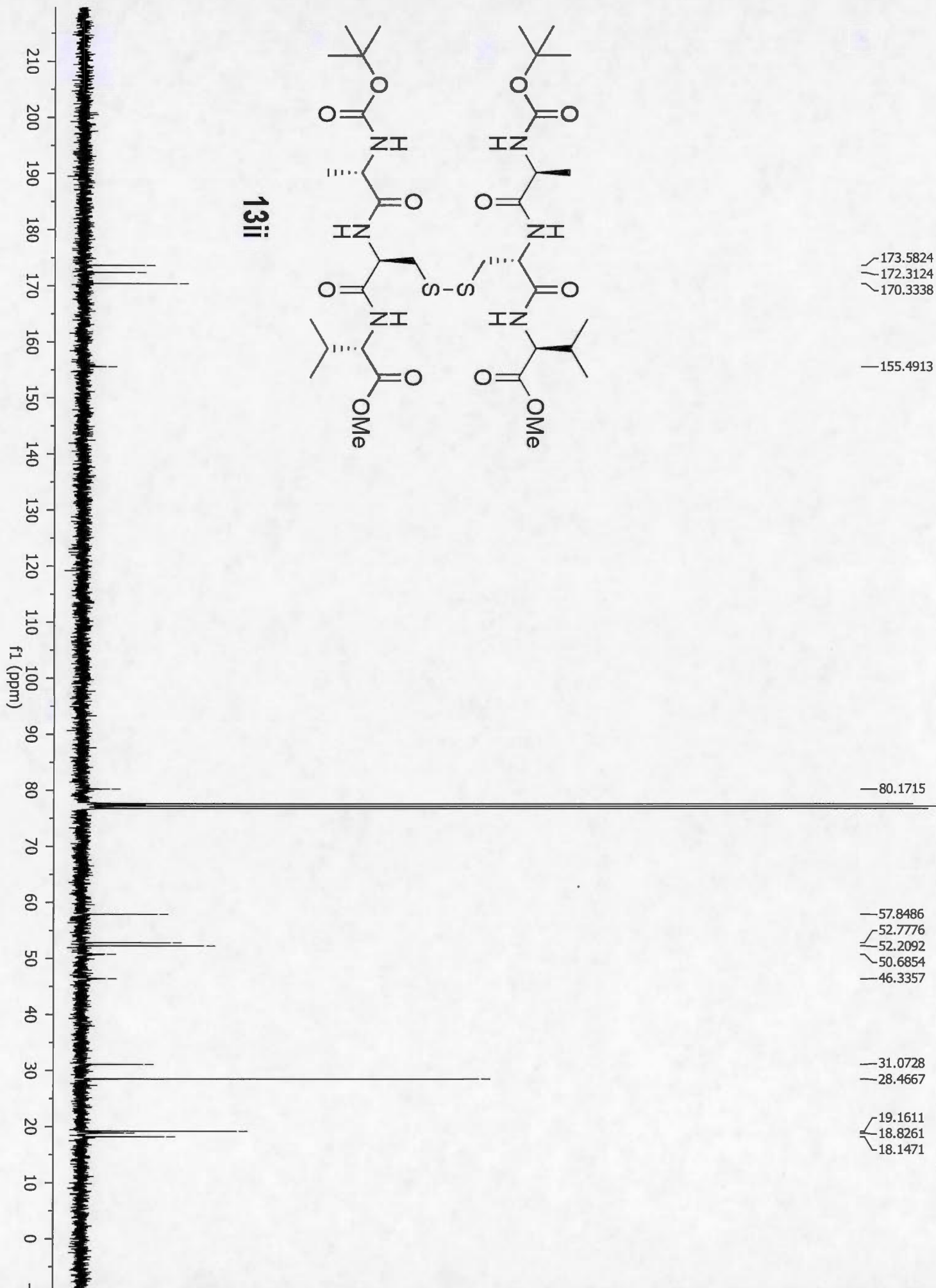






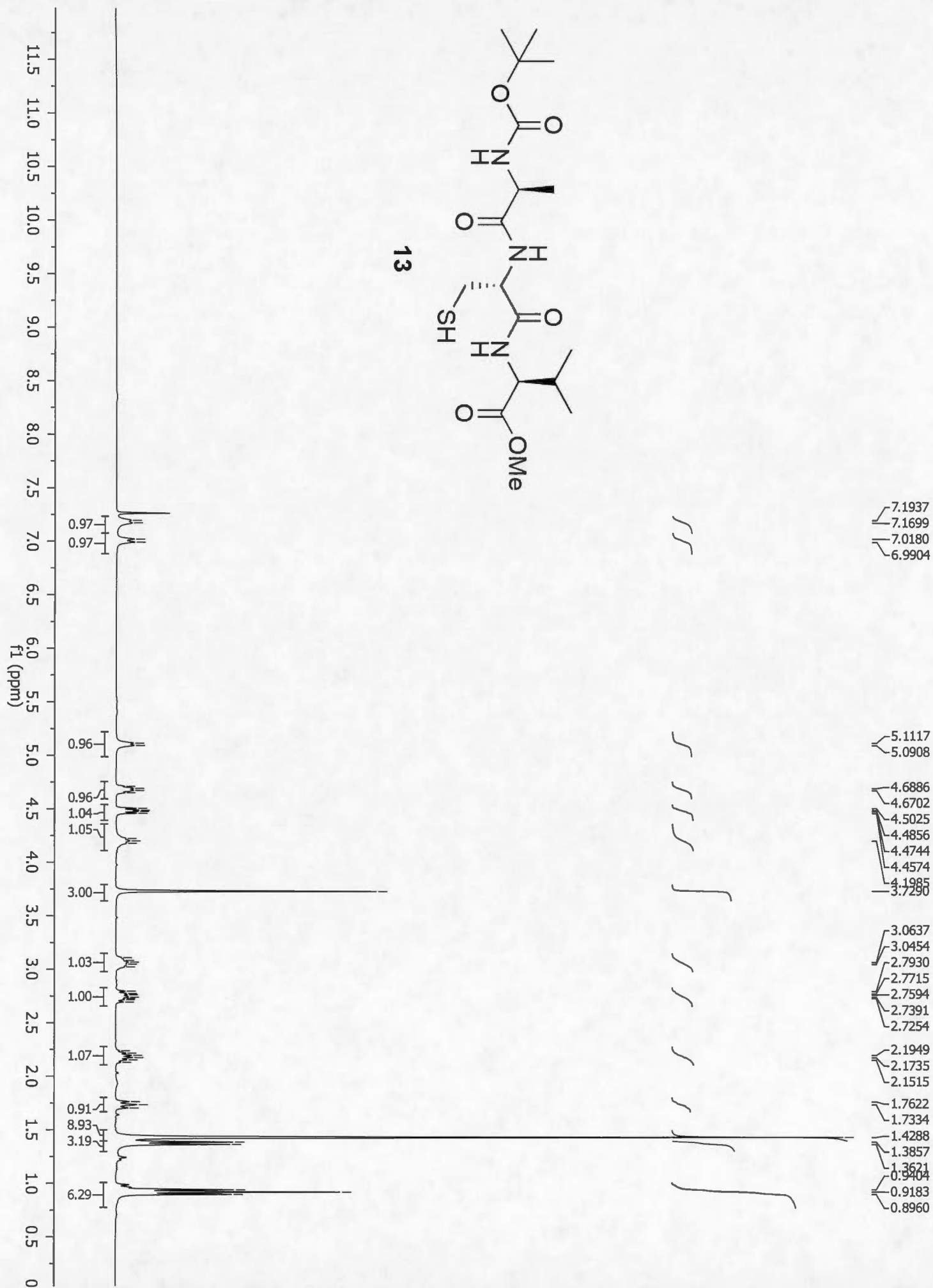
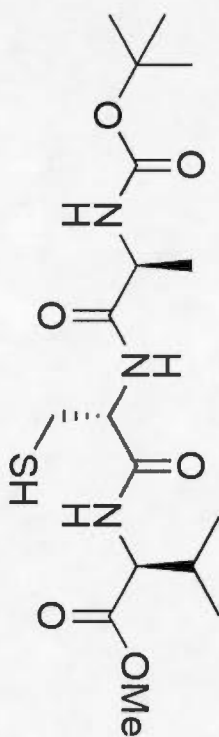
13H



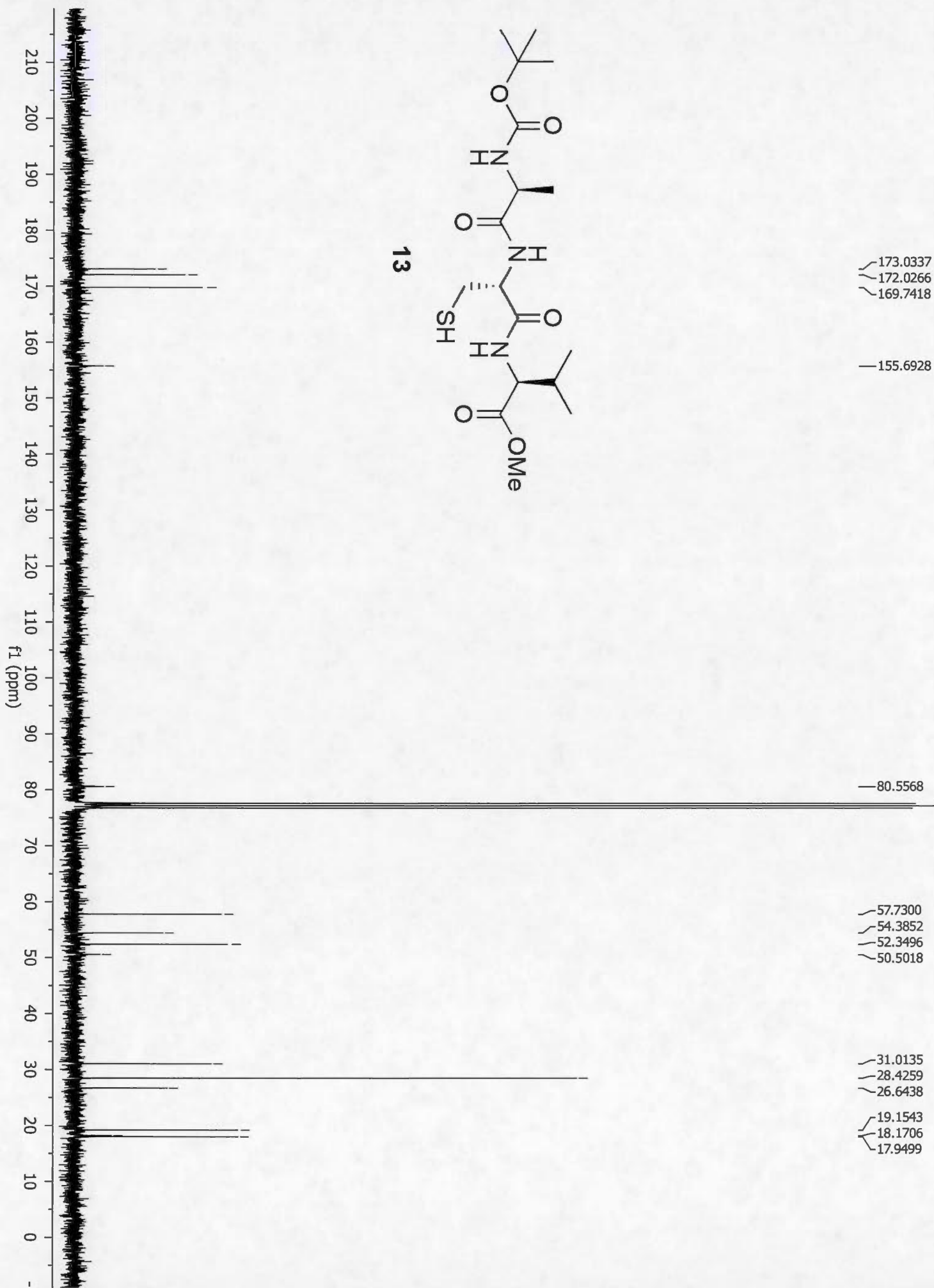
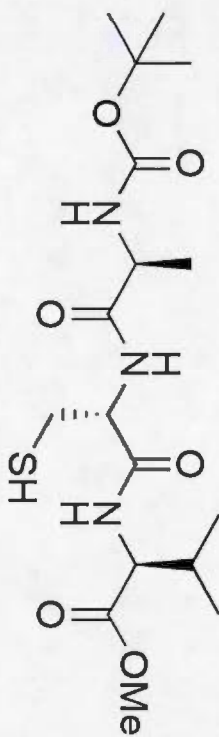


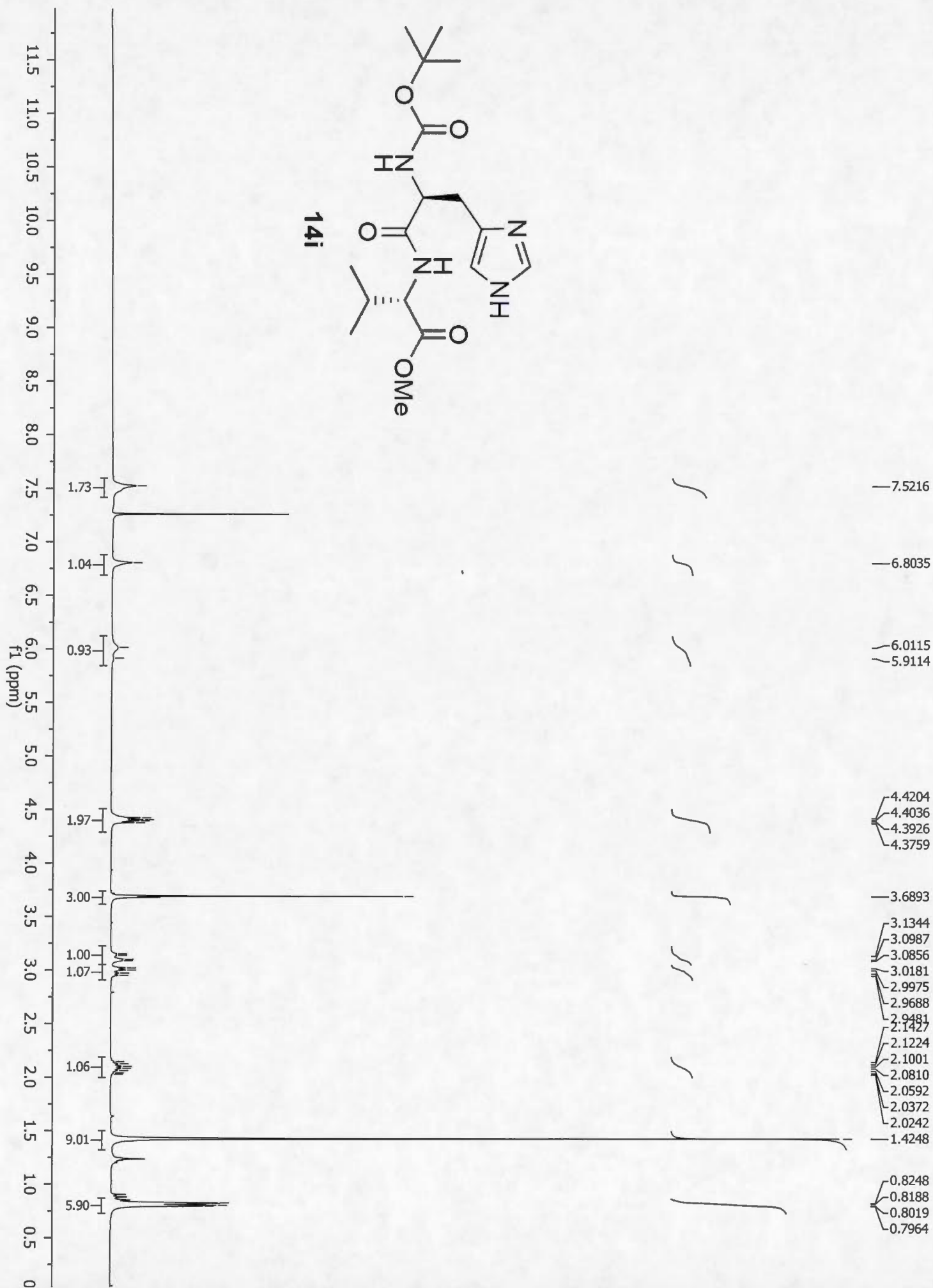
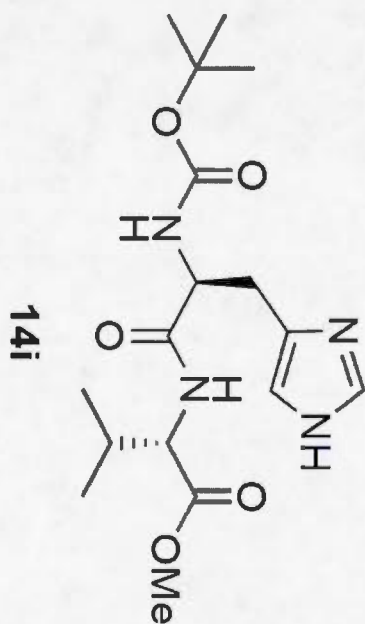


13

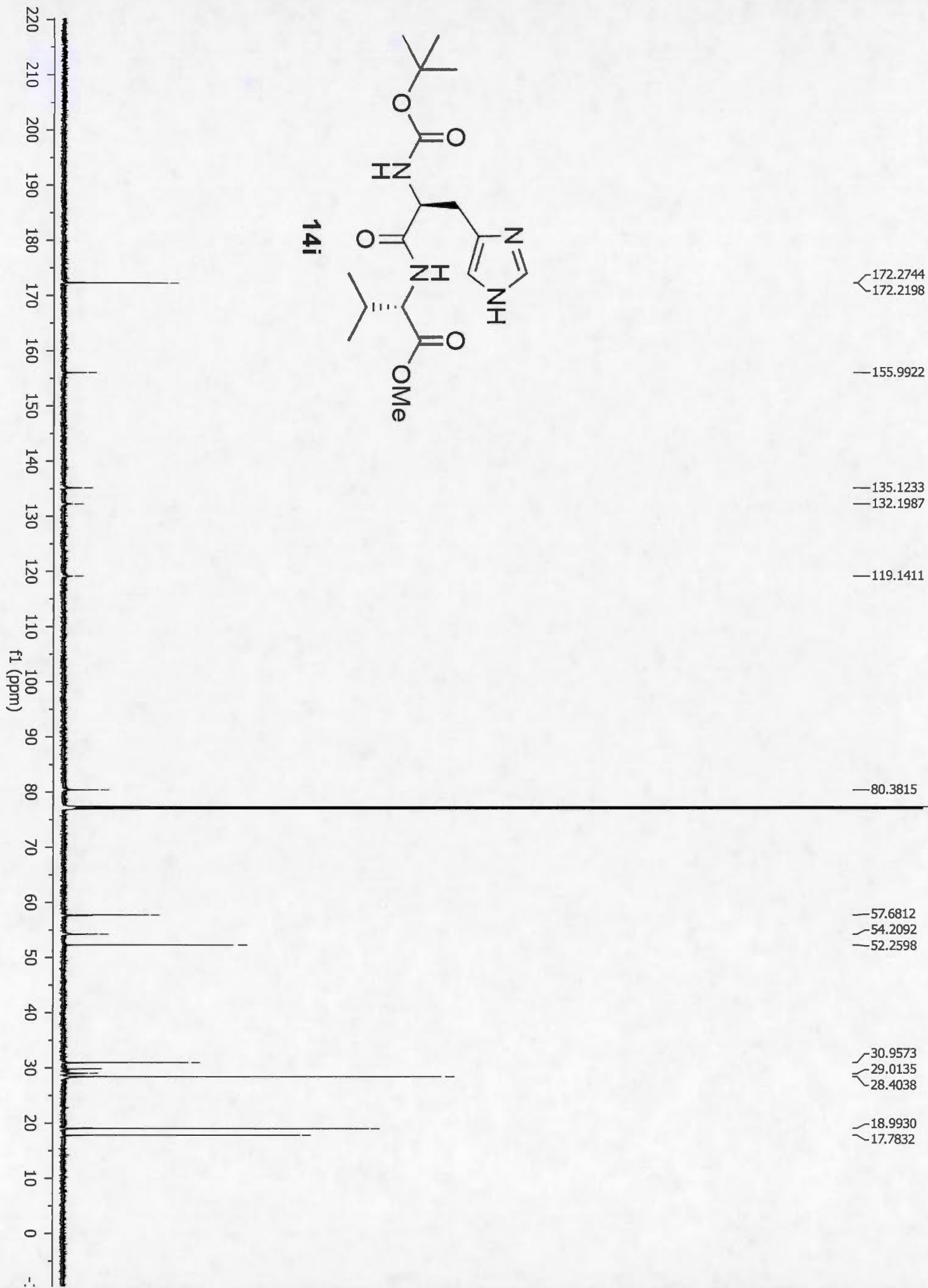
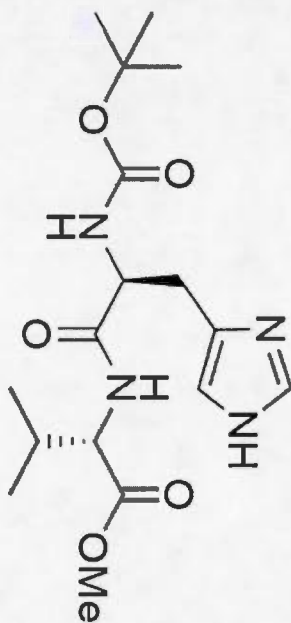


13



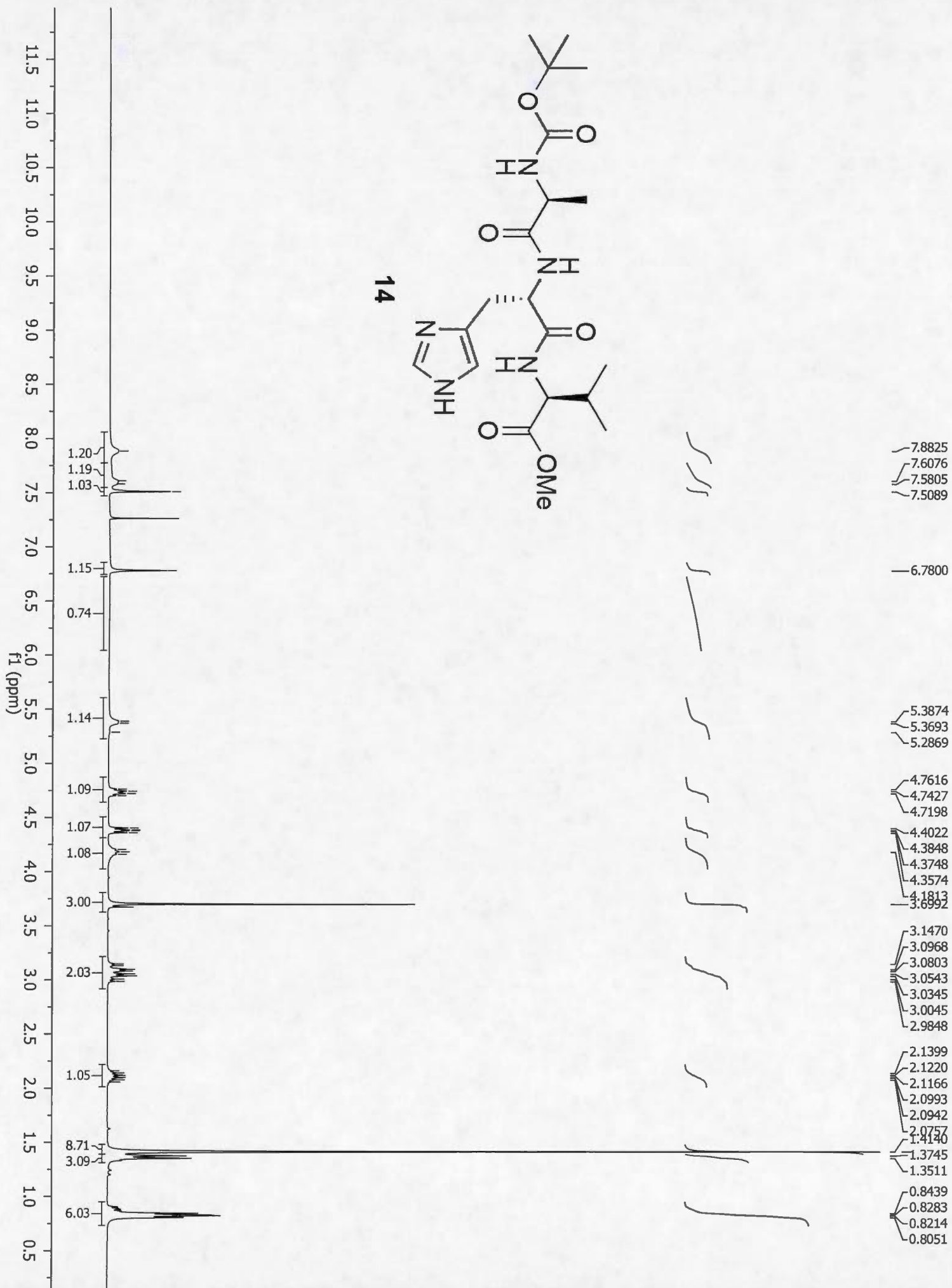
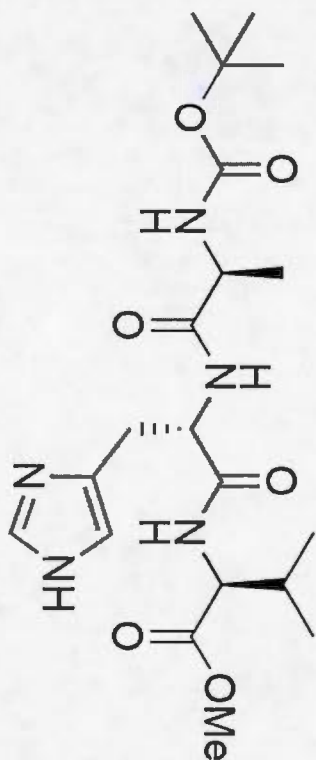


14i

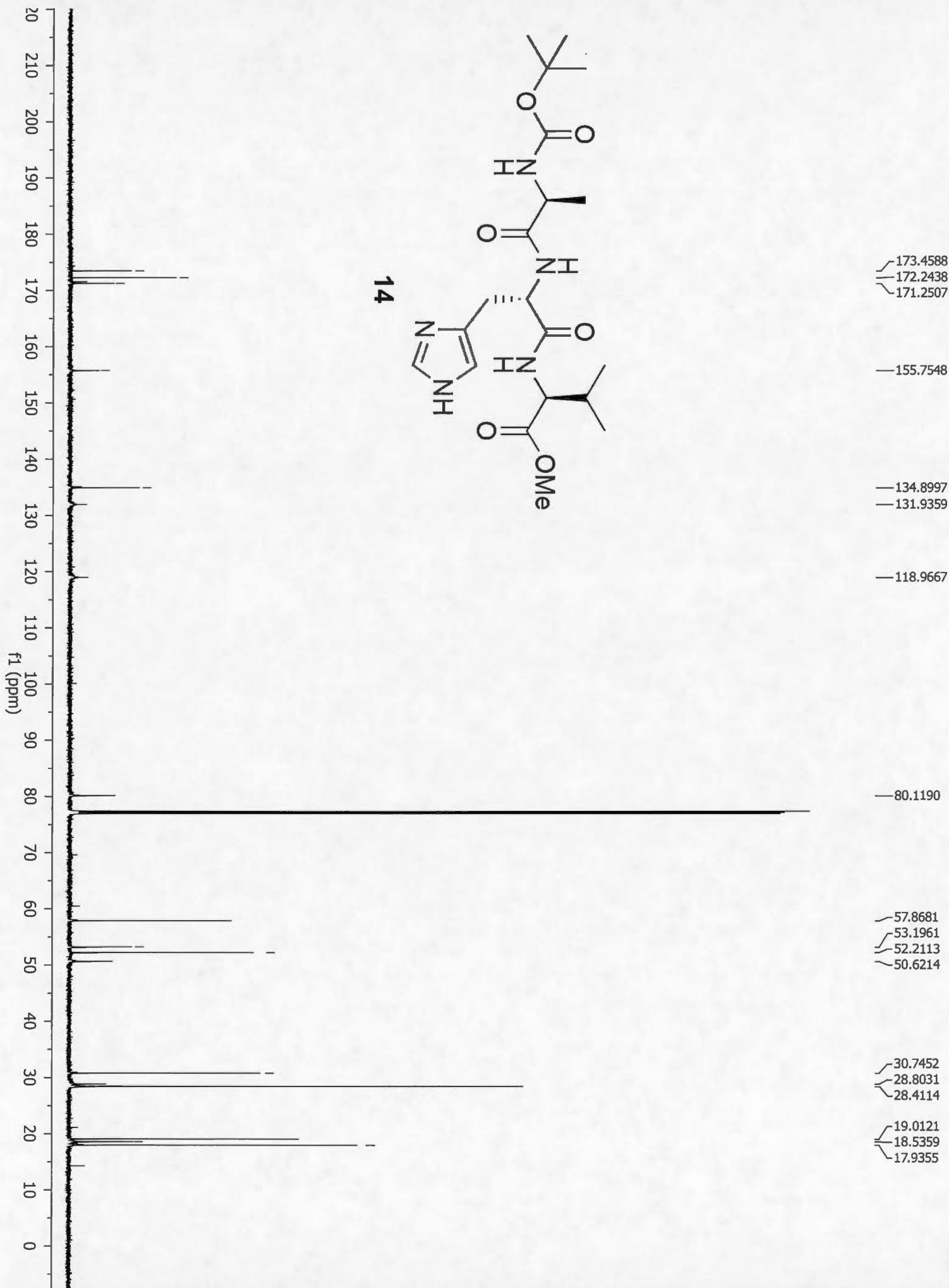
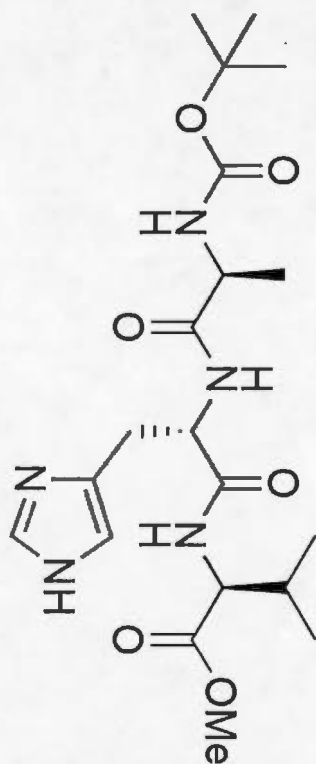


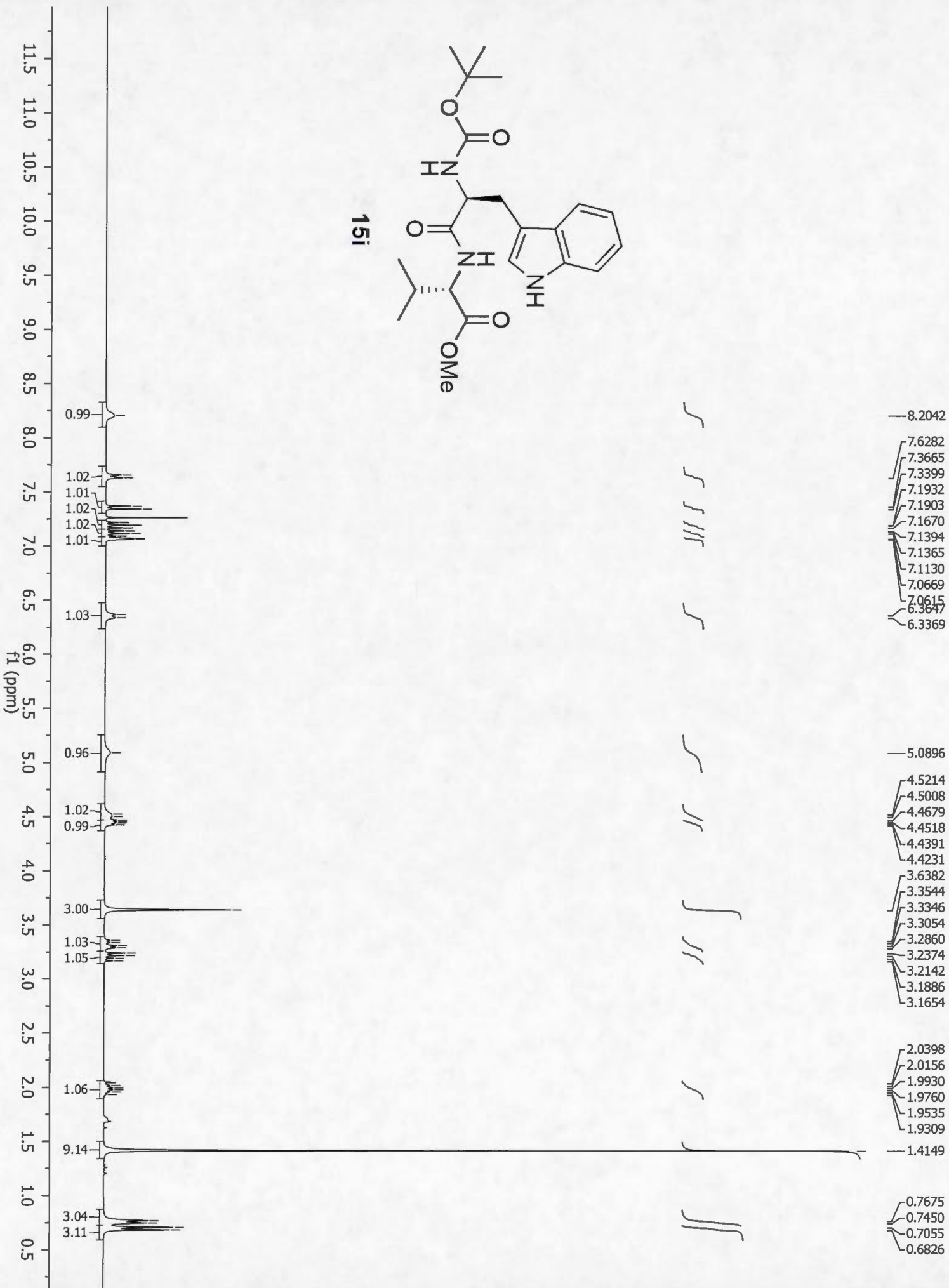


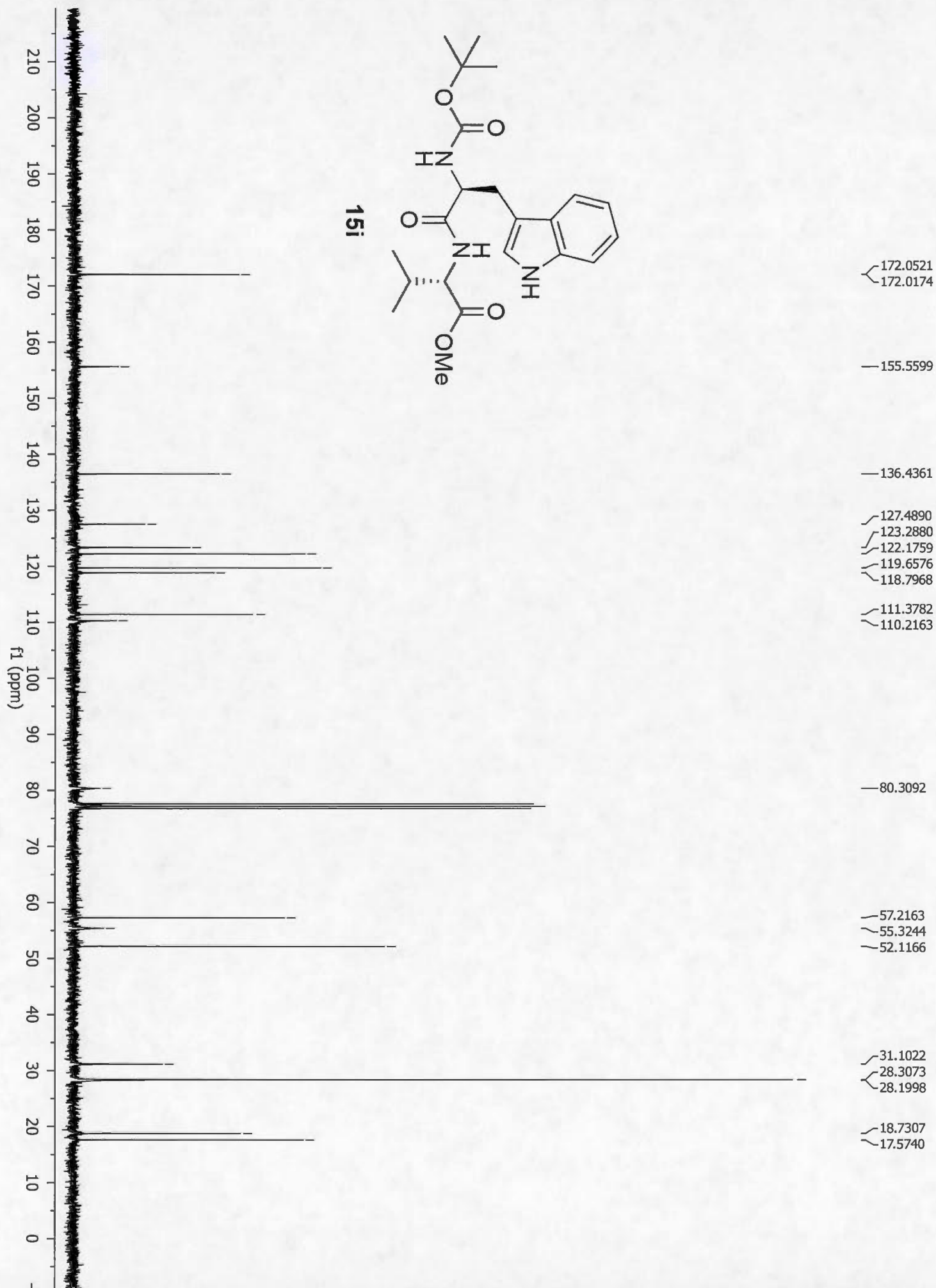
14



14

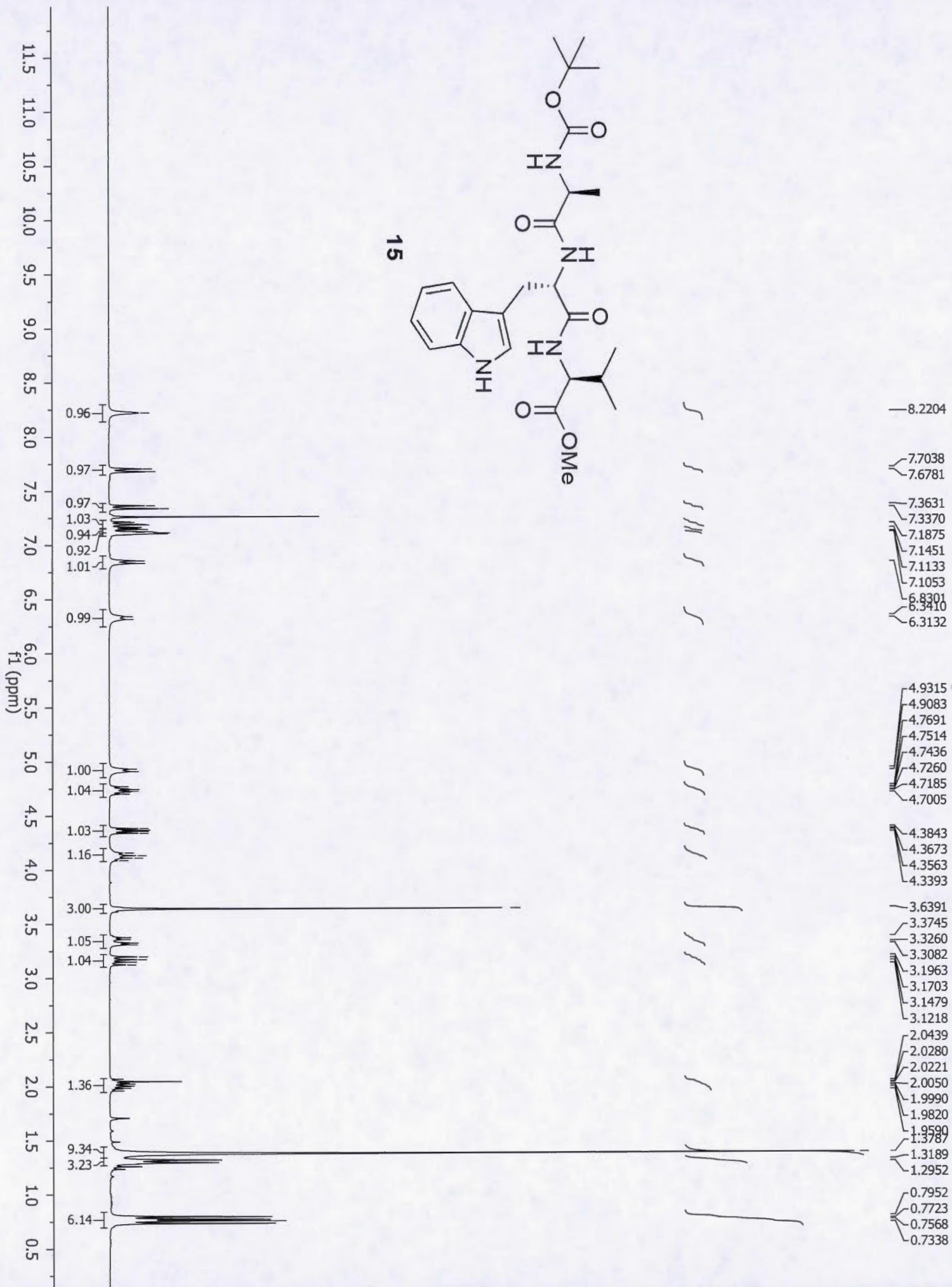
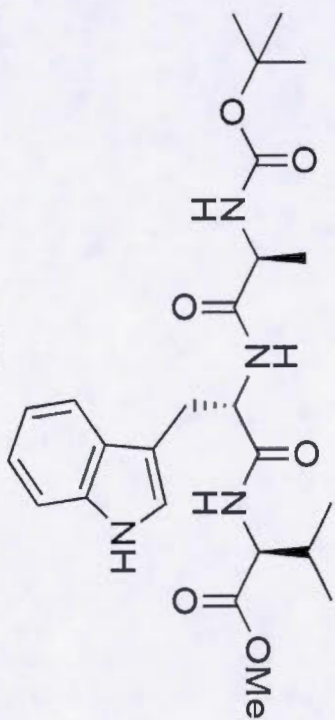


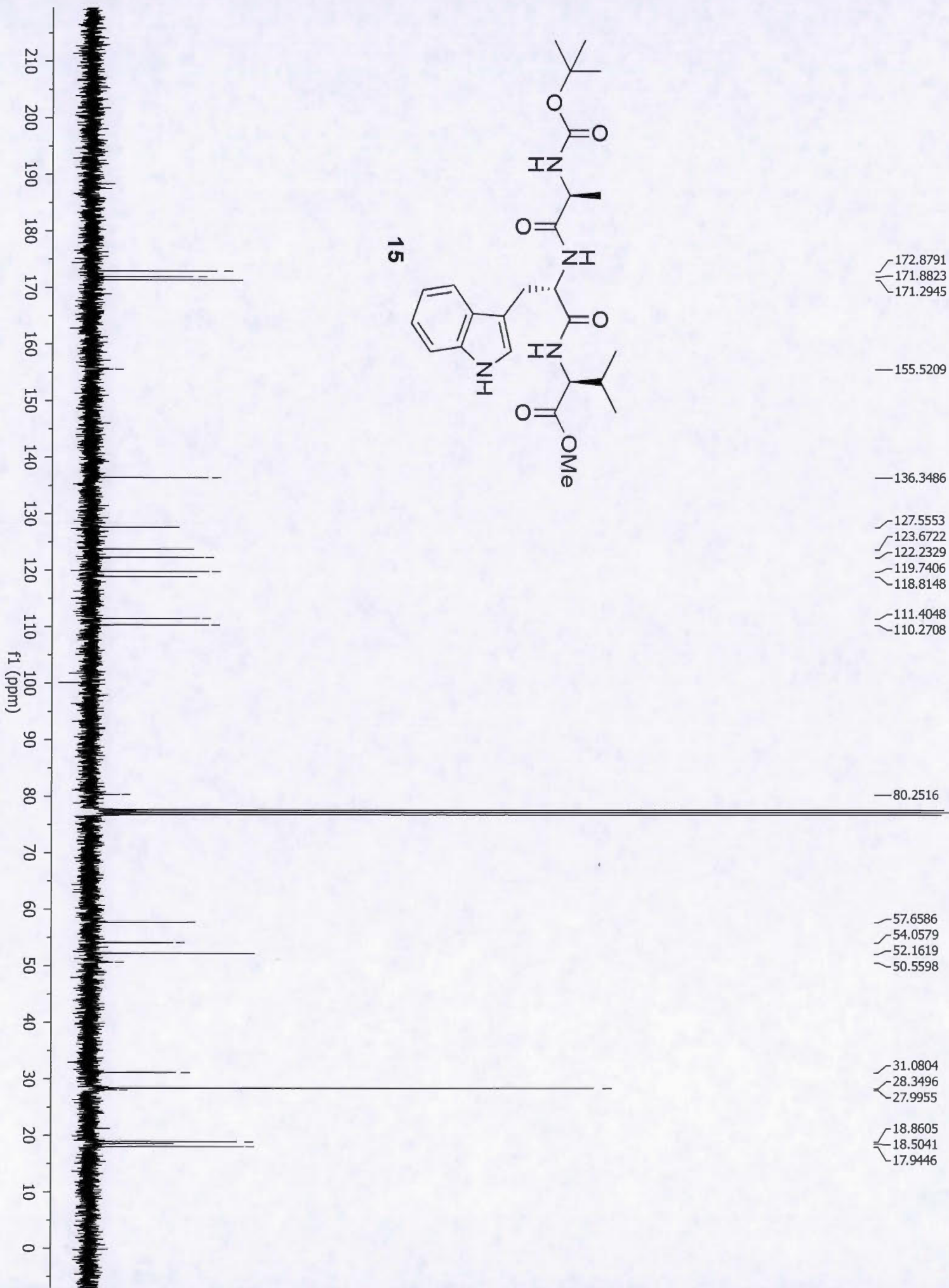




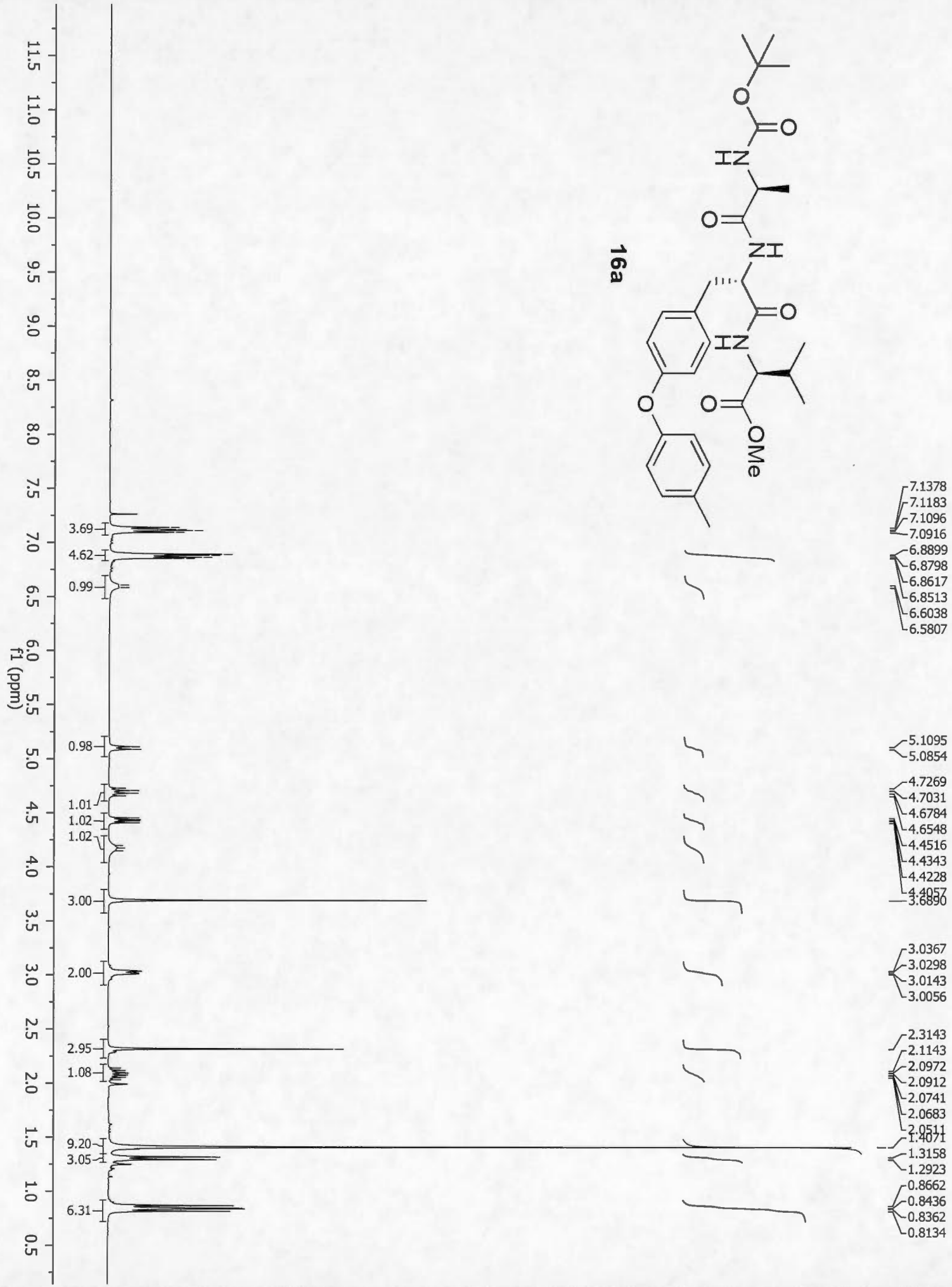
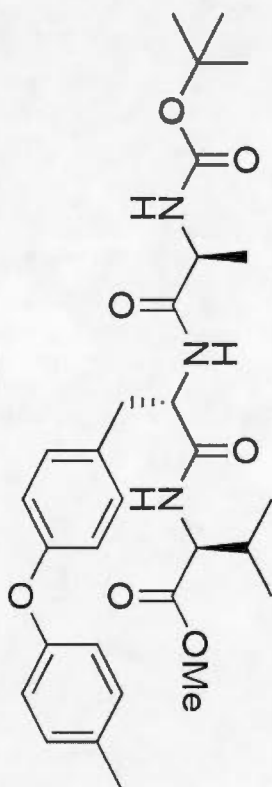


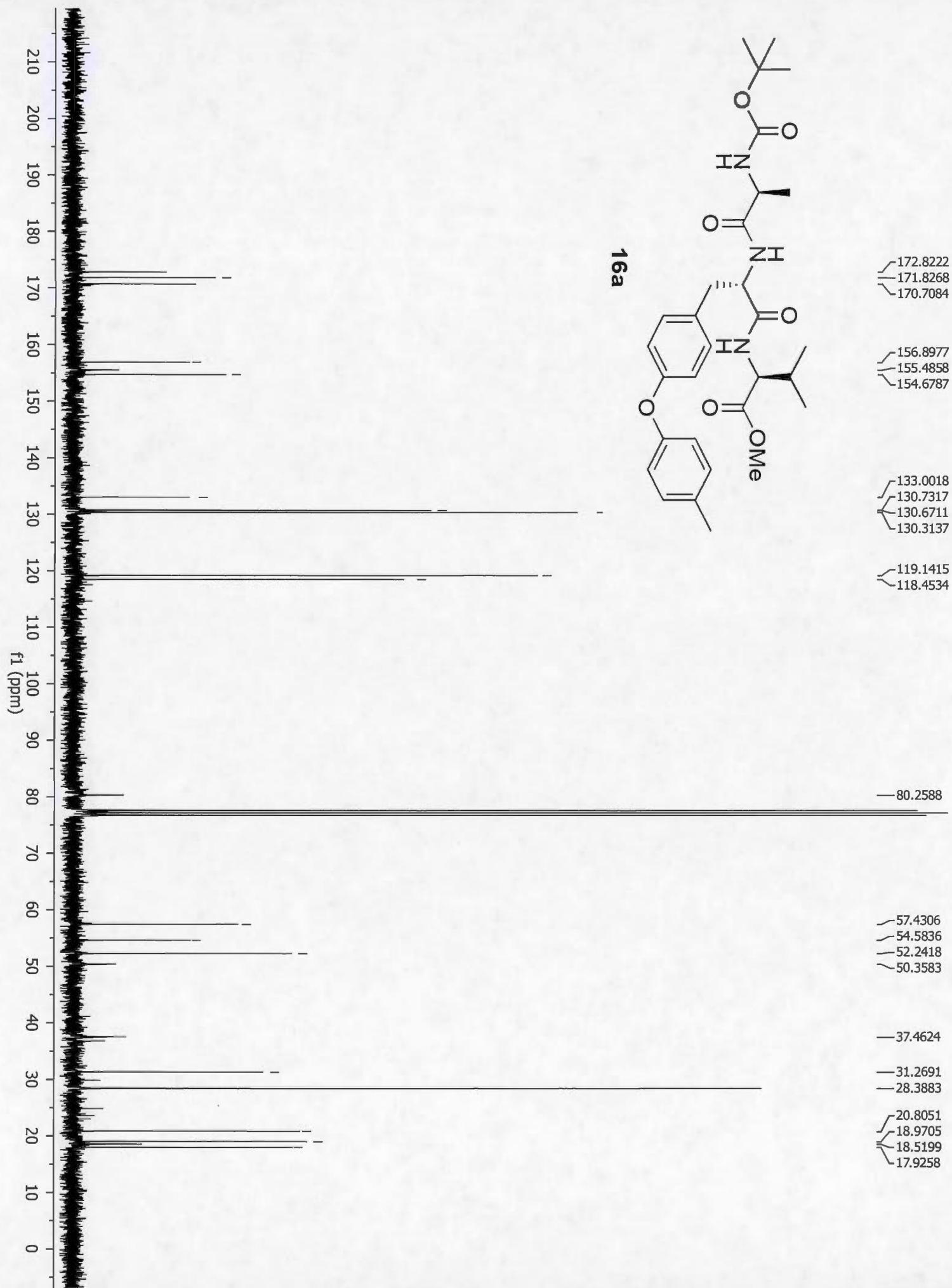
15





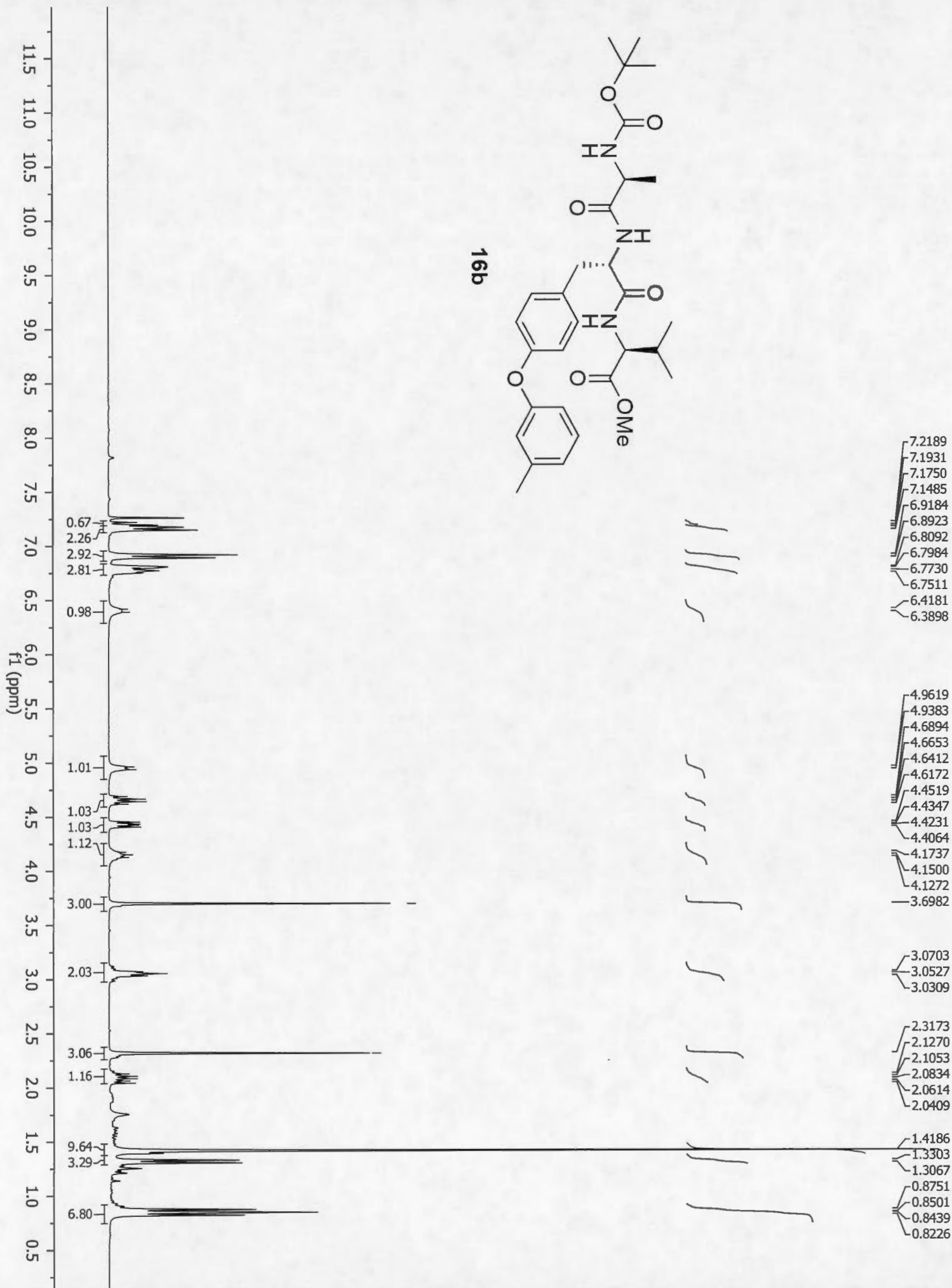
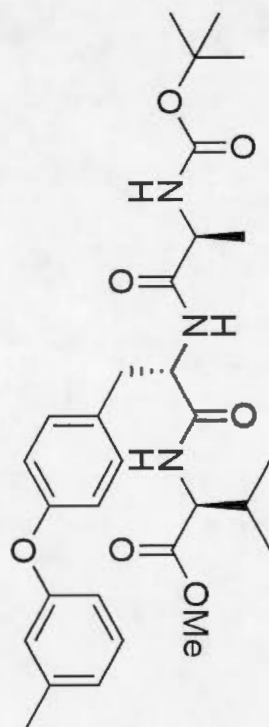
16a



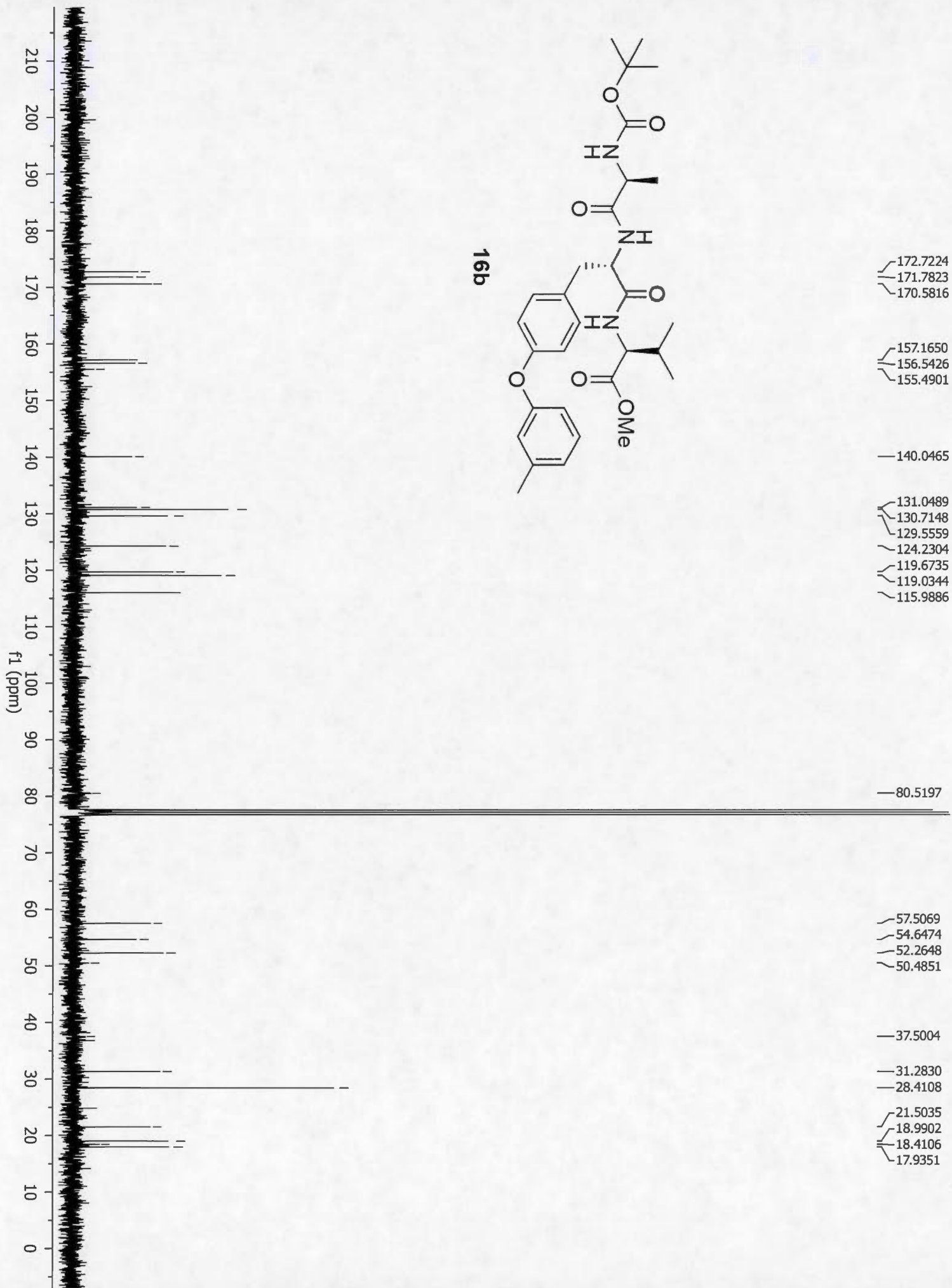
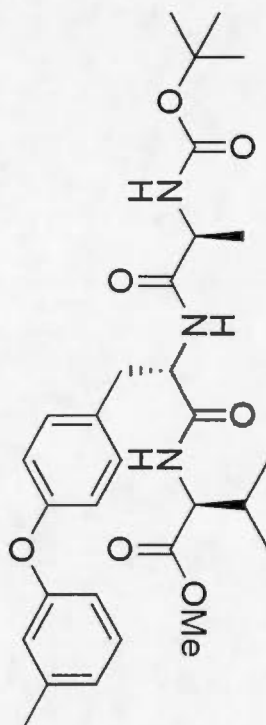




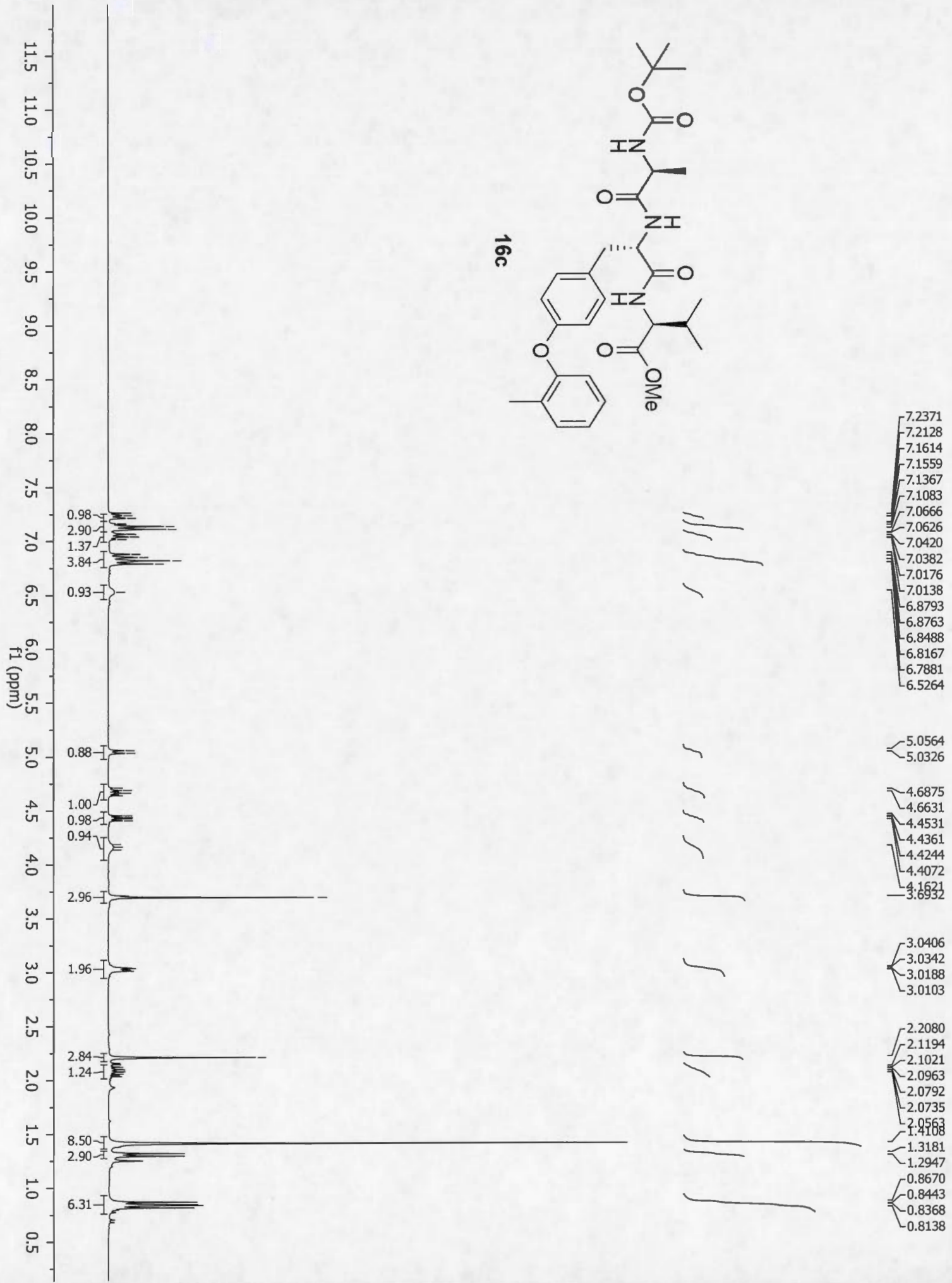
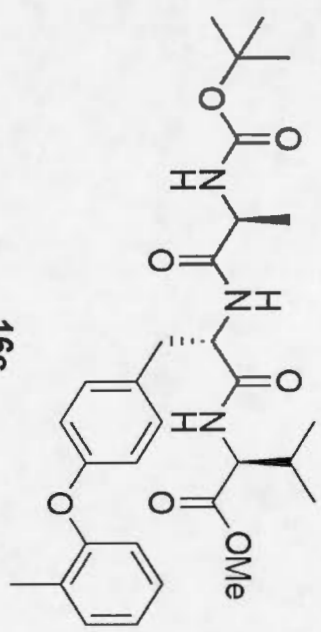
16b

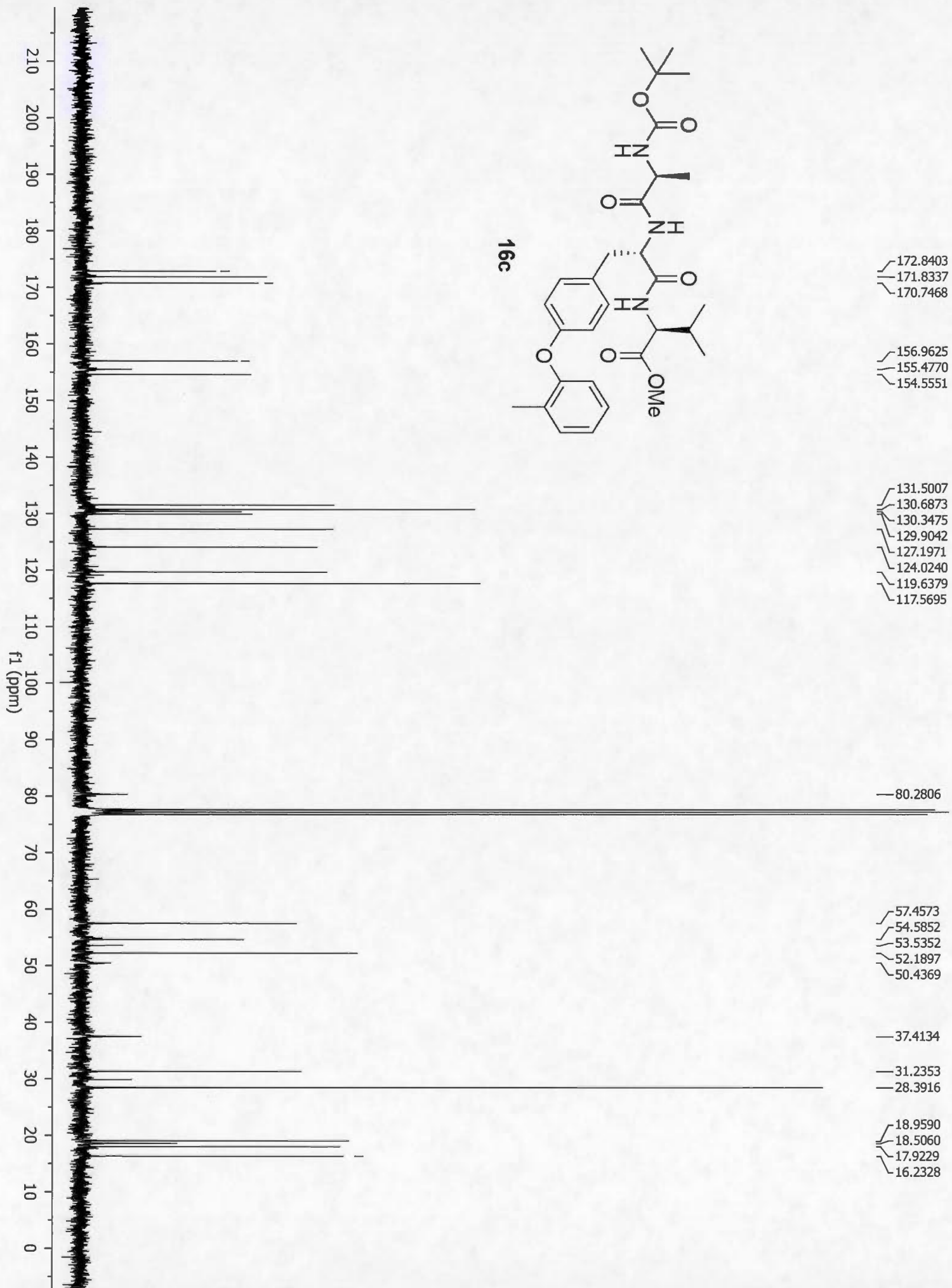


16b

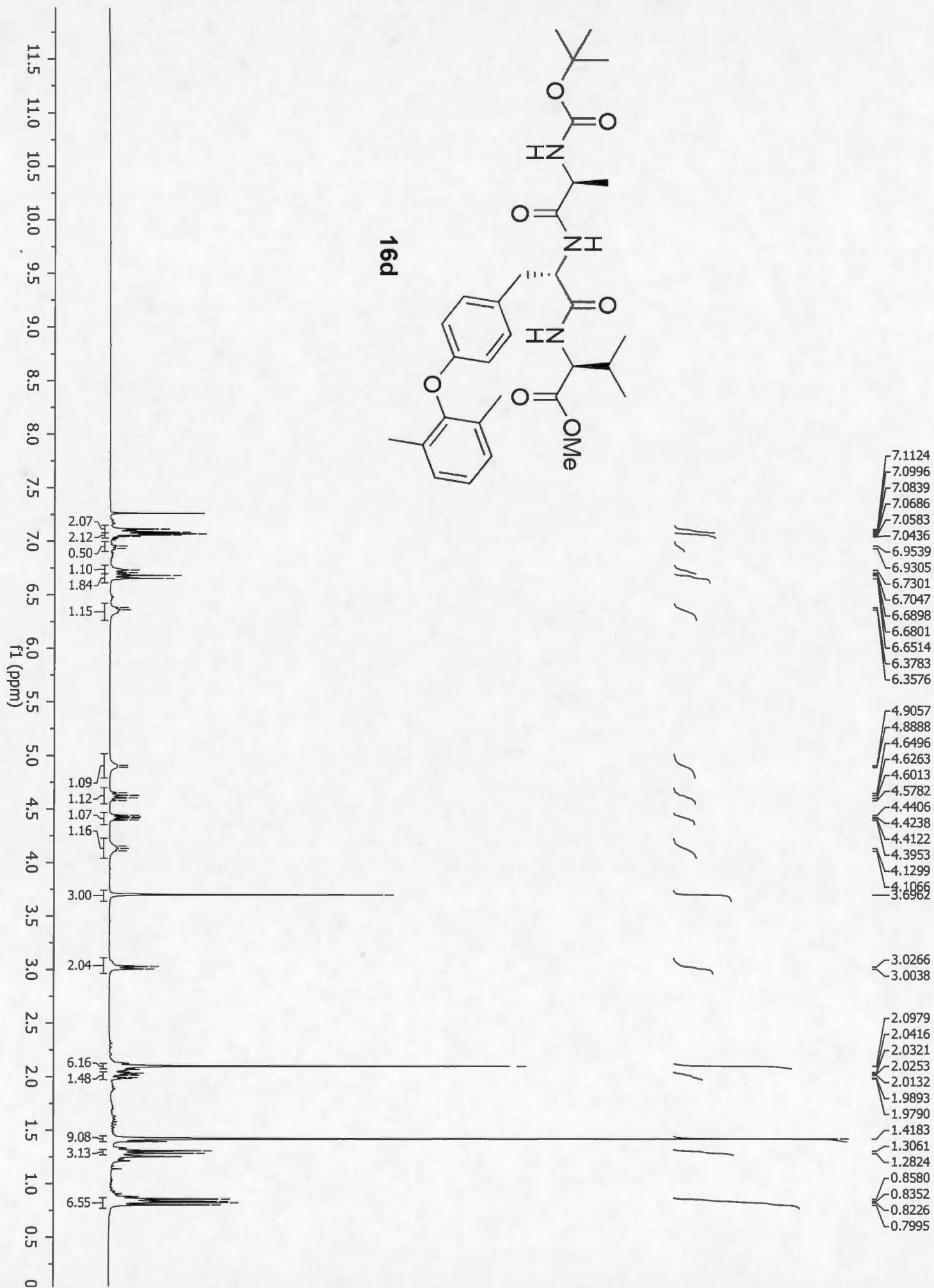
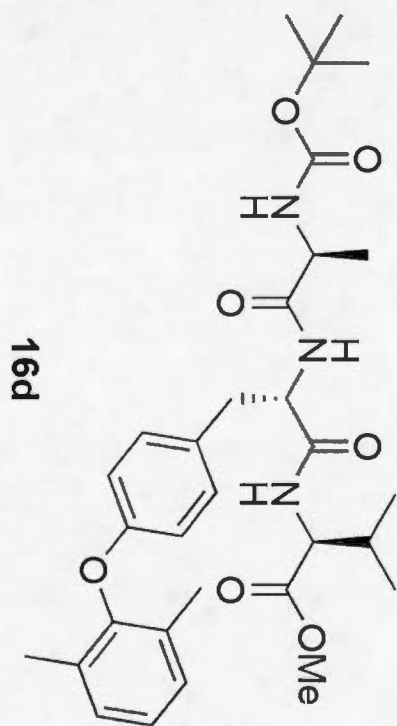


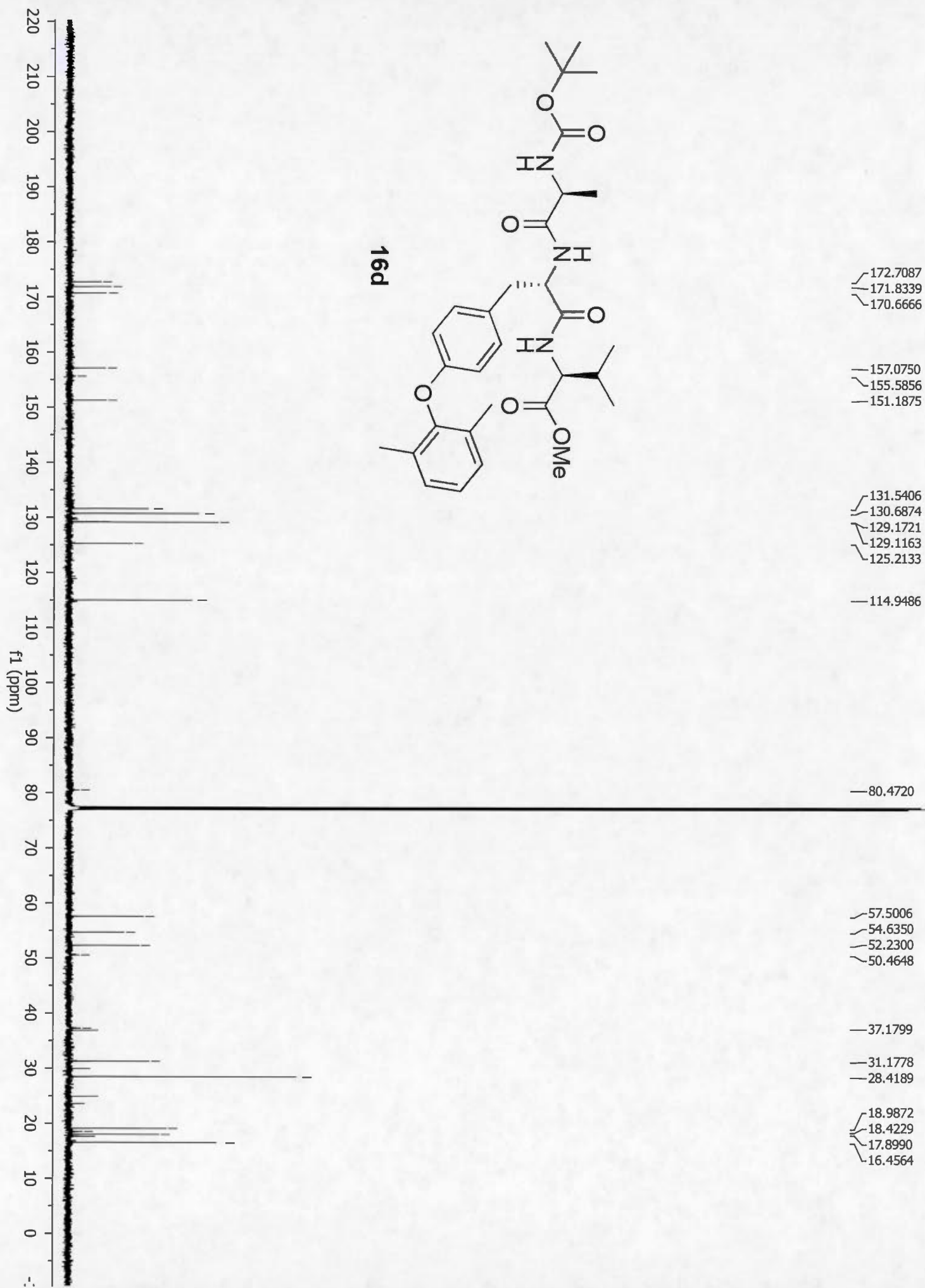
16c



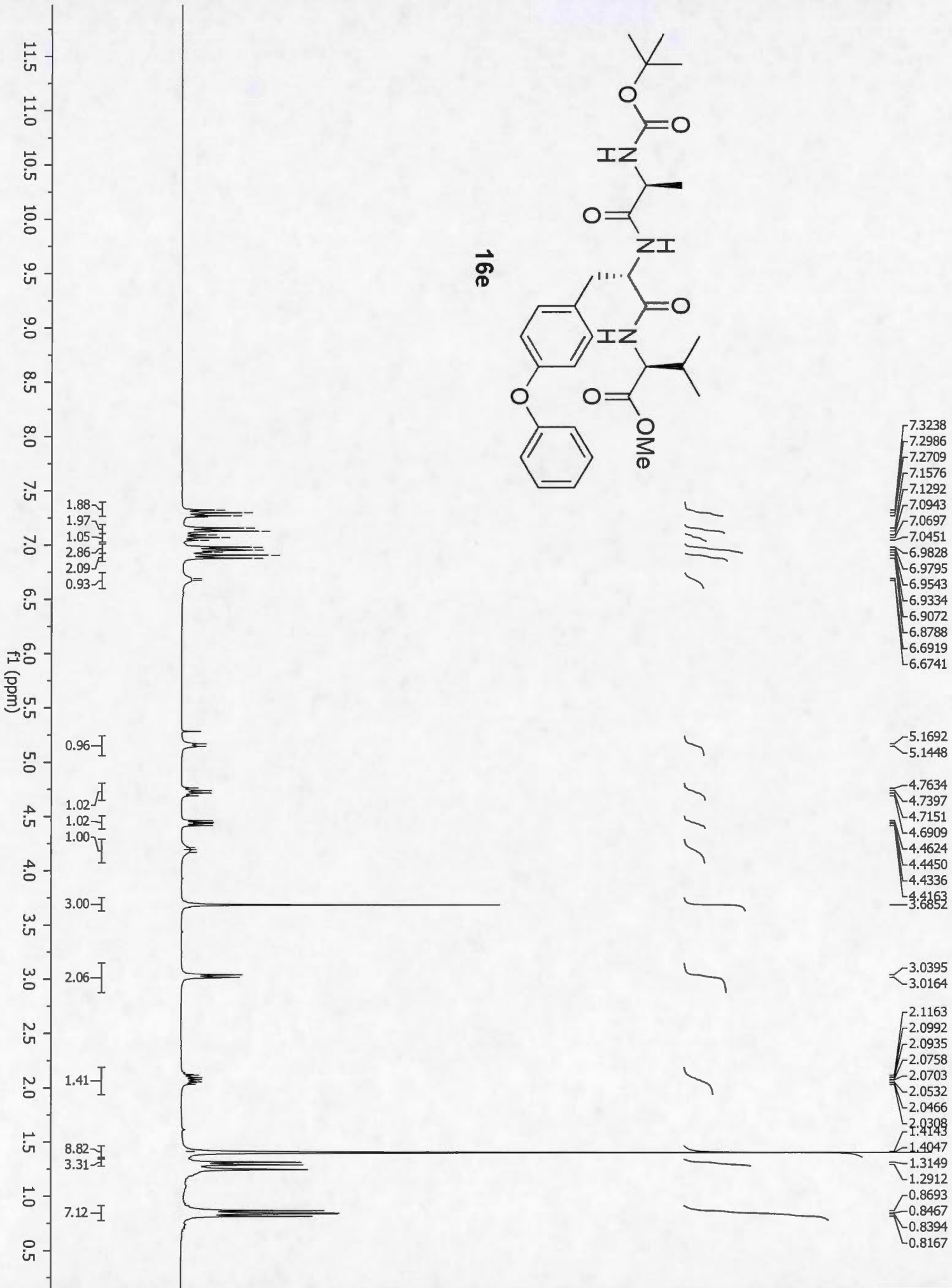
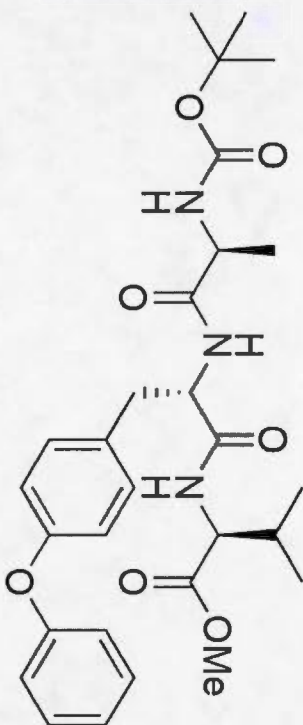


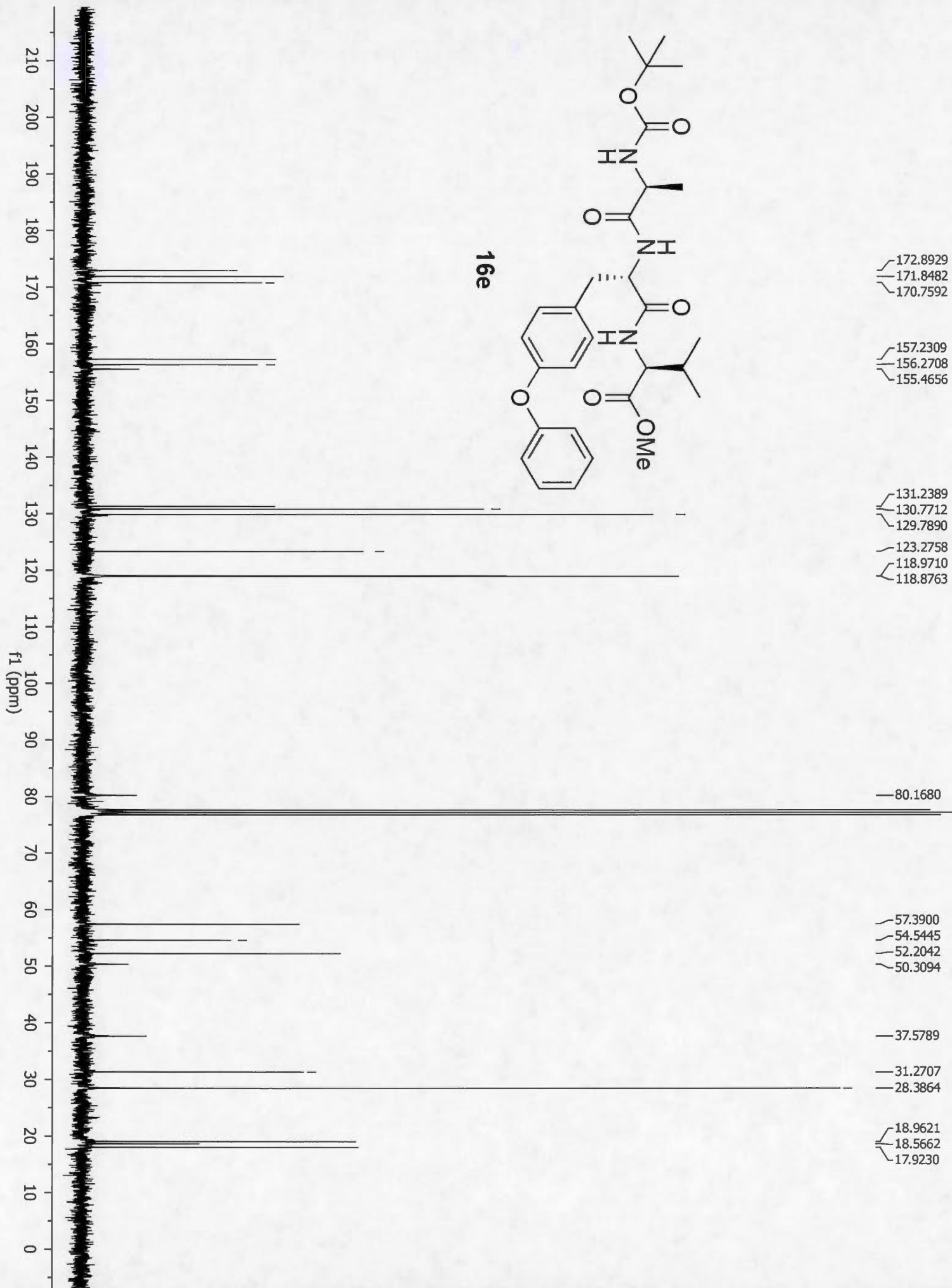




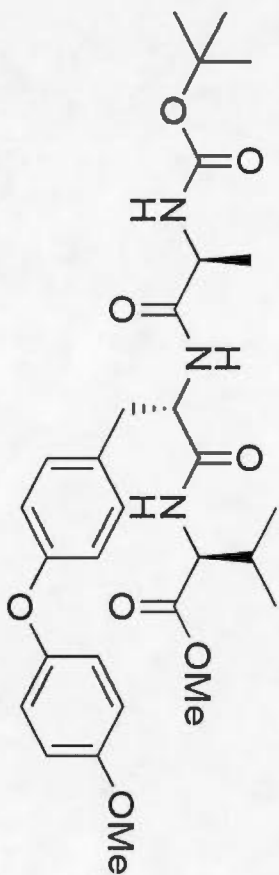


16e

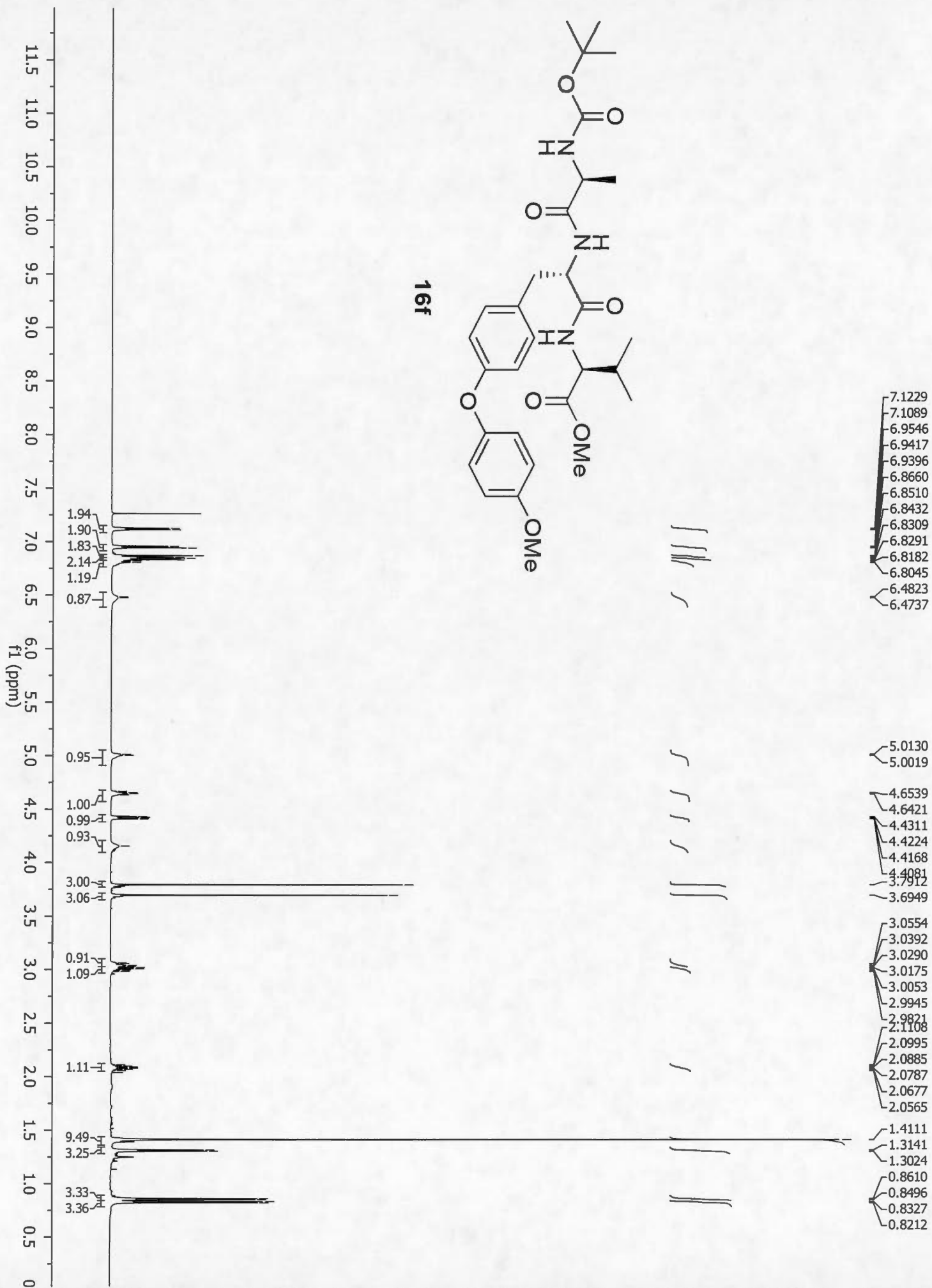


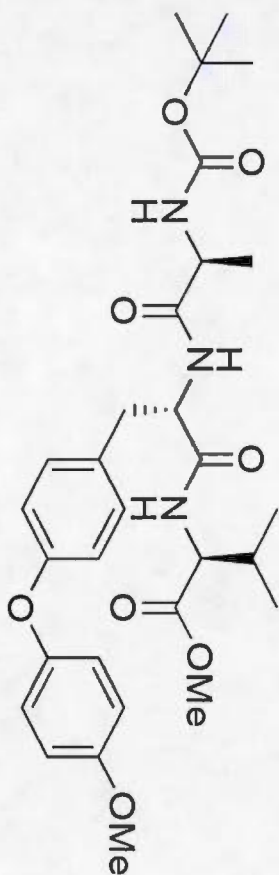




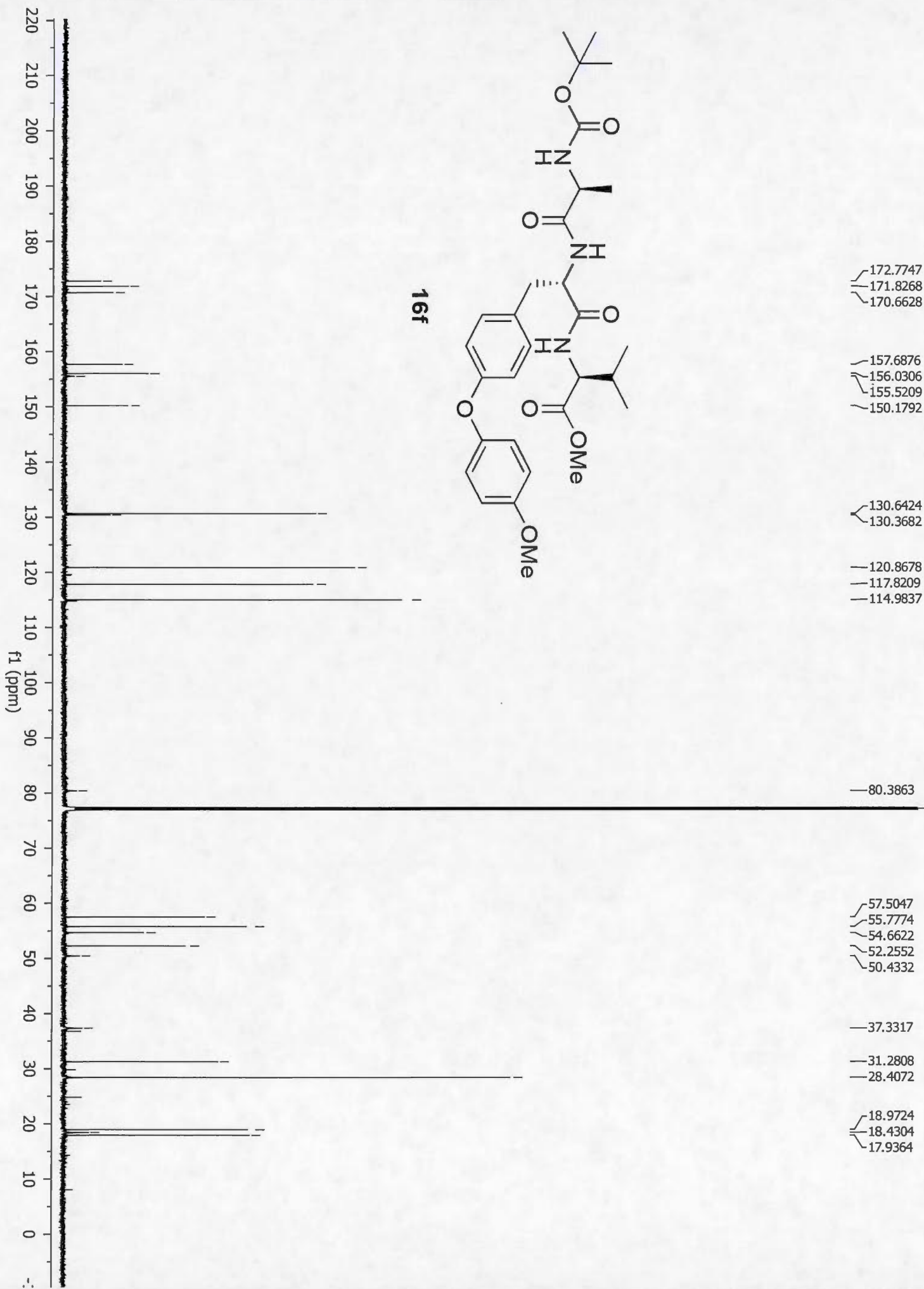


16f

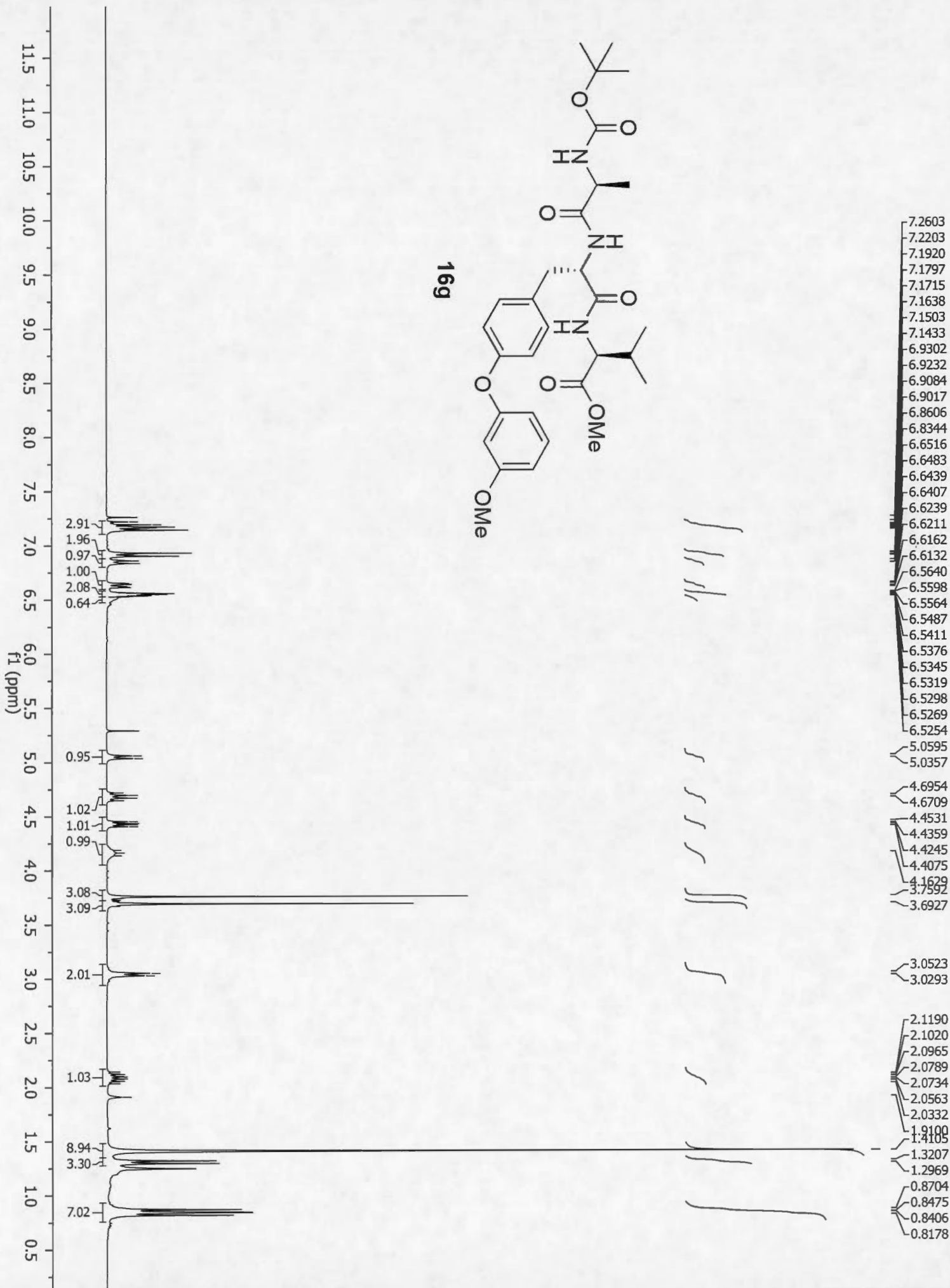
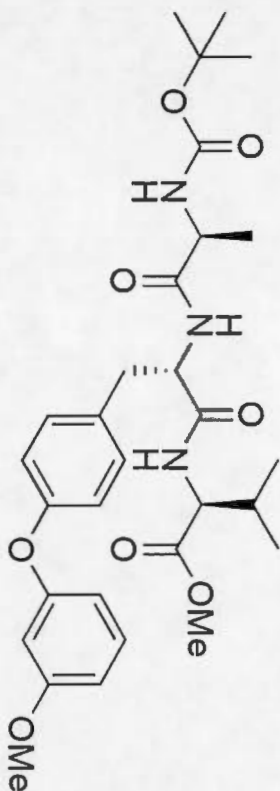




16f

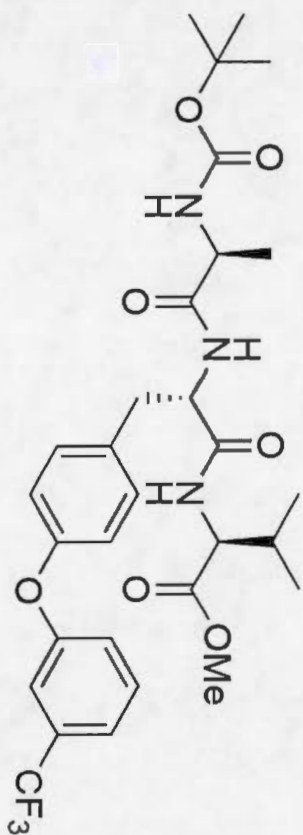


16g

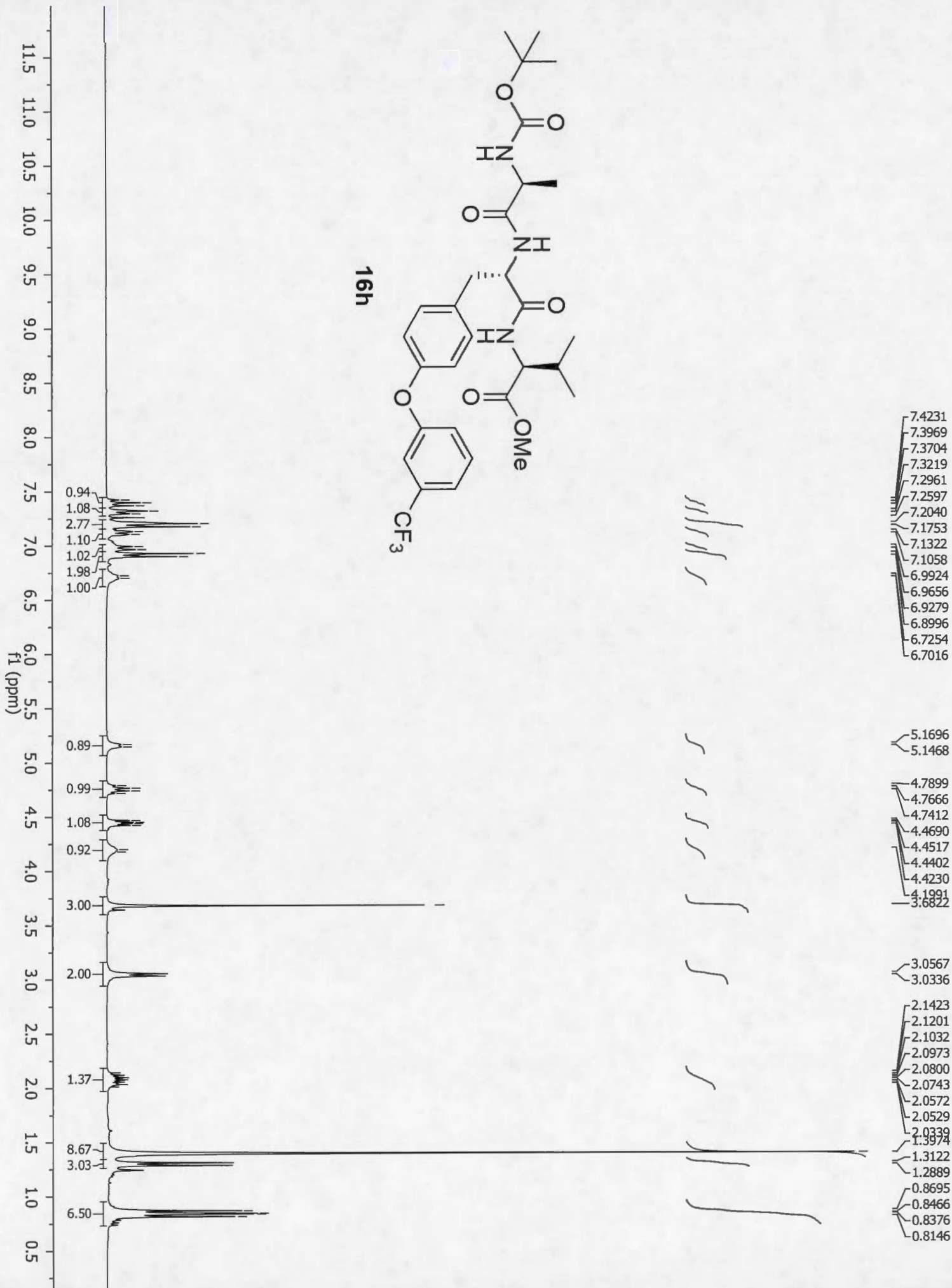


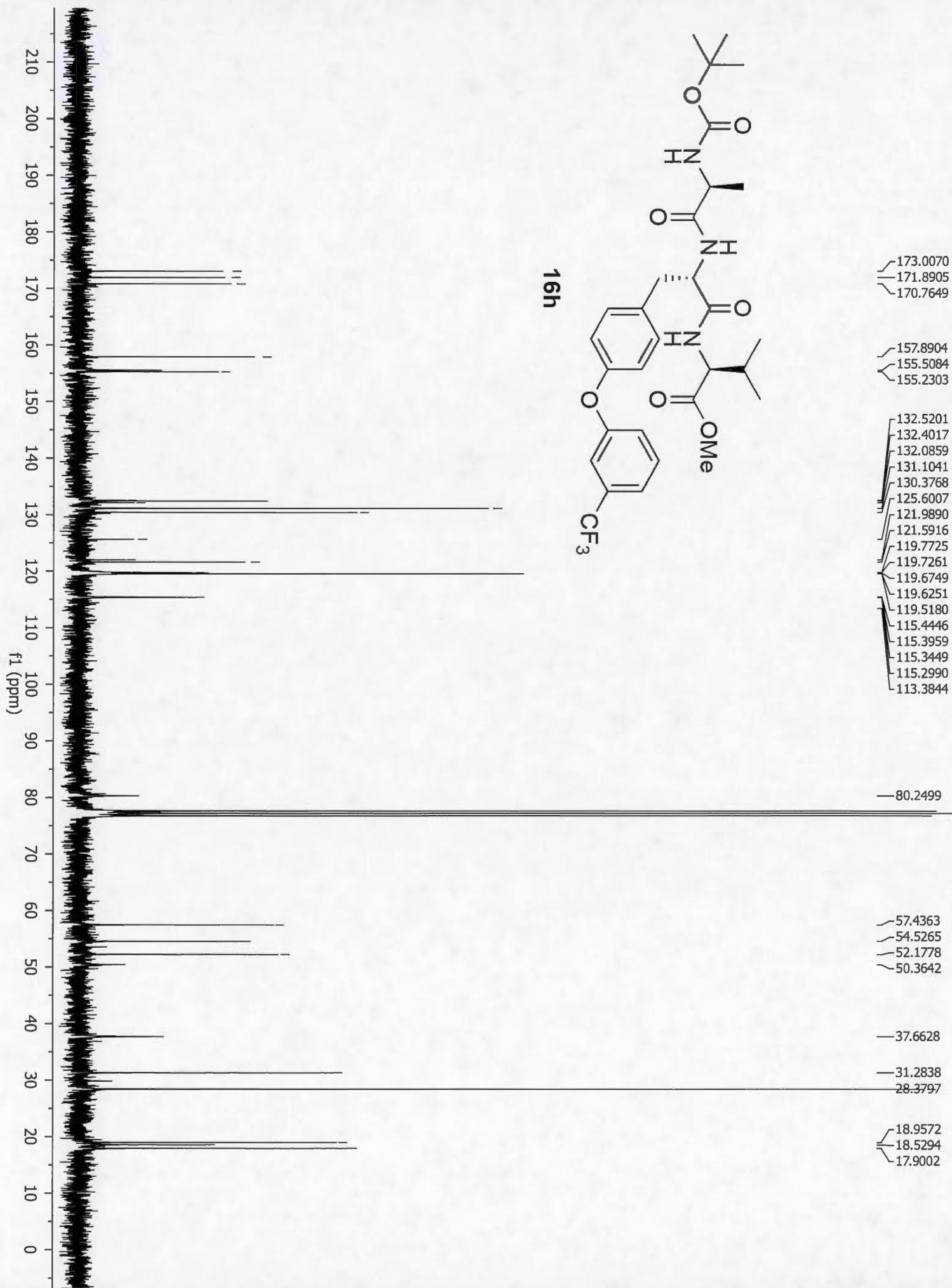




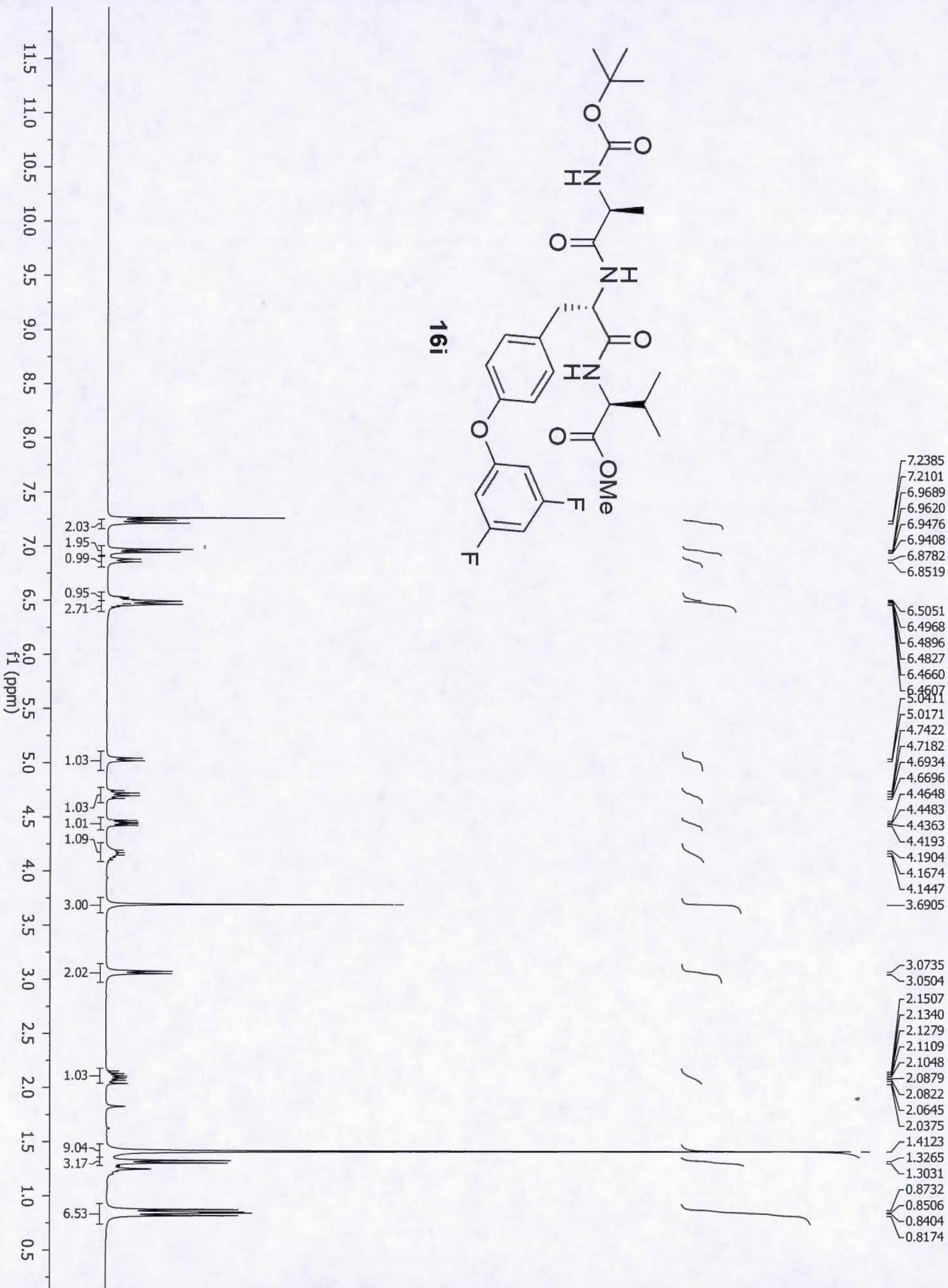
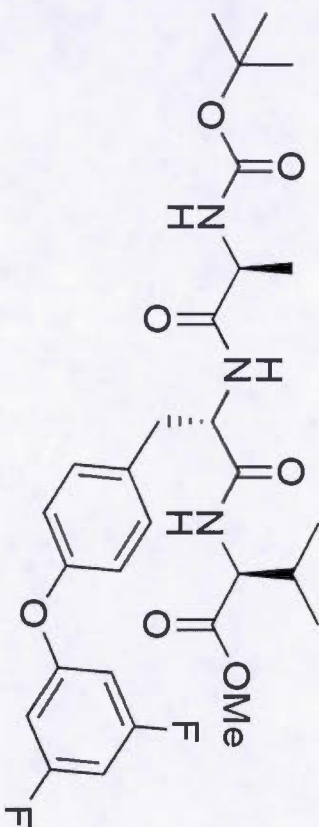


16h

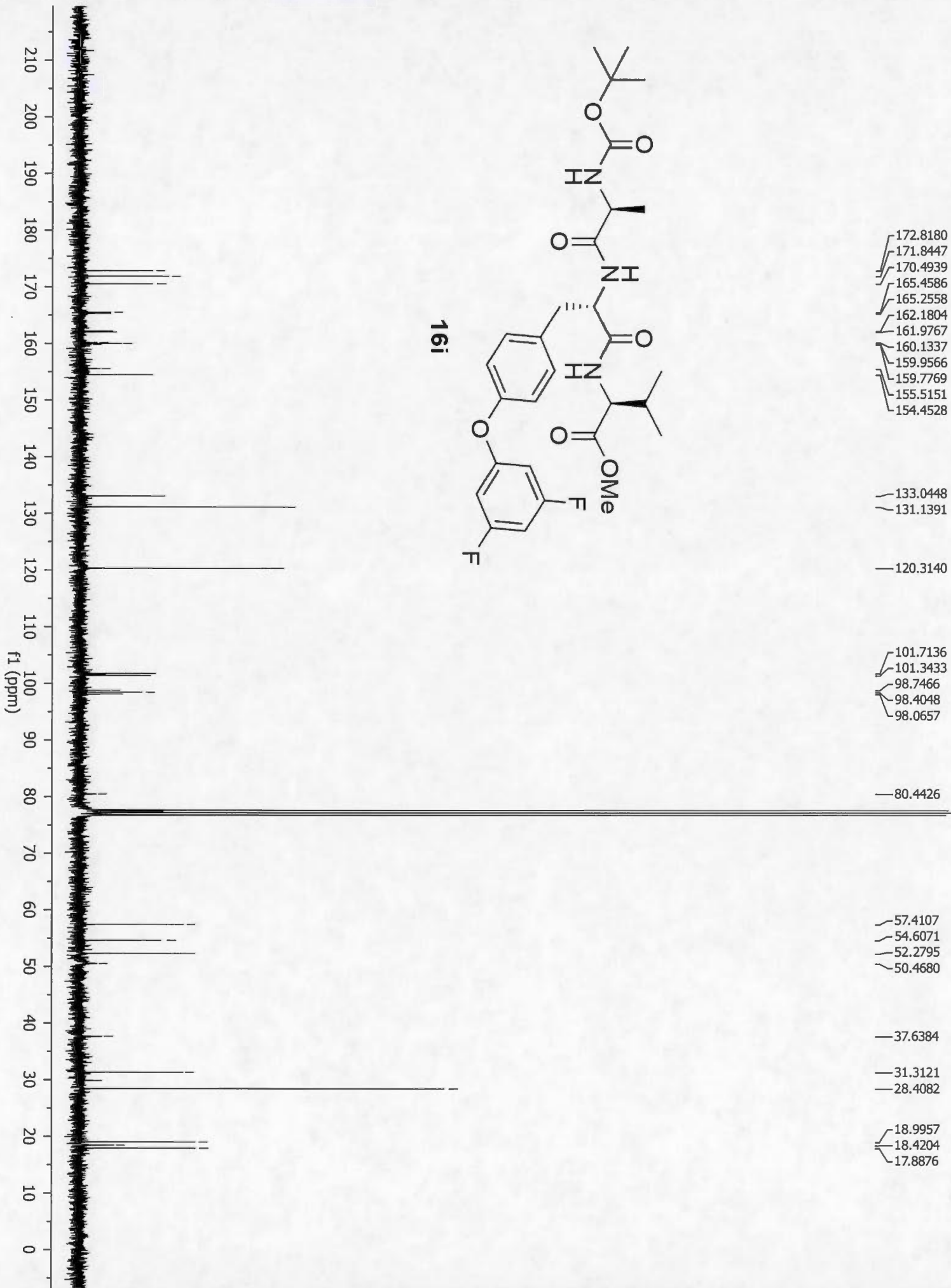
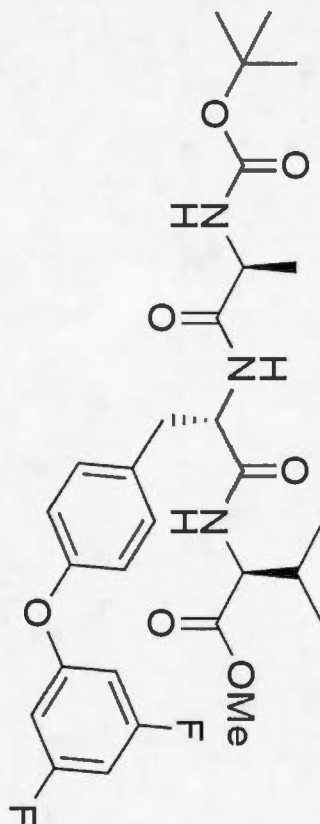




16i

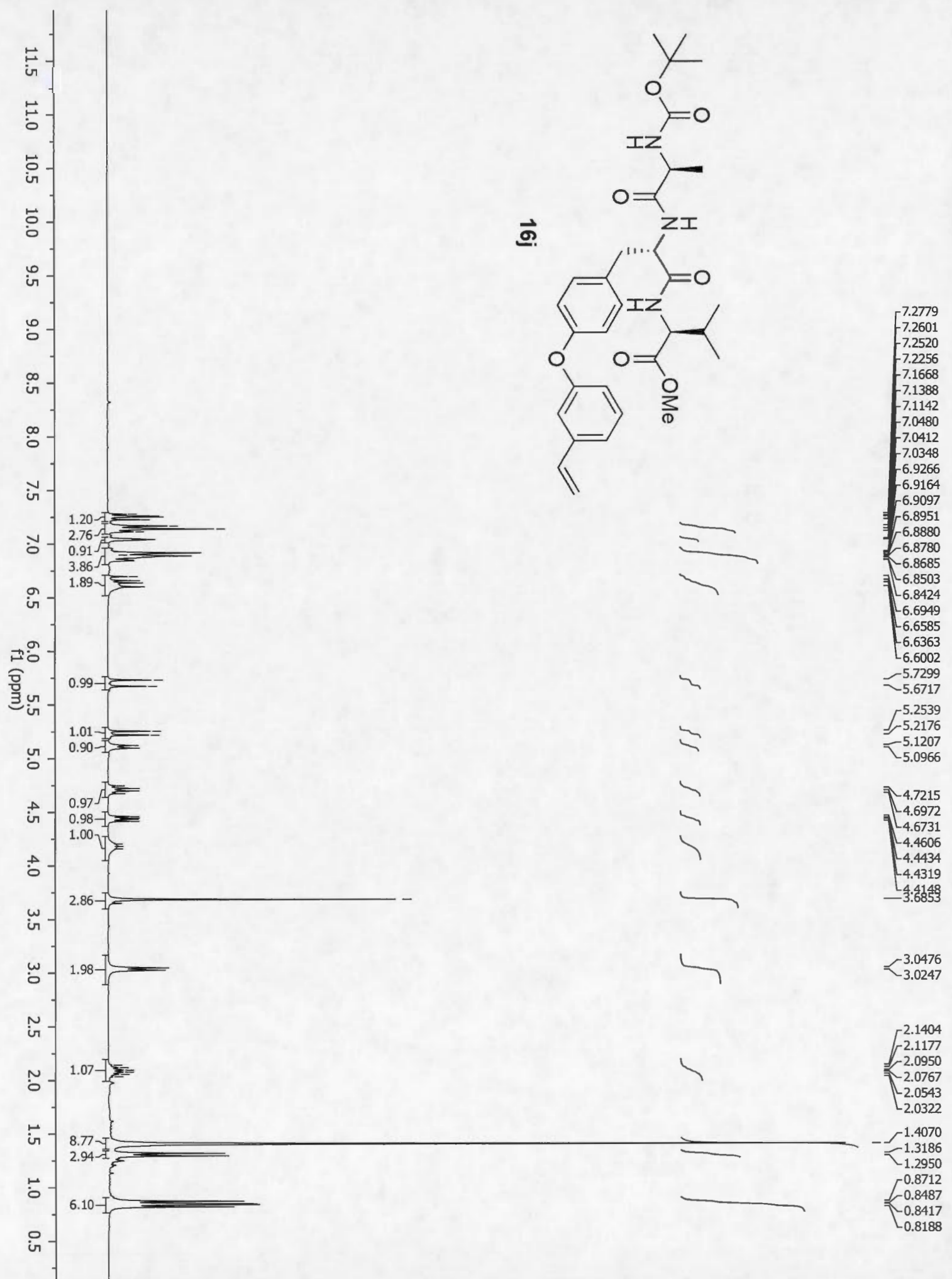
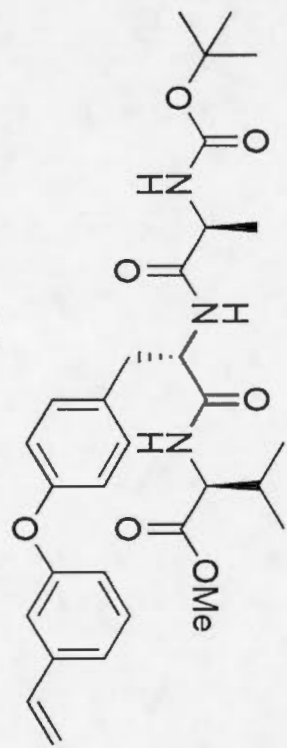


16i

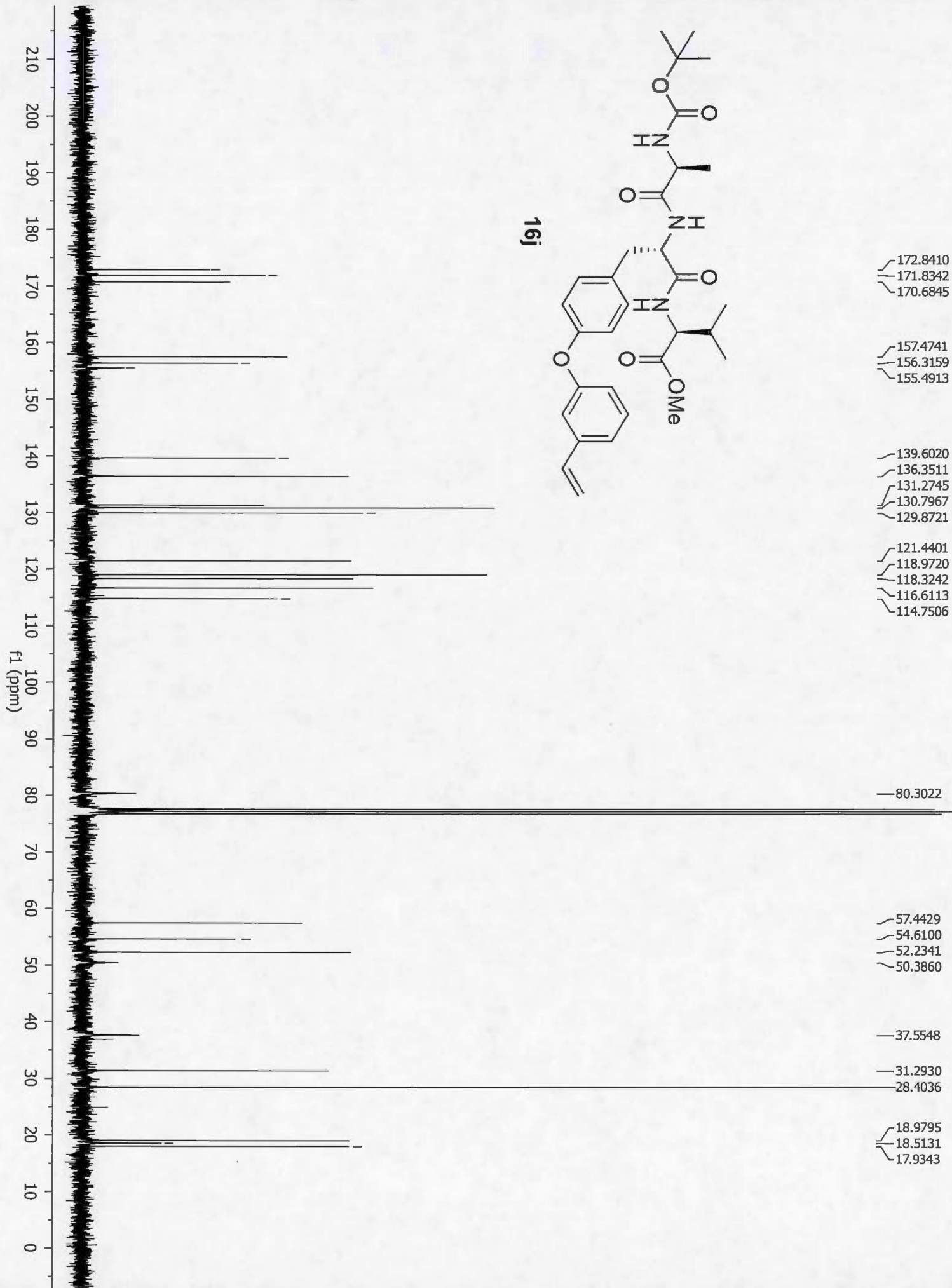
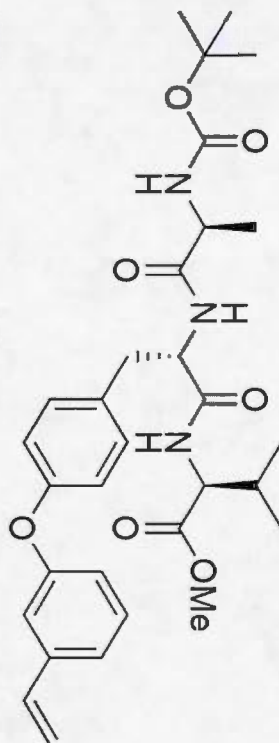




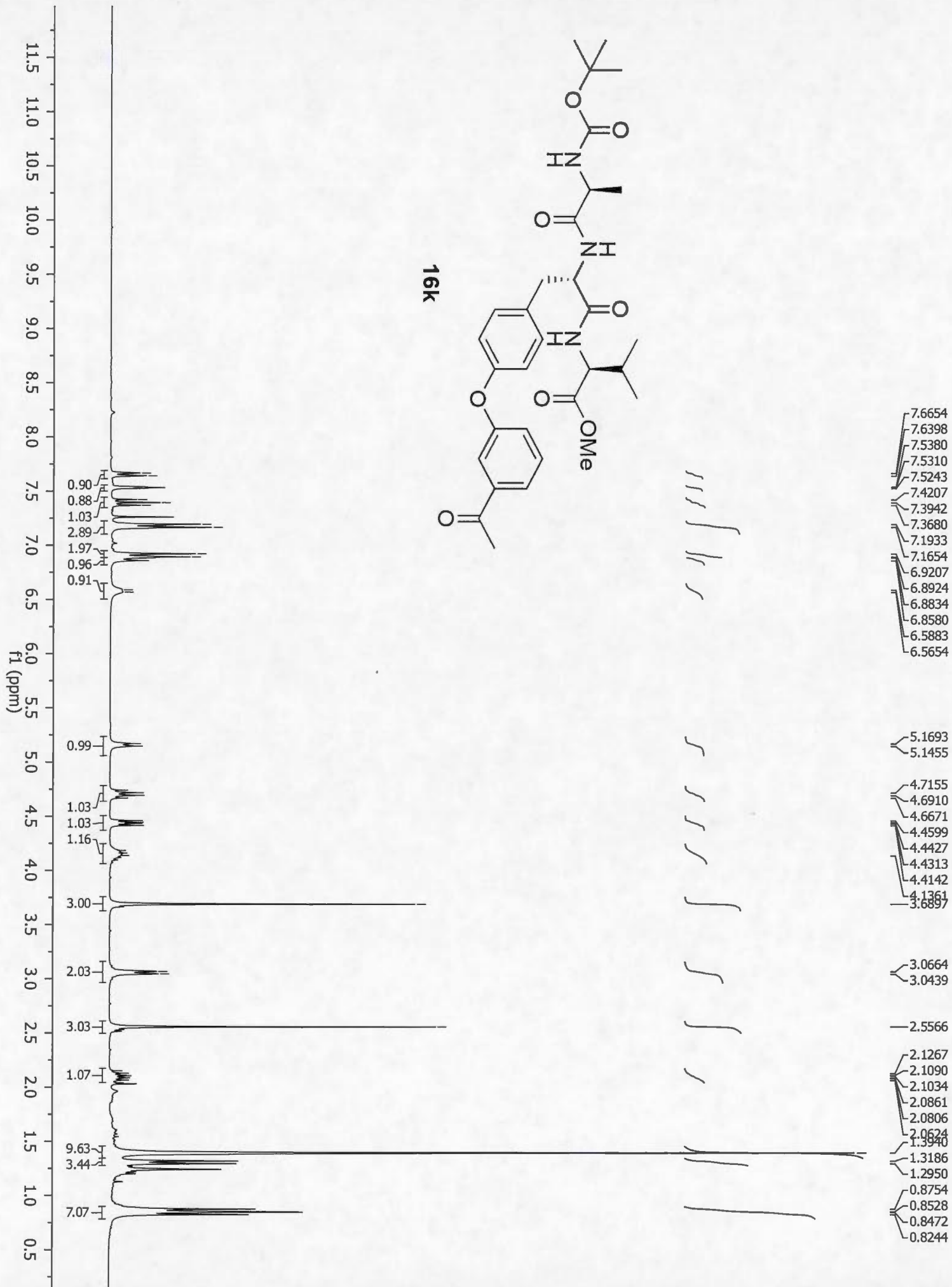
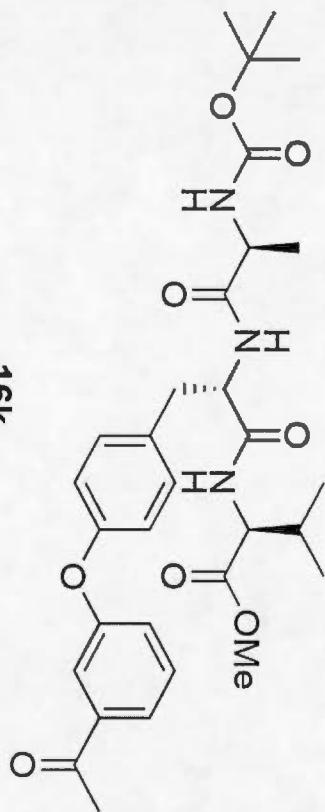
16j

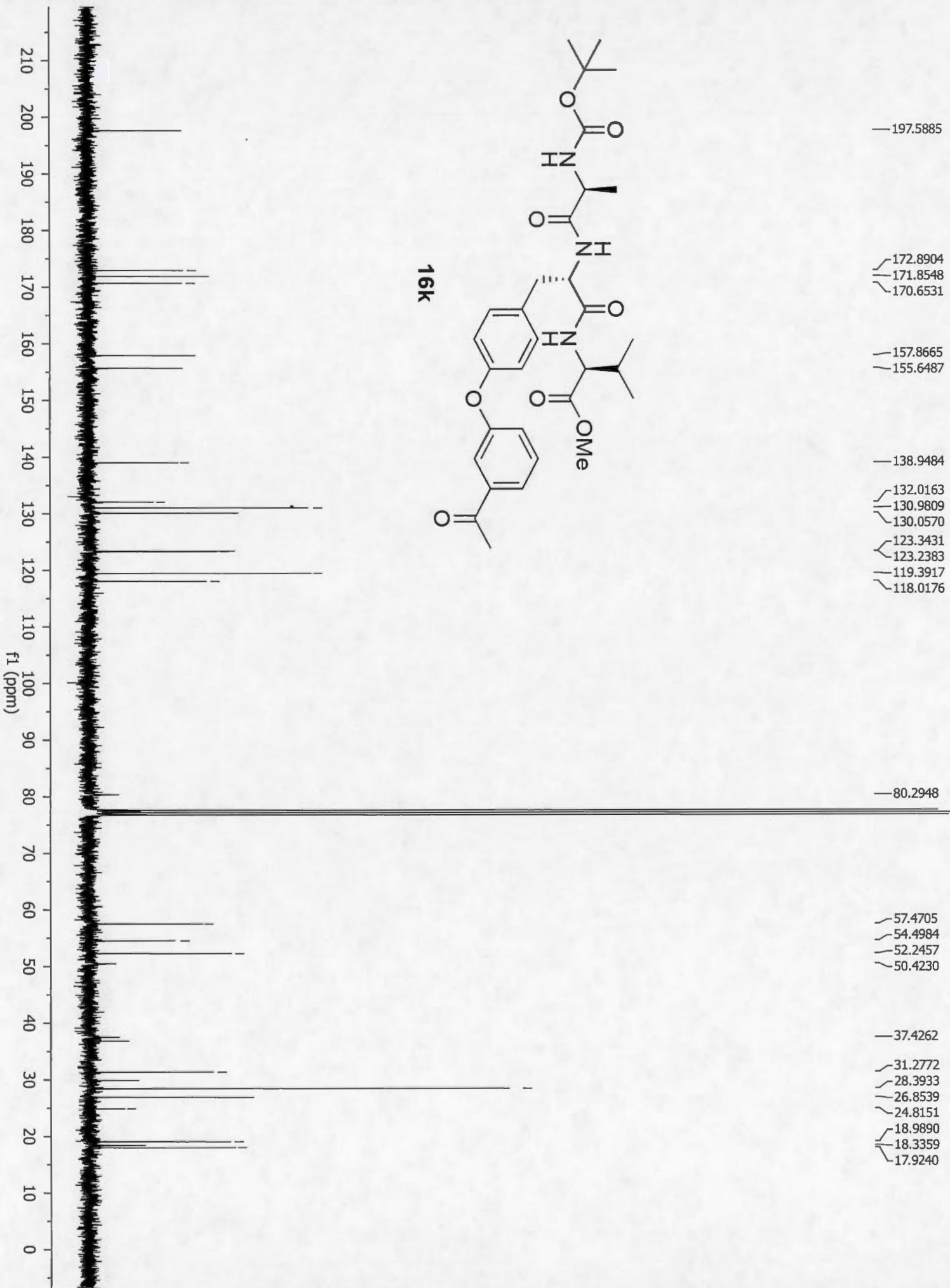


16j



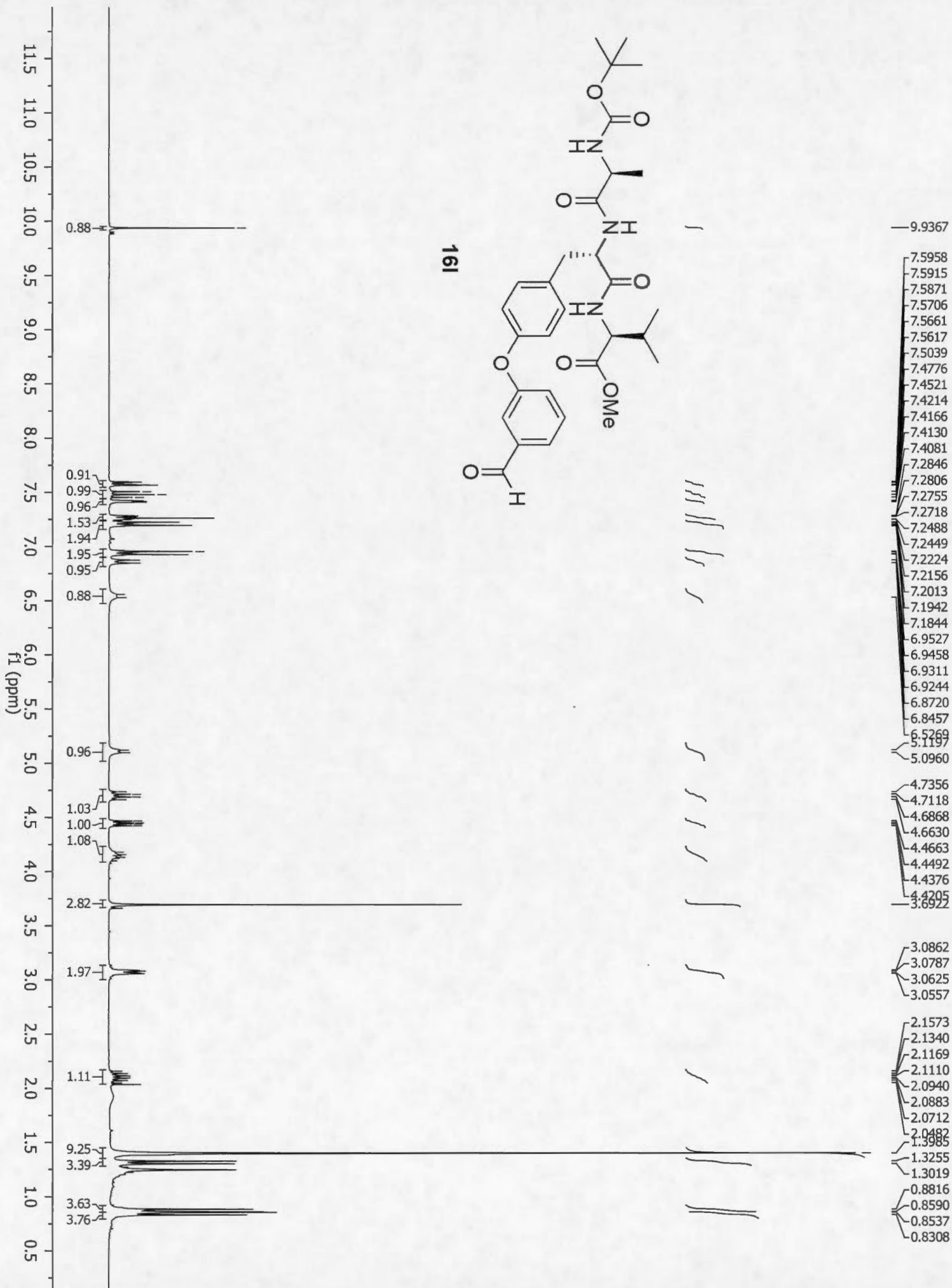
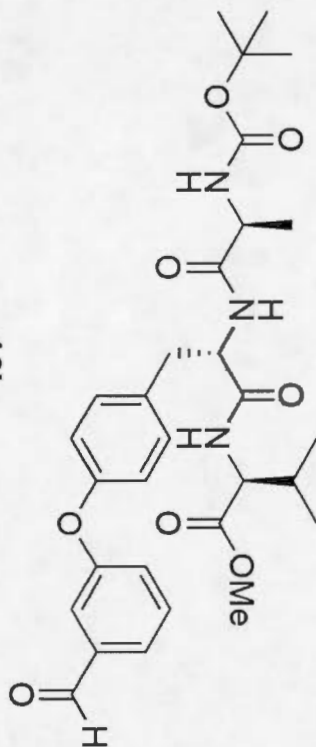
16k

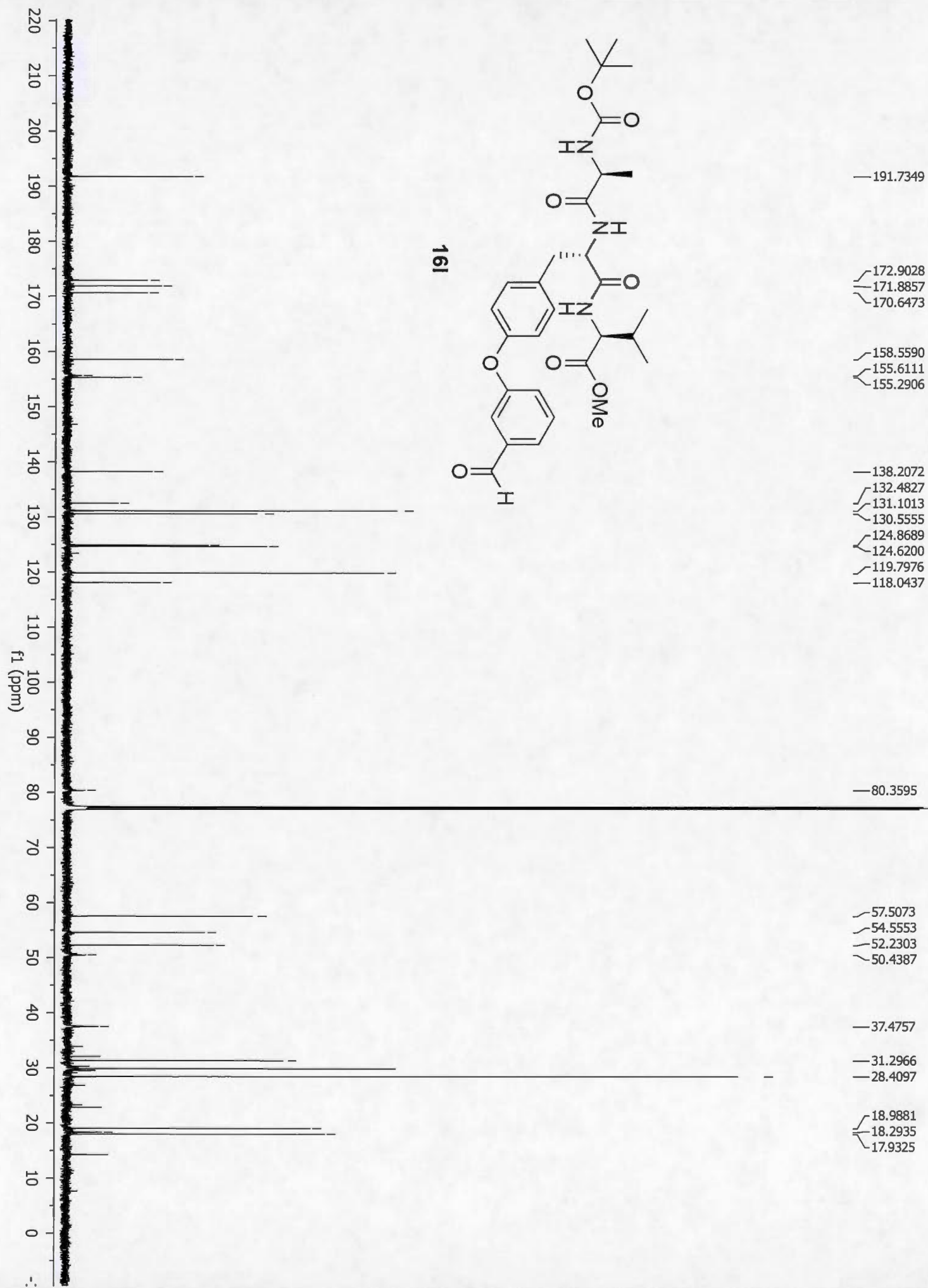


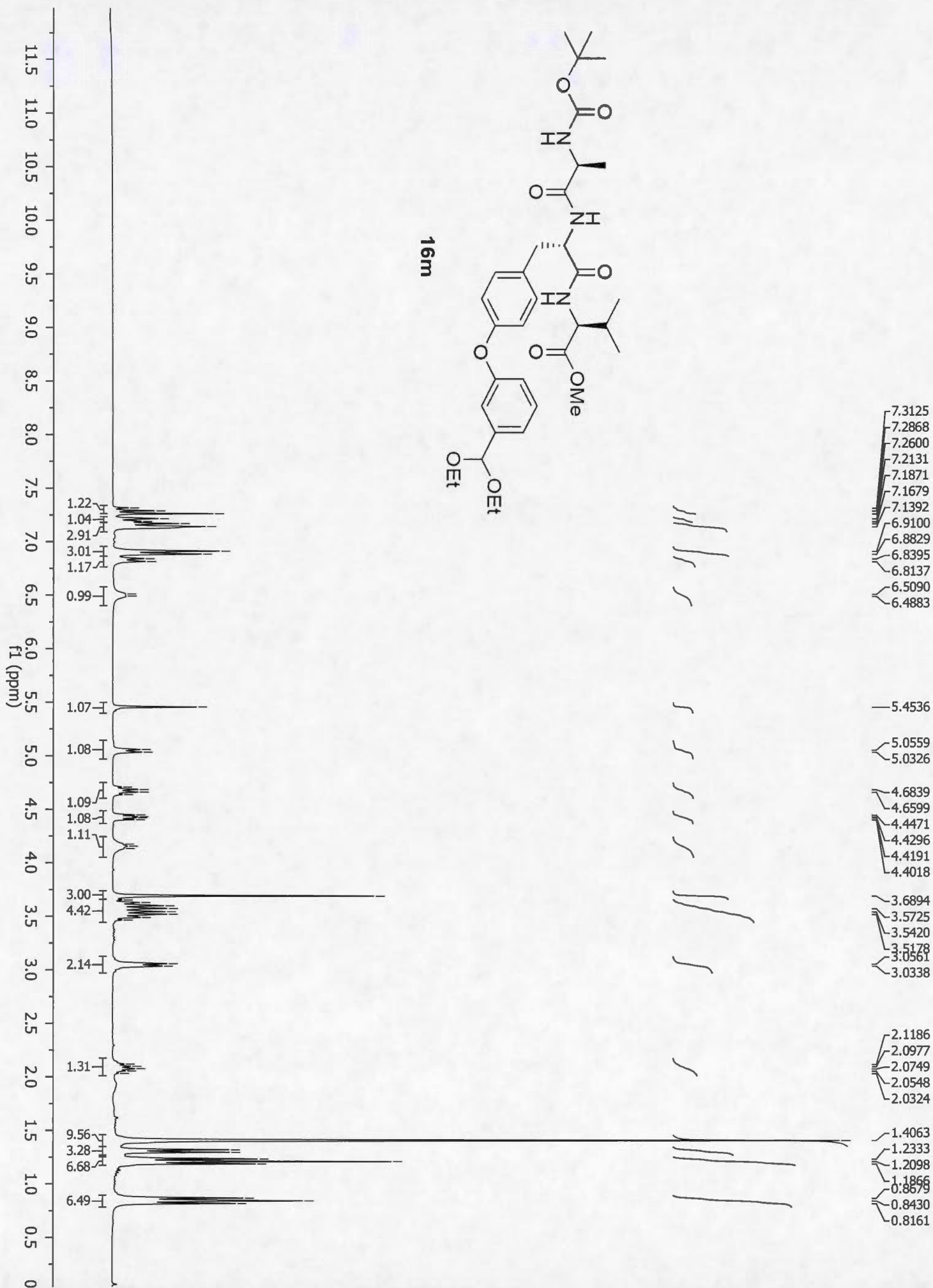
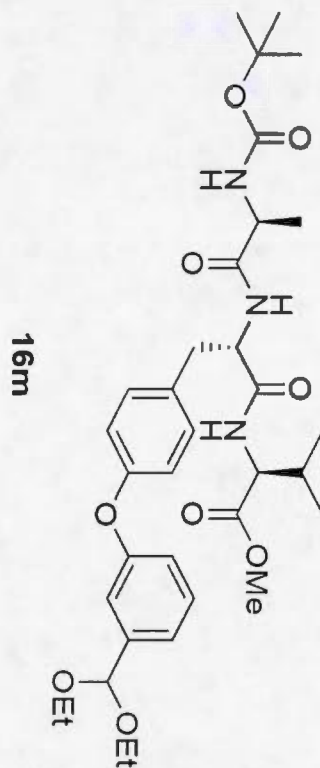


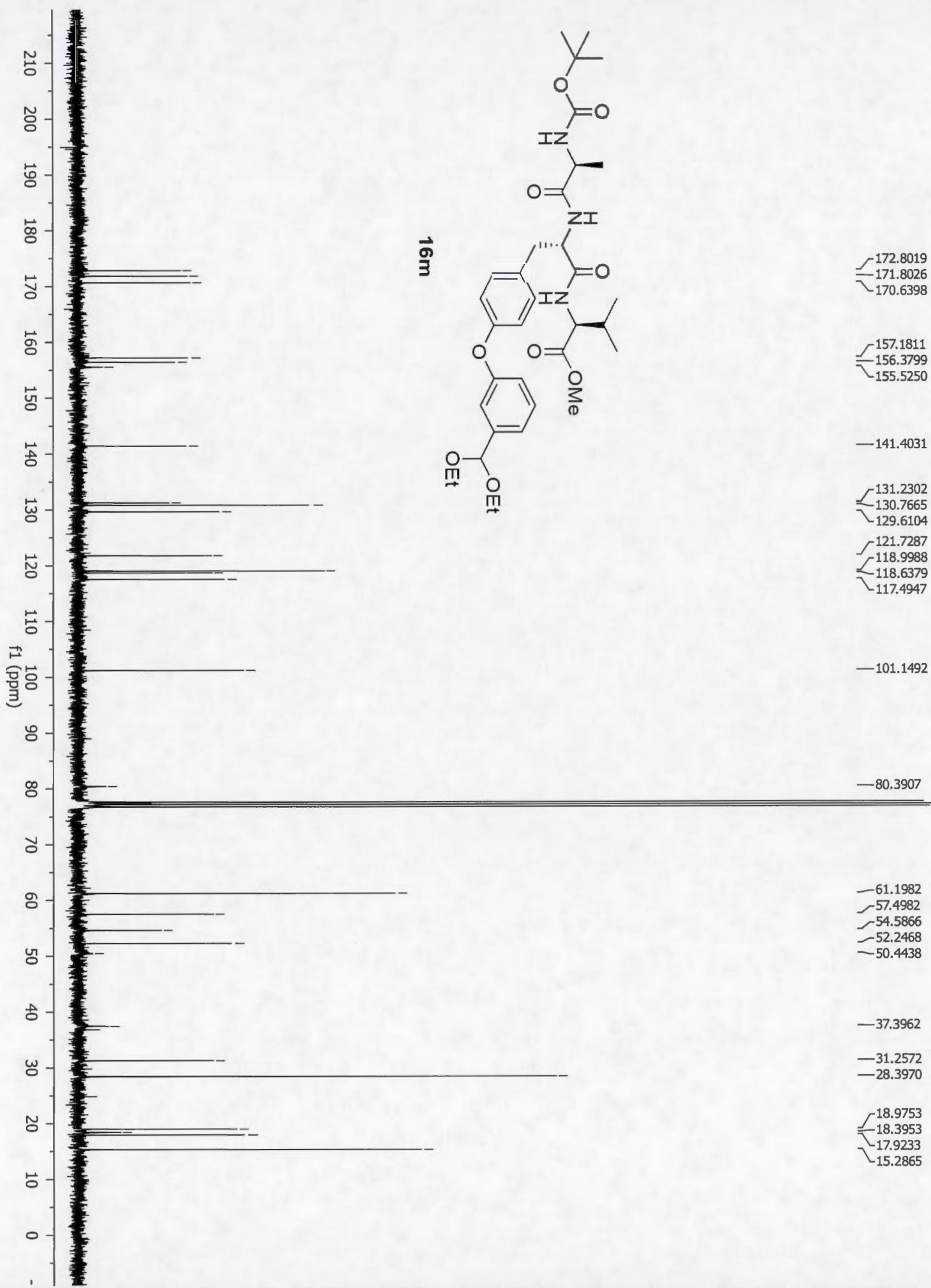


161



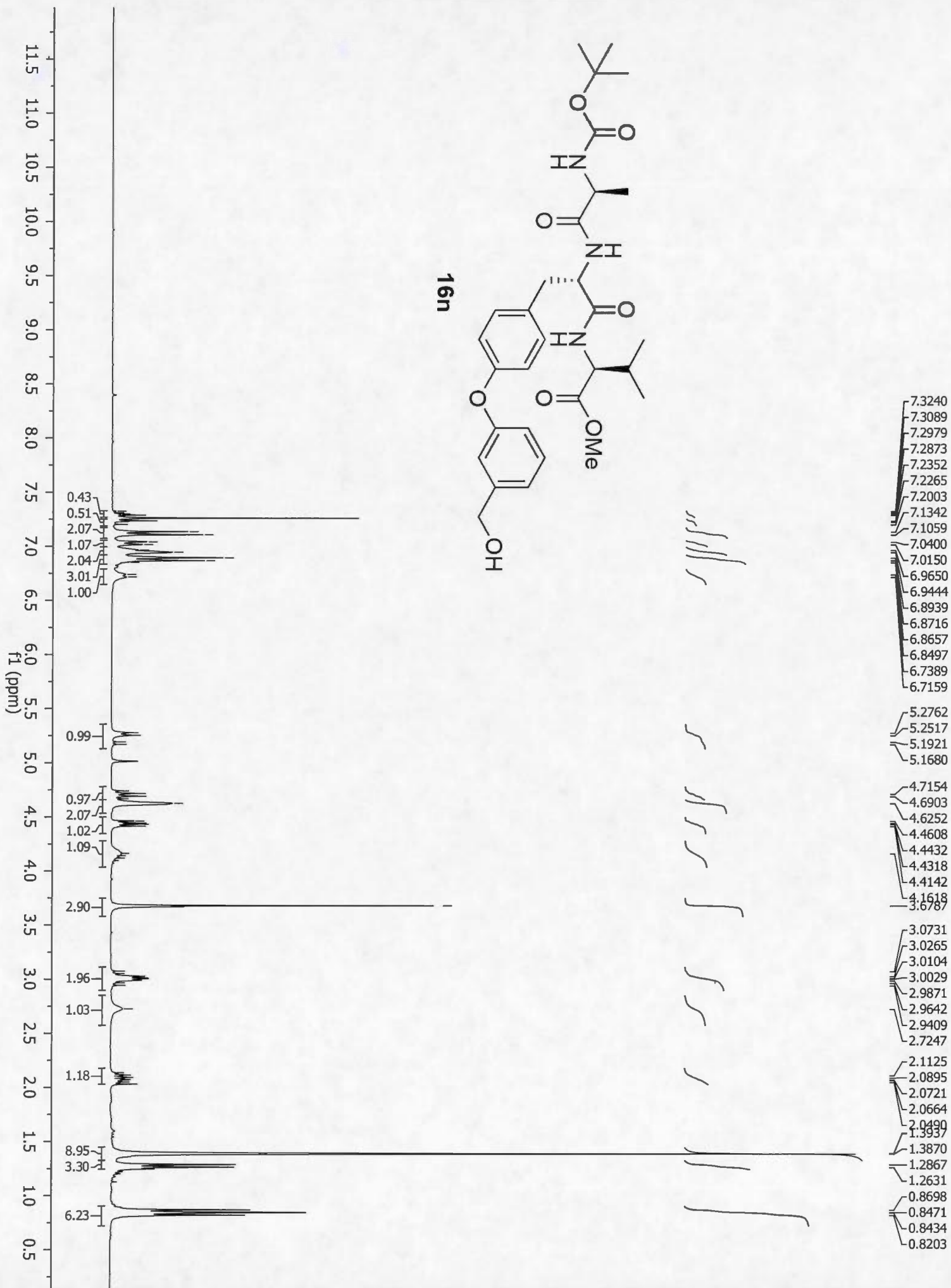
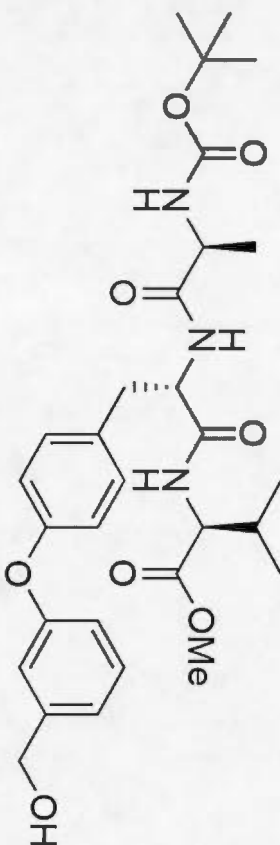


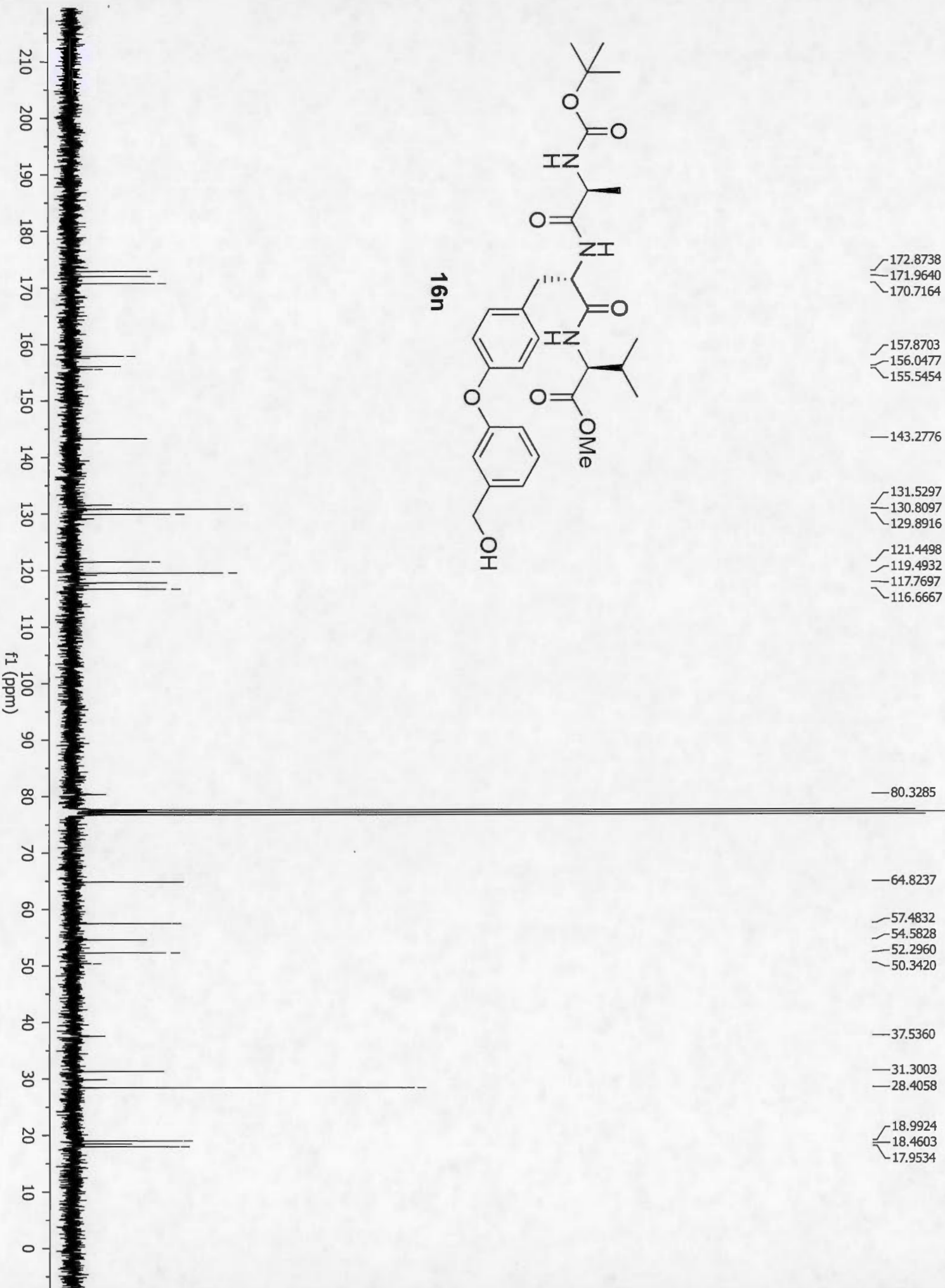


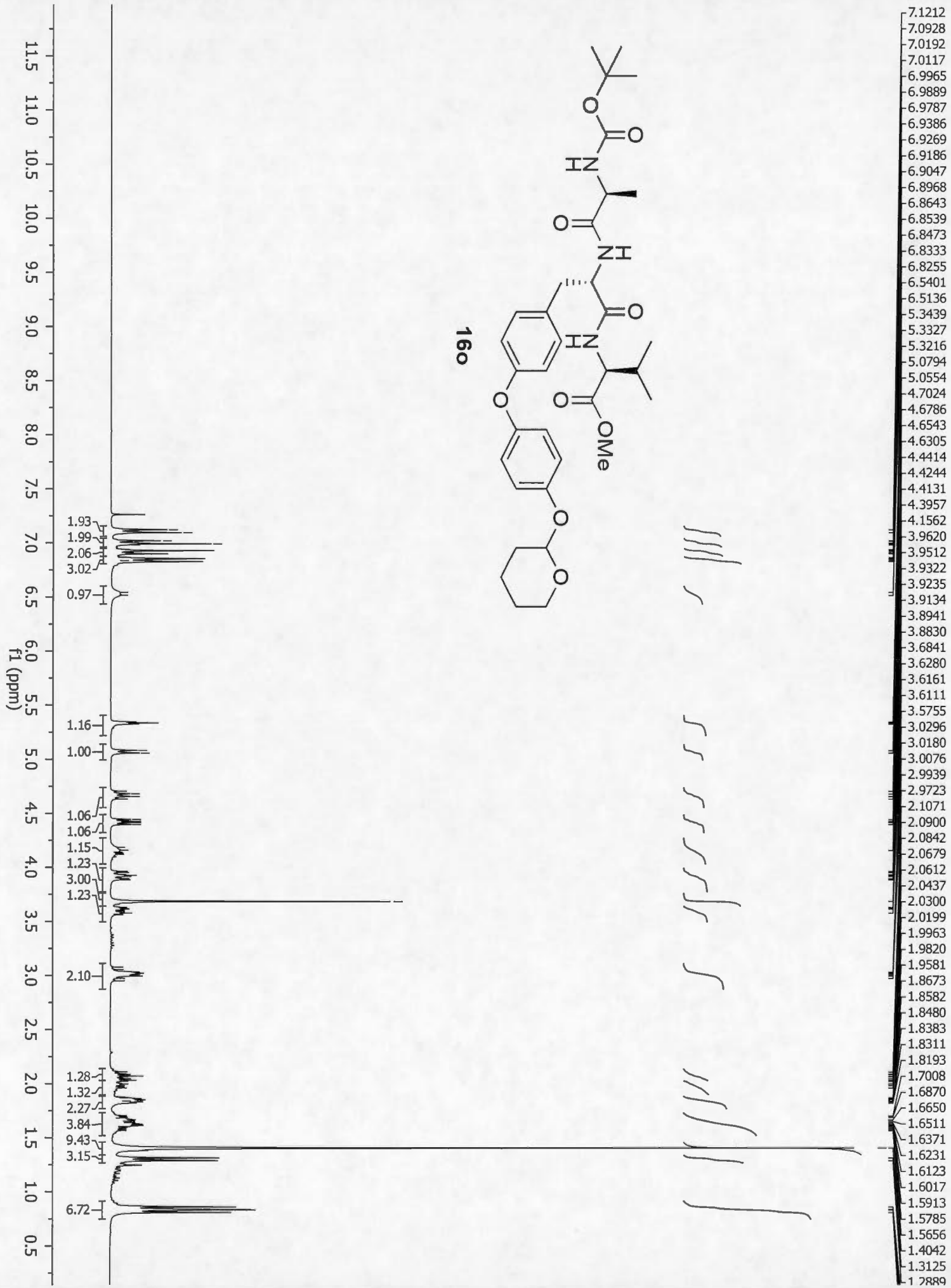
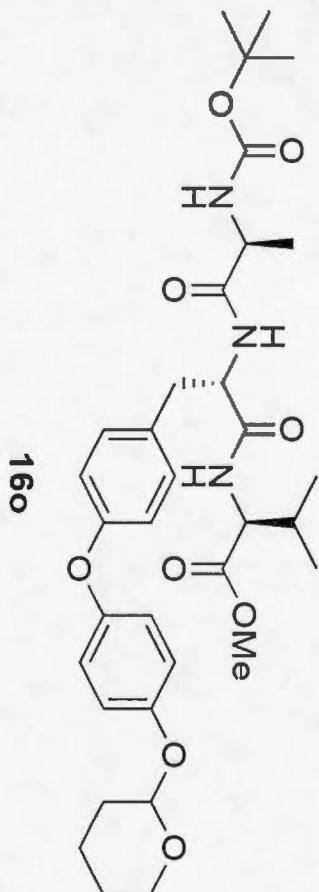




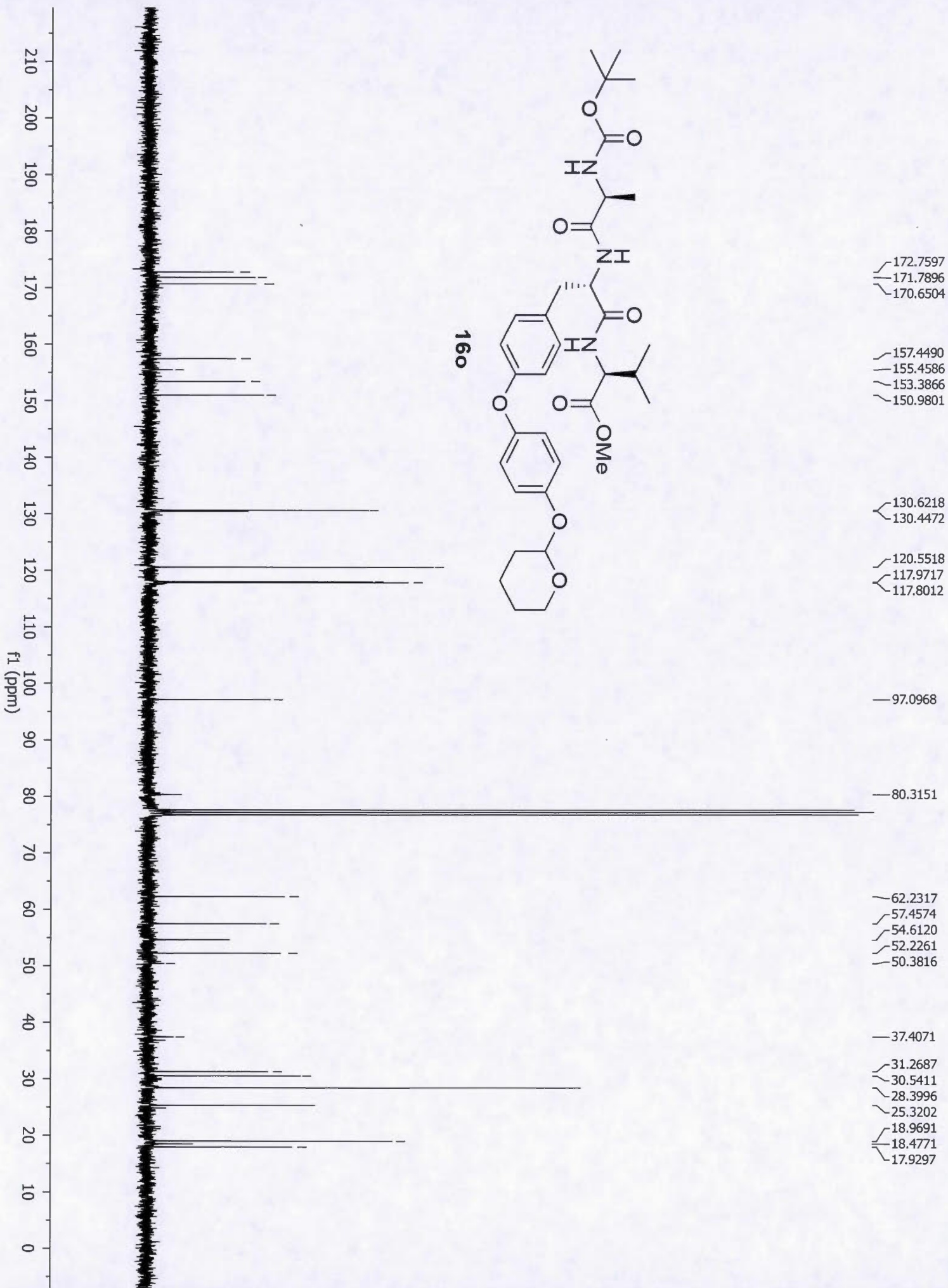
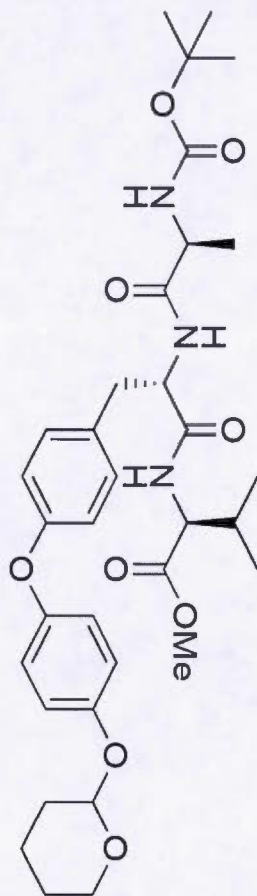
16n



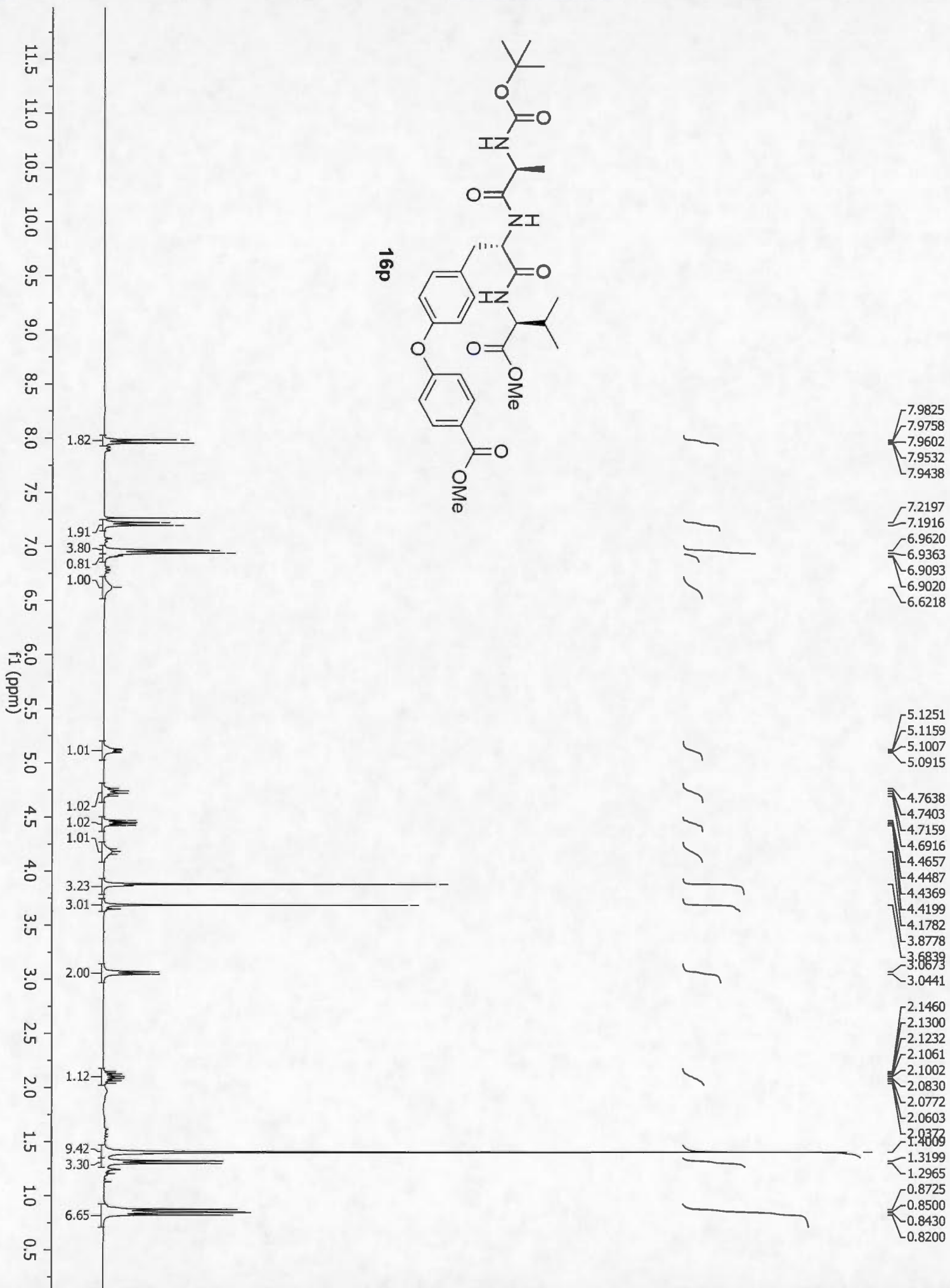
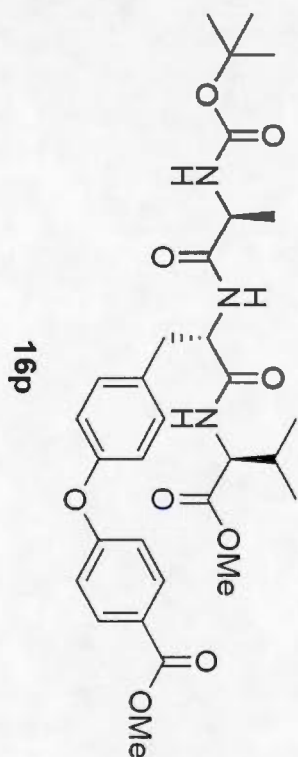




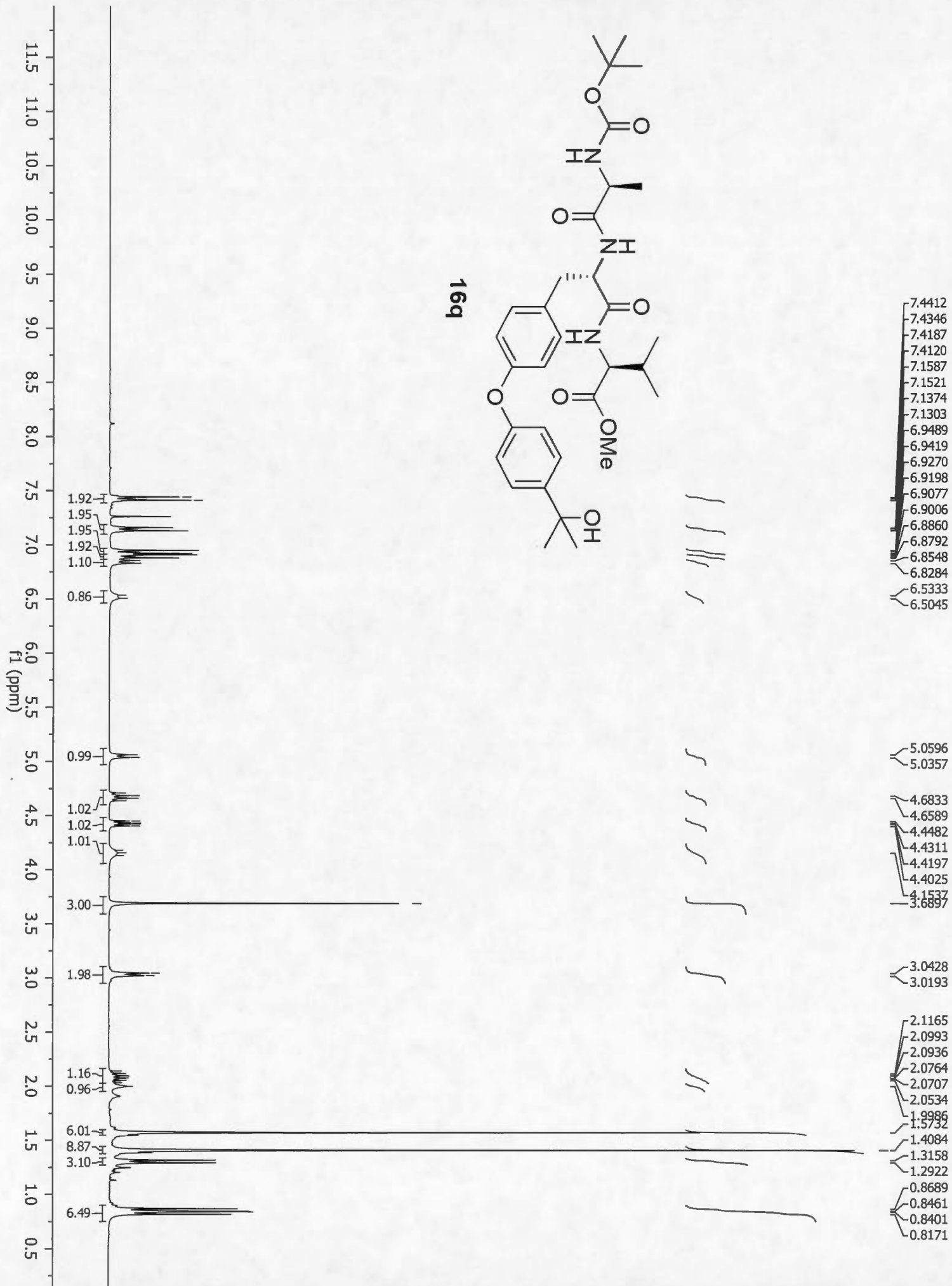
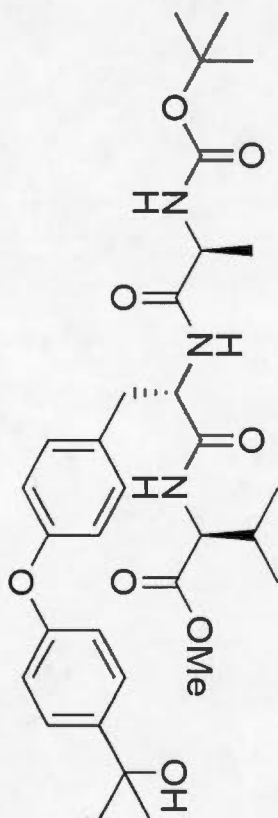
160



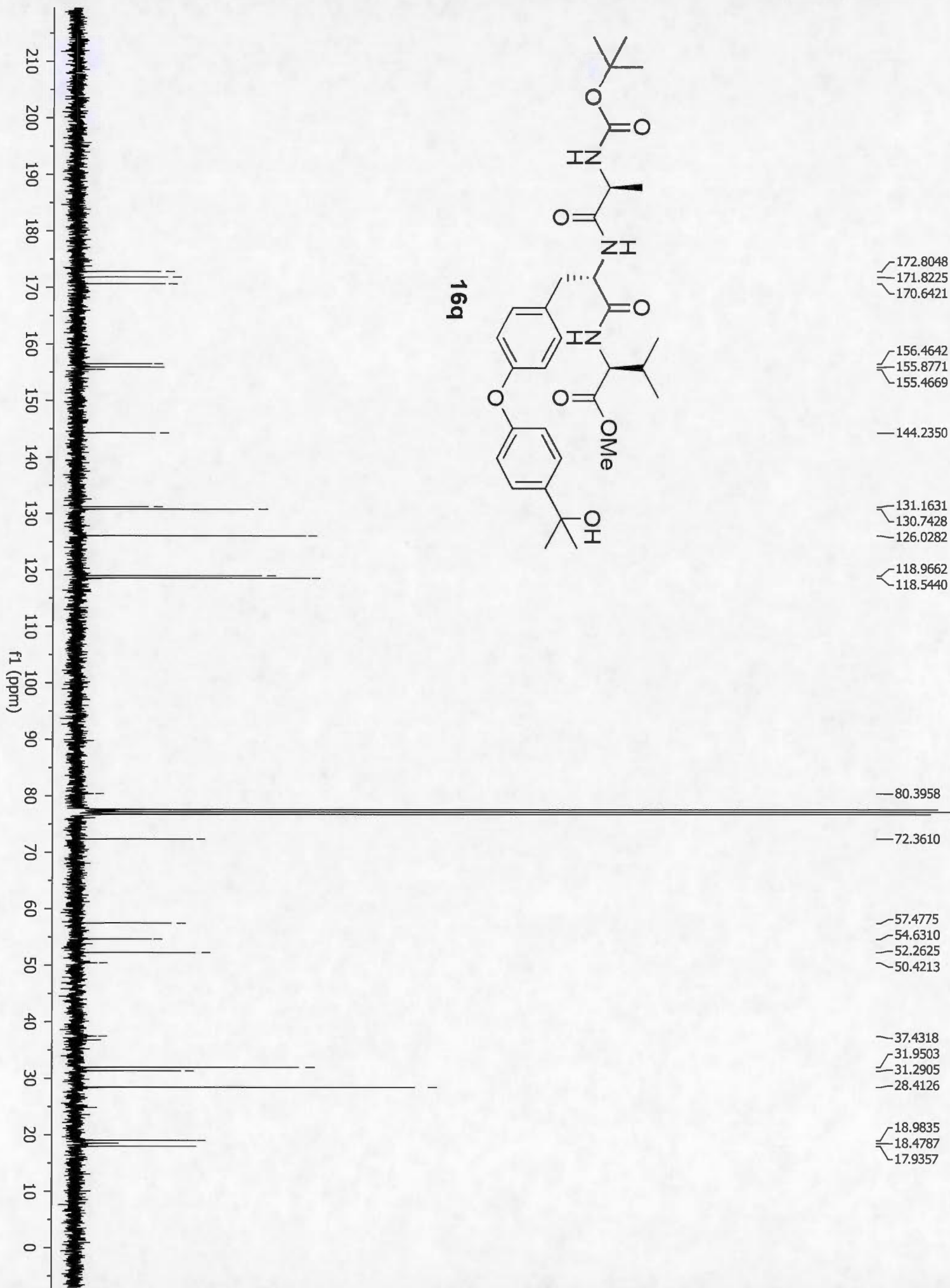
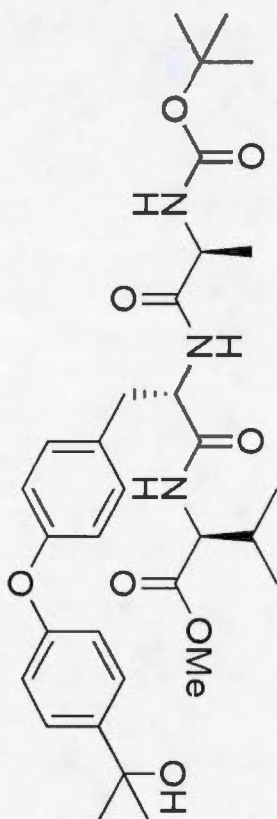






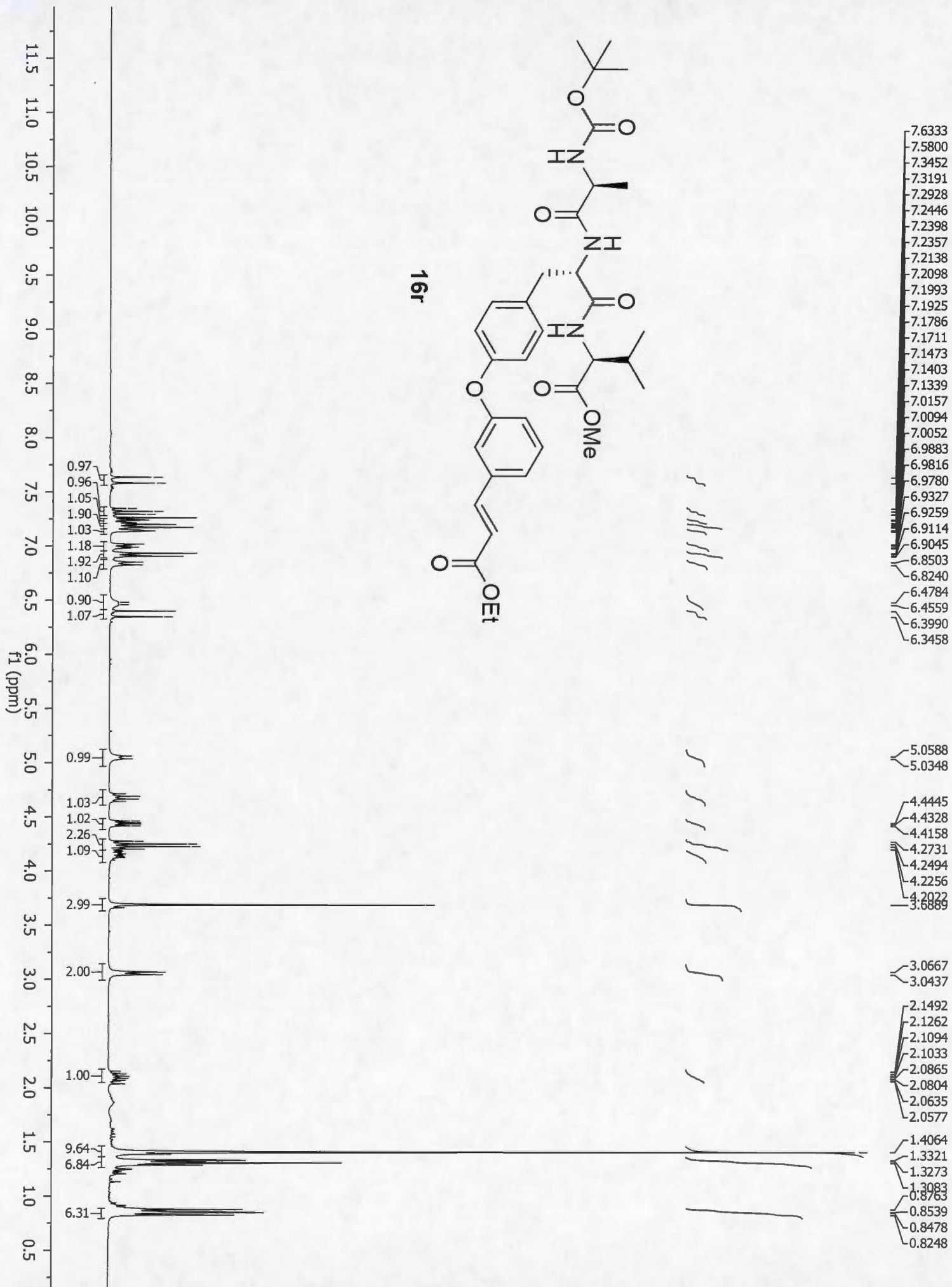
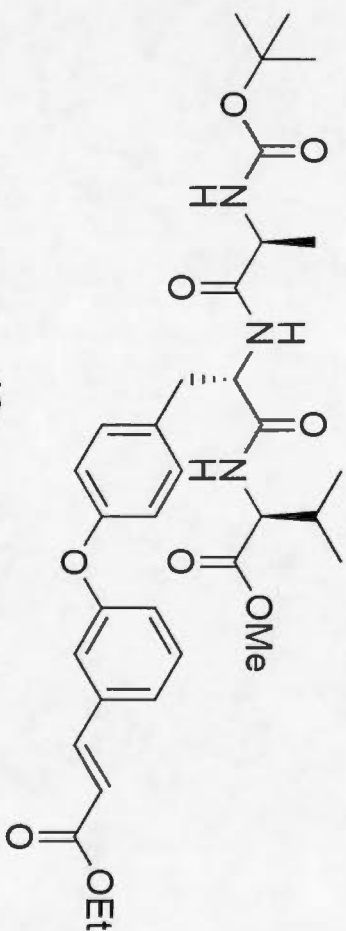


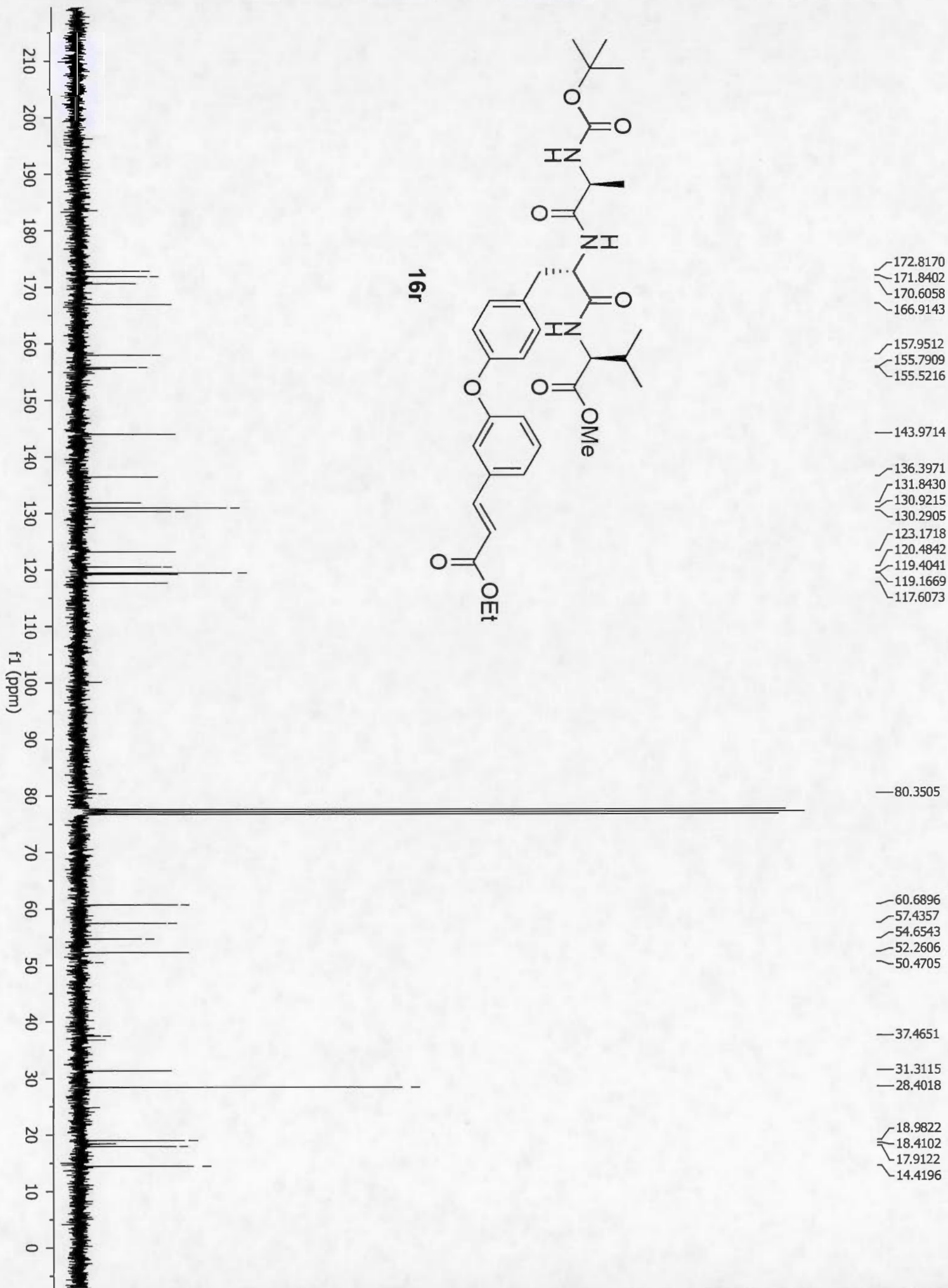
16q



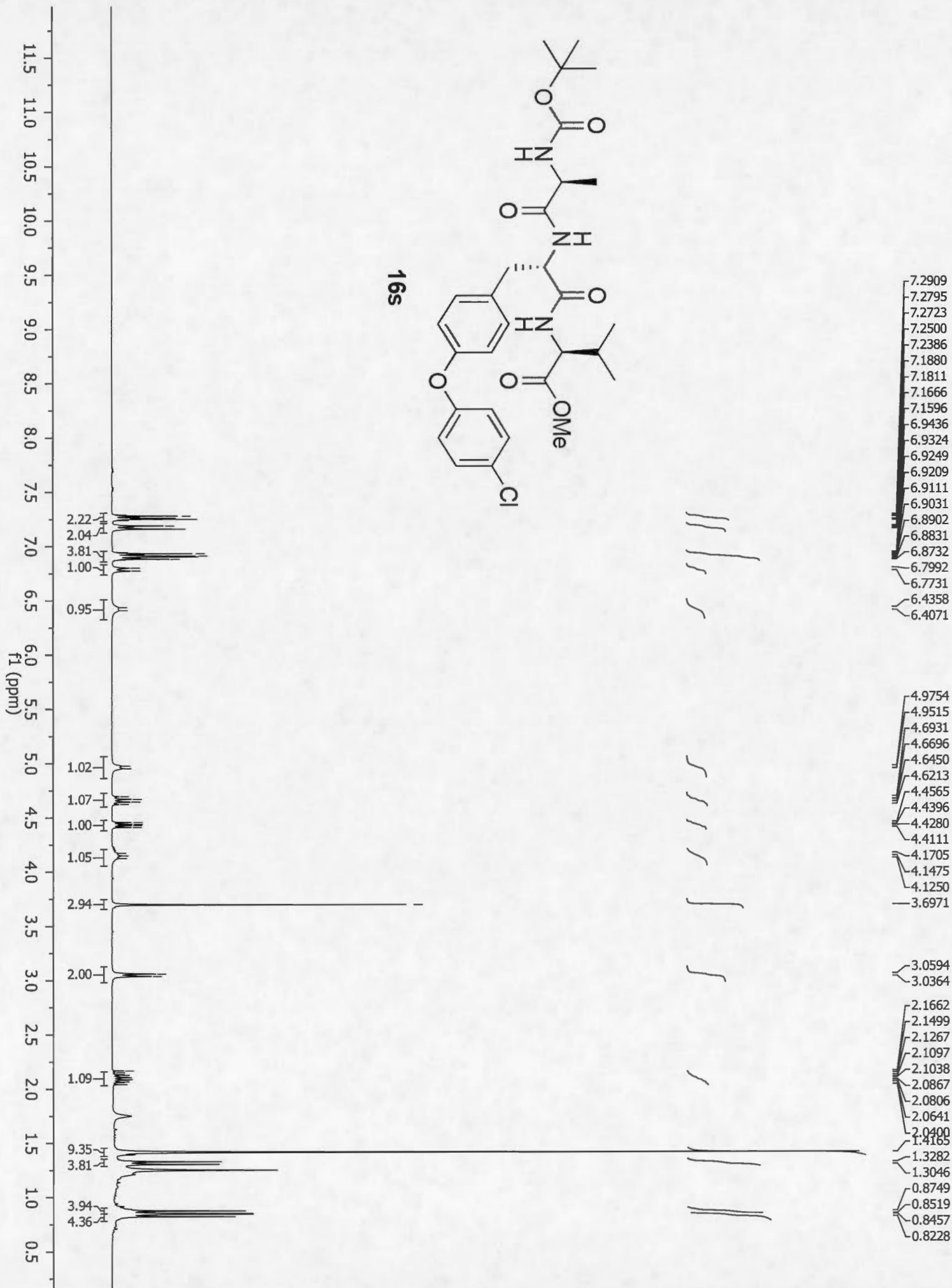
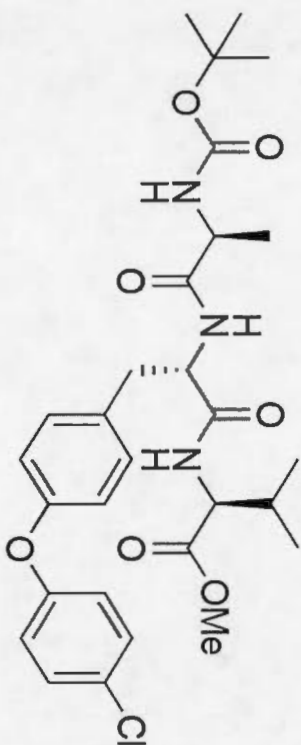


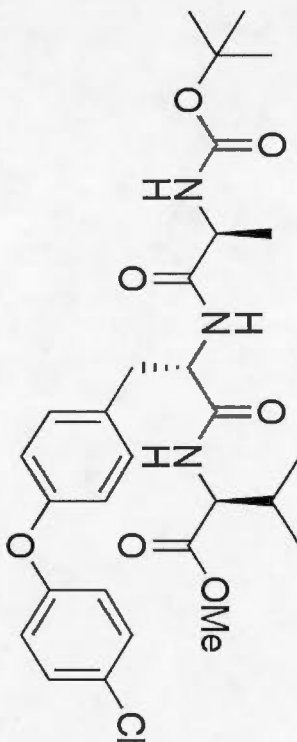
16r



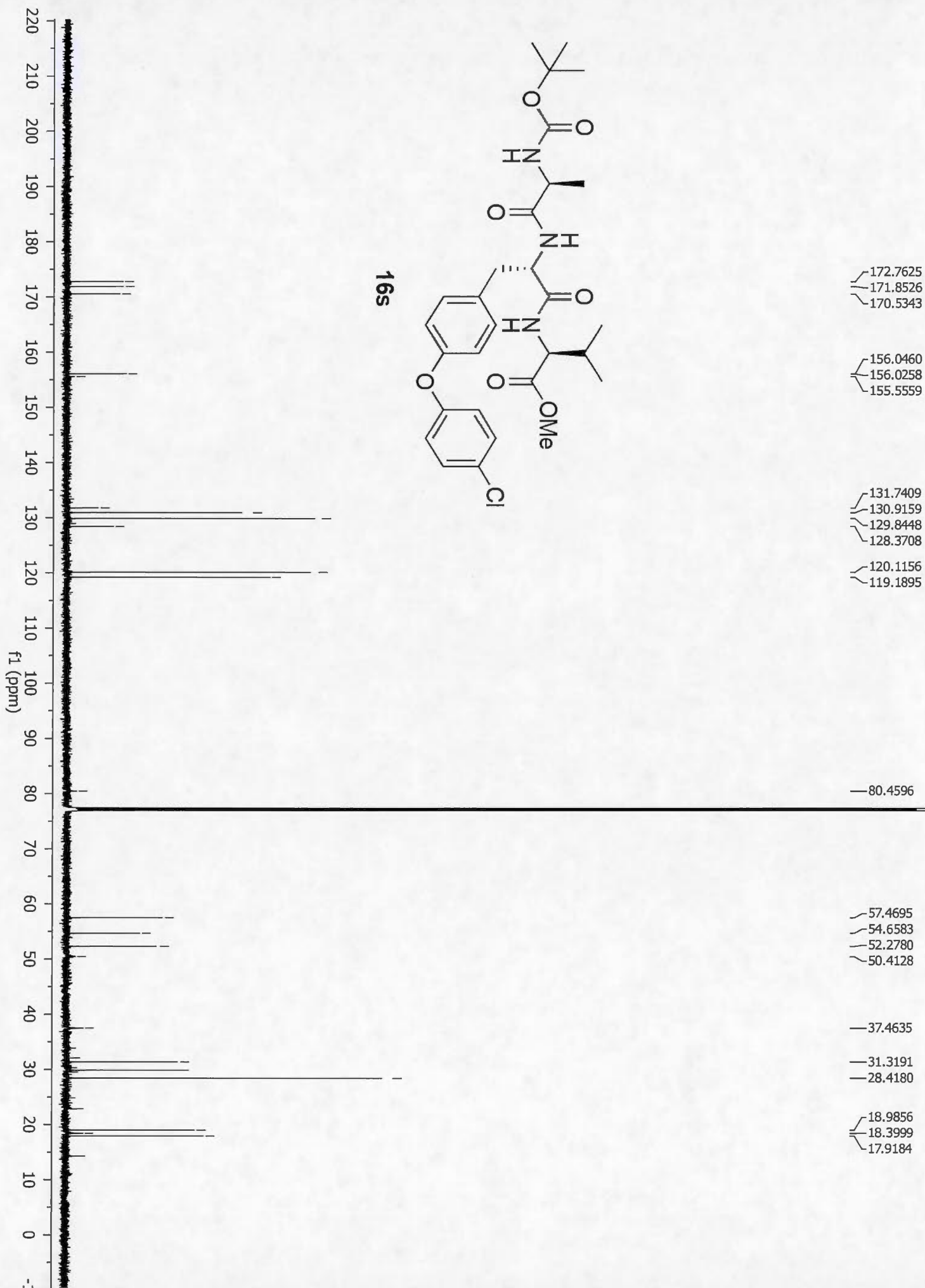


16s



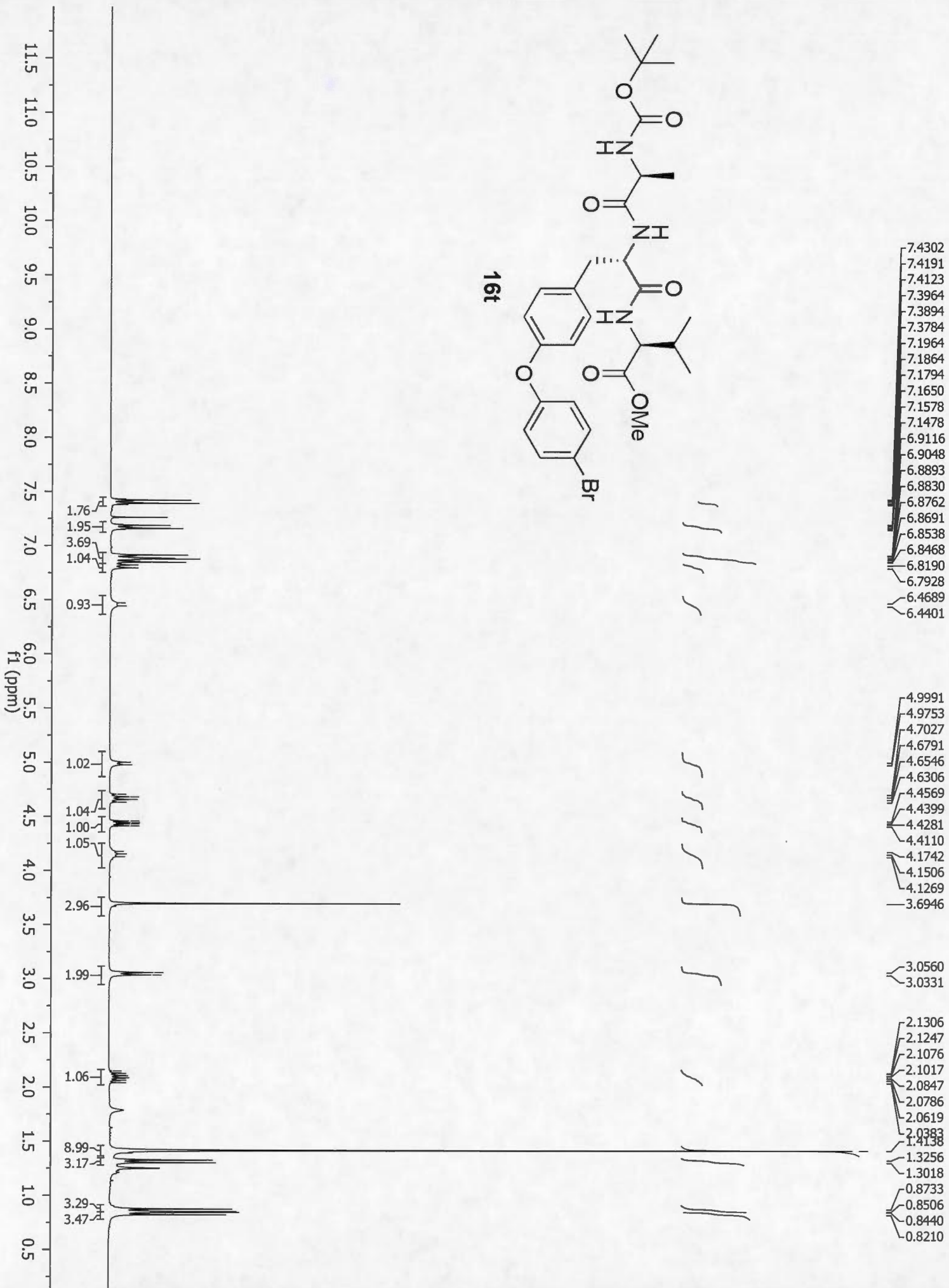
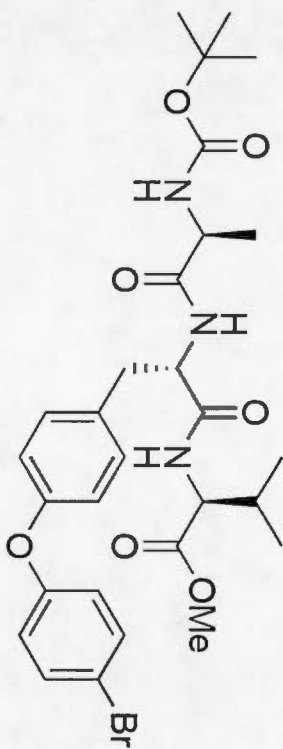


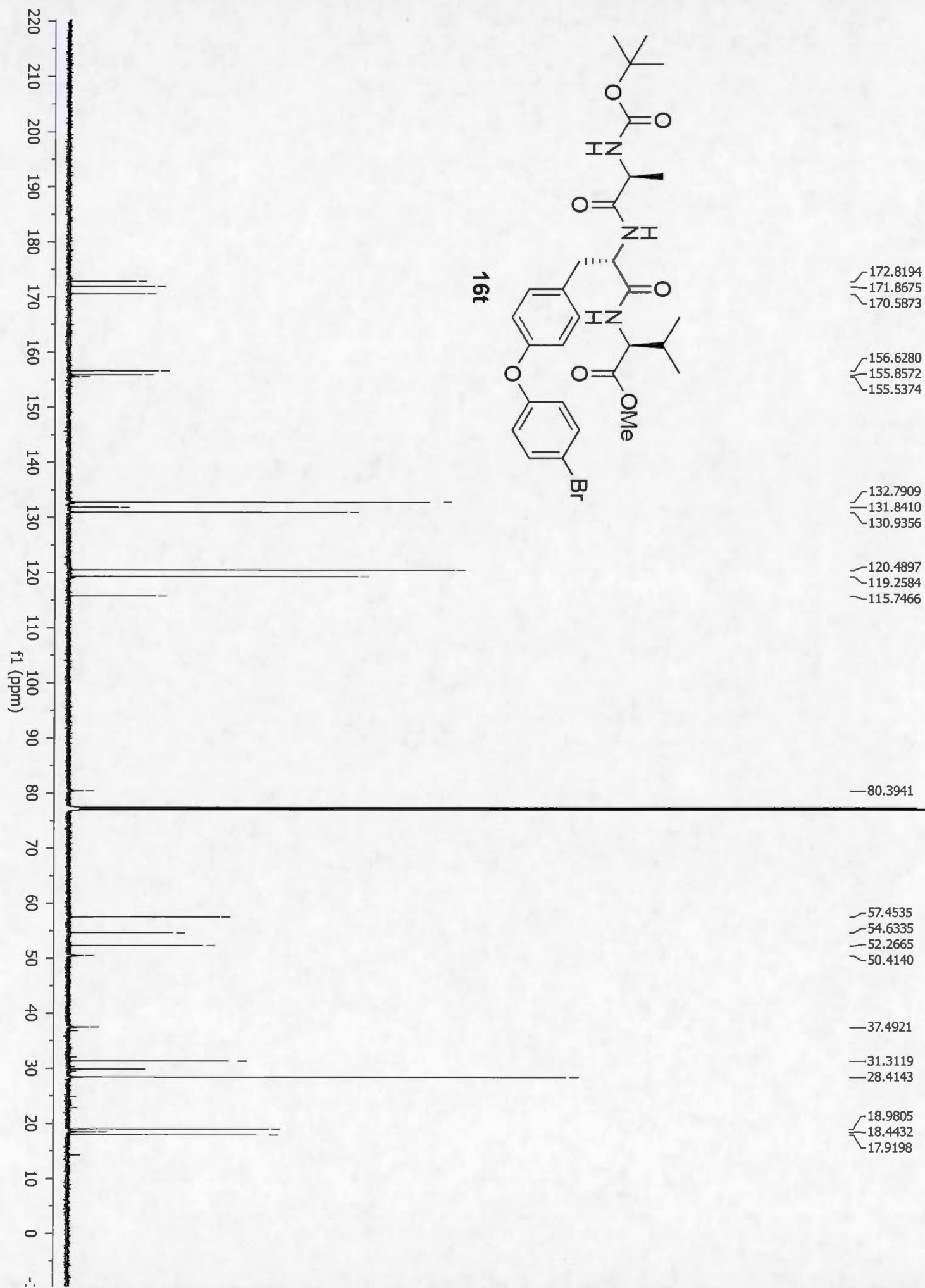
16s



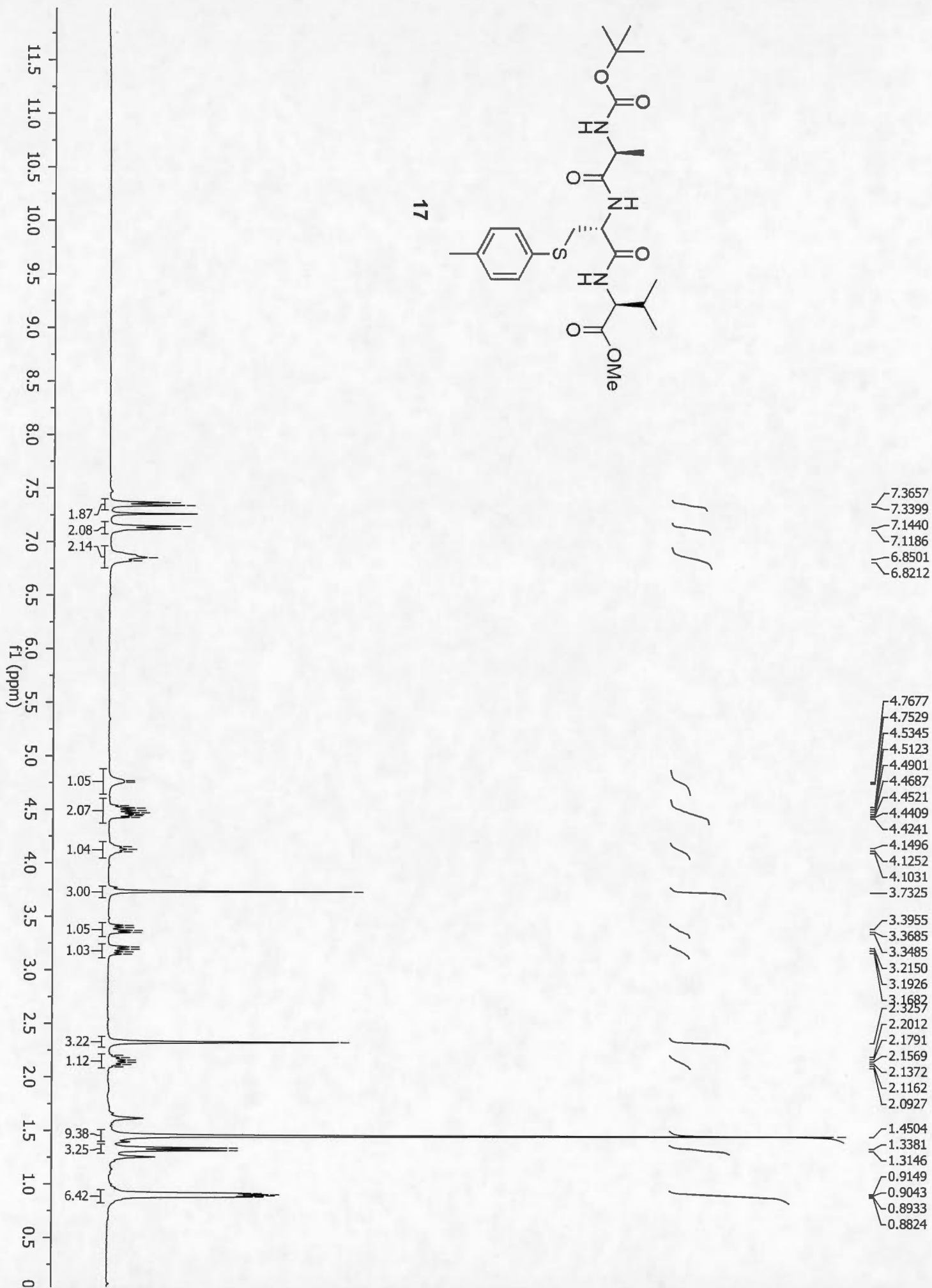
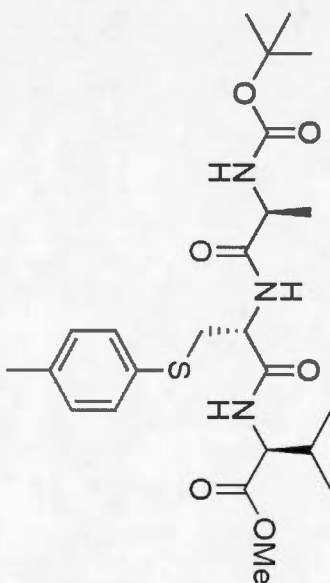


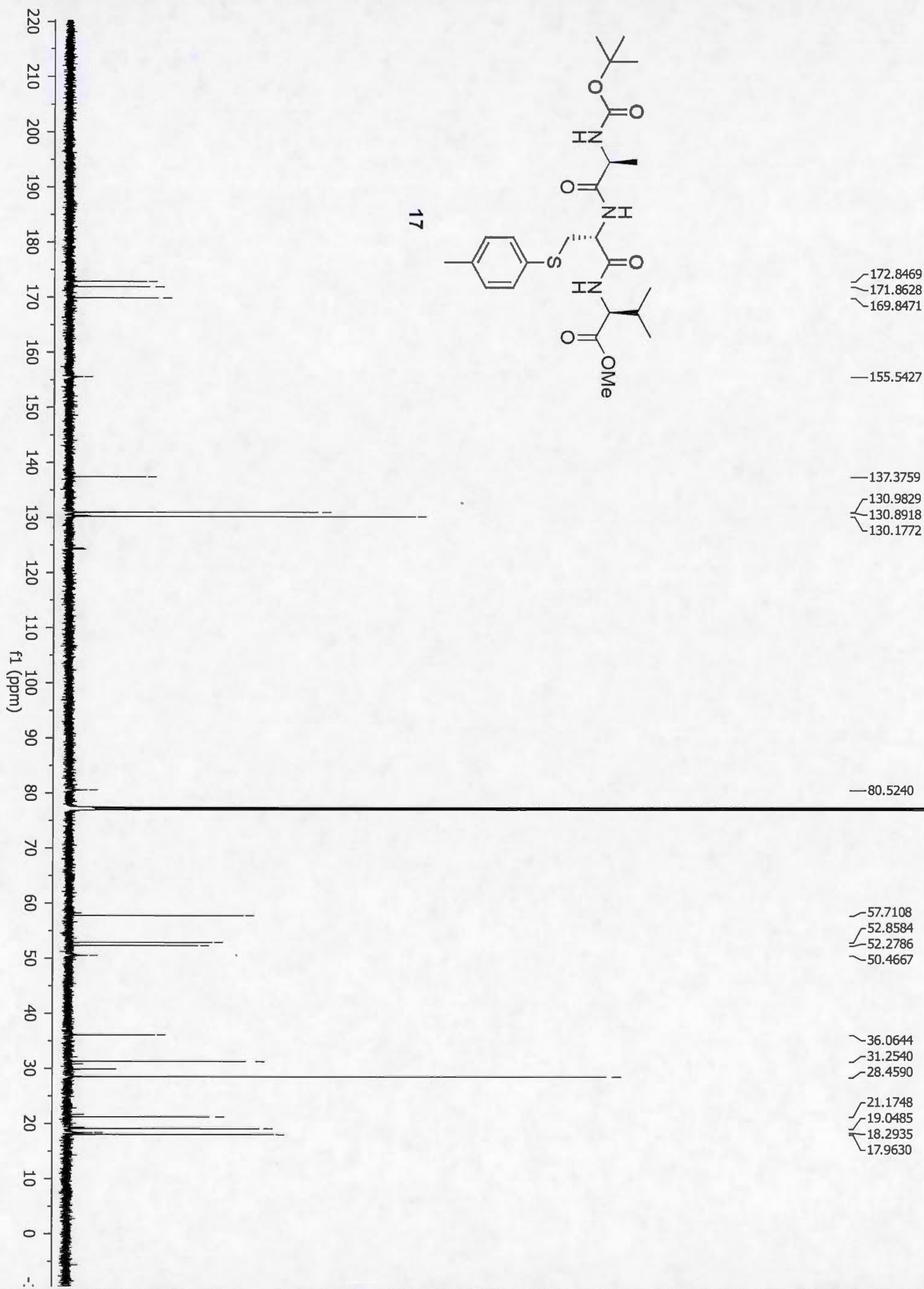
16t



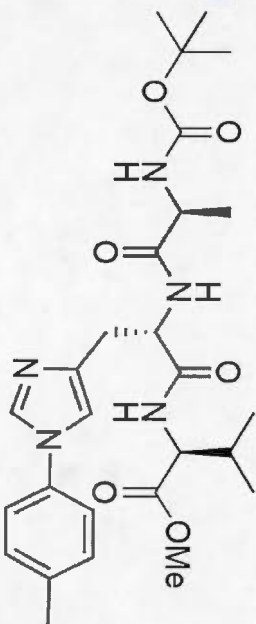


17

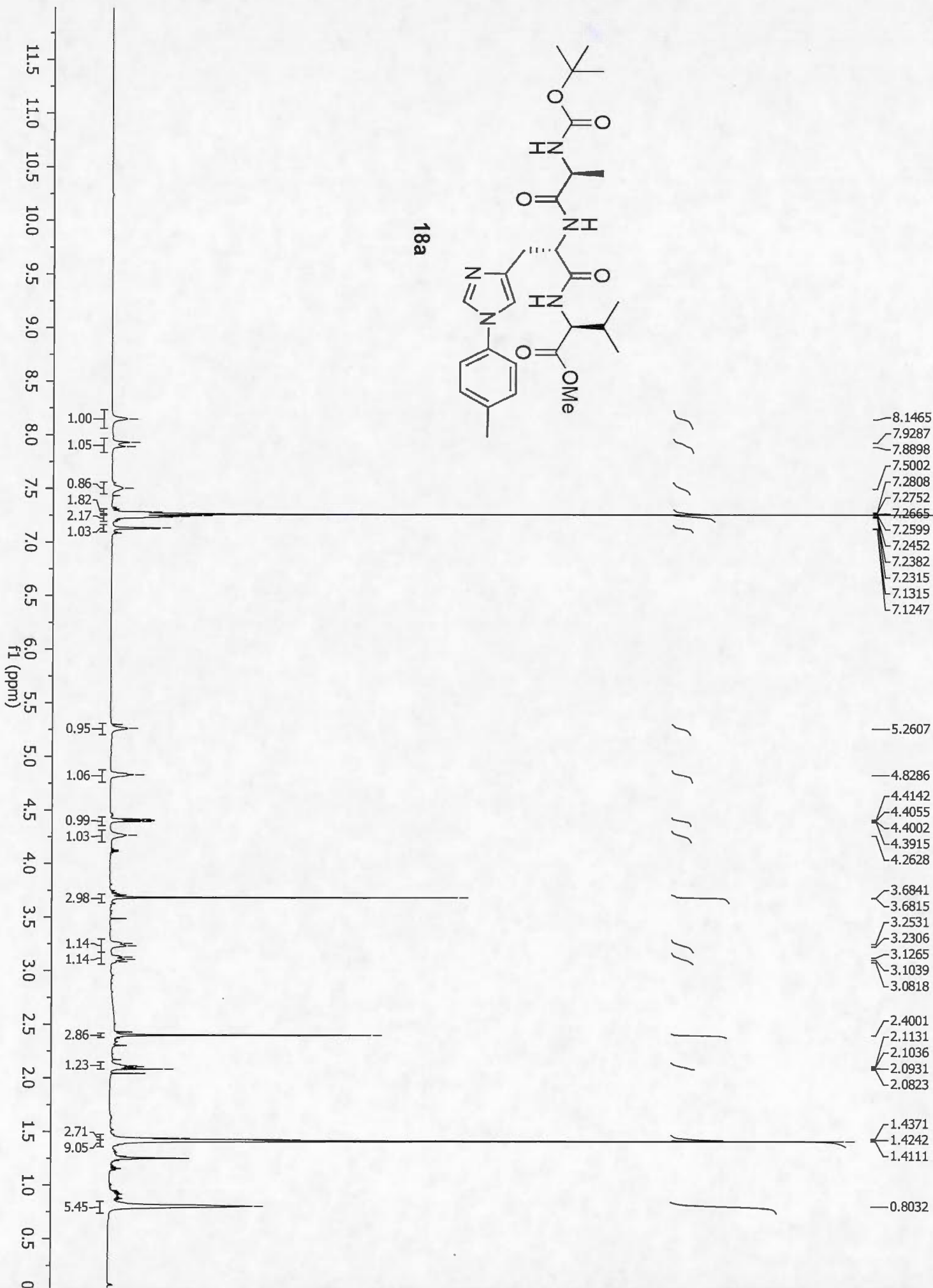


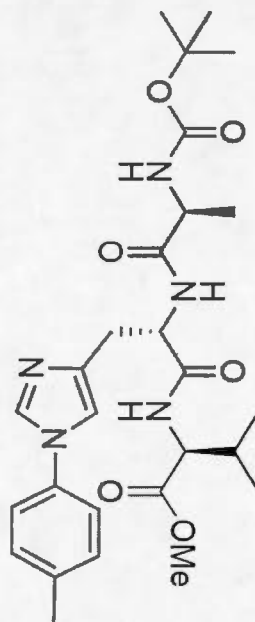




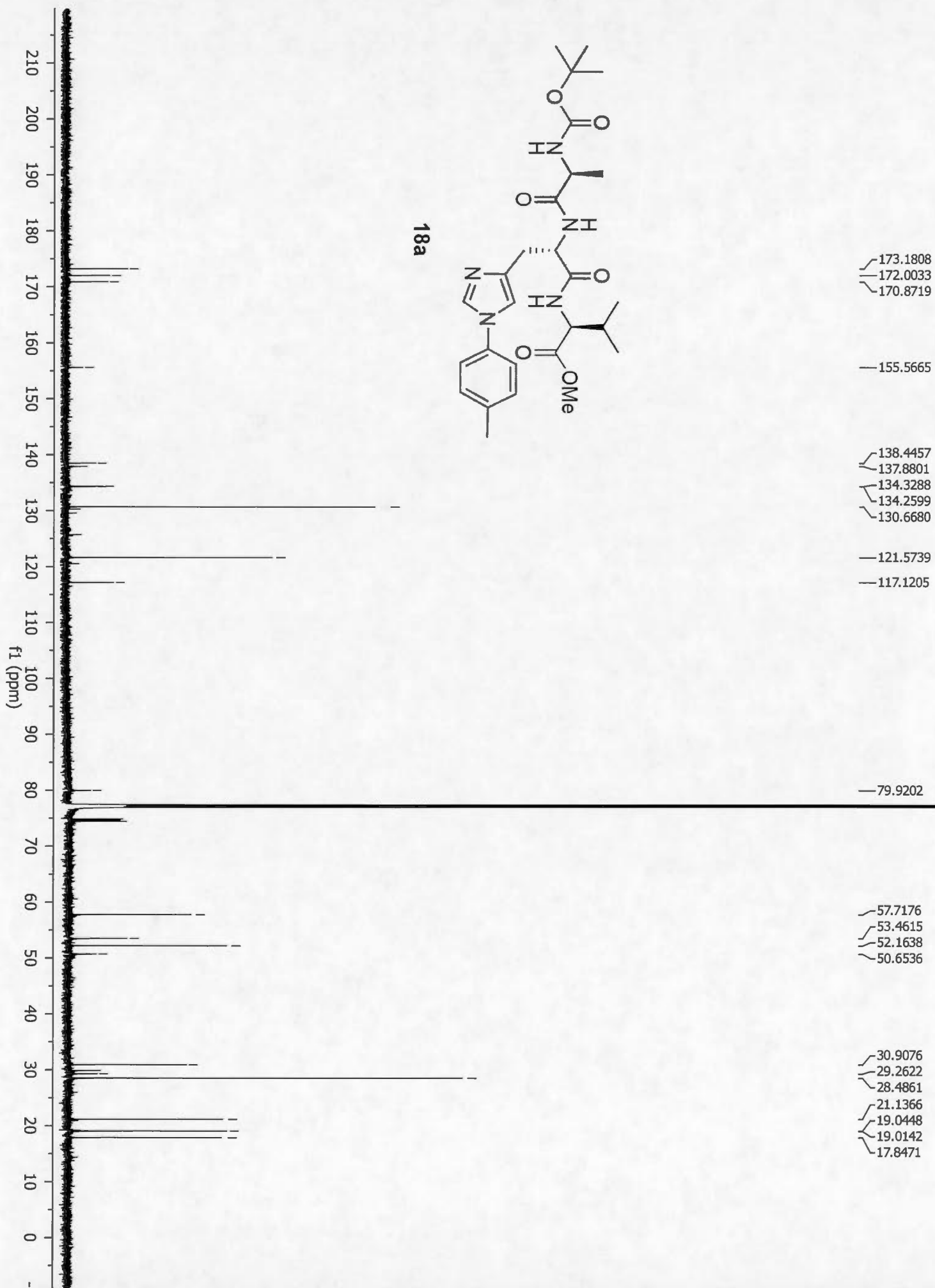


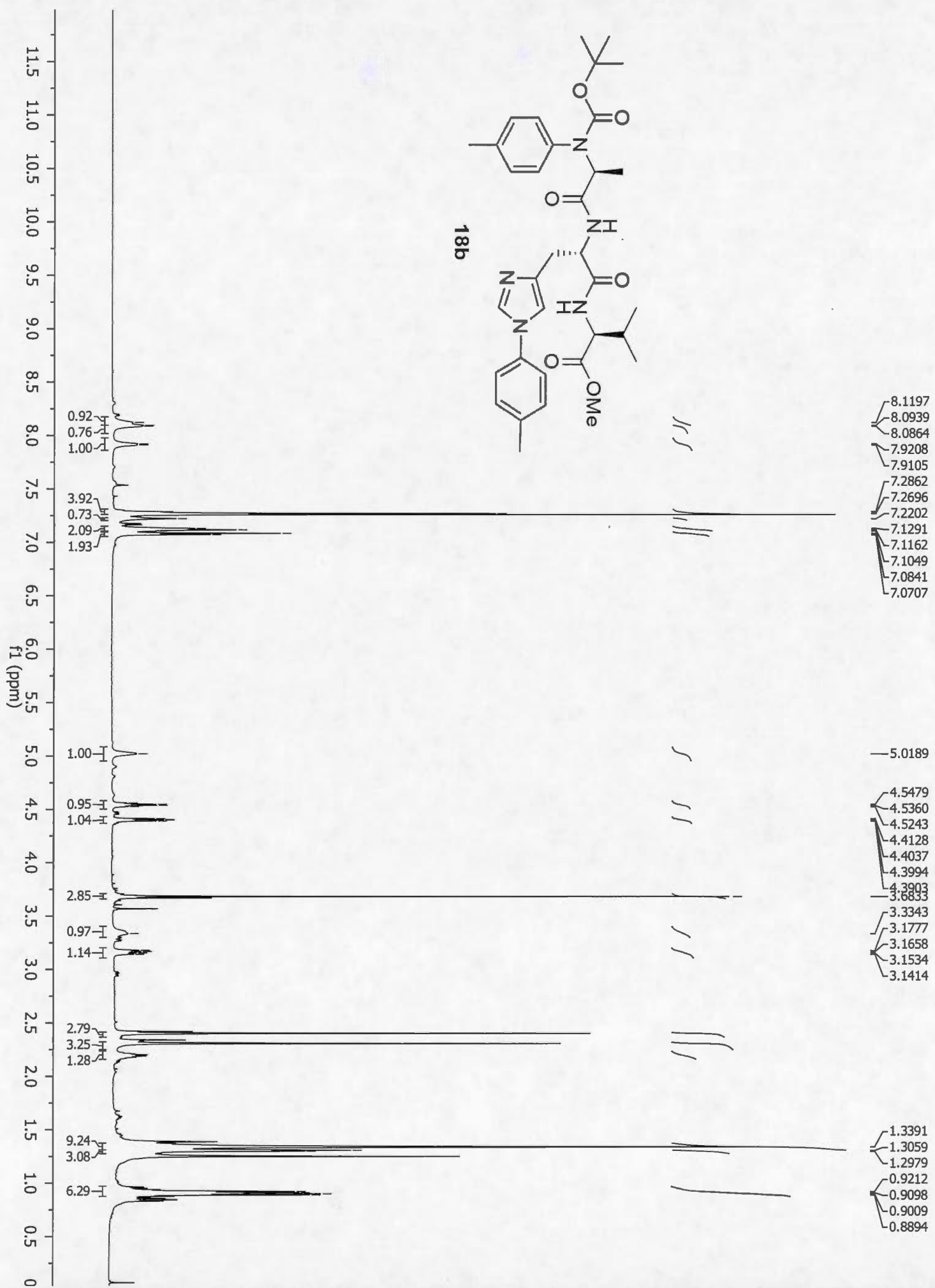
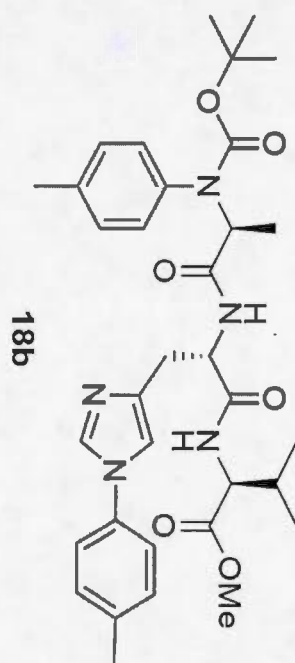
18a

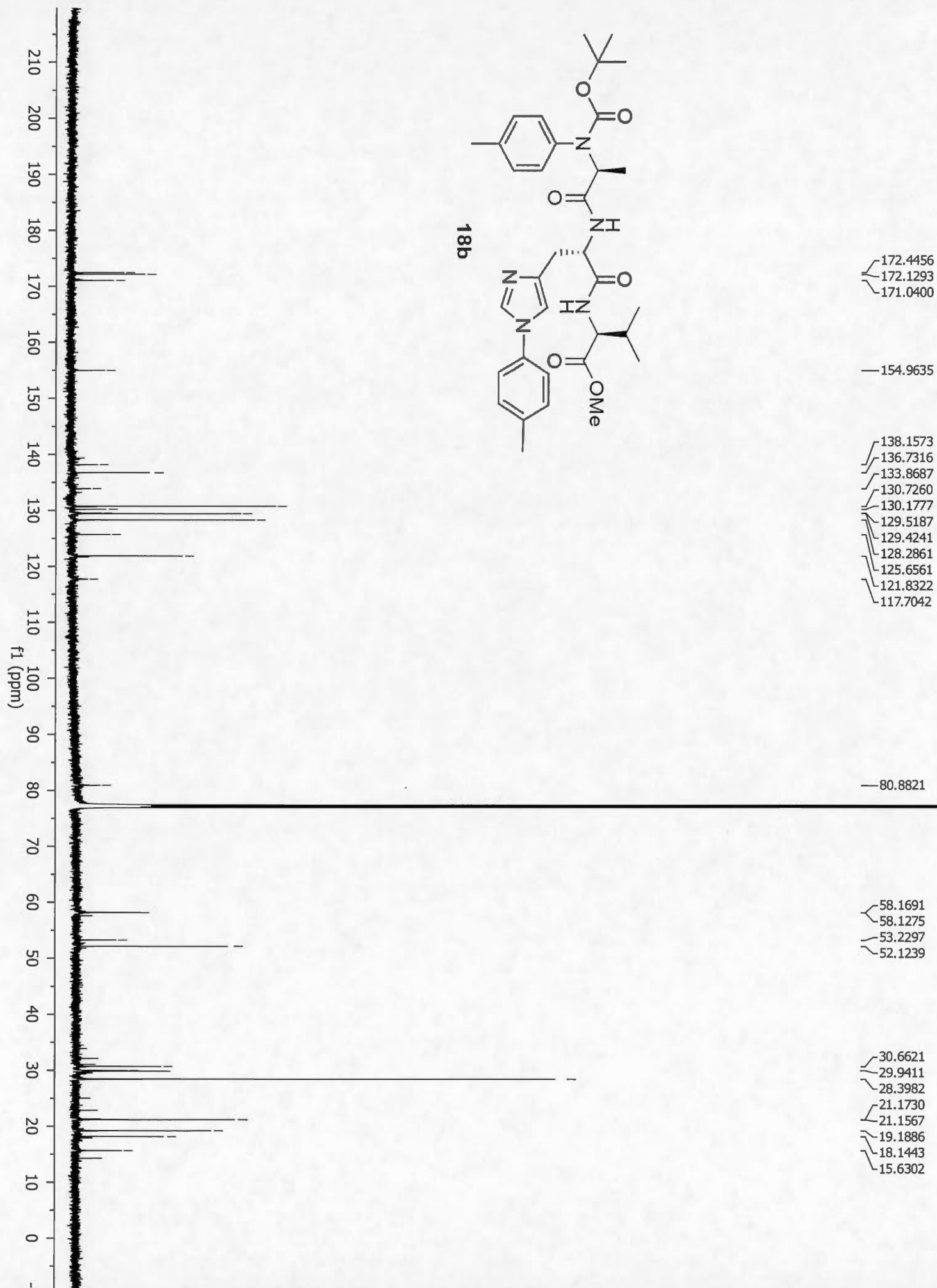




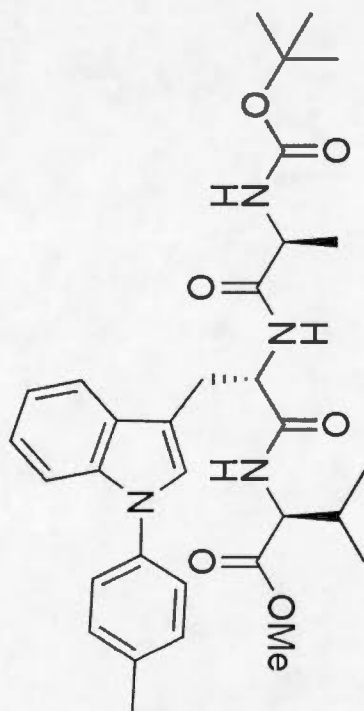
18a



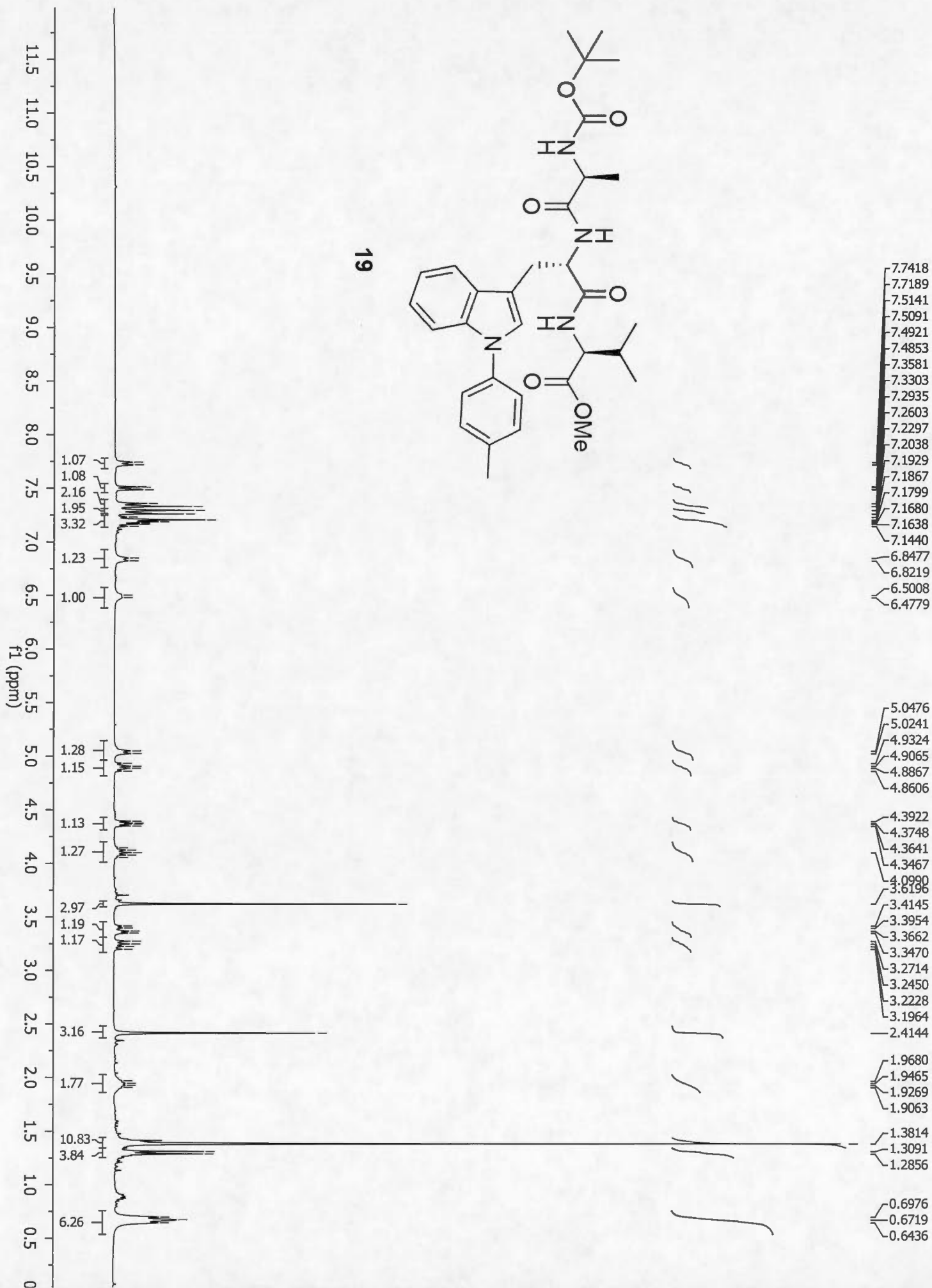


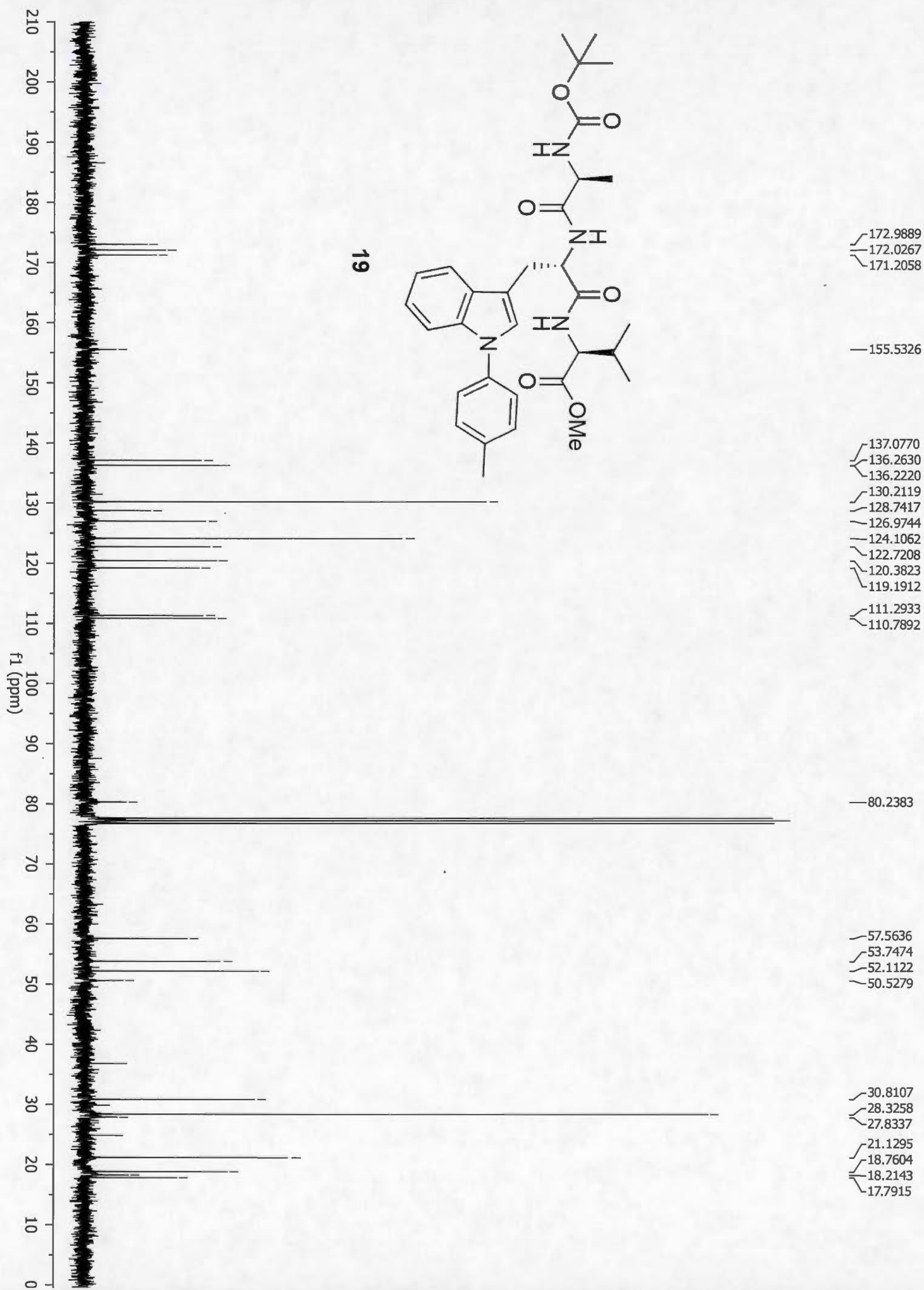


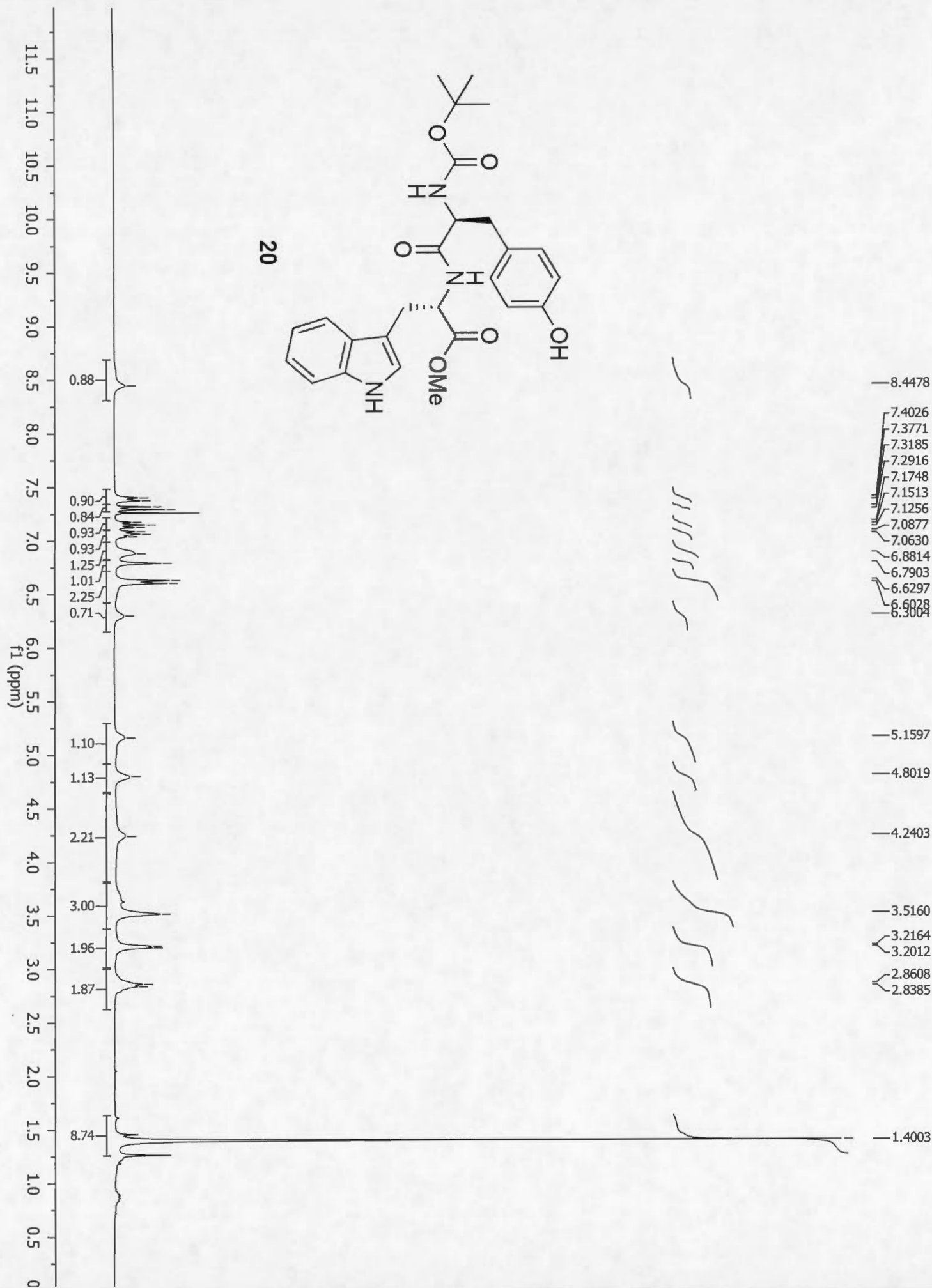


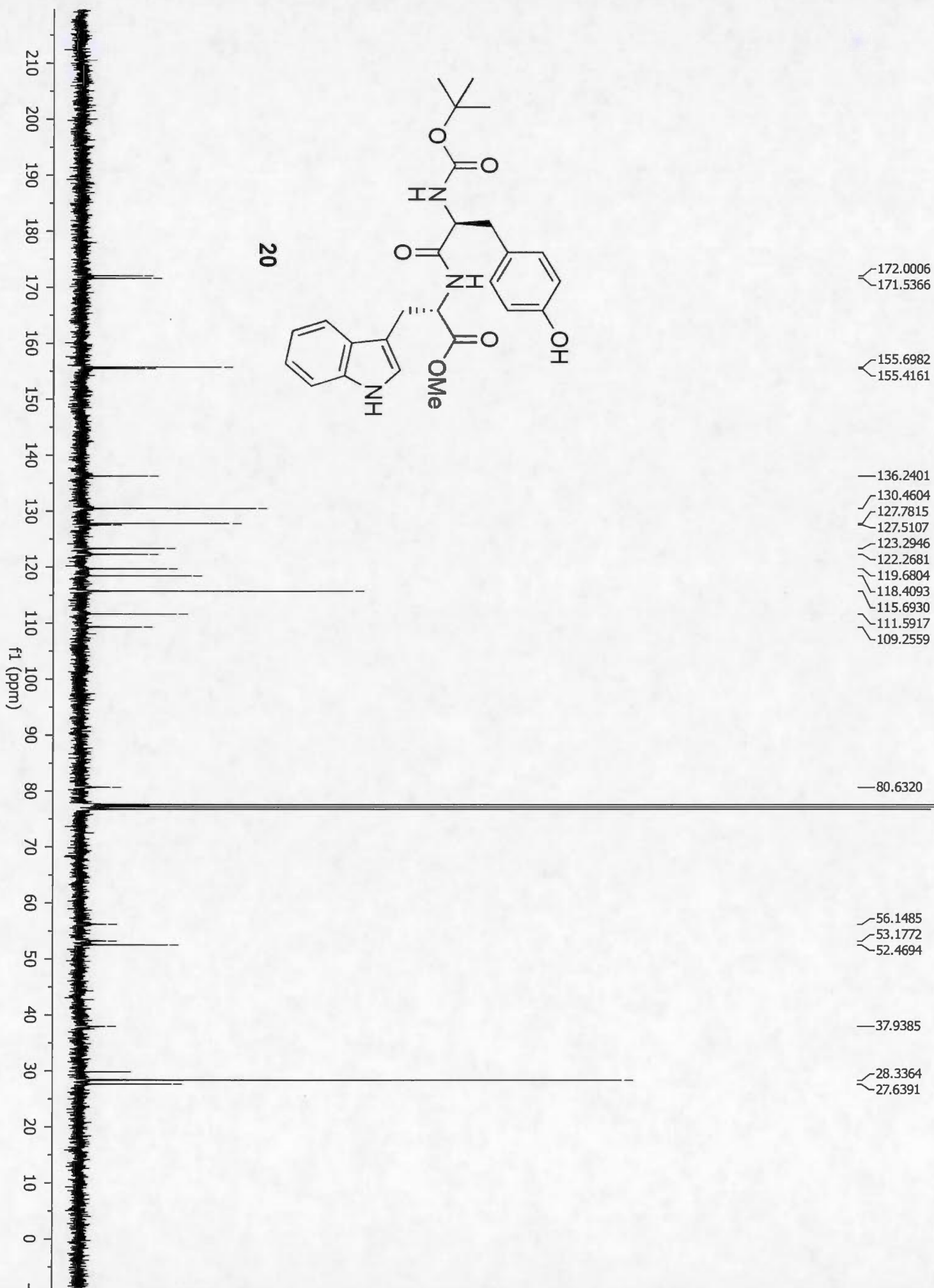


19



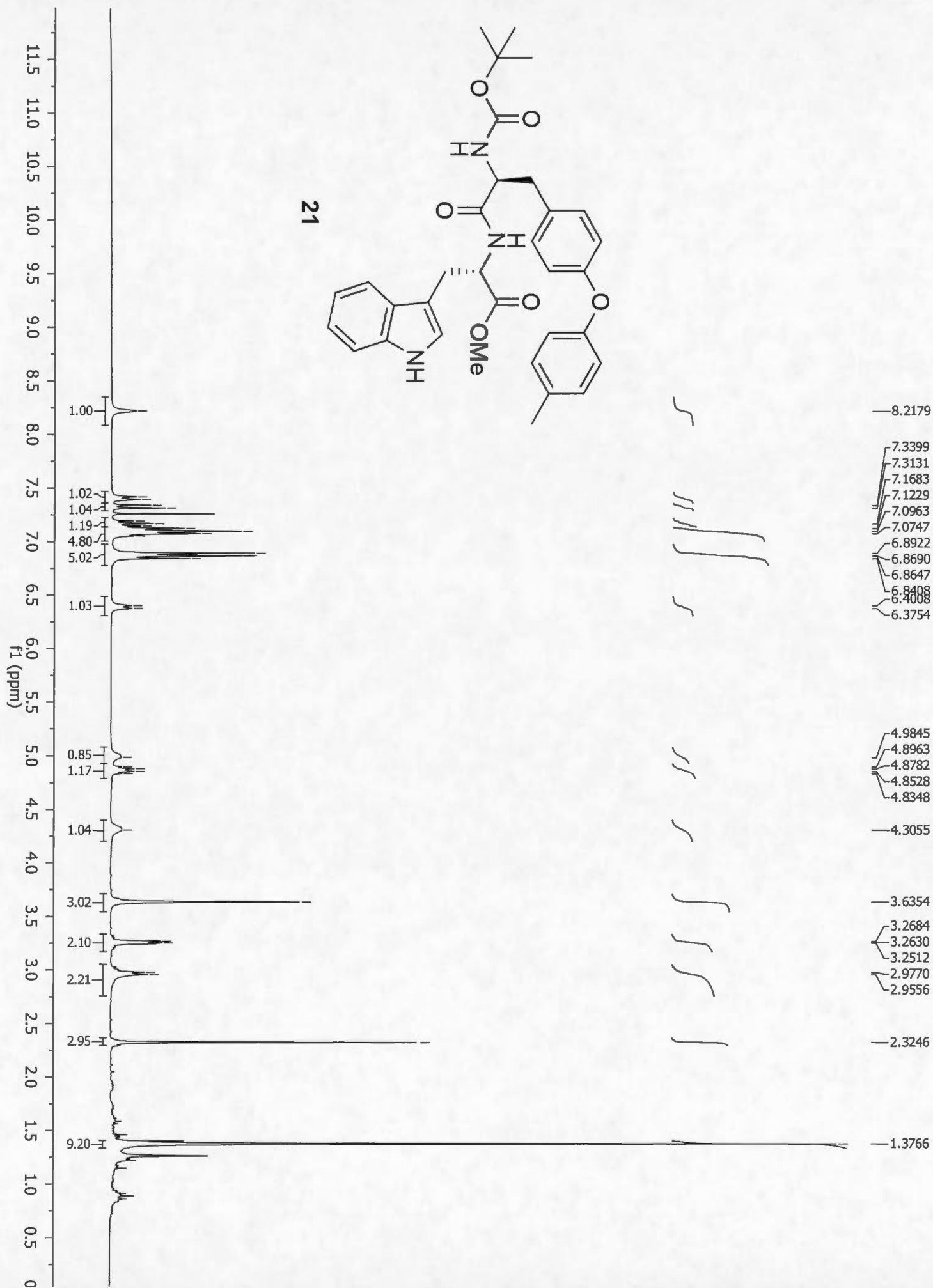
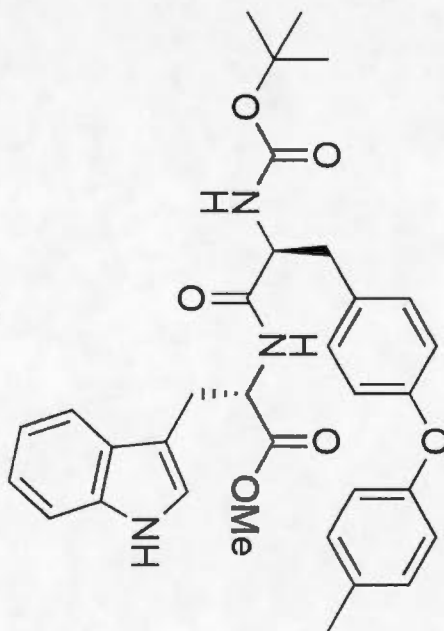


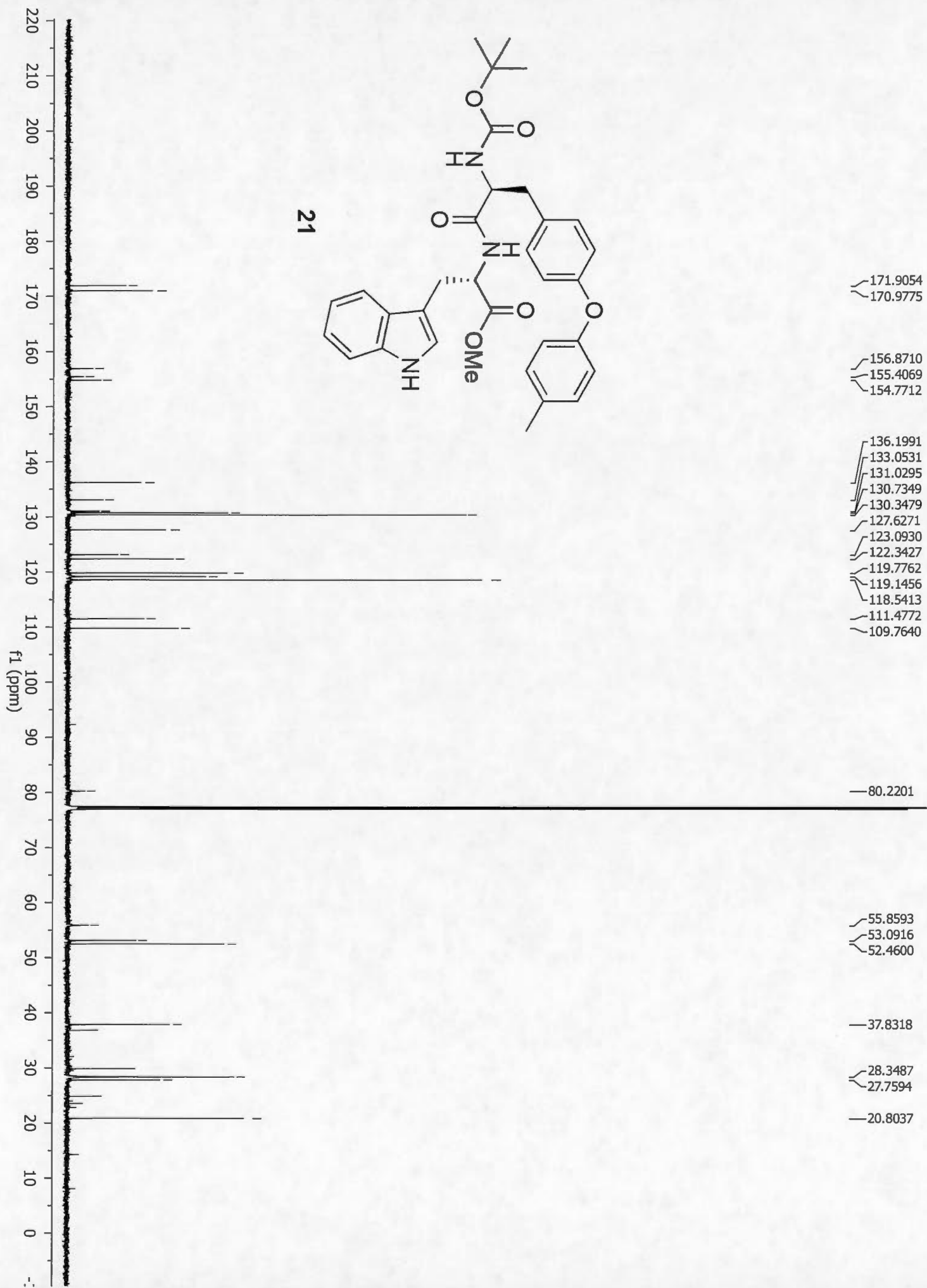


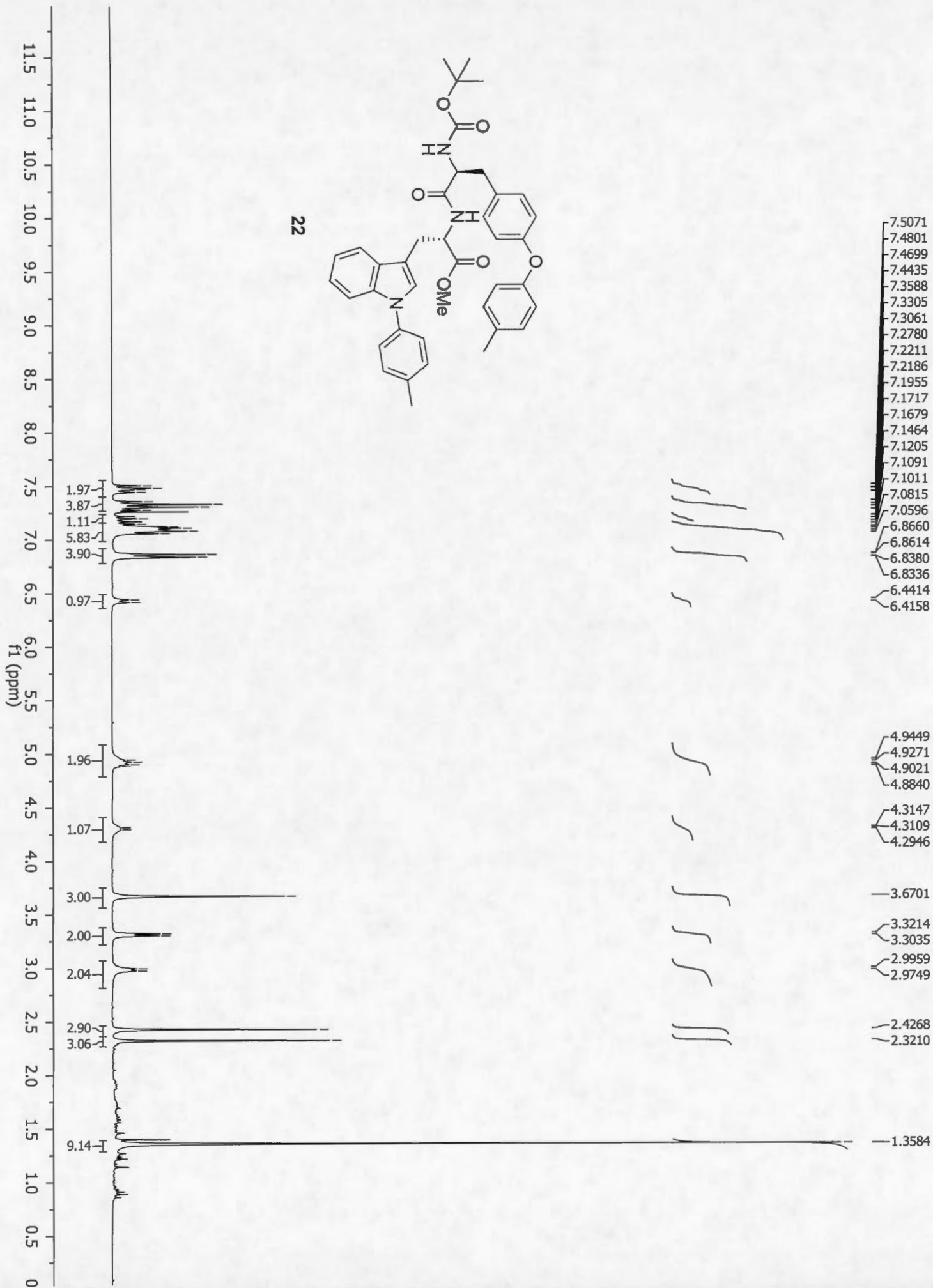


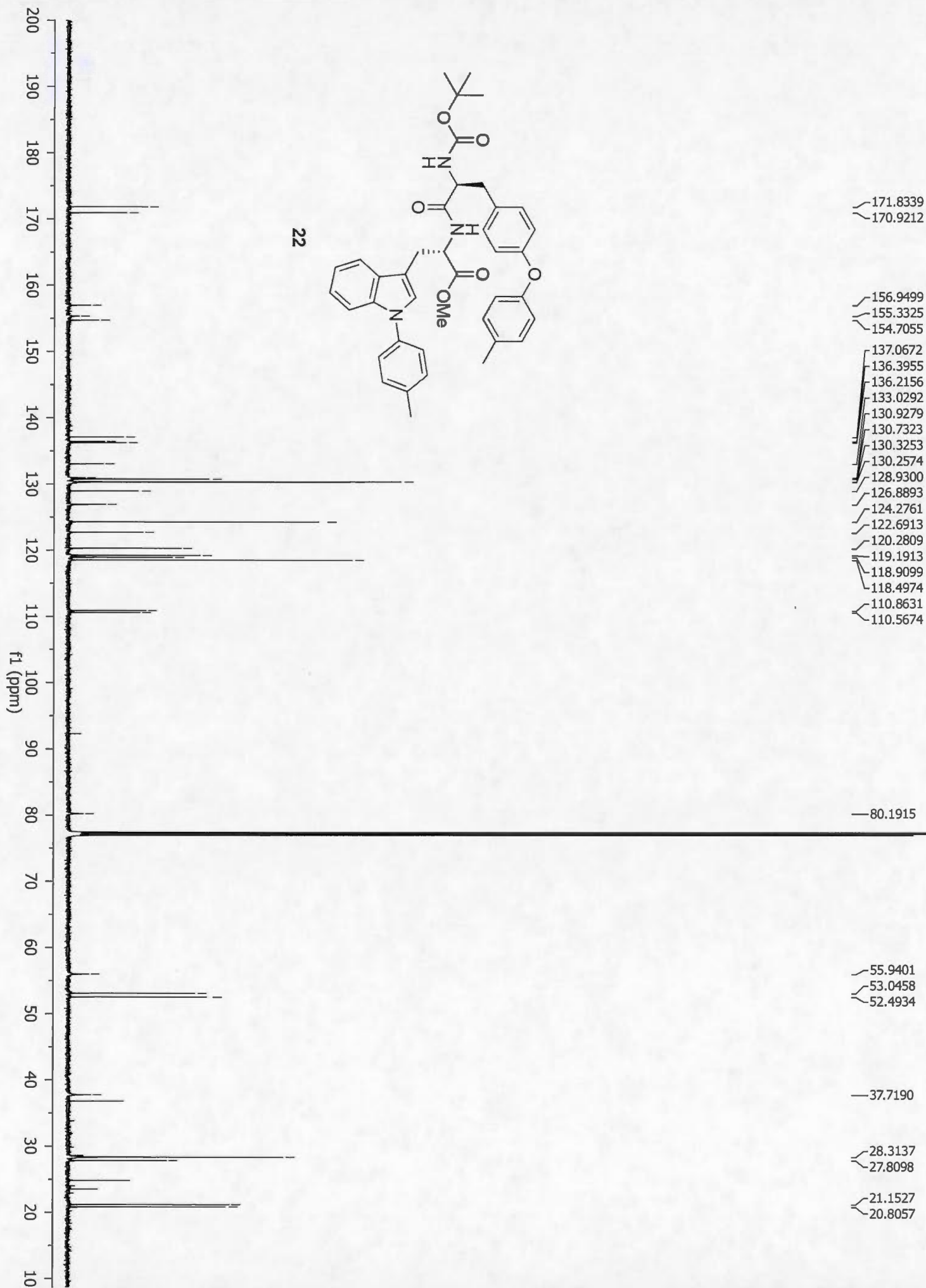


21



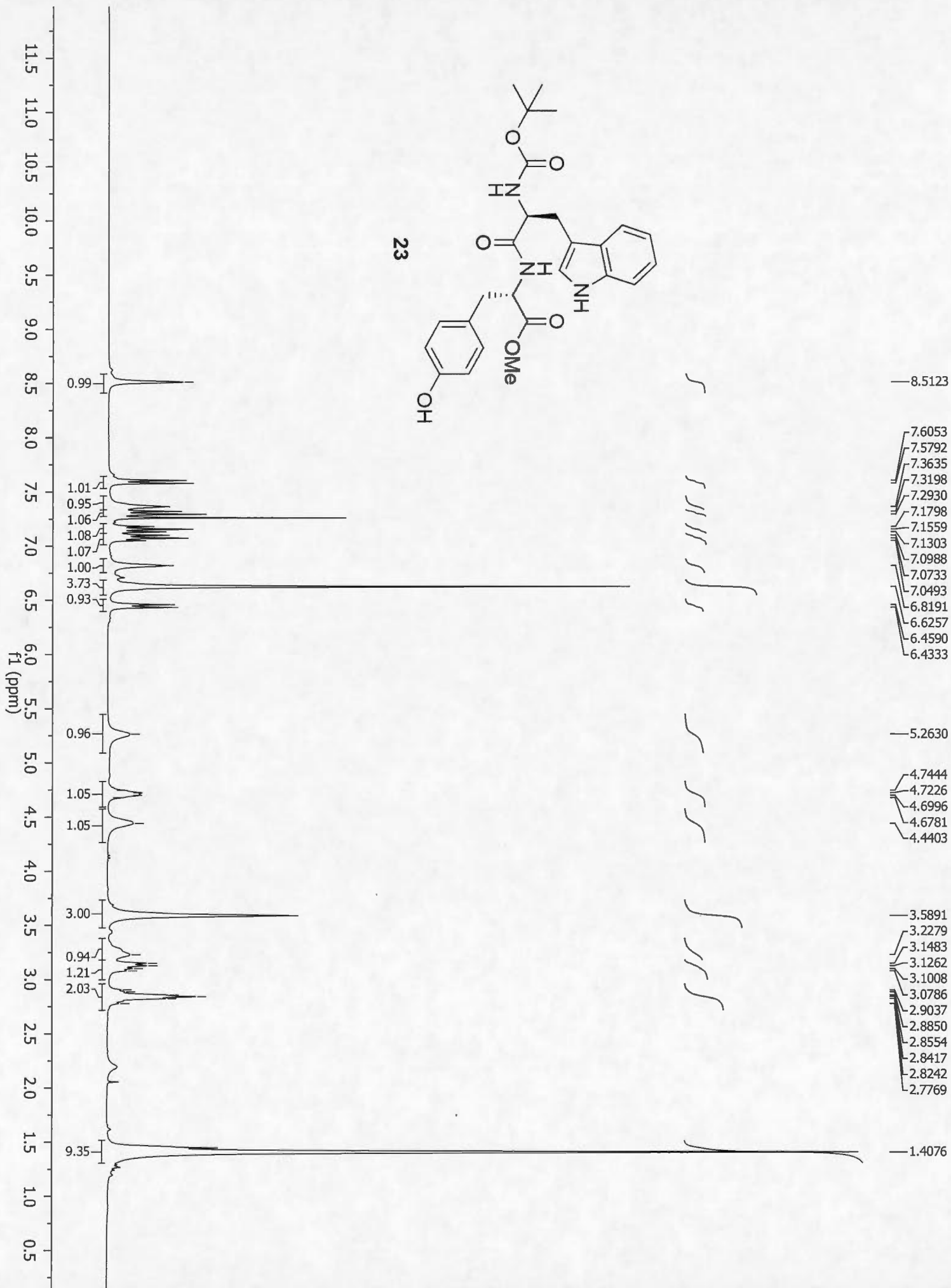
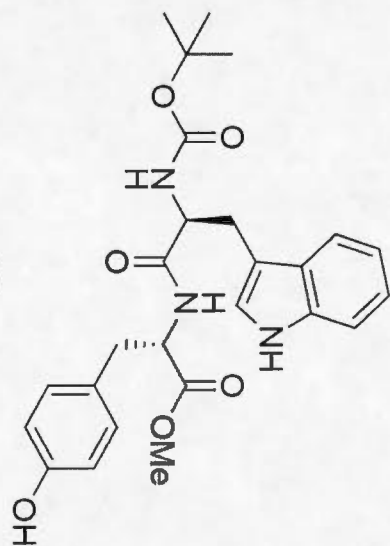


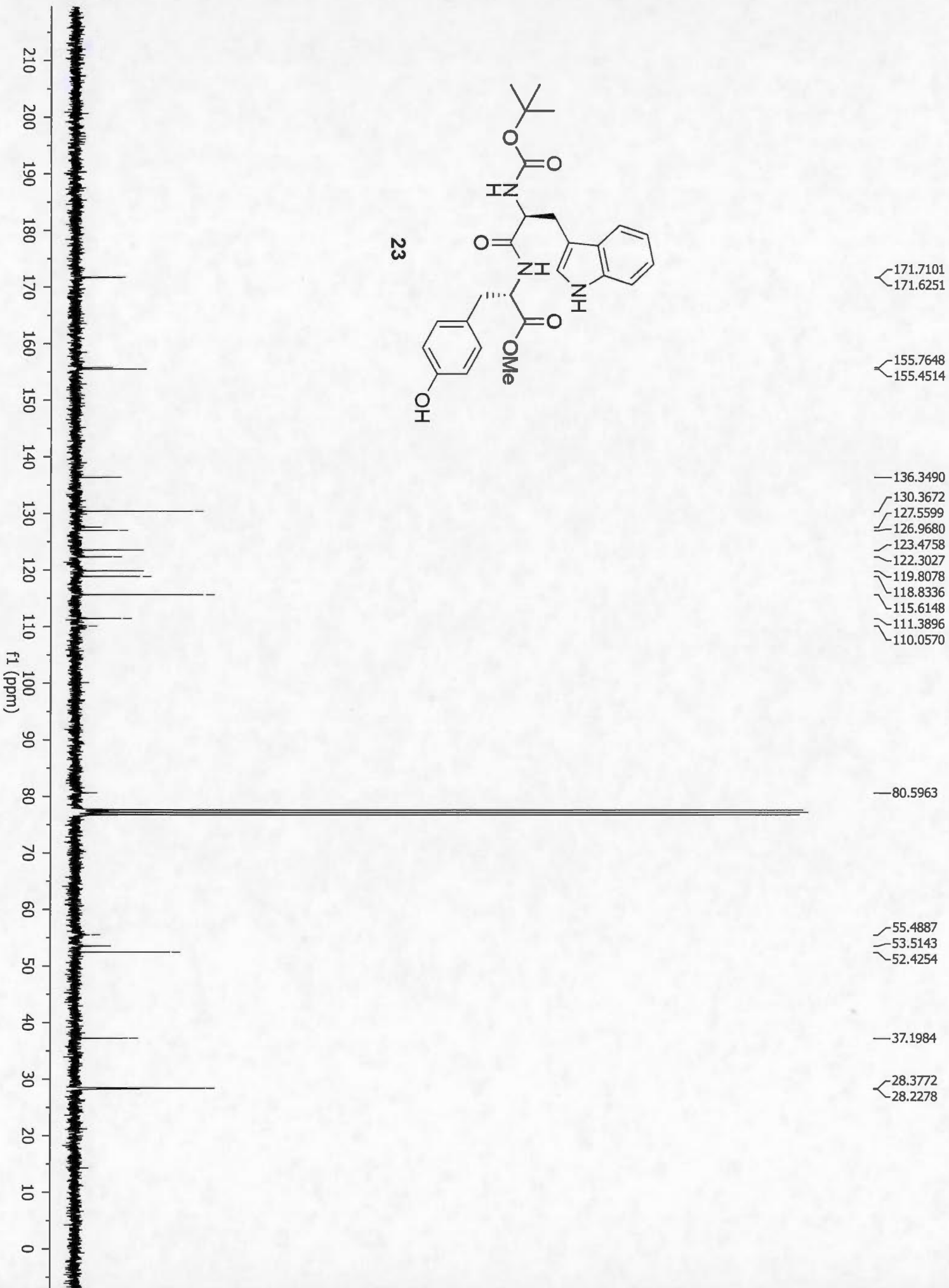


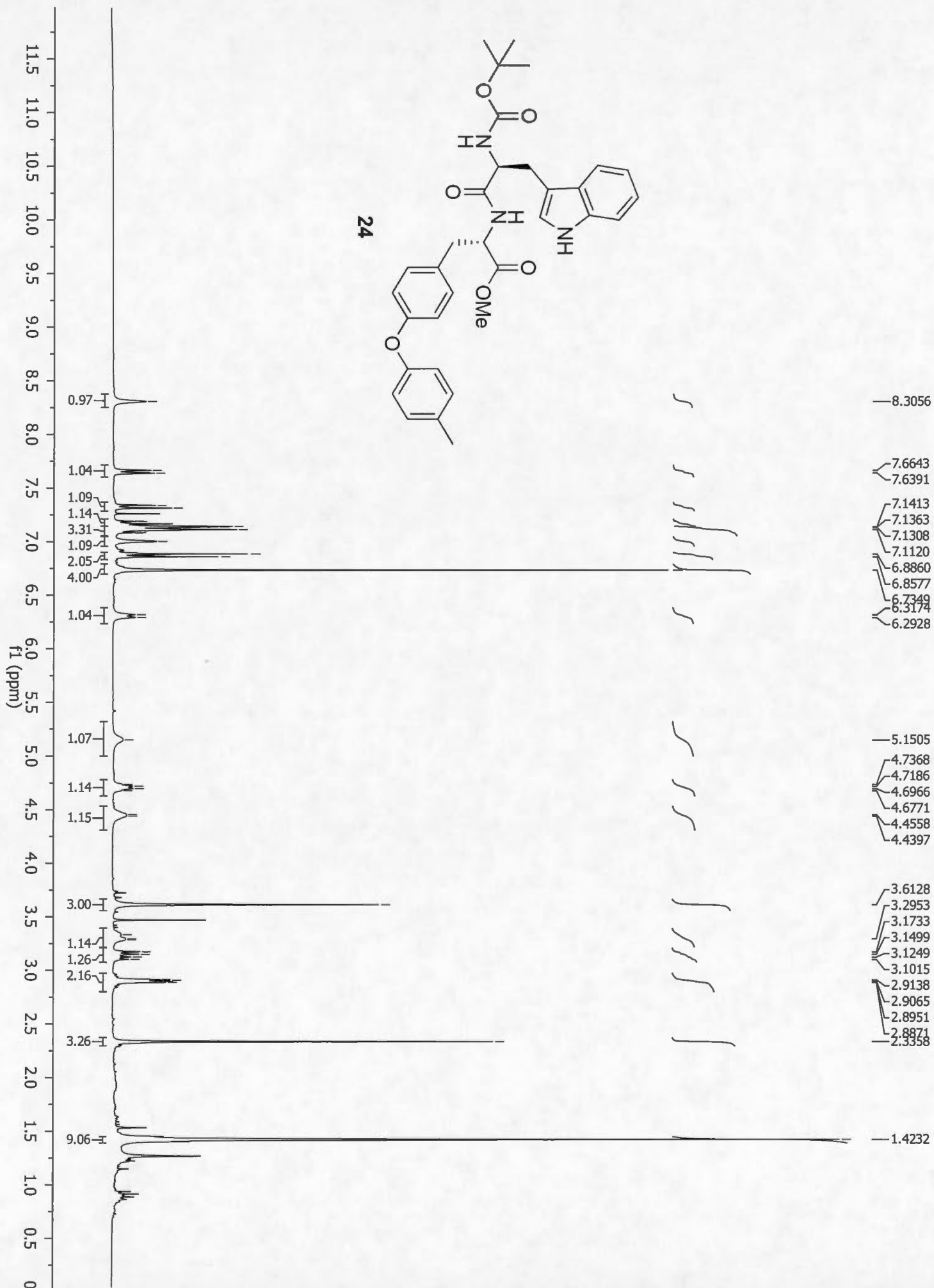


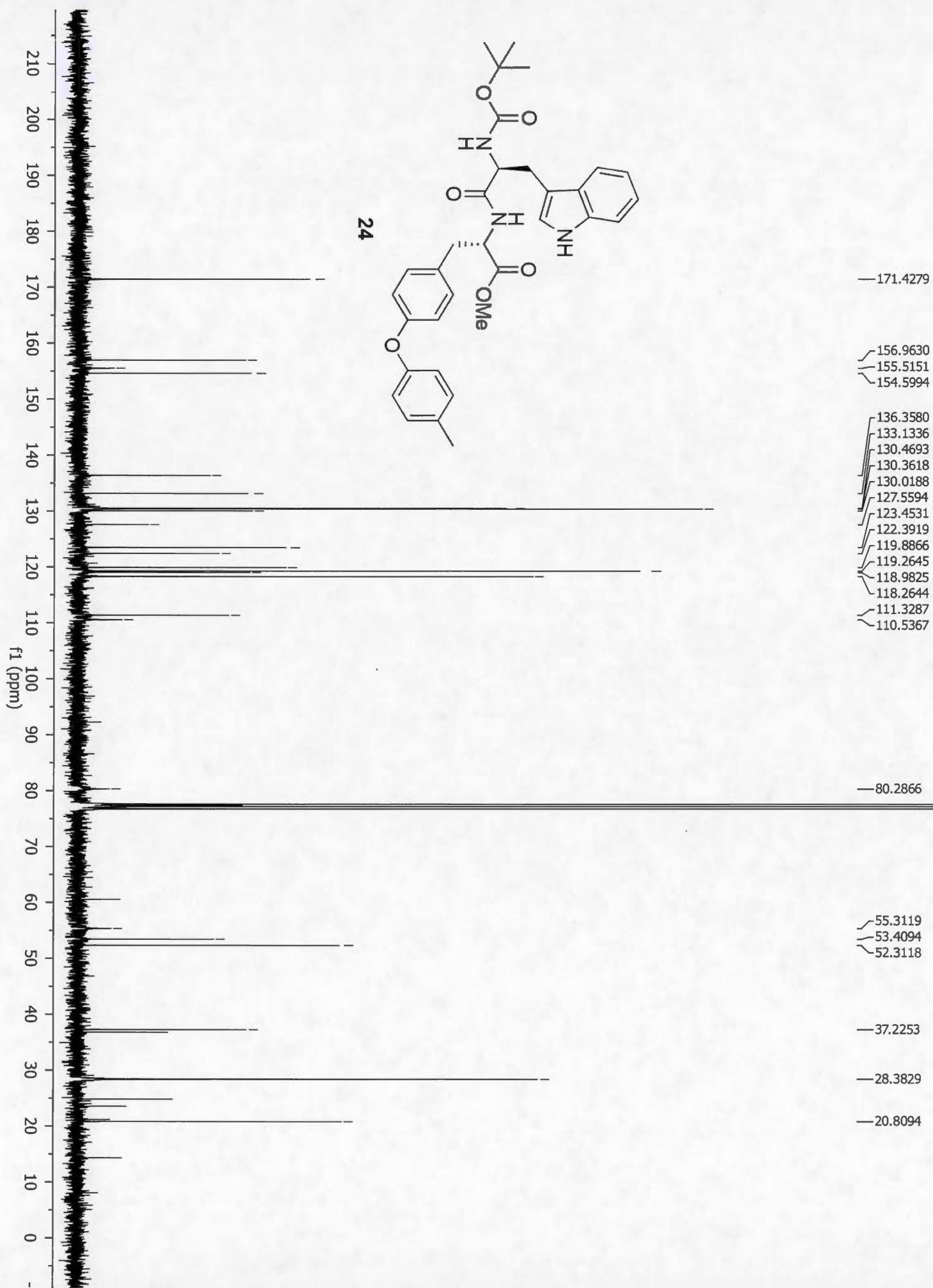


23











## BIBLIOGRAPHIE

- 1) Finet, J.-P. *Chem. Rev.* **1989**, *89*, 1487-1501.
- 2) Moham, R. *Nat. Chem.* **2010**, *2*, 336.
- 3) Löwig, C.; Schweizer, E. *Liebigs Ann. Chem.* **1850**, *75*, 315-355.
- 4) Michaëlis, A.; Polis, A. *Ber Dtsch. Chem Ges.* **1887**, *20*, 54-57.
- 5) Wittig, G.; Clauss, K. *Liebigs Ann. Chem.* **1952**, *578*, 136-146.
- 6) Petiot, P.; Gagnon, A. *Heterocycles* **2014**, *88*, 1615-1624.
- 7) Gagnon, A.; St-Onge, M.; Little, K.; Duplessis, M.; Barabé, F. *J. Am. Chem. Soc.* **2007**, *129*, 44-45.
- 8) Hong, Y.; Lai, Y.-T.; Chan, G. C.-F. Sun, H. *PNAS* **2015**, *112*, 3211-3216.
- 9) (a) Kobiki, Y.; Kawaguchi, S.-i.; Ogawa, A. *Org. Lett.* **2015**, *17*, 3490-3493; (b) Wang, T.; Sang, S.; Liu, L.; Qiao, H.; Gao, Y.; Zhao, Y. *J. Org. Chem.* **2014**, *79*, 608-617; (c) Kobiki, Y.; Kawaguchi, S.-i.; Ohe, T.; Ogawa, A. *Beilstein J. Org. Chem.* **2013**, *9*, 1141-1147; (d) Sueda, T.; Oshima, A.; Teno, N. *Org. Lett.* **2011**, *13*, 3996-3999; (e) Yasuike, S.; Nishioka, M.; Kakusawa, N.; Kurita, J. *Tetrahedron Lett.* **2011**, *52*, 6403-6406; (f) Sato, I.; Toyota, Y.; Asakura, N. *Eur. J. Org. Chem.* **2007**, 2608-2610; (g) Ikegai, K.; Fukumoto, K.; Mukaiyama, T. *Chem. Lett.* **2006**, *35*, 612-613; (h) Mukaiyama, T.; Sakurai, N.; Ikegai, K. *Chem. Lett.* **2006**, *35*, 1140-1141; (i) Koech, P. K.; Krische, M. J. *Tetrahedron* **2006**, *62*, 10594-10602; (j) Koech, P. K.; Krische, M. J. *J. Am. Chem. Soc.* **2004**, *126*, 5350-5351; (k) Ooi, T.; Goto, R.; Maruoka, K. *J. Am. Chem. Soc.* **2003**, *125*, 10494-10495; (l) Lermontov, S. A.; Rakov, I. M.; Zefirov, N. S.; Stang, P. J. *Tetrahedron Lett.* **1996**, *37*, 4051-4054; (m) Cho, C. S.; Yoshimori, Y.; Uemura, S. *Bull. Chem. Soc. Jpn.* **1995**, *68*, 950-957.

- 10) (a) Nicolaou, K. C.; Sarlah, D.; Wu, T. R.; Zhan, W. *Angew. Chem. Int. Ed.* **2009**, *48*, 6870-6874; (b) Krawczuk, P. J.; Schöne, N.; Baran, P. S. *Org. Lett.* **2009**, *11*, 4774-4776.
- 11) (a) Benjamin, S. L.; Reid, G. *Coord. Chem. Rev.* **2015**, *297-298*, 168-180; (b) Braunschweig, H.; Cogswell, P.; Schwab, K. *Coord. Chem. Rev.* **2011**, *255*, 101-117.
- 12) Yamago, S. *Chem. Rev.* **2009**, *109*, 5051-5068 et les références qui y sont citées.
- 13) (a) Yang, Y.; Ouyang, R.; Xu, L.; Guo, N.; Li, W.; Feng, K.; Ouyang, L.; Yang, Z.; Zhou, S.; Miao, Y. *J. Coord. Chem.* **2015**, *68*, 379-397; (b) Briand, G. G.; Burford, N. *Chem. Rev.* **1999**, *99*, 2601-2657.
- 14) (a) Banfi, A.; Bartoletti, M.; Bellora, E.; Bignotti, M.; Turconi, M. *Synthesis* **1994**, 775-776; (b) Marafuji, T.; Nishio, K.; Nagasue, M.; Tanabe, A.; Aono, M.; Sugihara, Y. *Synthesis* **2000**, 1208-1210; (c) Ludwig, C.; Dolny, M.; Götze, H.-J. *Spectrochimica Acta Part A* **1997**, *53*, 2363-2372; (d) De Ketelaere, R. F.; Delbeke, F. T.; Van Der Kelen, G. P. *J. Organomet. Chem.* **1971**, *30*, 365-368; (e) Matano, Y.; Kinoshita, M.; Suzuki, H. *Bull Chem. Soc. Jpn.* **1992**, *65*, 3504-3506.
- 15) Urgin, K.; Aubé, C.; Pichon, C.; Pipelier, M.; Blot, V.; Thobie-Gautier, C.; Léonel, E.; Dubreuil, D.; Condon, S. *Tetrahedron Lett.* **2012**, *53*, 1894-1896.
- 16) (a) Otero, A.; Royo, P. *J. Organomet. Chem.* **1979**, *171*, 333-336; (b) Li, X.-W.; Lorberth, J.; Massa, W.; Wocadlo, S. *J. Organomet. Chem.* **1995**, *485*, 141-147; (c) Lemus, A.; Sharma, P.; Cabrera, A.; Rosas, N.; Alvarez, C.; Gomez, Sharma, M.; Cespedes, C.; Hernandez, S. *Main Group Met. Chem.* **2001**, *24*, 835-840.
- 17) Dohle, W.; Lindsay, D. M.; Knochel, P. *Org. Lett.* **2001**, *18*, 2871-2873.
- 18) (a) Boymond, L.; Rottländer, M.; Cahiez, G.; Knochel, P. *Angew. Chem.* **1998**, *110*, 1801-1804; (b) Boymond, L.; Rottländer, M.; Cahiez, G.; Knochel, P. *Angew.*

*Chem. Int. Ed. Engl.* **1998**, *37*, 1701-1703; (c) Jensen, A. E.; Dohle, W.; Sapountzis, I.; Lindsay, D. M.; Vu, V. A.; Knochel, P. *Synthesis* **2002**, 565-569; (d) Knochel, P.; Dohle, W.; Gommermann, N.; Kneisel, F. F.; Kopp, F.; Kom, T.; Sapountzis, I.; Vu, V. A. *Angew. Chem.* **2003**, *115*, 4438-4456; (e) Knochel, P.; Dohle, W.; Gommermann, N.; Kneisel, F. F.; Kopp, F.; Kom, T.; Sapountzis, I.; Vu, V. A. *Angew. Chem. Int. Ed. Engl.* **2003**, *42*, 4302-4320.

19) (a) Andreev, P. V.; Somov, N. V.; Kalistratova, O. S.; Gushchin, A. V.; Chuprunov, E. V. *Crystallography Reports* **2015**, *60*, 517-520; (b) Ong, Y. C.; Blair, V. L.; Kedzierski, L.; Tuck, K. L.; Andrews, P. C. *Dalton Trans.* **2015**, *44*, 18215-18226; (c) Imachi, S.; Mukaiyama, T. *Chem. Lett.* **2007**, *36*, 718-719; (d) Kumar, I.; Bhattacharya, P.; Whitmire, K. H. *Organometallics* **2014**, *33*, 2906-2909; (e) Sharutin, V. V.; Sharutina, O. K.; Senchurin, V. S. *Russ. J. Inorg. Chem.* **2013**, *58*, 1470-1474; (f) Sharutin, V. V.; Senchurin, V. S.; Sharutina, O. K. *Russ. J. Inorg. Chem.* **2011**, *56*, 1565-1567; (g) Malysheva, Y. B.; Moiseev, D. V.; Gushchin, A. V.; Dodonov, V. A. *Russ. J. Gen. Chem.* **2005**, *75*, 1766-1770; (h) Ong, Y. C.; Blair, V. L.; Kedzierski, L.; Andrews, P. C. *Dalton Trans.* **2014**, *43*, 12904-12916; (i) Combes, S.; Finet, J.-P. *Syn. Commun.* **1996**, *26*, 4569-4575; (j) Arnould, T.; Barton, D. H. R.; Doris, E. *Tetrahedron Lett.* **1997**, *38*, 365-366; (k) Dodonov, V. A.; Starostina, T. I.; Kuznetsova, Y. L.; Gushcin, A. V. *Russ. Chem. Bull.* **1995**, *44*, 151-153.

20) Suzuki, H.; Komatsu, N.; Ogawa, T.; Murafuji, T.; Ikegami, T.; Matano, Y. (2001). *Organobismuth Chemistry (1<sup>st</sup> Ed.)*. Amsterdam : Elsevier Science B. V.

21) Goel, R. G.; Prasad, H. S. *Can. J. Chem.* **1970**, *48*, 2488-2493.

22) (a) Hassan, A.; Breeze, S. R.; Courtenay, S.; Deslippe, C.; Wang, S. *Organometallics* **1996**, *15*, 5613-5621 (b) Rasmussen, L. K.; Begtrup, M.; Ruhland, T. *J. Org. Chem.* **2004**, *69*, 6890-6893 (c) Combes, S.; Finet, J.-P. *Tetrahedron* **1998**, *54*, 4313-4318.



- 23) Ullmann, F. *Ber. Dtsch. Chem. Ges.* **1901**, *34*, 2174-2185.
- 24) (a) Ullmann, F. *Ber. Dtsch. Chem. Ges.* **1903**, *36*, 2382-2384; (b) Ullmann, F.; Sponagel, P. *Ber. Dtsch. Chem. Ges.* **1905**, *36*, 2211-2212; (c) Ullmann, F.; Maag, R. *Ber. Dtsch. Chem. Ges.* **1906**, *39*, 1693-1696.
- 25) Goldberg, I. *Ber. Dtsch. Chem. Ges.* **1906**, *39*, 1691-1692.
- 26) Evano, G.; Blanchard, N.; Toumi, M. *Chem. Rev.* **2008**, *108*, 3054-3131.
- 27) Hartwig, J. F. (2010). *Organotransition Metal Chemistry: From Bonding to Catalysis (1<sup>st</sup> Ed.)*. Sausalito: University Science Books.
- 28) Lam, P. Y. S.; Vincent, G.; Bonne, D.; Clark, C. G. *Tetrahedron Lett.* **2003**, *44*, 4927-4931.
- 29) (a) Sheppard, G. S. *Synlett* **1999**, 1207-1210; (b) Harada, S.; Hayashi, D.; Sato, I.; Hirama, M. *Synlett* **2012**, 405-408; (c) Combes, S.; Finet, J.-P. *Tetrahedron* **1998**, *54*, 4313-1318.
- 30) Mukaiyama, T.; Sakurai, N.; Ikegai, K. *Chem. Lett.* **2006**, *35*, 1140-1141.
- 31) Harada, S.; Hayashi, D.; Sato, I.; Hirama, M. *Synlett* **2012**, *23*, 405-408.
- 32) (a) Petiot, P.; Dansereau, J.; Hébert, M.; Khene, I.; Ahmad, T.; Samaali, S.; Leroy, M.; Pinsonneault, F.; Legault, C. Y.; Gagnon, A. *Org. Biomol. Chem.* **2015**, *13*, 1322-1327; (b) Barton, D. H. R.; Finet, J.-P.; Pichon, C. *J. Chem. Soc., Chem. Commun.* **1986**, 65-66.
- 33) Crifar, C.; Petiot, P.; Ahmad, T.; Gagnon, A. *Chem. Eur. J.* **2014**, *20*, 2755-2760.
- 34) Sorenson, R. J. *J. Org. Chem.* **2000**, *65*, 7747-7749.
- 35) Barton, D. H. R.; Finet, J.-P.; Khamisi, J. *Tetrahedron Lett.* **1987**, *28*, 887-890.



- 36) (a) Loog, O.; Mäeorg. *Synlett* **2004**, 2537-2540; (b) Loog, O.; Mäeorg, U.; Ragnarsson, U. *Synthesis* **2004**, 1591-1597.
- 37) Starkov, P.; Zemskov, I.; Sillard, R.; Tšubrik, O.; Mäeorg, U. *Tetrahedron Lett.* **2007**, 48, 1155-1157.
- 38) (a) Mandal, P. K.; Morlacchi, P.; Knight, J. M.; Link, T. M.; Lee, G. R. IV.; Nurieva, R.; Singh, D.; Dhanik, A.; Kavraki, L.; Corry, D. B.; Ladbury, J. E.; McMurray, J. S. *J. Med. Chem.* **2015**, 58, 8970-8984; (b) Morlacchi, P.; Mandal, P. K.; McMurray, J. S. *ACS Med. Chem. Lett.* **2014**, 5, 69-72; (c) Chan, D. M. T. *Tetrahedron Lett.* **1996**, 37, 9013-9016.
- 39) (a) Petiot, P.; Dansereau, J.; Gagnon, A. *RSC Adv.* **2014**, 4, 22255-22259; (b) Fedorov, A. Y.; Finet, J.-P. *Tetrahedron Lett.* **1999**, 40, 2747-2748.
- 40) (a) Miyaura, N.; Yamada, K.; Suzuki, A. *Tetrahedron Lett.* **1979**, 20, 3437-3440; (b) Miyaura, N.; Sugimoto, H.; Suzuki, A. *Tetrahedron Lett.* **1981**, 22, 127-130; (c) Miyaura, N.; Yamada, K.; Sugimoto, H.; Suzuki, A. *J. Am. Chem. Soc.* **1985**, 107, 972-980.
- 41) Baba, S.; Negishi, E. *J. Am. Chem. Soc.* **1976**, 98, 6729-6731.
- 42) López-Pérez, A.; Adrio, J.; Barretero, J. C. *Org. Lett.* **2009**, 11, 5514-5517.
- 43) Stille, J. K. *Pure Appl. Chem.* **1985**, 57, 1771-1780.
- 44) (a) Rao, M. L. N.; Giri, S.; Jadhav, D. N. *Tetrahedron Lett.* **2009**, 50, 6133-6138; (b) Rao, M. L. N.; Venkatesh, V.; Jadhav, D. N. *J. Organomet. Chem.* **2008**, 693, 2494-2498; (c) Barton, D. H. R.; Ozbalik, N. Ramesh, M. *Tetrahedron* **1988**, 44, 5661-5668.
- 45) (a) Rao, M. L. N.; Banerjee, D.; Dhanorkar, R. J. *Tetrahedron* **2010**, 66, 3623-3632; (b) Huang, X.; Wu, J. L. *Chin. Chem. Lett.* **1997**, 8, 759-762.

- 46) Rao, M. L. N., Yamazaki, O.; Shimada, S.; Tanaka, T.; Suzuki, Y.; Tanaka, M. *Org. Lett.* **2001**, *3*, 4103-4105.
- 47) (a) Rao, M. L. N.; Jadhav, D. N.; Banerjee, D. *Tetrahedron* **2008**, *64*, 5762-5772; (b) Rao, M. L. N.; Banerjee, D.; Giri, S. *J. Organomet. Chem.* **2010**, *695*, 1518-1525; (c) Rao, M. L. N.; Jadhav, D. N.; Dasgupta, P. *Org. Lett.* **2010**, *12*, 2048-2051; (d) Rao, M. L. N.; Dhanorkar, R. J. *Tetrahedron* **2015**, *71*, 338-349; (e) Rao, M. L. N.; Dhanorkar, R. J. *RSC Adv.* **2013**, *3*, 6794-6798; (f) Rao, M. L. N.; Dhanorkar, R. J. *Eur. J. Org. Chem.* **2014**, 5214-5228; (g) Rao, M. L. N.; Banerjee, D.; Dhanorkar, R. J. *Synlett* **2011**, 1324-1330; (h) Rao, M. L. N.; Awasthi, D. K.; Talode, J. B. *Synlett* **2012**, 1907-1912.
- 48) Hao, W.; Liu, H.; Yin, L.; Cai, M. *J. Org. Chem.* **2016**, *81*, 4244-4251.
- 49) (a) Zhu, Y.; Bauer, M.; Ackermann, L. *Chem. Eur. J.* **2015**, *21*, 9980-9983; (b) Ruiz-Rodríguez, J.; Albericio, F.; Lavilla, R. *Chem. Eur. J.* **2010**, *16*, 1124-1127; (c) Preciado, S.; Mendive-Tapia, L.; Albericio, F.; Lavilla, R. *J. Org. Chem.* **2013**, *78*, 8129-8135; (d) González, R.; Mendive-Tapia, L.; Pastrian, M. B.; Albericio, F.; Lavilla, R.; Cascone, O.; Iannucci, N. B. *J. Pep. Sci.* **2016**, *22*, 123-128; (e) Preciado, S.; Mendive-Tapia, L.; Torres-García, C.; Zamudio-Vásquez, R.; Soto-Cerrato, V.; Pérez-Tomás, R.; Albericio, F.; Nicolás, E.; Lavilla, R. *Med. Chem. Comm.* **2013**, *4*, 1171-1174.
- 50) Zhang, X.; He, G.; Chen, G. *Org. Biomol. Chem.* **2016**, *14*, 5511-5515.
- 51) Barton, D. H. R.; Finet, J.-P.; Khamisi, J. *Tetrahedron Lett.* **1989**, *30*, 937-940.
- 52) Sharma, K. K.; Sharma, S.; Kudwal, A.; Jain, R. *Org. Biomol. Chem.* **2015**, *13*, 4637-4641.
- 53) Khatib, M. E.; Molander, G. A. *Org. Lett.* **2014**, *16*, 4944-4947.

- 54) DalZotto, C.; Michaux, J.; Martinand-Lurin, E.; Campagne, J.-M. *Eur. J. Org. Chem.* **2010**, 3811-3814.
- 55) Yue, W.; Lewis, S. I.; Koen, Y. M.; Hanzlik, R. P. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 1637-1640.
- 56) (a) Elliott, G. I.; Konopelski, J. P.; *Org. Lett.* **2000**, *2*, 3055-3057; (b) Cappuccio, J. A.; Ayala, I.; Elliott, G. I.; Szundi, I.; Lewis, J.; Konopelski, J. P.; Barry, B. A.; Einarsdottir, O. *J. Am. Chem. Soc.* **2002**, *124*, 1750-1760; (c) White, K. N.; Sen, I.; Szundi, I.; Landaverry, Y. R.; Bria, L. E.; Konopelski, J. P.; Olmstead, M. M.; Einarsdottir, O. *Chem. Commun.* **2007**, 3252-3254; (d) Landaverry, Y. R.; White, K. N.; Olmstead, M. M.; Einarsdottir, O.; Konopelski, J. P. *Heterocycles* **2006**, *70*, 147-152.
- 57) (a) Handlogten, M. W.; Kiziltepe, T.; Alves, N. J.; Bilgicer, B. *Am. Chem. Soc., Chem. Biol.* **2012**, *7*, 1796-1801; (b) Wilson, D. L.; Meininger, I.; Strater, Z.; Steiner, S.; Tomlin, F.; Wu, J.; Jamali, H.; Krappmann, D.; Götz, M. G. *Am. Chem. Soc., Med. Chem. Lett.* **2016**, *7*, 250-255; (c) Boger, D. L.; Zhou, J.; Borzilleri, R. M.; Nukui, S. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 1089-1092; (d) Pérez-González, M.; Jackson, R. F. W. *Chem. Commun.* **2000**, 2423-2424; (e) Fotouhi, N.; Galakatos, N. G.; Kemp, D. S. *J. Org. Chem.* **1989**, *54*, 2803-2817; (f) Arnusch, C. J.; Ippel, J. H.; Kooijman, H.; Spek, A. L.; Liskamp, R. M. J.; Kemmink, J.; Pieters, R. J. *Eur. J. Org. Chem.* **2010**, *23*, 4501-4507; (g) Barra, M.; de Rossi, R. H. *J. Org. Chem.* **1989**, *54*, 5020-5025; (h) Nolasco, L.; Gonzalez, M. P.; Caggiano, L.; Jackson, R. F. W. *J. Org. Chem.* **2009**, *74*, 8280-8289; (i) Jayabharathi, J.; Thanikachalam, V.; Srinivasan, N.; Perumal, M. V. *Spectrochimica Acta Part A* **2011**, *84*, 233-237; (j) Liu, Z.; Sayre, *Tetrahedron* **2004**, *60*, 1601-1610.
- 58) (a) Höfling, S. B.; Bartuschat, A. L.; Heinrich, M. R. *Angew. Chem. Int. Ed.* **2010**, *49*, 9769-9772; (b) Wang, A.; Gui, L.; Lu, S.; Zhou, L.; Zhou, J.; Wei, S. *Dyes and*



*Pigments* **2016**, *126*, 239-250; (c) Lasch, R.; Fehler, S. K.; Heinrich, M. R. *Org. Lett.* **2016**, *18*, 1586-1589; (d) Jasch, H.; Höfling, S. B.; Heinrich, M. R. *J. Org. Chem.* **2012**, *77*, 1520-1532.

59) (a) Pearson, A. J.; Bruhn, P. R.; Hsu, S.-Y. *J. Org. Chem.* **1986**, *51*, 2139-2142; (b) Pearson, A. J.; Bruhn, P. R. *J. Org. Chem.* **1991**, *56*, 7092-7097.

60) (a) Smirnova, J.; Engman, L.; Andersson, C.-M.; Malm, J. *J. Organomet. Chem.* **2005**, *690*, 1784-1792; (b) Pearson, A. J.; Park, J. G.; Zhu, P. Y. *J. Org. Chem.* **1992**, *57*, 3583-3589.

61) Janetka, J. W.; Rich, D. H. *J. Am. Chem. Soc.* **1997**, *119*, 6488-6495.

62) (a) Cai, Q.; Zou, B.; Ma, D. *Angew. Chem. Int. Ed.* **2006**, *45*, 1276-1279; (b) Cai, Q.; He, G.; Ma, D. *J. Org. Chem.* **2006**, *71*, 5268-5273; (c) Nicolaou, K. C.; Boddy, C. N. C.; Natarajan, S.; Yue, T.-Y.; Li, H.; Bräse, S.; Ramanjulu, J. M. *J. Am. Chem. Soc.* **1997**, *119*, 3421-3422; (d) Nicolaou, K. C.; Ramanjulu, J. M.; Natarajan, S.; Bräse, S.; Li, H.; Boddy, C. N. C.; Rübsam, F. *Chem. Commun.* **1997**, *19*, 1899-1900.

63) (a) Ghosh, S.; Kumar, A. S.; Mehta, G. N.; Soundararajan, R.; Sen, S. *ARKIVOK* **2009**, *7*, 72-78; (b) Hitotsuyanagi, Y.; Ishikawa, H.; Naito, S.; Takeya, K. *Tetrahedron Lett.* **2003**, *44*, 5901-5903; (c) Ghosh, S.; Kumar, A. S.; Mehta, G. N.; Soundararajan, R. *Synthesis* **2009**, *19*, 3322-3326; (d) Kilitoglu, B.; Arndt, H.-D. *Synlett* **2009**, *5*, 720-723; (e) Ghosh, S.; Kumar, A. S.; Mehta, G. N.; Soundararajan, R. *Synth. Commun.* **2010**, *40*, 2389-2396; (f) Hitotsuyanagi, Y.; Odagiri, M.; Kato, S.; Kusano, J.-i.; Husada, T.; Fukawa, H.; Takeya, K. *Chem. Eur. J.* **2012**, *18*, 2839-2846.

64) Chapman, C. J.; Matsuno, A.; Frost, C. G.; Willis, M. C. *Chem. Commun.* **2007**, 3903-3905.

65) Jalalian, N.; Petersen, T. B.; Olofsson, B. *Chem. Eur. J.* **2012**, *18*, 14140-14149.



- 66) (a) Boutureira, O.; Bernardes, G. J. L. *Chem. Rev.* **2015**, *115*, 2174-2195; (b) Spicer, C. D.; Davis, B. G. *Nat. Commun.* **2014**, *5*, 4740.
- 67) Antos, J. M.; Francis, M. B. *Curr. Opin. Chem. Biol.* **2006**, *10*, 253-262.
- 68) McFarland, J. M.; Francis, M. B. *J. Am. Chem. Soc.* **2005**, *127*, 13490-13491.
- 69) Tilley, S. D.; Francis, M. B. *J. Am. Chem. Soc.* **2006**, *128*, 1080-1081.
- 70) Joshi, N. S.; Whitaker, L. R.; Francis, M. B. *J. Am. Chem. Soc.* **2004**, *126*, 15942-15943.
- 71) Antos, J. M.; Francis, M. B. *J. Am. Chem. Soc.* **2004**, *126*, 10256-10257.
- 72) Goddard, D. R.; Michaelis, L. *J. Biol. Chem.* **1935**, *112*, 361-371.
- 73) Nielsen, M. L.; Vermeulen, M.; Bonaldi, T.; Cox, J.; Moroder, L.; Mann, M. *Nat. Methods* **2008**, *5*, 459-460.
- 74) Sunbul, M.; Nacheva, L.; Jäschke, A. *Bioconjugate Chem.* **2015**, *26*, 1466-1469.